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## Antioxidant Defenses in the Ocular Surface

YING CHEN, PHD<sup>†</sup>, GAURAV MEHTA, MVS<sup>†</sup>, and VASILIS VASILIOU, PHD<sup>\*</sup>

Molecular Toxicology and Environmental Health Sciences Program, Department of Pharmaceutical Sciences, University of Colorado Denver, Aurora, CO, USA

### Abstract

The human eye is subjected constantly to oxidative stress due to daily exposure to sunlight, high metabolic activities, and oxygen tension. Reactive oxygen species generated from environmental insults and pathological conditions render the human eye particularly vulnerable to oxidative damage. The ocular surface composed of the tear film, the cornea, and the aqueous humor forms the first physical and biochemical barrier of the eye and plays a pivotal role in combating free radicals. These ocular compartments are enriched in certain antioxidants in the form of metabolic enzymes or small molecules. Such an antioxidant defense system in the ocular surface is essential for the maintenance of redox homeostasis in the eye and protection against oxidative damage. Herein, we review the properties and functions of key constituent antioxidants of the ocular surface.

### Keywords

age-related eye disease; antioxidants; free radicals; ocular defense system; reactive oxygen species; vitamin A; vitamin E

## I. INTRODUCTION

Daily exposure to sunlight and atmospheric oxygen challenges the eye with an intense burden of oxygen free radicals, making it highly vulnerable to oxidative damage. Generation of reactive oxygen species (ROS) and resultant oxidative stress have been implicated in the pathogenesis of numerous forms of eye disease, including photokeratoconjunctivities,<sup>1</sup> photokeratitis,<sup>2</sup> pingueculae and pterygia,<sup>3</sup> cataract,<sup>4–6</sup> glaucoma, and macular degeneration.<sup>7,8</sup> The cornea covers the outermost layer of the eye, serving two fundamental functions: 1) It is the initial barrier of the eye that protects the inner ocular tissues (such as lens and retina) against external insults; and 2) together with the lens, it constitutes the “refraction unit” of the eye to permit light to enter and focus on the retina.<sup>9</sup>

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Single-copy reprint requests to Vasilis Vasiliou, PhD. Corresponding author: Vasilis Vasiliou, PhD, Department of Pharmaceutical Sciences, University of Colorado Denver, Aurora, CO 80045, USA. Phone: (303) 724-3520; fax: (303) 724-2666; vasilis.vasiliou@ucdenver.edu.

<sup>†</sup>These authors have contributed equally to this work and should be considered as first authors.

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The cornea consists of three cellular layers, including a stratified squamous epithelium; a thick stroma containing collagen fibers, proteoglycans, glycosaminoglycans and keratocytes; and a posterior single layer of endothelium. Being a unique avascular tissue, the cornea depends on the tear film and the aqueous humor to provide its anterior and posterior surfaces, respectively, with nutrients and protective molecules. The ocular surface, including the tear film, the cornea, and the aqueous humor, is the first line to encounter environmental insults and play a pivotal role in protecting the inner ocular tissues against oxidative damage. In doing so, the ocular surface has developed a range of defense mechanisms in the form of enzymatic and non-enzymatic antioxidant molecules. Some of these molecules act to scavenge or neutralize ROS, while others function to repair the damage caused by these free radicals.

## II. FORMATION OF REACTIVE OXYGEN SPECIES IN THE EYE

ROS derive from diatomic oxygen ( $O_2$  [Figure 1]) and are produced as byproducts during cellular metabolism. As such, the generation of ROS normally correlates with cellular metabolic rate.<sup>10</sup> The addition of one electron to dioxygen forms the superoxide anion radical ( $O_2^{\cdot-}$ ), which occurs mostly during the process of mitochondrial respiration.<sup>11</sup> The dismutation of this molecule by the enzymes superoxide dismutases forms hydrogen peroxide ( $H_2O_2$ ).<sup>12</sup>  $H_2O_2$  is mostly scavenged efficiently by the enzymes glutathione peroxidase in the mitochondria<sup>13</sup> or by the enzyme catalase in peroxisomes.<sup>14</sup> Although hydrogen peroxide is less reactive than superoxide, the breakdown of this molecule by various transition metals ( $Fe^{2+}$ ,  $Cu^+$ , and others) via Fenton reaction can generate the highly reactive hydroxyl radical ( $OH^{\cdot}$ ).<sup>15</sup> The reaction of superoxide or hydroxyl radical with polyunsaturated fatty acids generates the peroxy radical ( $LOO^{\cdot}$ ).<sup>16</sup> In addition to the mitochondrial origin, ROS in the form of superoxide anion and hydrogen peroxide can be generated through the activities of cytoplasmic oxidases, such as xanthine oxidase, histamine oxidase, and monoamine oxidase.<sup>17</sup>

