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Patterns of Molecular Response to and Relapse After Combination of Sorafenib, Idarubicin, and Cytarabine in Patients With *FLT3* Mutant Acute Myeloid Leukemia

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Abstract

Background—FMS-like tyrosine kinase 3 (*FLT3*) is a class III receptor tyrosine kinase involved in hematopoietic progenitor cell development. Mutations of *FLT3* have been reported in about a third of patients with acute myeloid leukemia (AML), and inhibitors of *FLT3* are of clinical interest. Sorafenib is an orally active multikinase inhibitor with potent activity against *FLT3* and the Raf/ERK/MEK kinase pathway.

Methods—We studied the patterns of molecular response and relapse in 18 patients with mutated *FLT3* treated with the combination of sorafenib, idarubicin, and cytarabine.

Results—The median follow-up was 9 months. Sixteen patients achieved complete remission (CR), and the other 2 patients achieved CR but lacked platelet recovery for an overall response rate of 100%. Ten patients had their *FLT3*-mutated clone eradicated, with 6 patients who showed some residual *FLT3*-mutated cells, and 2 patients who showed persistent *FLT3*-mutated cells. The elimination of *FLT3*-mutated population at the time of morphologic CR, however, was not predictive of relapse. After a median follow-up of 9 months (range, 1–16 months), 10 (55%) patients had relapsed, with a median CR duration of 8.8 months (range, 1–9.5 months). By DNA sequencing, there was no evidence of an acquired *FLT3* point mutation at the time of relapse in 7 patients tested, which suggested the presence of other mechanisms of sorafenib resistance.

Conclusion—Sorafenib, combined with chemotherapy, is effective in attaining CR, but relapses still occur.

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Disclosure

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Keywords

Acute Myeloid Leukemia AML; *FLT3* mutation; Sorafenib

Introduction

Acute myeloid leukemia (AML) is the most common acute leukemia, and its effective therapy remains challenging. Induction with cytarabine and an anthracycline remains the standard of care. New regimens are incorporating target-specific agents to improve both complete remission (CR) and survival rates. These novel agents typically target protein products of mutated genes and balanced translocations, such as upregulated or constitutively activated tyrosine kinases, in leukemia cells.

FMS-like tyrosine kinase-3 (*FLT3*) is a member of class III receptor tyrosine kinase and is involved in normal hematopoietic progenitor cell development.¹⁻³ It is a membrane-bound receptor with an intrinsic tyrosine kinase domain.⁴ Autocrine signaling by the *FLT3* ligand through wild-type (WT) *FLT3* can lead to enhanced kinase activity.^{5,6} Mutations of *FLT3*, such as an internal tandem duplication (ITD) or tyrosine kinase domain (TKD) point mutations, occur in >30% of patients with AML and lead to constitutive activation of the *FLT3* kinase.^{6,7} The presence of *FLT3*-ITD is associated with reduced disease-free and overall survival (OS).⁷⁻⁹ Whether *FLT3*-TKD has a similar effect on the outcome has been the subject of conflicting reports.¹⁰⁻¹³

FLT3-ITD occurs in approximately 25% of younger patients with AML, with a variable length of duplicated DNA (between 3 and >400 base pairs in the juxtamembrane domain).¹⁴ The juxtamembrane domain is a negative regulatory domain that inhibits the activation loop from adopting the active conformation. Several studies have documented the prognostic impact of these genetic alterations with patients with the ITD having a poor prognosis.^{7-9,15,16} High mutant to WT allele burden, the number of mutants of different size in the same patient, and the size of DNA insertions, have all been linked to a worse prognosis.¹⁷⁻²⁰

Sorafenib is an oral, small-molecule, multikinase inhibitor that has been approved for the treatment of renal and hepatocellular cancers at a dose of 400 mg twice daily. It inhibits *FLT3* and its downstream Raf/ERK/MEK pathway.²¹ In preclinical studies, it caused dephosphorylation of MEK1/2 and ERK, and induced apoptosis in AML cells.²² This effect was seen preferentially in *FLT3*-mutated cells compared with cells with WT *FLT3* by >1000–3000-fold. Sorafenib has demonstrated clinical activity in phase I studies in patients with *FLT3*-ITD AML.²³⁻²⁵

Several potential mechanisms of resistance have been entertained in patients treated with *FLT3* inhibitors.^{26,27} These mechanisms include plasma protein binding, bypass activation of downstream STAT5 and MAP kinase signaling, limited specificity against target *FLT3*, and secondary TKD mutations that interfere with drug activity.²⁸⁻³² In in vitro culture studies, exposure of AML cell lines to continuous *FLT3* inhibitors (including sorafenib) induces kinase domain mutations that confer resistance.³⁰ The objective of this report was to

determine the pattern of molecular response and relapse in patients with previously untreated AML who received induction therapy with the combination of sorafenib and chemotherapy, and to determine potential mechanisms of resistance. An earlier report of this clinical trial, including patients treated on the phase I portion of the study, has been previously published.³³ In this report, we focus only on the 18 patients with *FLT3* mutation (including 3 patients not included in the previous report) treated on the phase II portion of the study.

