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## Functional Body Composition and Related Aspects in Research on Obesity and Cachexia

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### Abstract

The 12<sup>th</sup> Stock Conference addressed body composition and related functions in two extreme situations, obesity and cancer cachexia. The concept of “functional body composition” integrates body components into regulatory systems relating the mass of organs and tissues to corresponding *in vivo* functions and metabolic processes. This concept adds to an understanding of organ/tissue mass and function in the context of metabolic adaptations to weight change and disease. During weight gain and loss there are associated changes in individual body components while the relationships between organ and tissue mass are fixed. Thus, an understanding of weight regulation involves an examination of organ-tissue regulation rather than of individual organ mass. The between organ/tissue mass relationships are associated with and explained by cross-talk between organs and tissues mediated by cytokines, hormones, and metabolites that are coupled with changes in body weight, composition, and function as observed in obesity and cancer cachexia. In addition to established roles in intermediary metabolism, cell function and

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inflammation, organ-tissue cross-talk mediators are determinants of body composition and its' change with weight gain and loss. The 12<sup>th</sup> Stock Conference supported Michael Stocks' concept of gaining new insights by integrating research ideas from obesity and cancer cachexia. The conference presentations provide an in-depth understanding of body composition and metabolism.

### Keywords

Body composition; Metabolism; Mathematical Model; Nutritional Assessment; Skeletal Muscle; Adipose tissue; Bone mass; Obesity; Cachexia

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## INTRODUCTION

### From organ and tissue masses to functional relationships between organs and tissues

Manfred J. Müller

Although anatomy is central in medicine, it is only recently that the relationships between body composition and metabolic and functional characteristics came into the focus of research. During the last 20 years there were fascinating methodological and technical achievements in *in vivo* assessment of body composition and major whole body components have been characterized at different levels, i.e. the whole body (body mass) to tissues and organs (adipose tissue, brain, liver, skeletal muscle, bone), cells (fat and non-fat cell mass, extracellular mass, extracellular solids), molecular (fat, protein, minerals, water) and elements (e.g. whole body nitrogen and carbon content; 1). The descriptive concepts have been extended to the concept of functional body composition that integrates body components into regulatory systems by relating body components to their corresponding *in vivo* function and metabolic processes (2). Suitable applications of body composition analysis are (i) interpretation of body functions (e.g., fat-free mass (FFM) as the major determinant of energy expenditure) and their disturbances in the context of body components (e.g. insulin resistance related to ectopic fat accumulation in liver, skeletal muscle and pancreas) and *vice versa*, and (ii) interpretation of the meaning of individual body components in the context of their functional consequences (e.g., adaptation of energy expenditure to weight loss is related to fat mass and body water).

Accurate tools can now be used to assess body composition for risk prediction, 'phenotyping' the obese as well as the malnourished patients and their related co-morbidities (3). Individual body components such as fat mass are under hormonal and genetic control; they are also affected by environmental factors, lifestyle, and diseases. Regulation of body weight is a multiple (and at least in part) integrated control of individual body components. Since body components are inter-related and the relationships between individual body components are stable with weight changes, control of body weight seems to be on relationships between tissues and organs rather than on individual components or masses themselves.

Evidence for the idea comes from Benedicts' early starvation experiments of as well as Ancel Keys' seminal semi-starvation study which both have been re-analyzed more recently (4–6). In these studies, the ratio between losses in fat mass and losses in FFM remained

constant throughout a longitudinal weight loss protocol. There was some inter-individual variance in the so-called  $p$ -ratios (i.e., the ratio of protein energy mobilized to total energy mobilized) and baseline body composition was shown to be its major determinant (7,8). These coordinated changes in organ and tissue masses in response to caloric restriction go in parallel with mass- and tissue-independent changes in their specific metabolic rates (9).

Faced with the stable relations between individual organs, inter-organ and inter-tissue cross-talks are a challenging area of research. The present discussion is mainly about cross-talks of adipose tissue with other organs such as skeletal muscle, liver and brain brought about by a still increasing number of secretory products derived from the adipocyte. In addition, liver, skeletal muscle, kidney, bone, immune cells and the gastrointestinal tract have also been characterized as endocrine organs with a huge number of secretory products contributing to various between organ and tissue cross-talks and feedback control signals.

As for concepts, cross-talks may have different functional characteristics. In electronics, a cross-talk is characterized by an interaction of signals (10). Then, a cross-talk may happen at the near end such as between different fat depots and skeletal muscle. Alternatively, a sum near end cross-talk brings together multiple adjacent pairs such as the sum of signals generated in adipose tissue, muscle or immune cells acting on endothelial cells. In addition a far end cross-talk interferes between a peripheral organ and tissue and a central control unit, as for example between adipose tissue and the hypothalamus. Cross-talks may also be attenuated or compensated as might occur with the generation of brown fat cells within white adipose tissue as related to and affects the adipose tissue-skeletal muscle cross-talk. Finally, there may be interferences between different cross-talks as in the case when muscle-adipose tissue cross-talk interferes with the muscle-bone cross-talk.