Harboring unpaired electron(s), ROS provide a great degree of reactivity to induce reduction-oxidation (redox) reactions, thereby having the potential to attack important biomolecules. Under physiological conditions, cellular redox homeostasis is maintained by a delicate balance between ROS generation and antioxidant systems (Figure 1). Oxidative stress occurs in biological systems when such balance is disrupted so that ROS is overproduced. Excessive ROS can cause damage to DNA, cellular lipids, and proteins, thereby inhibiting their biological functions.<sup>18</sup> As the most reactive molecule, hydroxyl radical has the potential to react with each component of the DNA molecule, damaging the purine, pyrimidine bases and the deoxyribose backbone<sup>19</sup>; such permanent modification of DNA initiates the process of mutagenesis and carcinogenesis. The amino acid residues of proteins, especially cysteine and methionine residues, are prone to oxidation by ROS; oxidations of structurally or functionally important sites result in protein inactivation or misfolding.<sup>20</sup> The cellular membrane component is another target of ROS; lipid peroxidation occurs after the formation of peroxy radicals and generates cytotoxic products, such as 4-hydroxy-2-nonenal (HNE) and malondialdehyde (MDA).<sup>16</sup>

Given its intense exposure to light, robust metabolic activity, and high oxygen tension, the human eye is constantly subjected to oxidative stress. Solar ultraviolet radiation (**UVR**) is the major environmental inducer of ROS formation in the eye. UVR consists of UVA (315–400 nm), UVB (280–315 nm), and UVC (100–280 nm). All UVC and most UVB are absorbed by the cornea, whereas UVA is primarily absorbed by the lens (Figure 2). No UVC or UVB and very little UVA (<1%) reach the retina.<sup>21</sup> Absorption of UVR by these ocular tissues, particularly associated with the shorter and higher energy wavelengths of UVC and UVB, lead to photochemically generated ROS, including singlet oxygen ( $^1\text{O}_2$ ), superoxide anion, hydroxyl and peroxy radicals.<sup>21</sup>

All three spectrums of UVR have been shown to cause strand breakage, pyrimidine and thymine dimer formation, and protein cross-linking.<sup>22,23</sup> These UVR-induced molecular modifications in the eye have been associated with eye pathologies, including cataract formation and corneal and retinal degeneration.<sup>24</sup> In addition, oxidative stress is a common threat to ocular tissues in certain pathological conditions of the eye, such as inflammation and diabetes, where a substantial amount of ROS can be produced.<sup>25,26</sup> Xanthine oxidase-mediated ROS formation in the tear film has been suggested to contribute to the hypoxia-reoxygenation injury of the cornea associated with prolonged wearing of hydrophilic contact lenses.<sup>27</sup> Facing such challenge, the ocular surface, with its robust antioxidant defense systems, functions crucially in maintaining the redox homeostasis of the eye.

### III. ANTIOXIDANTS IN THE CORNEA

#### A. Enzymatic Antioxidants

To date, the enzymatic antioxidants that have been documented to be present in the cornea include superoxide dismutases (**SODs**), catalase (**CAT**), glutathione peroxidases (**GPXs**) and reductase (**GR**), and glucose-6-phosphate dehydrogenase (**G6PD** [Table 1]). It is reported that, in normal corneal epithelium, the total level of SOD activity is much higher than CAT activity, which is higher than that of GPX.<sup>28</sup> In addition, certain members of aldehyde dehydrogenase (**ALDH**) superfamily that are expressed abundantly in the cornea are recognized to act as antioxidants.

**1. Superoxide Dismutases**—SODs (EC 1.15.1.1) catalyze the reaction for the formation of less reactive hydrogen peroxide from superoxide radical (Figure 1).<sup>29</sup> Three different SOD isozymes have been found to be constitutively expressed in the cornea of various species, including human: the extracellular SOD containing copper-zinc (**EC-SOD**, homotetrameric with a 26kDa subunit), the cytosolic SOD containing copper-zinc (**CuZn-SOD**, homodimeric with a 16kDa subunit), and the mitochondrial SOD containing manganese (**Mn-SOD**, homotetrameric with a 21–25kDa subunit).<sup>30,31</sup> More importantly, these enzymes are biologically functional in the corneal epithelial and endothelial layers.<sup>32</sup>

The activity of SOD in the cornea seems to vary depending on species, ranging from high in rabbits and guinea pigs to low in pigs and almost absent in cattle.<sup>33</sup> Behndig et al have reported a comparable level of activity (~85 U/mg protein) for EC-SOD and CuZn-SOD in human corneas, whereas Mn-SOD has the lowest level among the three (~5.7 U/mg protein).<sup>31</sup> Three individual mouse lines harboring the deficiency in *Sod1*, *Sod2* or *Sod3*, the

gene encoding the cytosolic CuZn-SOD, mitochondrial Mn-SOD or extracellular EC-SOD, respectively, have been generated.<sup>34–37</sup> Homozygous mutants of these mouse lines reveal distinct eye phenotypes (Table 2), indicating a differential role for the SOD isozymes in eye physiology. In particular, the protective role of extracellular SOD in the cornea is evidenced in EC-SOD knockout animals, which show increased spontaneous age-related loss of endothelial cells and increased susceptibility to oxidative stress and inflammatory damage.<sup>34</sup>