Patients and Methods

Patient Eligibility

Patients 18–60 years old with previously untreated AML (based on the World Health Organization [WHO] criteria) were eligible for treatment on this phase II study. Patients 61–65 years old also were eligible if they had a low probability of 8-week mortality with intensive chemotherapy based on adverse risk factors (cytogenetics, ECOG PS [Eastern Cooperative Oncology Group performance status], antecedent hematologic diseases, and organ function).³⁴ All the patients had to have adequate cardiac, renal, and hepatic function, with an ECOG PS of 0, 1, 2, and 3 (left ventricular ejection fraction $\geq 50\%$, creatinine ≤ 2.0 mg/dL, bilirubin ≤ 2.0 mg/dL, and liver transaminases < 3 times the institutional upper limit of normal). All the patients signed an informed consent approved by the institutional review board. Only patients with *FLT3* mutations (ITD, TKD, or both) were included in this report.

Treatment Regimen

Induction consisted of sorafenib 400 mg orally (p.o.) twice daily for 7 days combined with cytarabine 1.5 g/m² by continuous intravenous (I.V.) infusion daily for 4 days (patients > 60 years of age received 3 days only) in addition to idarubicin 12 mg/m² I.V. over 1 hour daily for 3 days. The patients who did not achieve CR after 1 course could receive another induction course. For consolidation, patients in CR received up to 5 cycles of idarubicin 8 mg/m² I.V. daily for 2 days with cytarabine 0.75 g/m² I.V. over 24 hours for 3 days in addition to sorafenib 400 mg p.o. twice daily for 28 days. The cycles were repeated every 4–6 weeks based on toxicity and recovery of counts. Patients who completed consolidation received up to 1 year of sorafenib as maintenance therapy unless they underwent stem cell transplantation. The dose of all agents could be reduced during consolidation and maintenance based on the available guidelines related to the various adverse effects.

Response Criteria

CR was defined by the presence of $< 5\%$ blasts in the bone marrow (BM) with $> 1 \times 10^9/L$ neutrophils and $> 100 \times 10^9/L$ platelets in the peripheral blood. CR duration was calculated from the time of CR until relapse. Relapse was defined by the recurrence of $> 5\%$ blasts in BM aspirate not related to recovery or the development of extramedullary disease. OS was calculated from the time of diagnosis until death.

FLT3 Mutation Detection

FLT3-ITD and codon 835/836 TKD mutation status were determined in DNA from initial, postinduction, follow-up, and relapsed unsorted BM aspirate samples by a polymerase chain

reaction (PCR) based method, with an analytical sensitivity of 1%–2% mutation-bearing cells. *FLT3* allele burden was determined by the ratio of the area under the mutated and unmutated PCR amplicon peaks as detected software calls after capillary electrophoresis on a 3100 or 3130 Genetic Analyzer (Applied Biosystems, Foster City, CA). A manual 400-cell differential performed on smears and multicolor flow cytometry on aspirate samples was used to track the levels of residual leukemia blasts. When leukemic blasts were detected, ratios of *FLT3* mutation were normalized to blast count. Relapsed BM samples (or skin biopsy in 1 case) were analyzed for kinase domain mutations in exons 16, 17, 20, and 21 by using PCR-based Sanger sequencing of genomic DNA (sensitivity level of 20% mutation-bearing cells). In diagnostic BM samples, exon 1 and 2 *KRAS* and *NRAS* point mutations were assessed by using PCR-based pyrosequencing of genomic DNA (sensitivity level of 5% mutation-bearing cells) and *NPM1* exon 12 mutation and/or duplications were assessed by using a PCR-based capillary electrophoresis method (sensitivity level of 2% mutation-bearing cells) as previously described.^{35,36}

Statistical Analysis

The main objective of this trial was to provide an assessment of the efficacy of adding sorafenib to combination chemotherapy. Survival was calculated by the Kaplan-Meier method, and different categories were compared by the log-rank test. Differences in subgroups by different covariates were evaluated by using the χ^2 test for nominal values (high vs. low *FLT3* mutation burden) and the Mann-Whitney *U* test and Fisher exact test for continuous variables (*FLT3* burden and outcome).

Results

Patient Characteristics

From October 2007 to September 2009, 64 patients with AML were enrolled into this phase II clinical study; 18 had mutated *FLT3* and are subjects of this report (Table 1). These patients are the patients who participated in the phase II portion of the study previously reported, including 3 patients not reported in the previous publication.³³ Their median age was 54 years (range, 20–65 years); 10 were women. Fifteen patients had *FLT3*-ITD, 2 had *FLT3*-TKD, and one had mutations at both sites. Five patients had a low *FLT3* mutation burden (ie, blast-normalized mutation ratio of <25%) consistent with presence in only a subset of the blast population. Two patients also had *RAS* mutations (both were *NRAS* exon 12), whereas 5 others had *NPM1* mutations. Most patients (83%) had a PS of 0 and 1 compared with only 3 (17%) patients with a PS of 2 and 3. Ten (55%) of 18 patients had diploid cytogenetics, and 8 had various abnormalities (deletions or additions but no translocations). The most common FAB subtype was M1 in 8 (44%) patients and M4 in 4 (22%) of them.