It is challenging to discuss these new concepts in two opposite situations, obesity and cachexia. The feature common to both is perturbation in energy balance. As Michael Stock and others have already noted, both areas may provide possibilities for an integrated approach (11). Integration of research ideas and joint discussion of similarities and extremes will benefit future understanding and research in obesity and cachexia (11,12). If present, a malignant tumor adds an additional dimension both in terms of the specific metabolic rate of the tumor mass as well as its secretory products which may override normal physiological controls (see Cachexia section).

## CONCEPTS

### A model of functional body composition in humans

Kevin D. Hall

Dynamic changes in functional body composition result from an imbalance between the metabolizable energy content of the diet and the body's energy demands. More specifically, body composition changes at the chemical level result from imbalances between the macronutrients absorbed from the diet and the metabolic fuels oxidized to meet energy requirements. Since energy expenditure and metabolic fuel utilization are both strongly

influenced by body composition, there results a complex dynamic interplay between these variables.

Mathematical modeling provides a useful approach to dealing with this complexity (8). These models can be envisioned by analogy to a “*flex-fuel automobile*” that can run on an arbitrary mixture of different fuels (as occurs in humans with carbohydrate, protein, and fat). Such a “*flex-fuel vehicle*” would allow the driver to fill the tank with whatever fuel was cheaper or more readily available, regardless of what mixture is already in the tank. Designing such a vehicle would be a significant engineering challenge, but imagine how much more difficult it would be if the vehicle was not allowed to have a fuel tank. Rather, the vehicle itself must be composed of its fuel and would be continually breaking down and reconstructing its components. Despite the daily turnover of its components and fluctuations of fuel delivery, the composition of the vehicle must remain relatively stable and maintain similar performance characteristics. The human body accomplishes this remarkable engineering feat by the use of three dietary macronutrients, carbohydrate, protein, and fat to both fuel metabolism and provide substrates for body constituents.

While the physiological mechanisms underlying the regulation of human macronutrient metabolism and body composition dynamics are exceedingly complex, the whole-body system obeys thermodynamic laws that make the overall system amenable to mathematical modeling. These conservation constraints form the basis of mathematical models that relate macronutrient imbalances between dietary intake and metabolic utilization with changes in stored glycogen, protein, and fat. The changes in these chemical constituents and the fluid shifts associated with these changes allow for mathematical models of functional body composition dynamics at multiple levels of organization ranging from the chemical to tissue and organ levels.

Model predictions have been validated against the data from independent controlled feeding studies in humans (4,5,13,14) and were found to match the model predictions (8,15,16). This was not only true for prediction of weight and body composition changes, but also resting energy expenditure, carbohydrate and fat oxidation, nitrogen balance, and individual substrate fluxes, e.g. de novo lipogenesis, protein degradation and synthesis as well as gluconeogenesis (13–17). This can be extended by taking into account metabolic adaptations, i.e. the thermic effect of feeding, adaptive thermogenesis in response to underfeeding, and the effects of physical activity and exercise on energy expenditure and substrate oxidation rates. Applying models to available experimental data allows for exploration of unaccounted for variance and the potential to infer unmeasured quantities and reveal new metabolic findings (18).

Simplification of complex macronutrient balance models can be achieved by quantifying how energy imbalances are partitioned between either fat mass or fat free mass (or body protein) during weight loss and weight gain, and most models of human body composition dynamics have used such simplifications (8). It is also possible to derive energy partitioning relationships between different body fat compartments. For example, visceral adipose tissue (VAT) changes as fraction of the change in whole body fat mass (FM) is strongly correlated with the initial VAT/FM-ratio (19). Data suggest that there is a preferential loss in VAT

with a higher initial VAT/FM ratio and that the same allometric relationship describes the VAT response to exercise, diet, diet + exercise, and weight loss after bariatric surgery in both men and women.

Dynamic mathematical models provide an integrative framework to help design, predict, and interpret the results of human experiments and in clinical practice. For example, modeling weight loss in response to diet and exercise provides a sound basis in treating obese patients (17). Furthermore, mathematical modeling is beginning to help better understand the metabolic and body composition derangements that occur in diseases such as cancer cachexia (20).

### **Regulation of body composition: body components - brain feedback in weight control**

Abdul Dulloo

The regulation of body composition in response to energy deficit and energy surplus can be conceptualized as being brought about through control systems that operate via the control of body energy partitioning between lean and fat tissues, via compensatory changes in energy intake and via adaptive changes in thermogenesis. Despite considerable advances made over the past decades towards establishing the existence and operational modes of these control systems, a mechanistic explanation of body composition regulation in humans remains largely fragmentary. Fundamental questions have been raised and addressed from the analysis of data from longitudinal studies of experimentally-induced weight loss/recovery or weight gain/recovery. These findings have been integrated into conceptual models of body composition auto regulation and they pinpoint some of the important issues and gaps in knowledge about various components of feedback loops between changes in body composition and compensatory changes in energy intake and energy expenditure (6,21,22).