## 2. Glutathione Peroxidases and Glutathione Reductase—GPXs (EC 1.11.1.9)

represent a family of enzymes that catalyze the reduction of H<sub>2</sub>O<sub>2</sub> or organic hydroperoxides to water or corresponding alcohols using glutathione (**GSH**) as the electron donor.<sup>13,34</sup> By catalyzing this reaction, GPXs bring about an end to the peroxide-dependent chain reaction of free radical generation and resultant membrane damage of ocular tissues.<sup>38</sup> GR functions to recycle GSH from its oxidized form (**GSSG**), thereby maintaining the level of this important nonenzymatic antioxidant (see below) in the cornea. In addition, GR can reduce protein thiols to their native state. It has been reported that corneal GR activity is elevated significantly in inflamed corneas, suggesting a role of GR in combating inflammation-derived ROS.<sup>39</sup> The activities of GPX and GR have been found in both corneal epithelium and endothelium.<sup>40,41</sup> Deficiency of GPX1, the isoform identified in the cytosol, nucleus, and mitochondria, results in increased oxidative damage and associated pathology in lens and retina in a gene-knockout mouse model<sup>42</sup>; however, no corneal pathology has been reported in this animal model (Table 2).

## 3. Catalase—CAT (EC 1.11.1.6), a homotetramer with a 60kDa subunit, is an important scavenger of hydrogen peroxide present outside of mitochondria (Figure 1). It functions not only to protect ocular tissues against hydrogen peroxide, but also to protect superoxide dismutases from inactivation.<sup>43</sup> This enzyme in its bioactive form has been detected in normal corneal epithelium and endothelium.<sup>39,44</sup> In corneal cell cultures, overexpression or inhibition of CAT confers protection or susceptibility, respectively, to ROS-induced injury.<sup>45,46</sup> Interestingly, Heck et al proposed a novel function of this enzyme in response to UVB exposure in skin keratinocytes.<sup>47</sup> They hypothesized that CAT absorbs UVB light directly and, as a product of this absorption, it mediates the production of ROS that can be detoxified by other antioxidants. It is worth speculating that such a UVB-filtering property of CAT may contribute to its protective role in the cornea aside from the antioxidant function of this enzyme. Catalase null mice display increased sensitivity to ROS-mediated tissue injuries; however, no eye pathology has been reported in the mutant mice.<sup>48</sup>

## 4. Glucose-6-phosphate Dehydrogenase—G6PD (EC 1.1.1.49) contributes to the enzymatic defense mechanisms of the cornea against oxidation by maintaining the cellular reductive potential in the form of nicotinamide adenine dinucleotide phosphate (**NADPH**, see below) through the pentose phosphate pathway.<sup>49</sup> This enzyme supplies NADPH as the electron donor for GR-mediated reduction of GSSG and consequently acts to balance the cellular pool of GSH. The important role of GR activity in corneal defense against UV-induced oxidative stress is implicated by enhanced GR activity in porcine corneas exposed to UVA or a small dose of UVC.<sup>50</sup> Nevertheless, no eye pathologies have been reported in human subjects or rodent models with G6PD deficiencies.<sup>51,52</sup>

**5. Aldehyde Dehydrogenases 3A1 and 1A1**—ALDH3A1 (EC 1.2.1.5) and ALDH1A1 (EC 1.2.1.36) belong to a superfamily of NAD(P)<sup>+</sup>-dependent enzymes that catalyze the oxidation of a wide variety of endogenous and exogenous aldehydes to their corresponding acids.<sup>53</sup> ALDH3A1 accumulates in high levels in the cornea of most mammals, including human, making up about 5–50% of the total water-soluble corneal proteins, depending on species.<sup>54</sup> The rabbit cornea, on the other hand, exceptionally expresses ALDH1A1 instead of ALDH3A1.<sup>55</sup> These enzymes are considered “corneal crystallins” and are believed to contribute to the transparent and refractory properties of the cornea.<sup>55</sup>