Response and Outcome

All 18 (100%) patients achieved CR or CR but lacked platelet recovery (CRp) (16 CR and 2 CRp). All but 2 patients responded after one cycle of induction; 2 patients achieved CRp after 2 cycles of induction. After a median follow-up of 9 months (range, 1–16 months), 10 (55%) patients had relapsed, with a median CR duration of 8.8 months (range, 1–9.5

months) The CR duration was independent of *FLT3* mutation burden at diagnosis ($P = .63$) (Figures 1, 2). Twelve (67%) patients proceeded to allogeneic stem cell transplantation; 5 in CR1, and 7 after relapse. Four patients died (3 of whom had matched unrelated allogeneic donor stem cell transplantation in either CR1 [$n = 1$] or CR2 [$n = 2$]), with a median OS of 14.8 months (range, 1–16 months). Survival duration was independent of *FLT3* mutation burden at diagnosis ($P = .9$) (Figures 3, 4). Three of the 4 deceased patients had a high mutant allele burden.

***FLT3* Mutation Levels After Initial Therapy**

Samples from all the patients after induction (approximately day 30) were collected and analyzed for molecular response (Figure 5). Ten (56%) patients showed no evidence of the mutated *FLT3* clone by PCR analysis, whereas 6 (33%) showed partial regression of the *FLT3* mutated clone, and 2 (11%) patients had persistent molecular evidence of the disease, with no preferential regression of the mutated clone. The degree of regression of the mutated *FLT3* clone did not predict relapse (the relapse rate in patients with complete regression was 6/10 [60%] vs. 5/8 [63%] in the rest). We also compared the regression of *FLT3* mutated clone in the study group with 67 patients with *FLT3-ITD* treated at our institution with standard anthracycline-cytarabine regimens without sorafenib. Overall, partial or complete *FLT3* regression after the first cycle of therapy was evident in 89% of the study patients compared with 6% among patients treated with regimens that lacked sorafenib ($P < .001$, χ^2 test).

Pattern of *FLT3* Mutation After Relapse

Nine of the 10 patients who relapsed had recurrence of *FLT3* mutated clone, although one had no evidence of mutated *FLT3* in the sample obtained at relapse. The relapse rate was independent of the level of *FLT3* mutation at diagnosis ($P = .63$). In 7 patients at relapse, sequencing of the TKD domain (which covers exons 16, 17, 20, and 21) revealed no additional mutation in *FLT3* in BM specimens ($n = 6$) or skin biopsy ($n = 1$).

Discussion

Patients with *FLT3*-mutated AML have a worse prognosis than those with WT *FLT3*. Predictors of outcome such as the length of the tandem duplication mutation as well as the ratio of WT to mutant allele have been reported.^{17–20,37} Several novel agents targeted against *FLT3* kinase have been evaluated in clinical trials.^{33,38–44} An improved response in patients with *FLT3*-mutated disease has been reported in these studies, but further studies are needed to demonstrate any effect on survival. Combining these agents with chemotherapy is preferred because a synergistic effect has been shown in in vitro studies.⁴⁵ This inhibitory effect is present in both WT- and mutant-*FLT3* cells in vitro but varies with dependence on *FLT3* signaling.⁴⁶

In this study, we observed an excellent response, with all patients achieving CR or CRp, which suggests a beneficial on-target effect. However, the majority of the patients relapsed suggesting the limited efficacy of the regimen to produce long-term remissions. Sorafenib was administered in induction and consolidation, and for maintenance for about 1 year. We

were unable to identify any secondary mutations that, in 6 of 10 patients who relapsed, would confer resistance to sorafenib as has previously been reported and as has been demonstrated for BCR-ABL inhibitors,^{26,29,30,32} which suggests that other mechanisms of resistance may exist, although there are several limitations associated with this conclusion. For instance, when looking for secondary mutations, we only sequenced 4 exons and not the whole *FLT3* gene. In addition, no pharmacokinetic data were collected in this phase II trial to demonstrate that adequate serum sorafenib levels were achieved. Finally, the small number of patients whose samples were sequenced further limits the validity of any conclusions.