For example, the adipocyte-releasing hormone leptin – which acts on brain areas to induce satiety and enhance sympathetic control of thermogenesis – is often integrated in the lipostatic theory of weight regulation. Yet, the role of leptin as a circulating ‘adipostatic’ signal controlling body fat is questionable in view of the poor correlation between the kinetics of circulating leptin and dynamic changes in body fat in response to energy deprivation and refeeding. Furthermore, a feedback loop between body fat depletion and the brain circuitries controlling food intake cannot alone explain why human subjects recovering weight after starvation continue to overeat well after body fat has been restored to pre-starvation values, thereby resulting in ‘fat overshooting’. Indeed, a detailed re-analysis of the classic Minnesota Experiment of semistarvation and refeeding suggests that the autoregulatory component of the hyperphagic response to energy deprivation goes beyond an explanation based solely on the lipostatic theory since, in addition to the depletion of fat mass, the reduction in FFM also contributes to the compensatory hyperphagia (23).

The integrated outcome of this reanalysis suggests that the critical event that eventually leads to the prolongation of hyperphagia beyond the complete recovery of fat mass resides in the suppression of thermogenesis which drives fat recovery at a rate that is greater than that determined by the partitioning characteristic of the individual (24). As this enhanced

metabolic efficiency that drives fat acceleration is a function of fat depletion, and the prolongation of hyperphagia (after complete recovery of fat mass) is a function of depleted FFM still to be recovered, the extent of fat overshooting would therefore depend upon the extent to which both fat mass and FFM are depleted. This in turn would depend upon the partitioning characteristic of the individual which is known to be dictated primarily by the initial (prestarvation) adiposity. Indeed, the extent of fat overshooting can be shown to increase exponentially with decreasing initial %body fat (22). From a perspective of autoregulation of body composition therefore, lean dieters are at greater risk for fat overshooting than the obese dieters. Given the increasing prevalence of dieting among those in the normal-weight range (due to pressure for a slim image, body dissatisfaction or athletic performance), together with accumulating evidence suggesting increased cardiometabolic risks associated with weight fluctuations in the non-obese population groups (25), the notion that large fluctuations in body weight may predispose to increased fatness and the metabolic syndrome warrants greater experimental scrutiny and deserves greater public health concern than so far acknowledged.

Overall, the available evidence suggests that feedback signals from both fat and lean tissues operate in the regulation of body composition through their central effects on food intake and thermogenesis, although the various key components of these adipostatic and proteinostatic feedback mechanisms remain undefined. However, the discovery that a multiplicity of factors are secreted by adipocytes and myocytes opens new avenues in the search for adipostatic and proteinostatic feedback signals to the brain in the regulation of body composition, with major implications for the pathogenesis and management of obesity and cachexia. Furthermore, there is emerging evidence of a role for brain-muscle interactions in the mechanisms by which inflammatory signals mediate the loss of skeletal muscle mass during cachexia. In addition to the direct effects of inflammatory cytokines in inducing skeletal muscle atrophy, the integration of inflammatory cytokines signaling pathways within the central nervous system has been shown to play a critical role in muscle wasting via activation of the hypothalamic–pituitary–adrenal axis (26). Future investigations along such brain-muscle interactions are bound to provide powerful insights into the mechanisms of muscle wasting during cachexia and in the pathogenesis of sarcopenic obesity.

### **What is an appropriate energy expenditure for body composition?**

Steven B. Heymsfield

Determining if a subject's resting energy expenditure (REE) is low or high for its' body size is a pervasive question in clinical nutrition research (27,28). High or low REE values can signal variance in the composition of FFM (i.e. the relative proportion of high, metabolic rate organs like brain, heart, liver and kidneys), underlying metabolic disease, predisposition to weight gain or loss, or metabolic processes that are unrelated to body size. Early investigators applied body mass and height as size measures, all that they had available at the time, and formulated the Surface Law ( $REE \propto \text{Mass}^{0.66}$ ) and Kleiber's Law ( $REE \propto \text{Mass}^{0.75}$ ), although each has limitations when adjusting REE (29).



Body composition methods first developed and then introduced during the mid-twentieth century provided the first opportunity to identify ‘homogeneous’ metabolically active compartments (29). REE is highly correlated with the body size-composition measures and the adjusted value of REE should be independent of body size. Consider the simple regression model

$$\text{REE} = m\text{FFM} + b, \quad (1)$$

with  $m$  being the slope and  $b$  (REE) the y-axis intercept. For the ratio REE/FFM to be independent of FFM, the y-axis intercept must be zero so that

$$\text{REE} = m\text{FFM} \text{ and, thus,} \quad (2)$$

$$\text{REE/FFM} \quad (3)$$

is a stable value (i.e.  $m$ ) across all healthy adults. In fact, when REE is regressed against FFM, the  $b$  intercept term is positive and differs significantly in magnitude from zero. REE for men and women plotted against FFM results in a regression line with a significant intercept term of 410.4 kcal d<sup>-1</sup> (27,29).

FFM has remained the main compartment used by investigators over the past several decades to adjust REE for body size (27,28), although the need to remove low metabolic rate compartments other than fat from body mass as a means of estimating ‘meta-bolically active’ body mass was recognized.