In addition to serving a structural role, ALDH3A1 and ALDH1A1 have been shown in rodent models to confer protection against lens oxidative damage and cataract formation (Table 2),<sup>56</sup> which is associated with UVR exposure. Similarly, it has been found that decreased ALDH3A1 activity is associated with pathologic corneas.<sup>57</sup> Given the abundant expression of these enzymes in the cornea, it is believed that ALDH3A1 and ALDH1A1 play a key role in protecting the eye from UV-induced damage by multifactorial mechanisms<sup>58</sup>: 1) acting as antioxidants by scavenging directly UV-induced free radicals or by producing the antioxidant NADPH, 2) direct absorption of UV light, and 3) metabolism of toxic aldehydes produced by UV-induced lipid peroxidation. Consistent with an antioxidant role for ALDH3A1 is the observation that overexpression of human ALDH3A1 prevents GSH depletion caused by oxidative agents in rabbit corneal fibroblast cells and protects human corneal epithelial cells from oxidant-induced DNA damage.<sup>59,60</sup>

## B. Nonenzymatic Antioxidants

**1. Ascorbic Acid**—Among the nonenzymatic antioxidants present in the cornea, ascorbic acid (vitamin C) is believed to be the most important one.<sup>9,33,61,62</sup> This water soluble molecule is a strong electron donor, readily reacting with a broad spectrum of free radicals, including superoxide anion and hydroxyl and peroxy radicals. Dehydroascorbate, the oxidized product of ascorbate, is then reduced back to ascorbate, using GSH or NADPH as electron donors.<sup>63</sup>

Ascorbate is abundant in various ocular compartments in diurnal animals that do or do not possess *de novo* ascorbate synthesis,<sup>64</sup> emphasizing a significant role for this antioxidant in the eye. The accumulation of ascorbate in the eye is achieved primarily by active transport through the iris-ciliary body into aqueous humor and subsequent transport into the cornea and lens.<sup>65</sup> The distribution of ascorbate in the cornea has been investigated in the bovine eye.<sup>66</sup> The highest ascorbate concentration (~1.56 mg/g) was found in the corneal epithelium, particularly in the central region covering the papillary area. A much lower level (~0.2 mg/g), although exceeding that of serum, was observed in the corneal stroma and endothelium. Based on this pattern of ascorbate distribution in the cornea and the contribution of corneal fractions to UV absorption, it is proposed that ascorbate may absorb the UVB spectrum of light between 280–310 nm, thereby protecting internal ocular structures against UVR-induced damage.<sup>66</sup> The difference in the levels of ascorbate in corneas of diurnal and nocturnal animals also supports an important role of this antioxidant in protecting the eye from light-induced stress and tissue damage.<sup>67</sup> It is suggested that

ambient radiation plays a role in sustaining the high ascorbate concentration in the corneal epithelium.<sup>68</sup> Despite all these facts, however, oral supplementation of ascorbate that increases ascorbate concentration of lens by 53% is not protective against UVB-induced cataract in guinea pigs.<sup>69</sup> It should be noted that ascorbate serves as an essential cofactor for collagen synthesis by corneal fibroblasts, by which ascorbate promotes healing of damaged corneal tissues and reduces the incidence of corneal ulceration and perforation.<sup>70</sup>

**2. Glutathione**—GSH, a tripeptide composed of glutamate, cysteine, and glycine, is ubiquitously synthesized in all cell types.<sup>71</sup> It is the most abundant cellular non-protein thiol, and it pairs with GSSG to function as the major cellular redox buffer. GSH scavenges hydroxyl radical and superoxide directly and serves as a cofactor for the enzyme GPXs in metabolizing hydrogen peroxide, as well as lipid peroxides (Figure 1).<sup>72</sup> Through the action of the glutathione S-transferases (GSTs), GSH can be conjugated to a great variety of electrophilic endogenous compounds and foreign chemicals, resulting in efficient and safe elimination.<sup>73</sup> Furthermore, GSH is able to regenerate other important antioxidants, Vitamins C and E, back to their active forms. The cellular redox status of GSH (GSH/GSSG ratio) has been suggested to be an important determinant of cell fate, including proliferation, apoptosis, and senescence.<sup>74,75</sup>

In addition to its role as a major antioxidant, GSH has been shown to participate in other physiological processes, including nucleotide metabolism, formation of lipid second messengers, regulation of nitric oxide homeostasis, and post-translational modification of proteins.<sup>76</sup> Corneal GSH is in the mM range (4~7 mM), and it accumulates mostly in the epithelium, where GSH concentration is five-fold higher than that in the stroma.<sup>76,77</sup> Unlike ascorbate, GSH in the corneal cellular layers can be synthesized via its *de novo* biosynthetic pathway, which is mediated by two cytosolic enzymes, glutamate-cysteine ligase (GCL) and glutathione synthetase (GSS).<sup>78</sup> Under oxidative stress, fast turnover of corneal GSH via its synthetic pathway and recycling by GR are required to counteract free radicals. In the cornea, GSH plays a pivotal role in maintaining the normal hydration level,<sup>41</sup> protecting the integrity of the cellular membrane and degrading xenobiotics.