The lack of correlation of molecular response with relapse makes any conclusions on the role of sorafenib and other kinase inhibitors in this disease difficult. For instance, the presence of minimal residual disease by PCR did not predict for relapse. Two patients relapsed with WT-*FLT3* AML, which raises the possibility of clonal evolution (vs. selective inhibition of the mutated clone and the appearance of less competing clones). Achieving maximal or partial regression of the mutated clone did not predict relapse because 3 of 7 patients who achieved partial regression continued to be disease free until the date of this article. Monocytic phenotype (M4, M5) did not show any worse outcome compared with other FAB subtypes (3/5 patients were disease free) that did not support the report by Koh et al,¹⁵ which suggests clinical significance of *FLT3* in monocytic subtype only.

In conclusion, induction therapy with sorafenib, idarubicin, and cytarabine is effective in attaining remission and in reducing the mutated clone in patients with mutated *FLT3* but does not universally eradicate it. Continuous therapy with sorafenib in induction may be beneficial in further reducing the leukemic clone; however, the extent of suppression of the clone is not predictive of relapse. No secondary *FLT3* mutations were found in the evaluated patients at relapse, which suggests that factors other than mutations that confer resistance to sorafenib may be important.

References

1. Ravandi F, Talpaz M, Estrov Z. Modulation of cellular signaling pathways: prospects for targeted therapy in hematological malignancies. *Clin Cancer Res.* 2003; 9:535–50. [PubMed: 12576416]
2. Meshinchi S, Appelbaum FR. Structural and functional alterations of FLT3 in acute myeloid leukemia. *Clin Cancer Res.* 2009; 15:4263–9. [PubMed: 19549778]
3. Gilliland DG, Griffin JD. The roles of FLT3 in hematopoiesis and leukemia. *Blood.* 2002; 100:1532–42. [PubMed: 12176867]
4. Stirewalt DL, Radich JP. The role of FLT3 in haematopoietic malignancies. *Nat Rev Cancer.* 2003; 3:650–65. [PubMed: 12951584]
5. Zheng R, Levis M, Piloto O, et al. FLT3 ligand causes autocrine signaling in acute myeloid leukemia cells. *Blood.* 2004; 103:267–74. [PubMed: 12969963]
6. Yamamoto Y, Kiyoi H, Nakano Y, et al. Activating mutation of D835 within the activation loop of FLT3 in human hematologic malignancies. *Blood.* 2001; 97:2434–9. [PubMed: 11290608]
7. Frohling S, Schlenk RF, Breitruck J, et al. Prognostic significance of activating FLT3 mutations in younger adults (16 to 60 years) with acute myeloid leukemia and normal cytogenetics: a study of the AML Study Group Ulm. *Blood.* 2002; 100:4372–80. [PubMed: 12393388]
8. Kottaridis PD, Gale RE, Frew ME, et al. The presence of a FLT3 internal tandem duplication in patients with acute myeloid leukemia (AML) adds important prognostic information to cytogenetic risk group and response to the first cycle of chemotherapy: analysis of 854 patients from the United