At the tissue-organ level, the present availability of estimates of organ and tissue masses provided a new opportunity to develop physiological REE prediction formulas based upon established heat production rates of major body tissues (9,30,31). Tissue and organ mass can be estimated with great accuracy using CT or MRI; the energy expended by each tissue and organ can be calculated as the product of mass and mass-specific metabolic rate based on compiled from *in vivo* and *in vitro* observations. REE can be calculated as the sum of all tissue and organ metabolic rates. Viewing REE from the tissue-organ perspective the large contributions of four FFM components (brain, liver, heart and kidneys) to whole body REE becomes obvious. While <6% of representative body mass, these organs contribute to 60–70% of REE. REE is then calculated from

$$\begin{aligned} \text{REE(kcal/d)} \\ = (240 \times \text{brain mass}) + (200 \times \text{liver mass}) + (440 \times \text{heart mass}) + (440 \times \text{masses of kidneys}) + (13 \times \text{mass of skeletal m} \end{aligned} \quad (4)$$

Residual mass is the difference between the sum of measured body components and body weight. Today, physiological REE model terms for brown fat and the microbiome are not yet available.

Tissue-organ proportions vary with body size (i.e. between normal weight, overweight and obese subjects), in adolescents and adult age, across men and women, between race groups, and with stature (29,30,32). All tissues and organs scale to FFM with powers approaching 1.0, except brain, which has powers <0.2 for both men and women (29). Multiple regression

analysis was used to establish how skeletal muscle, heart, liver, kidney, brain and residual mass (weight minus the sum of other tissues, including adipose tissue and organs) scale to FFM. Tissue–organ mass was set as the dependent variable and FFM, age and adipose tissue mass as potential predictor variables in multiple regression models. The explained variance of traditional REE model with FFM alone as the predictor was 0.78 and SEE = 129.4 kcal d<sup>-1</sup>. Adding age and adipose tissue mass increased R<sup>2</sup> to 0.84; (SEE, 112.2 kcal d<sup>-1</sup>).

All these models had statistically significant intercepts and they resemble the general form of traditional FFM-based REE prediction models (29). However, brain, liver, skeletal muscle, bone, adipose tissue, residual mass and age added as significant covariates and the final R<sup>2</sup> reached 0.86 with lowest SEE (105.9 kcal d<sup>-1</sup>) of the developed series of REE prediction models with a non-significant intercept of 40.6 kcal d<sup>-1</sup>. Since REE links to tissue-organ level, the traditional REE-FFM model given in formula (1) is converted into

$$\text{REE(kcal/d)}=40.6+5.1\text{AT}+294.2\text{Br}+141.5\text{Li}+20.3\text{SM}+13.7\text{RM}-1.6\text{Age} \quad (5)$$

The newly defined compartments have improved correlations with REE estimates over body weight-height approaches, but all share a common limitation: REE-body composition ratios are not “constant” but vary across men and women and with race, age, and body size (27,28). The currently accepted alternative to ratio-based norms is to statistically adjust for predictors by applying regression models to calculate “residuals” that determine if a measured REE is relatively high or low (27,28).

The distinguishing feature of statistical REE-body composition models is a “non-zero” intercept, the cause of which is unknown. FFM is not a metabolically homogeneous compartment, but instead FFM varies systematically in tissue–organ proportions as a function of body size. The non-zero intercept observed with traditional REE models largely becomes non-significant when organ volumes (e.g., brain and/or liver mass) are included as model covariates (29). These new findings provide a context for future research aimed at establishing between subject differences in energy metabolism. To go beyond the whole body or tissue/organ mass is to directly measure specific energy expenditure of individual tissues and organs as well as their changes in response to changes in weight and body composition (9,31). These advances will likely improve our understanding of energy expenditure effects of disease, including cancer and other chronic conditions associated with cachexia,

## FUNCTIONAL ASPECTS OF ADIPOSE TISSUE AND MUSCLE

### Effects of over- and under-nutrition on body composition and metabolism of the mouse – what goes up doesn’t necessarily come down

John Speakman

When animals consume excess calories they gain weight, and when they consume insufficient calories relative to expenditure they lose weight. However these gross changes in body weight mask the details of the changes in body composition that accompany the responses to over- and under-nutrition. These relations were studied in adult mice (5 months



old) with stable body weights who were fed two graded levels of over nutrition (high fat diets with differing % fat up to 60%) and five graded levels of caloric restriction (from 0 to 40% less calories than their individual baseline intakes). As anticipated greater over nutrition led to greater total body weight gain and greater levels of under-nutrition led to greater levels of body weight loss.

The changes in 23 tissues assessed, however, were radically different. When animals gained weight they did so primarily by increasing the sizes of their fat stores in different body regions (i.e., subcutaneous, epididymal, retroperitoneal, omental, mesenteric and brown adipose tissue). The sizes of the vital organs (i.e., brain, liver, kidneys, heart, lungs, pancreas and spleen) were unaffected but structural organs (bone and skeletal muscle, pelage and tail) also increased in size.

During weight loss with caloric restriction there was also a loss in body fat, but the largest weight losses were actually among structural organs – like the skeletal muscle, skin and tail. The vital organs were also generally decreased but to a lesser extent than structural organs or fat. Testes mass decreased at the highest level of caloric restriction only, whereas the mass of accessory organs such as seminal vesicles linearly decreased with the degree of starvation. Some vital organs related to the alimentary tract (i.e., stomach, small intestine, caecum, large intestine) actually increased or remained unchanged in size under restriction.