Disrupted GSH homeostasis has been associated with various human diseases affecting cornea, such as viral infections and diabetes.<sup>79,80</sup> In rodent models with pharmacologically or genetically induced systemic GSH deficiency, ocular pathologies have been observed. For instance, in the mouse model with disrupted  $\gamma$ -glutamyl transpeptidase (*Ggt1*) gene encoding the membrane-bound enzyme that cleaves GSH to recycle precursor amino acids for GSH re-synthesis, lens GSH levels decrease by 90% and cataracts develop at an earlier age ubiquitously compared to normal mice (Table 2).<sup>81</sup> In a preliminary study on a mouse model with combined deficiency in GSH and ascorbate, pathologies in the cornea and lens have been noted (Chen and Vasiliou, unpublished observation). These studies provide convergent evidence to support a protective role of GSH against oxidative damage in the ocular tissues.

**3. Reduced Nicotinamide-Adenine Dinucleotide Phosphate**—NADPH and its oxidized product NADP form the other important cellular redox buffer. The cellular concentration of NADPH is in the  $\mu$ M range, but has been found to be higher in ocular tissues, probably because of the presence of abundant NADPH-dependent “enzyme-

crystallins” in the cornea and lens.<sup>82</sup> NADPH functions as an antioxidant in the cornea via multiple mechanisms: 1) It is a coenzyme of GR in the GPX/GR system for the regeneration of GSH from GSSG<sup>83</sup>; 2) It may act as a direct antioxidant by reducing glutathyl, tyrosyl, and peroxy nitrite radicals generated during oxidative stress<sup>84</sup>; 3) It is a UVR filter<sup>85</sup>; 4) It may protect other antioxidant enzymes from ROS-induced inactivation<sup>86</sup>; and, 5) It maintains a reducing potential for pyridine nucleotide-dependent redox-active enzymes, such as isocitrate dehydrogenase, malic dehydrogenase and 6-phosphate dehydrogenase, which are involved in protecting eye tissues.<sup>87</sup>

**4.  $\alpha$ -Tocopherol and Retinol**— $\alpha$ -Tocopherol (the most active form of vitamin E) is a fat-soluble antioxidant, and it functions uniquely to break the chain reaction of lipid peroxidation by quenching peroxy radicals.<sup>88,89</sup> This action of  $\alpha$ -tocopherol is crucial for the maintenance of cellular membrane integrity. In addition,  $\alpha$ -tocopherol can regenerate other antioxidants, including GSH and ascorbate.<sup>90</sup> A protective role of vitamin E in the eye is implicated by studies showing significant correlation between high plasma  $\alpha$ -tocopherol levels and lesser prevalence of cataract.<sup>91</sup> In agreement with this, vitamin E-enriched diet has been reported to protect UVR-induced damage in the cornea and lens.<sup>92</sup>

Retinol (vitamin A) not only plays a physiological role in normal vision, but also acts as a hydrophobic antioxidant by reducing oxidative stress and preventing apoptosis in corneal endothelial cells. It has been shown that vitamin A supplementation in the culture medium provides considerable protection in murine corneal endothelial cells against lipid peroxidation and oxidative damage induced by iron overload; this effect of vitamin A is greater than that of the combination of vitamin C and Vitamin E.<sup>93</sup> The antioxidant role of vitamin A is believed to be partially due to the polyene hydrophobic unit, which can quench singlet oxygen species, neutralize radicals, and stabilize peroxy radicals.<sup>94</sup>

**5. Ferritin**—In the presence of free iron, UVR generates the hydroxyl radical, which triggers lipid peroxidation and ocular tissue injury. Considering the amount and duration of UVR exposure of the cornea during a lifetime, it is highly necessary that the level of free iron is tightly regulated in this tissue and kept at a safe low level. This sequestering of iron in the cytoplasm is brought about by the iron-binding protein ferritin.<sup>22</sup> Cai et al showed that ferritin is a developmentally regulated protein in avian corneal epithelial cells and is expressed in the nucleus.<sup>95</sup> This nuclear ferritin protein is indistinguishable from the cytoplasmic form and can sequester iron, and, thus, acts as an antioxidant.<sup>22,95,96</sup> This function of nuclear ferritin in corneal epithelial cells has been suggested to be protective against UVR-induced DNA damage.<sup>97</sup>

**6. Albumin**—Serum albumin diffuses from peripheral blood vessels present around the cornea toward its center. Levels of albumin at 12–25% of total water soluble proteins have been reported in mammalian corneas,<sup>98</sup> suggesting an important role for this protein in the cornea. Albumin is distributed exclusively in the stromal layer of the cornea. Aside from being a binding and transporting protein, albumin has been shown to act as an antioxidant by scavenging hydrogen peroxide.<sup>98</sup>