- Kingdom Medical Research Council AML 10 and 12 trials. *Blood*. 2001; 98:1752–9. [PubMed: 11535508]
9. Schnittger S, Schoch C, Dugas M, et al. Analysis of FLT3 length mutations in 1003 patients with acute myeloid leukemia: correlation to cytogenetics, FAB subtype, and prognosis in the AMLCG study and usefulness as a marker for the detection of minimal residual disease. *Blood*. 2002; 100:59–66. [PubMed: 12070009]
 10. Bacher U, Haferlach C, Kern W, Haferlach T, Schnittger S. Prognostic relevance of FLT3-TKD mutations in AML: the combination matters—an analysis of 3082 patients. *Blood*. 2008; 111:2527–37. [PubMed: 17965322]
 11. Whitman SP, Ruppert AS, Radmacher MD, et al. FLT3 D835/I836 mutations are associated with poor disease-free survival and a distinct gene-expression signature among younger adults with de novo cytogenetically normal acute myeloid leukemia lacking FLT3 internal tandem duplications. *Blood*. 2008; 111:1552–9. [PubMed: 17940205]
 12. Mead AJ, Linch DC, Hills RK, Wheatley K, Burnett AK, Gale RE. FLT3 tyrosine kinase domain mutations are biologically distinct from and have a significantly more favorable prognosis than FLT3 internal tandem duplications in patients with acute myeloid leukemia. *Blood*. 2007; 110:1262–70. [PubMed: 17456725]
 13. Mead AJ, Gale RE, Hills RK, et al. Conflicting data on the prognostic significance of FLT3/TKD mutations in acute myeloid leukemia might be related to the incidence of biallelic disease. *Blood*. 2008; 112:444–5. [PubMed: 18606888]
 14. Schlenk RF, Dohner K, Krauter J, et al. Mutations and treatment outcome in cytogenetically normal acute myeloid leukemia. *N Engl J Med*. 2008; 358:1909–18. [PubMed: 18450602]
 15. Koh Y, Park J, Ahn KS, et al. Different clinical importance of FLT3 internal tandem duplications in AML according to FAB classification: possible existence of distinct leukemogenesis involving monocyte differentiation pathway. *Ann Hematol*. 2009; 88:1089–97. [PubMed: 19296110]
 16. Kayser S, Schlenk RF, Londono MC, et al. Insertion of FLT3 internal tandem duplication in the tyrosine kinase domain-1 is associated with resistance to chemotherapy and inferior outcome. *Blood*. 2009; 114:2386–92. [PubMed: 19602710]
 17. Meshinchi S, Alonzo TA, Stirewalt DL, et al. Clinical implications of FLT3 mutations in pediatric AML. *Blood*. 2006; 108:3654–61. [PubMed: 16912228]
 18. Thiede C, Steudel C, Mohr B, et al. Analysis of FLT3-activating mutations in 979 patients with acute myelogenous leukemia: association with FAB subtypes and identification of subgroups with poor prognosis. *Blood*. 2002; 99:4326–35. [PubMed: 12036858]
 19. Gale RE, Green C, Allen C, et al. The impact of FLT3 internal tandem duplication mutant level, number, size, and interaction with NPM1 mutations in a large cohort of young adult patients with acute myeloid leukemia. *Blood*. 2008; 111:2776–84. [PubMed: 17957027]
 20. Stirewalt DL, Kopecky KJ, Meshinchi S, et al. Size of FLT3 internal tandem duplication has prognostic significance in patients with acute myeloid leukemia. *Blood*. 2006; 107:3724–6. [PubMed: 16368883]
 21. Wilhelm SM, Carter C, Tang L, et al. BAY 43-9006 exhibits broad spectrum oral antitumor activity and targets the RAF/MEK/ERK pathway and receptor tyrosine kinases involved in tumor progression and angiogenesis. *Cancer Res*. 2004; 64:7099–109. [PubMed: 15466206]
 22. Zhang W, Konopleva M, Shi YX, et al. Mutant FLT3: a direct target of sorafenib in acute myelogenous leukemia. *J Natl Cancer Inst*. 2008; 100:184–98. [PubMed: 18230792]
 23. Delmonte J Jr, Kantarjian HM, Andreeff M, et al. Update of a phase I study of sorafenib in patients with refractory/relapsed acute myeloid leukemia or high-risk myelodysplastic syndrome. *ASH Annual Meeting Abstracts*. 2007; 110:893.
 24. Mori S, Cortes J, Kantarjian H, Zhang W, Andreeff M, Ravandi F. Potential role of sorafenib in the treatment of acute myeloid leukemia. *Leuk Lymphoma*. 2008; 49:2246–55. [PubMed: 19052971]
 25. Metzelder S, Wang Y, Wollmer E, et al. Compassionate use of sorafenib in FLT3-ITD-positive acute myeloid leukemia: sustained regression before and after allogeneic stem cell transplantation. *Blood*. 2009; 113:6567–71. [PubMed: 19389879]
 26. Chu SH, Small D. Mechanisms of resistance to FLT3 inhibitors. *Drug Resist Updat*. 2009; 12:8–16. [PubMed: 19162530]