Calculations reveal that tissue loss contributes to about 20% of the energy shortfall explaining 52 to 81% of the reduction in whole body energy expenditure. This is associated with by decreases in circulating IGF-1 and resistin levels.

Two hypotheses emerge about the implications of organ and tissue mass loss during caloric restriction. First, animals oxidize the energy derived from those tissues. Second, reducing organ-tissue mass lowers energy expenditure and thus contributes to metabolic adaptation. These changes reflect functional responses of the animals that attempt to minimize expenditure while preserving vital functions and sustaining food intake.

With overfeeding, weight gain is mostly explained by expansion of white adipose tissue. In addition, there were small changes in structural body components. By contrast, masses of vital organs and brown adipose tissue remained unchanged.

### **Functional correlates of fat mass - fat-free mass relationships during underfeeding and refeeding in humans**

Anja Bosy-Westphal

Short-term voluntary perturbations in energy balance lead to metabolic and neuro-endocrine adaptations that counteract weight changes. For example, regain of weight and fat mass may exceed prior weight loss, the so-called “*catch-up fat phenomenon*” (5,23). Fat regain at an intense rate can also occur in patients with cancer cachexia during short term remission of their disease, suggesting that the impetus for catch-up is also present in disease (33). Adaptive thermogenesis and reduced energetic efficiency of physical activity add to compensate body weight whereas insulin sensitivity and a low metabolic flexibility

contribute to the partitioning of weight regain (i.e. as gain of either fat mass or FFM (34–35). Beyond energy and macronutrient balances, diet composition (e.g., glycemic load, GL; glycemic index, GI; protein content) contributes to these adaptations.

When compared with high GI- diets with a lower protein content a low GI diet with a higher protein content resulted in improved weight loss maintenance in weight-reduced obese patients (36). These findings point to insulin secretion and/or insulin sensitivity as determinants of weight gain. However, insulin sensitivity had no effect on weight gain (37) but in Pima Indians, insulin resistance was associated with reduced risk of weight gain (38). By contrast, improved insulin sensitivity with energy restriction was associated with lower weight regain after previous weight loss (39).

Using a controlled dietary intervention study protocol to investigate whether diets differing in carbohydrate content and GI affect regain in fat and FFM under conditions of fixed energy intake and physical activity, 32 healthy, normal weight, young men followed 1-week overfeeding protocol (mean weight gain of 1.8 kg), 3-weeks of caloric restriction (mean weight loss of about 6.0 kg) and 2 weeks of hypercaloric re-feeding at  $\pm 50\%$  energy requirement (mean weight re-gain of 3.4 kg; for details see 40–43). During refeeding four study groups differing in carbohydrate intake (50%CHO, 65%CHO) and GI (low GI, high GI) were formed. Changes in fat mass were measured by quantitative magnetic resonance (EchoMRI; Houston, TX, USA; 43) and adjusted for predicted values using mathematical modeling of energy balance (15).

Both, GI and carbohydrate content affected insulin sensitivity, metabolic flexibility (i.e., the increase in respiratory ratio (RQ) with re-feeding and thus the capacity to utilize lipid and carbohydrate fuels and to transition between them; 44), and the partitioning of weight regain. Re-feeding was associated with impaired fasting fat oxidation and thus augmented fat regain. Energetic efficiency at low work intensities was reduced in response to caloric restriction. The higher the fall in energetic efficiency during low level exercise the greater the regain of body weight.

As to the determinants of weight regain, (i) decreases in plasma leptin levels, (ii) reduced energetic efficiency at low work intensities during preceding caloric restriction as well as (iii) a limited activation of the sympathetic nervous system (SNS) with refeeding were significantly associated with re-gain in body mass. Weight regain as either fat mass or FFM was associated with a reduced insulin sensitivity and low metabolic flexibility during re-feeding (40,41).

Taken together, during caloric restriction, the decrease in plasma leptin levels, the increase in energetic efficiency at low grade physical activity and a relatively low SNS activity are associated with re-gain in body weight during subsequent refeeding. During refeeding, insulin resistance and a low fat oxidation add to regain of fat mass.

### **Skeletal muscle crosstalk in response to exercise and nutrition**

Jürgen Eckel

Regular physical activity has beneficial effects on whole body metabolism, while a sedentary lifestyle is a major risk factor for developing metabolic diseases such as type 2 diabetes mellitus. Besides their established roles in work performance, energy and protein stores, skeletal muscle and adipose tissue are endocrine organs. Skeletal muscle cells and adipocytes both secrete a broad range of proteins and cytokines, generally termed adipokines in the case of adipocytes, and myokines for muscle cells. Myokines, have been shown to affect muscle physiology and additionally exert systemic effects on other tissues and organs.

Several myokines are regulated by contraction, like angiopoietin-related protein 4, fibroblast growth factor 21, interleukin (IL)-6, IL-7, IL-15, leukemia inhibitory factor, myonectin, myostatin and vascular endothelial growth factor (45). The beneficial effects of physical activity are often considered due to an improved energy metabolism; however, myokines are also thought to be involved since skeletal muscle secretes higher levels of myokines in response to contraction (45).