#### IV. ANTIOXIDANTS IN THE TEAR FILM AND AQUEOUS HUMOR

The tear film and aqueous humor are important components of defense mechanisms in the ocular surface. The tear film covers the anterior surface of the cornea and is the first line of defense against external insults. It creates a smooth refractive surface on the cornea and protects the cornea from dehydration and environmental damage. It also functions in nourishing the anterior cornea and concentrating locally released biochemical metabolites. Consistent with a protective role against oxidative stress, the tear film contains both nonenzymatic and enzymatic antioxidants. In human tears, ascorbic acid (665  $\mu\text{M}$ ) and uric acid (328  $\mu\text{M}$ ) account for ~50% of the total antioxidant activity, with ascorbic acid being the most abundant and uric acid the second; other small molecules including GSH (107  $\mu\text{M}$ ), *L*-cysteine (48  $\mu\text{M}$ ) and *L*-tyrosine (45  $\mu\text{M}$ ) make up the rest.<sup>99</sup> The only reported antioxidant enzyme in the tear film is SOD, which displays a activity at 1~32 U/mg protein.<sup>31,100</sup>

The aqueous humor secreted by ciliary bodies is a clear and slightly alkaline liquid that occupies the space between the cornea and lens. It plays a crucial role in nourishing and protecting the corneal endothelium and the anterior epithelial lining of the lens. It also removes the metabolic wastes and biochemical products generated by the cornea and lens. Thus, ROS can be continuously generated in the aqueous humor in the form of hydrogen peroxide, superoxide anion, singlet oxygen, and peroxy radicals. The antioxidant profile of the aqueous humor resembles that of the tear film.<sup>101</sup> In the order of abundance in human aqueous humor, non-enzymatic antioxidants include ascorbic acid (530  $\mu\text{M}$ ), *L*-tyrosine (78  $\mu\text{M}$ ), uric acid (43  $\mu\text{M}$ ), *L*-cysteine (14.3  $\mu\text{M}$ ) and glutathione (5.5  $\mu\text{M}$ ). Quite differently in nocturnal rat, the aqueous humor is concentrated in thiol antioxidant GSH (125  $\mu\text{M}$ ) and *L*-cysteine (63  $\mu\text{M}$ ). The SOD activity at  $2.2 \pm 0.3$ ,  $2.7 \pm 1.2$  and  $<0.2$  U/ml for EC-SOD, CuZn-SOD and Mn-SOD, respectively, has been reported in the aqueous humor; it is believed that this trace amount of SOD activity does not contribute significantly to the antioxidant defense mechanisms of the aqueous humor.<sup>31</sup>

The high concentration of ascorbic acid in aqueous humors of diurnal species, likely resulting from more efficient sequestering of this molecule,<sup>102</sup> strongly supports an important role of ascorbic acid in protecting against light (UVR)-mediated damage. Indeed, systemic administration of ascorbic acid that increases the ascorbic acid level in the aqueous humor by 30-fold has been shown to protect lens epithelium against UVR-induced DNA damage in rat.<sup>103</sup> It is believed that three different mechanisms are involved in such a protective effect of ascorbic acid compartmentalized in the aqueous humor<sup>104</sup>: direct absorption of UVR; quenching the fluorescence of biomolecules; and controlling the fluorescence-mediated biotransformation. This quenching of fluorescence also facilitates the decrease of light scattering.<sup>104</sup> Amino acid *L*-tyrosine is electrochemically active and scavenges hydroxyl radicals and singlet oxygen species.<sup>101,105</sup> Uric acid is a purine metabolite and its antioxidant property has been documented in extracellular fluids, including the tear film and the aqueous humor.<sup>106</sup> The concentration of uric acid in human ocular fluids is only a fraction of the plasma concentration.<sup>101,107</sup> This water-soluble molecule has high reactivity toward singlet oxygen and hydroxyl radicals, serving as a potent scavenger for these ROS.<sup>107,109</sup> In addition, it has been proposed that uric acid

regulates the redox state of GSH-ascorbate system.<sup>107,108</sup> Amino acid L-cysteine replenishes the tissue GSH pool by supplying the rate-limiting substrate for GSH biosynthesis<sup>71</sup>; it also acts as an antioxidant directly via the thiol group.

Taken together, the extracellular fluid in the ocular surface, namely the tear film and the aqueous humor, is replete with water-soluble and low-molecular-weight antioxidants, which enrich the defense mechanisms of the cornea against oxidative stress.