27. Ravandi F, Jilani I, Estey E, et al. Soluble phosphorylated fms-like tyrosine kinase III. FLT3 protein in patients with acute myeloid leukemia (AML). *Leuk Res.* 2007; 31:791–7. [PubMed: 17156841]
28. Piloto O, Wright M, Brown P, Kim KT, Levis M, Small D. Prolonged exposure to FLT3 inhibitors leads to resistance via activation of parallel signaling pathways. *Blood.* 2007; 109:1643–52. [PubMed: 17047150]
29. Bagrintseva K, Geisenhof S, Kern R, et al. FLT3-ITD-TKD dual mutants associated with AML confer resistance to FLT3 PTK inhibitors and cytotoxic agents by overexpression of Bcl-x(L). *Blood.* 2005; 105:3679–85. [PubMed: 15626738]
30. Williams A, Nguyen B, Levis M, Brown P, Small D. Mutations in FLT3/ITD produce varying levels of resistance to FLT3 tyrosine kinase inhibitors. *ASH Annual Meeting Abstracts.* 2009; 114:3776.
31. Heidel F, Solem FK, Breitenbuecher F, et al. Clinical resistance to the kinase inhibitor PKC412 in acute myeloid leukemia by mutation of Asn-676 in the FLT3 tyrosine kinase domain. *Blood.* 2006; 107:293–300. [PubMed: 16150941]
32. Bagrintseva K, Schwab R, Kohl TM, et al. Mutations in the tyrosine kinase domain of FLT3 define a new molecular mechanism of acquired drug resistance to PTK inhibitors in FLT3-ITD-transformed hematopoietic cells. *Blood.* 2004; 103:2266–75. [PubMed: 14604974]
33. Ravandi F, Cortes JE, Jones D, et al. Phase I/II study of combination therapy with sorafenib, idarubicin, and cytarabine in younger patients with acute myeloid leukemia. *J Clin Oncol.* 2010; 28:1856–62. [PubMed: 20212254]
34. Kantarjian H, O'Brien S, Cortes J, et al. Results of intensive chemotherapy in 998 patients age 65 years or older with acute myeloid leukemia or high-risk myelodysplastic syndrome: predictive prognostic models for outcome. *Cancer.* 2006; 106:1090–8. [PubMed: 16435386]
35. Lin P, Jones D, Medeiros LJ, Chen W, Vega-Vazquez F, Luthra R. Activating FLT3 mutations are detectable in chronic and blast phases of chronic myeloproliferative disorders other than chronic myeloid leukemia. *Am J Clin Pathol.* 2006; 126:530–3. [PubMed: 16938665]
36. Chen W, Jones D, Medeiros LJ, Luthra R, Lin P. Acute myeloid leukaemia with *FLT3* gene mutations of both internal tandem duplication and point mutation type. *Br J Haematol.* 2005; 130:726–8. [PubMed: 16115128]
37. Whitman SP, Archer KJ, Feng L, et al. Absence of the wild-type allele predicts poor prognosis in adult de novo acute myeloid leukemia with normal cytogenetics and the internal tandem duplication of FLT3: a cancer and leukemia group B study. *Cancer Res.* 2001; 61:7233–9. [PubMed: 11585760]
38. Smith BD, Levis M, Beran M, et al. Single-agent CEP-701, a novel FLT3 inhibitor, shows biologic and clinical activity in patients with relapsed or refractory acute myeloid leukemia. *Blood.* 2004; 103:3669–76. [PubMed: 14726387]
39. Knapper S, Burnett AK, Littlewood T, et al. A phase 2 trial of the FLT3 inhibitor lestaurtinib (CEP701) as first-line treatment for older patients with acute myeloid leukemia not considered fit for intensive chemotherapy. *Blood.* 2006; 108:3262–70. [PubMed: 16857985]
40. Levis M, Ravandi F, Wang ES, et al. Results from a randomized trial of salvage chemotherapy followed by lestaurtinib for FLT3 mutant AML patients in first relapse. *ASH Annual Meeting Abstracts.* 2009; 114:788.
41. Stone RM, Fischer T, Paquette R, et al. A phase 1b study of midostaurin (PKC412) in combination with daunorubicin and cytarabine induction and high-dose cytarabine consolidation in patients under age 61 with newly diagnosed de novo acute myeloid leukemia: overall survival of patients whose blasts have FLT3 mutations is similar to those with wild-type FLT3. *ASH Annual Meeting Abstracts.* 2009; 114:634.
42. Stone RM, DeAngelo DJ, Klimek V, et al. Patients with acute myeloid leukemia and an activating mutation in FLT3 respond to a small-molecule FLT3 tyrosine kinase inhibitor, PKC412. *Blood.* 2005; 105:54–60. [PubMed: 15345597]
43. O'Farrell AM, Yuen HA, Smolich B, et al. Effects of SU5416, a small molecule tyrosine kinase receptor inhibitor, on FLT3 expression and phosphorylation in patients with refractory acute myeloid leukemia. *Leuk Res.* 2004; 28:679–89. [PubMed: 15158089]

44. Yee KW, Schittenhelm M, O'Farrell AM, et al. Synergistic effect of SU11248 with cytarabine or daunorubicin on FLT3 ITD-positive leukemic cells. *Blood*. 2004; 104:4202–9. [PubMed: 15304385]
45. Levis M, Pham R, Smith BD, Small D. In vitro studies of a FLT3 inhibitor combined with chemotherapy: sequence of administration is important to achieve synergistic cytotoxic effects. *Blood*. 2004; 104:1145–50. [PubMed: 15126317]
46. Knapper S, Mills KI, Gilkes AF, Austin SJ, Walsh V, Burnett AK. The effects of lestaurtinib (CEP701) and PKC412 on primary AML blasts: the induction of cytotoxicity varies with dependence on FLT3 signaling in both FLT3-mutated and wild-type cases. *Blood*. 2006; 108:3494–503. [PubMed: 16868253]