With contraction, muscle glucose uptake and fat oxidation rates increase and these effects are associated with the release of myokines such as IL-6, IL7 and BDNF (brain-derived neurotrophic factor). Up to now there are more than 540 myokines and IL-6 is the most prominent muscle-derived protein, which was demonstrated to be up-regulated in plasma after exercise (46). By contrast, contraction blocks TNF $\alpha$  release and signaling in muscle. As IL6 and BDNF result in increased lipid oxidation, IL7 regulates muscle cell development. Novel contraction-regulated myokines are YKL-40, irisin, and MCP-1.

Many of the contraction-regulated myokines described in the literature are additionally known to be secreted by adipocytes. These proteins were termed adipo-myokines. The current literature mainly describes a negative crosstalk between excess body fat and skeletal muscle, while the data of IL-6 and irisin indicate an additional crosstalk from the muscle to the adipose tissue (47,48).

Adipokines and myokines appear to have autocrine effects within skeletal muscle and adipose tissue. Additionally, they are involved in an endocrine crosstalk with other tissues such as liver. Depending on the serum level and the incubation time, adipo- myokines appear to have a beneficial or an adverse effect on the target tissue (49). Adipokines and myokines may also have inconsistent effects. It is presently unclear why pro-inflammatory adipokines are up-regulated in the obese state and thus may be associated with negative sequelae even though they also have beneficial effects after exercise.

Myokines have been proposed to increase energy expenditure, an effect explained by the development of brown-fat like cells of white adipocytes (47). At the cellular level, the transcriptional co-activator, PGC1 $\alpha$ , activated by FGF21 (fibroblast growth factor) generated in brown adipocytes stimulates glucose uptake, lipolysis and metabolic rate. PGC1 $\alpha$  expression in muscle increases the expression of FNDC5, a membrane protein that is cleaved to the myokine irisin. Irisin is induced in mice but its existence in humans is questionable (48). In contrast to mice, FNDC5 is not up-regulated by contractile activity in humans and irisin was not found in supernatants of human myotubes (48). Genome analysis revealed that irisin is a pseudogene in humans, the protein is not translated due to a mutation

in the start codon. In addition, irisin had no effect on the white-to-brown transition of human pre-adipocytes.

More recently, a novel cytokine, YKL-40 (also known as Chitinase-3-like protein1, CHI3L1) has been identified in the secretome of primary human skeletal muscle cells by mass spectroscopic analysis. This protein is expressed by many other cells (e.g., macrophages and hepatocytes). It lacks enzymatic activity and binds to PAR2 (Protease-activated receptor2). YKL-40 secretion goes down during differentiation and is up-regulated by inflammatory cytokines. YKL-40 reduces (i) TNF $\alpha$ - but not IL-1beta-induced NF-kB-activation and (ii) the TNF $\alpha$ -mediated secretion of MCP1, IL-8 and IL-6 secretion. In addition, YKL-40 protects from TNF $\alpha$ -mediated insulin resistance. Taken together YKL-40 reflects the versatility of an individual myokine

### Adipokines, Myokines and Tissue Cross-talk

Paul Trayhurn

White adipose tissue, which was traditionally considered to have little or no function beyond that of a fuel storage organ, is now recognized to have multiple roles. These include thermal insulation (particularly in marine mammals), mechanical protection and as a key signaling and endocrine organ. Our current perspective on the physiological roles of white fat follows from the discovery that white adipocytes are major secretory cells, releasing a multiplicity of metabolic and signaling factors (50–54). Adipocyte secretions, the quantitatively most important of which are fatty acids, comprise both lipids and proteins. Most attention has been focused on the protein factors secreted from fat cells – the adipokines. Nevertheless, there are a range of lipid moieties released from white adipocytes, and these include prostaglandins, endocannabinoids,  $\alpha$ -tocopherol, cholesterol and the active form of vitamin D<sub>3</sub> (51,52).

More than one hundred different adipokines have been clearly identified, based on gene expression and the demonstration that the encoded protein is secreted from adipocytes. However, proteomic studies indicate that there are several hundred adipokines in total (55). Many adipokines are linked to immunity and the inflammatory response (including both classical cytokines and chemokines), to insulin sensitivity and to the architecture of the extracellular matrix of adipose tissue (53–56). Others are involved in lipid metabolism, vascular hemostasis, the regulation of blood pressure or energy balance. Much attention has been focused on inflammation-related adipokines and the state of inflammation that develops in adipose tissue as tissue mass expands in obesity (53). This inflammatory state, which involves the recruitment of macrophages and other immune cells as well as the increased synthesis and release of inflammation-related adipokines, is widely considered to underpin the development of obesity-associated disorders – particularly insulin resistance and the other components of the metabolic syndrome (53–56).

Two particular adipokines – leptin and adiponectin – are major hormones that were first discovered in adipose tissue and which have multiple actions, both locally and distally. Indeed, it was the discovery of leptin in 1994 as the product of the ‘ob’ (now LEP) gene, a mutation in which leads to the profound obesity of the *ob/ob* mouse that has led to the

unambiguous recognition of white adipose tissue as an endocrine organ (57). Some adipokines may function locally, through autocrine or paracrine actions, rather than being endocrine factors. By definition, endocrine factors secreted from adipocytes are involved in communicating with other tissues and organs. One of the most potent examples of the endocrine action of an adipokine comes from the central effects of adipocyte-derived leptin on the neuroendocrine regulation of appetite in the hypothalamus. This demonstrates a direct communication from fat cells to the brain.