## V. CONCLUDING REMARKS

The ocular surface is equipped with a battery of redundant and diverse antioxidants to combat oxidative stress arising from environmental exposure. While serving their functions, these antioxidants are consumed by ROS and/or damaged by radiations.<sup>33,64,102</sup> As age progresses, the production of these antioxidants and their functioning tend to decrease in the eye and almost completely halt during the later stages of life.<sup>2,110,111</sup> It is during and at this crucial stage that the pro-oxidant species begin to advance, resulting in certain pathological conditions of the eye, such as cataract and glaucoma.<sup>2,93,110</sup> Based on these observations, the effect of pharmacological supplements, mainly GSH and vitamins, on aging-related eye diseases have been investigated in experimental animal models. Results derived from these studies are contradictory,<sup>69,80,112–116</sup> suggesting the complexity in the disease etiologies.

The antioxidant defense network in the ocular surface, as well as in other ocular compartments, is characterized by the cross-talk between pathways or individual antioxidants, which is essential in maintaining the overall cellular redox balance in the eye. Classical examples include GPX-GSH-GR-NADPH and GSH-VitC-VitE systems. The fact that deficiency of one antioxidant is not always associated with eye pathologies may be explained by the redundancy of the antioxidant defense system in the ocular surface. On the other hand, the distinct role of certain antioxidants in ocular surface defense is reflected by ocular phenotypes resulting from their disruption. For instance, *Aldh3a1* null mice develop cataracts and punctate opacities in lens cortex as early as in 1 month, whereas *Aldh1a1* null mice develop cataracts much later in life (6–9 months).<sup>56</sup> Given the abundant expression of ALDH3A1 in the cornea, the protective effect of ALDH3A1 is most likely due to its filtering property. On the other hand, ALDH1A1 may protect against cataract formation by detoxifying the products of lipid peroxidation in both the cornea and lens. Furthermore, the highest incidence and most severe lenticular phenotypes are observed in *Aldh3a1/1a1* null mice, indicating an additive effect of ALDH3A1 and ALDH1A1. Future studies aimed at understanding the interplay between these antioxidants and the underlying mechanisms will provide valuable information on the pathogenesis and therapeutic interventions of ocular diseases.

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## Abbreviations

<b>ALDH</b>	Aldehyde dehydrogenases
<b>CAT</b>	catalase
<b>CuZn-SOD</b>	Copper-zinc superoxide dismutases
<b>EC-SOD</b>	Extracellular superoxide dismutases
<b>G6PD</b>	Glucose-6-phosphate dehydrogenase
<b>GPXs</b>	glutathione peroxidases
<b>GR</b>	glutathione reductase
<b>GSH</b>	Glutathione
<b>GSSG</b>	Oxidized form of glutathione
<b>GST</b>	Glutathione S-transferases
<b>H<sub>2</sub>O<sub>2</sub></b>	Hydrogen peroxide
<b>LOO</b>	Peroxyl radical
<b>Mn-SOD</b>	Manganese-containing superoxide dismutases
<b>NADPH</b>	Nicotinamide adenine dinucleotide phosphate
<b>OH</b>	Hydroxyl radical
<b>ROS</b>	Reactive oxygen species
<b>SODs</b>	superoxide dismutases
<b>UVR</b>	ultraviolet radiation

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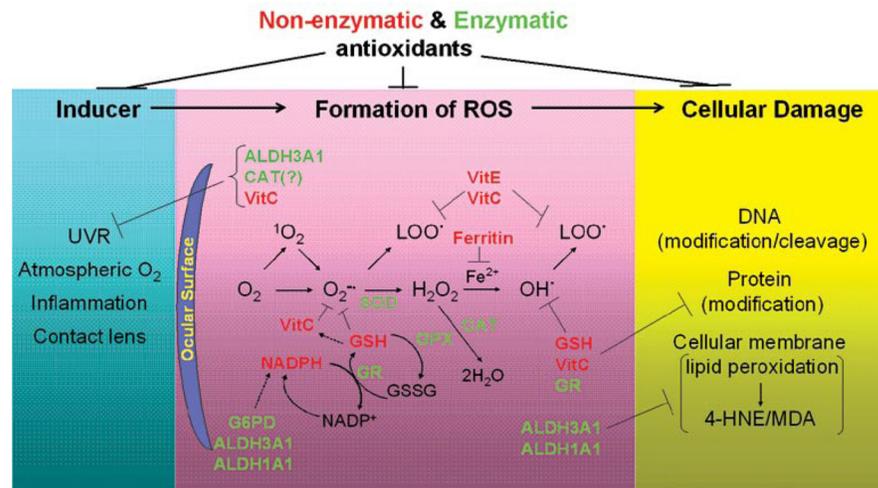
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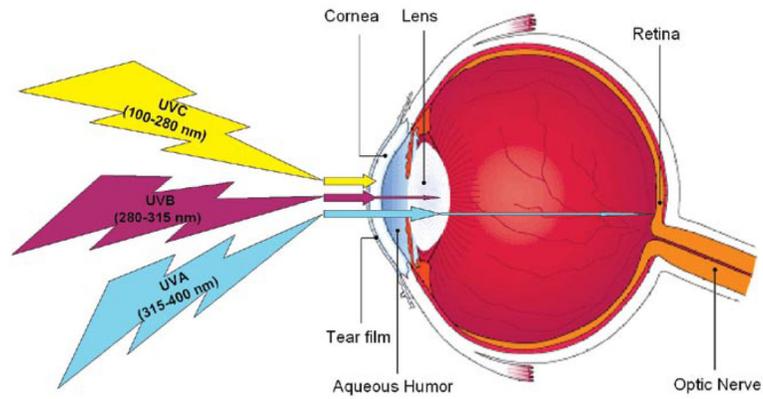
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**Figure 1.**