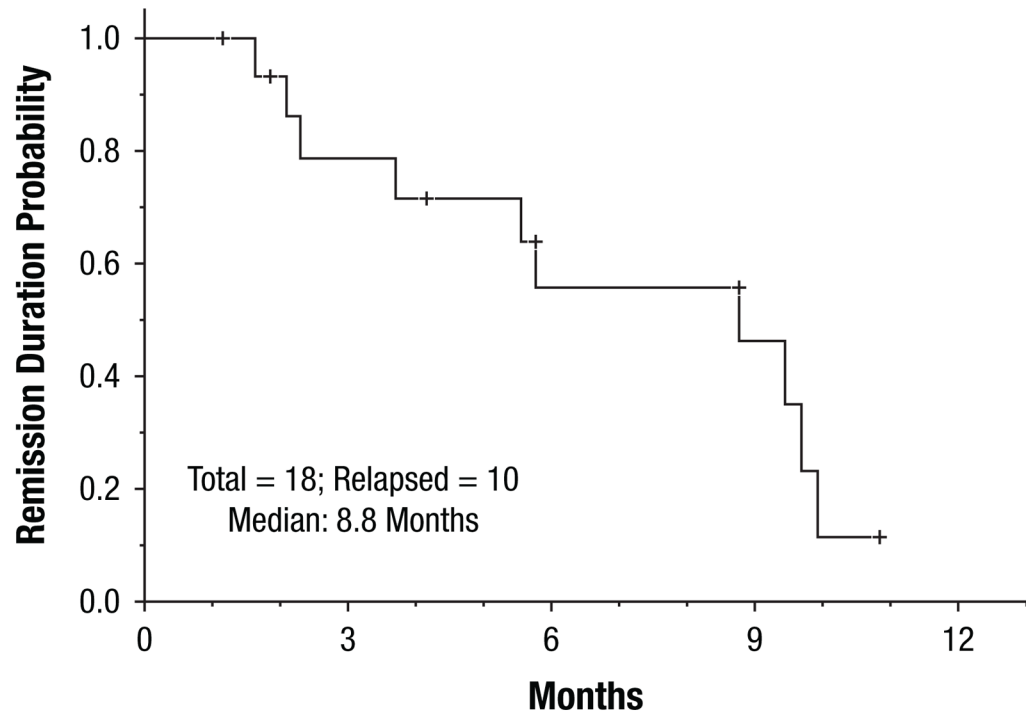


Figure 1.
Complete Remission Duration

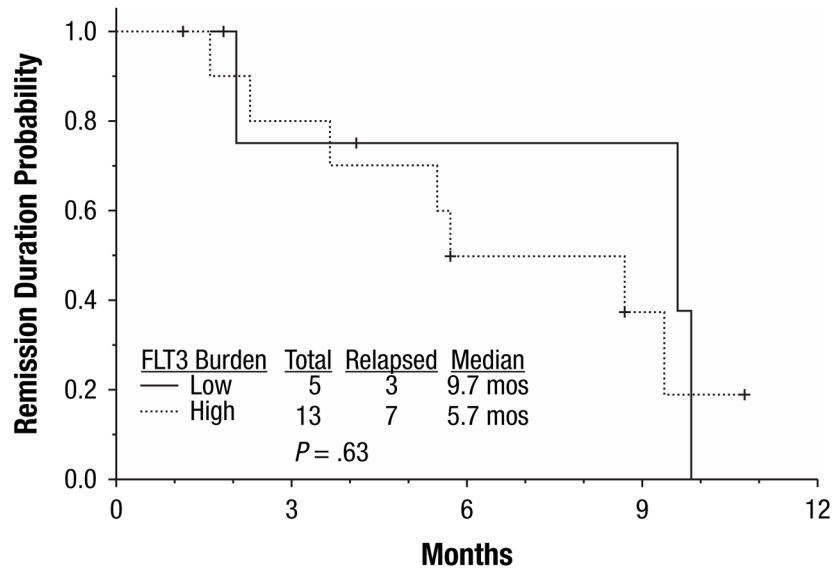


Figure 2.
Complete Remission Duration for Patients with High- and Low-*FLT3* Mutation Burden