While the synthesis and secretion of many key adipokines, including leptin and inflammation-related factors such as IL-1 $\beta$  and IL-6, rise with increasing adipose mass in obesity, the production and release of the other major adipocyte hormone, adiponectin, falls (58–59). Importantly, this has implications for inflammation and insulin sensitivity since the hormone has both anti-inflammatory and insulin-sensitizing actions (60–62). One of the conundrums in adipose tissue biology is why the production of many adipokines changes with tissue expansion in the obese. One proposed mechanism is that it is a response to hypoxia, mouse data demonstrating clearly that the O<sub>2</sub> tension is markedly lower in adipose tissue depots of obese than lean animals (51,64). Hypoxia may underpin the inflammatory response and other functional changes in adipocytes that occur with obesity. Indeed, in adipocytes in culture reduced O<sub>2</sub> tension leads to the stimulation of the expression and release of adipokines such as leptin, VEGF, IL-6 and Angptl4, while inhibiting the production of adiponectin (64).

The identification of the rapidly growing family of adipokines has revolutionized our understanding of the biology of white adipose tissue and the extent to which it is integrated into metabolic regulation and whole-body homeostasis. It has also increasingly served as a model for other tissues and organs which had not been previously considered to exhibit a significant secretory function. Perhaps the most potent example is skeletal muscle. While myostatin (also known as growth differentiation factor 8) had been recognized as a secretion from myocytes, inhibiting muscle differentiation and growth, the possibility that there are a range of protein secretory products from muscle cells appeared unlikely. However, the perspective began to change following the discovery that skeletal muscle releases large quantities of IL-6 into the circulation following exercise (65–67). Muscle contraction leads to the stimulation of the expression of the IL-6 gene in muscle cell cultures and the encoded protein is released. This has led to the development of the concept of myokines as secreted proteins from muscle, paralleling the adipokine paradigm.

Subsequent studies, taking in effect a candidate gene/protein approach, have identified several other myokines, including IL-7, IL-8, IL-15, CXCL-1 and LIF (65, 67–69). This has recently also been followed by proteomic approaches in which the muscle protein secretome has been investigated. Proteomics suggests that the number of myokines may be in the low hundreds (70–72), although caution has to be exercised in that there is the risk that cell damage leads to the leakage of proteins into the culture medium which would not otherwise be released – similar caution needs to be considered in the case of proteomic studies on adipocytes. Nevertheless, it is increasingly evident that the myokines as a group are not restricted to a small number of proteins. A recently identified myokine of particular interest is irisin. This is encoded by the FNDC5 gene, the immediate product of which is a

membrane protein that is subsequently cleaved and secreted as irisin (73). Irisin has the intriguing action of ‘browning’ of white adipose depots – that is, of driving the tissue towards a thermogenic, energy-dissipating profile (73,74). However, while this is the case in rodents, as with resistin, the situation appears different in humans, as noted above in the previous part of this manuscript.

Irisin provides a strong example of cross-talk between tissues, and between skeletal muscle and adipose tissue in particular. A further example is leptin which has actions on muscle as well as other organs, including the brain as noted earlier. There has been some emphasis on the role of muscle-derived IL-6 as a lipolytic factor, but this is increasingly viewed as an action within muscle itself rather than distally on adipose tissue (66).

## CACHEXIA

### Body composition and clinical course in patients with cancer cachexia

Vickie Baracos

The cardinal diagnostic criterion of cachexia is the appearance of involuntary weight loss, signaling alteration of the normally precise controls which serve to maintain body weight and body composition throughout adulthood. Skeletal muscle wasting is considered the central feature of cancer cachexia and a 2011 international consensus of experts (75) underscored that this muscle loss may occur with or without concurrent loss of fat mass. Clinical and basic research on cancer cachexia has a strong focus on the etiology and clinical implications of this muscle loss. Severe depletion of skeletal muscle, termed sarcopenia (Greek ‘*sarx*’ of flesh) is defined as a low level of muscle associated with statistically significant increases in health risks and an impaired health state (e.g. mortality, pharmaceutical drug toxicity, loss of strength and reduced physical disability (75). Cutoffs defining sarcopenia and its relation to risk of mortality which were solved using statistical approaches have been published for cancer patients (76,77). Sarcopenia is prevalent in older populations and is frequently seen in cancer patients. Cancer patients presenting with a loss of muscle mass may be underweight, normal weight, overweight or even obese (76, 77)

In clinical practice, sarcopenia may be camouflaged by overweight and obesity and, thus, there is need of detailed body composition analysis. Since cancer patients are routinely followed in clinical practice by computed tomography (CT) imaging there is a considerable opportunity for detailed quantification of different tissues with high specificity and precision. CT imaging has enormous potential within the field of body composition analysis. In clinical practice, abdominal scans are often available and images can be evaluated at the level of the 3<sup>rd</sup> lumbar vertebra where cross-sectional areas of muscle strongly correlate with whole body muscle mass. Using these measures (77) in a prospective clinical study of cancer patients with BMI >30 kg/m<sup>2</sup>, survival was about 11 months in sarcopenic cancer patients compared with 21 months in a group of patients with normal muscle mass and this was independent of cancer site, stage and performance status. This was subsequently confirmed and expanded (76) in a population cohort of patients crossing all categories of BMI. Sarcopenia also predicted poor outcomes of cancer surgery including infectious complications and use of inpatient rehabilitation (78). Cancer patients with sarcopenia



appear unusually prone to chemotherapy-associated severe toxicity across a wide array of cancer sites and cancer therapies and these studies have been summarized by (79)