Formation of reactive oxygen species and antioxidant defenses in the eye. Environmental exposures (such as solar ultraviolet radiation and high atmospheric oxygen) and certain pathological conditions (such as inflammation and prolonged contact lens wearing) induce the generation of reactive oxygen species (ROS) in ocular tissues. ROS are derived from diatomic oxygen (O<sub>2</sub>), including superoxide anion radical (O<sub>2</sub><sup>•-</sup>), hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), hydroxyl (OH<sup>•</sup>), and peroxyl radicals (LOO<sup>•</sup>). ROS have high potential to react with DNA, proteins and cellular membranes, resulting in modifications of these macromolecules and consequent cellular damage. The antioxidant defense systems in the ocular surface function to combat ROS and protect ocular tissues from oxidative damage. Superoxide dismutases (SODs), catalase (CAT), glutathione peroxidases (GPXs), glutathione reductase (GR) and aldehyde dehydrogenases (ALDH3A1 and ALDH1A1) represent enzymatic antioxidants. Glutathione (GSH), ascorbic acid (VitC), α-tocopherol (VitE), NADPH and ferritin represent nonenzymatic anti-oxidants. The properties and functions of each antioxidant are discussed in detail in the text.



**Figure 2.**

The absorption of solar ultraviolet radiation by the eye. Solar ultraviolet radiation (UVR) consists of UVA at 315–400 nm, UVB at 280–315 nm, and UVC at 100–280 nm. The cornea absorbs all UVC and most UVB, whereas UVA is primarily absorbed by the lens. No UVC or UVB and very little UVA (<1%) reach the retina.

**Table 1**

Antioxidants present in the ocular surface

Compartment	Antioxidants	
	Enzymatic	Nonenzymatic
Tear film	Superoxide dismutases	Uric acid Ascorbic Acid Glutathione <i>L</i> -tyrosine <i>L</i> -cysteine
Cornea	Superoxide dismutases Glutathione peroxidases Glutathione reductase Catalase Glucose-6-Phosphodehydrogenase Aldehyde dehydrogenase 3A1 Aldehyde dehydrogenase 1A1	Ascorbic Acid Glutathione NADPH $\alpha$ -tocopherol Retinol Ferritin Albumin
Aqueous humor	Superoxide dismutases	Ascorbic Acid <i>L</i> -tyrosine Uric Acid <i>L</i> -cysteine Glutathione

\* The properties and functions of each antioxidant are discussed in detail in the text.

**Table 2**

Antioxidant-deficient rodent models displaying eye phenotypes

Gene	Antioxidant deficiency	Eye Phenotypes
Superoxide dismutase 1 (Sod1)	Cytosolic CuZn-SOD	Homozygous mutants are sensitive to diabetes-induced cataracts formation. <sup>37</sup>
Superoxide dismutase 2 (Sod2)	Mitochondrial Mn-SOD	Homozygous mutants show retinal pathologies before they die by 2.5 weeks. <sup>35</sup>
Superoxide dismutase 3 (Sod3)	Extracellular CuZn-SOD	Homozygous mutants show age-related loss of corneal endothelial cells and increased susceptibility to LPS-induced inflammatory endothelial damage. <sup>34</sup>
Glutathione peroxidase 1 (Gpx1)	Cellular GPX	Homozygous mutants show progressive lens pathologies with age and develop mature cataracts after 15 months. <sup>42</sup>
Aldehyde dehydrogenase 3A1 (Aldh3a1)	ALDH3A1	Homozygous mutants develop cataracts and punctate opacities in lens cortex by 1 month. <sup>56</sup>
Aldehyde dehydrogenase 1A1 (Aldh1a1)	ALDH1A1	Homozygous mutants develop cataracts at 6–9 months of age. <sup>56</sup>
Aldh3a1 and Aldh1a1	ALDH3A1 and ALDH1A1	Homozygous double mutants develop cataracts and punctate opacities in lens cortex by 1 month. <sup>56</sup>
$\gamma$ -glutamyl transpeptidase1 (Ggt1)	GSH	Homozygous mutants develop bilateral cataracts by 2–3 months of age. <sup>81</sup>