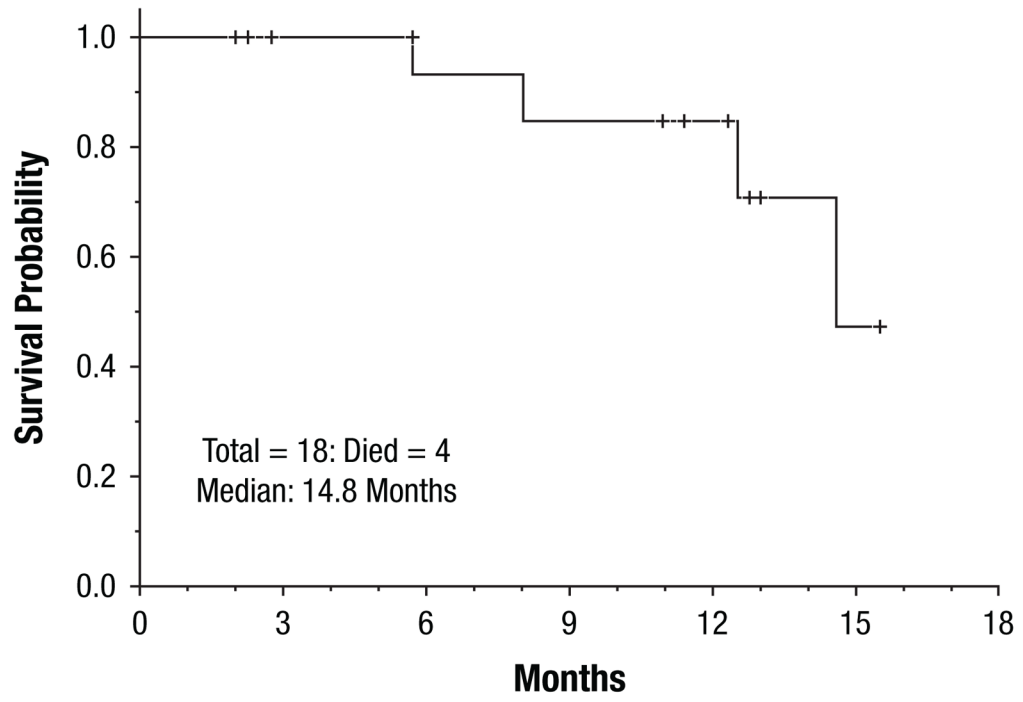


Figure 3.
Overall Survival

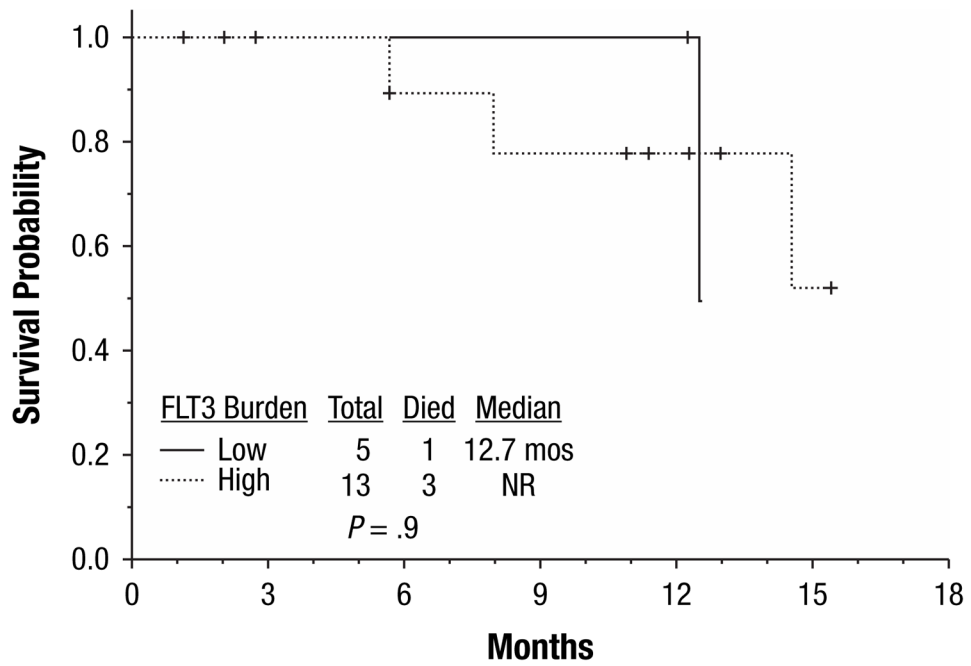


Figure 4.
Overall Survival for Patients with High and Low FLT3 Mutation Burden

	Dx	CR	3	6	9	12	15
ITD	H	+ P	+	+		+	†
	H	+ P	+	+			
	H				+	+	
	L					†	
	H			+			†
	H	+		+			†
	L					+	
	H	+ P					
	L					+	
	H						
	H	+	+				
	H	+ P	+	+	+		
	L						
	H	+ P					
	H						
ITD/TKD	H	+ P					
TKD	L						
	H			+			

Figure 5. Follow-up, Morphologic and Molecular Relapse
 Red Color = Positive for Leukemia by Flow Cytometry (FCM) and/or Morphology; Green Color = Negative for Leukemia by FCM and/or Morphology; Orange Color = Extramedullary Acute Myeloid Leukemia (AML); H = High Mutant *FLT3* Burden, L = Low Mutant *FLT3* Burden, † = Deceased, + = Positive Minimal Residual Disease by Polymerase Chain Reaction for Mutant *FLT3*; P = Partial Regression of Mutant *FLT3* clone

Table 1

Patient Characteristics

Patient Characteristics	
Median (Range) Age (y)	54 (20–65)
<i>FLT3</i> Mutation Type	
ITD	15 (83%)
TKD	2 (11%)
ITD/TKD	1 (6%)
<i>FLT3</i> Mutation Burden	
High ^a	13 (72%)
Low	5 (28%)
Eastern Cooperative Oncology Group Performance Status	
0	5 (28%)
1	10 (55%)
2	2 (11%)
3	1 (6%)
Median (Range) White Blood Cell Count ($\times 10^9/L$)	18 (1.4–196)
Median (Range) Hemoglobin (g/dL)	8.5 (6.9–10.8)
Median (Range) Platelet ($\times 10^9/L$)	51.5 (18–189)
Cytogenetics	
Diploid	10 (55%)
Other	8 (45%)

^a 25% blast-normalized mutation ratio (see Methods section).