An emerging set of findings relate to the presence of fatty infiltration of muscle, termed myosteatorsis. This is detected as reduced skeletal muscle x-ray attenuation in quantitative analysis of computed tomography (CT) (i.e., between -29 and +29 HU, low attenuation muscle; between +30 and +150 HU, normal attenuation muscle). Independently of classic prognostic factors in cancer such as disease stage, site and patient performance status, low attenuation values in muscle predicts increased mortality (76,80) however the underlying basis of the fatty infiltration of muscle and its relation to survival remain to be understood. Myosteatorsis is discussed in further detail by Dr. Fearon in the next section.

Longitudinal studies provide an opportunity to study the body composition changes of cachexia-over time (20, 33). Repeated measurements suggest that tumor progression is correlated with losses in adipose tissue and muscle mass and this would be expected given the high energy demand of tumor tissue (20,33). The progression of muscle and fat loss with tumor progression was exponential and appears similar in different cancers (lung, colorectal, pancreas cancer, and cholangiocarcinoma) (33) with the exception that patients with pancreatic cancer had higher overall incidence and rate of fat loss.

In patients with advanced cancer, the potential for positive energy balance skeletal muscle anabolism is not well characterized. In a longitudinal study (33) of a mixed population of patients with advanced solid tumors was studied over a period of ~12 months and each interval between 2 CT scans (~ 3 months) was assessed for loss, gain and stable behavior of fat and muscle. The overall frequency of muscle gain was 15.4% whereas muscle mass was stable in 45.6% of intervals between any 2 scans, making maintenance or gain of muscle the predominant behavior. Likewise adipose tissue was stable in 27.1% of intervals and gain occurred in 24.8% of intervals. These findings suggest that cachexia is not an unmitigated trajectory of loss, but rather a period dominated by the opposing forces of the cancer (catabolic) and treatment (anabolic) in a context where controls of body energy balance are functional and may result in periods of stability and regain, as well as loss. The clinical course of skeletal muscle wasting in advanced cancer and the window of possible muscle anabolism were assessed. Multinomial logistic regression revealed that being within 90 d (vs.> 90 d) from death was the principal risk factor for muscle loss [odds ratio (OR)=2.67] and muscle gain was correspondingly less likely (OR=0.37) at this time (33). Thus, a window of anabolic potential exists at defined early phases of the disease trajectory (>90d survival) creating an opportunity for intervention to stop or reverse muscle wasting. A variety of nutrition therapies (macro- and micronutrients, branched chain amino acids, leucine, *n*3-fatty acids) and drugs (e.g. selective androgen receptor agonists, antagonists of myostatin, cytokines and protein degradation) are under investigation to exploit and expand anabolic potential.

### **Adipose tissue muscle crosstalk in cancer cachexia**

Kenneth C.H. Fearon

Cancer cachexia is multifactorial characterized by an ongoing loss of skeletal muscle (with or without loss of fat mass) that cannot be reversed by conventional nutritional support (75). The pathophysiology is characterized by a negative protein and energy balance driven by reduced food intake, low physical activity, systemic inflammation and abnormal metabolism. Cachexia in patients with cancer is characterized by anorexia, increased or decreased energy expenditure, increased lipolysis, insulin resistance, reduced protein turnover and reduced physical activity. The degree of weight loss and the severity of the underlying metabolic changes are mainly driven by inflammation: Cancer patients with a positive acute phase response had a low energy intake at concomitantly increased resting energy expenditure.

Mediators of cachexia include cytokines, neuroendocrine hormones and tumor-specific factors. Presently, it is not known to what extent tissue cross-talk (via myokines or cytokines) contributes to these features in humans. Recent evidence does, however, raise the possibility that lipotoxicity from increased fatty acids may be important (81–83). In cachectic cancer patients, macro- and microscopic changes indicate skeletal muscle lipid infiltrations. Mean lipid droplet count in muscle tissue correlated positively with the severity of weight loss and increases further with loss of adipose mass in other body compartments (82).

With regard to muscle fiber type, type II fibers are targeted selectively with relative preservation of type I fibers (84). Protein synthesis and myogenic cell proliferation and protein degradation and apoptosis determine muscle mass. Faced with the clinical features of cancer cachexia, protein degradation and, thus, the activity of the proteasome is a driver of muscle wasting. In fact, an increased proteasome activity and increased autophagy markers in skeletal muscle have been described in cancer cachexia (84,85). By contrast, divergent effects on protein synthesis have been reported with expression profiles characterized by 1750 down regulated genes but 150 up-regulated genes (85). However, when compared to healthy controls, muscle transcriptome was indistinguishable in cancer patients 8 months after tumor resection.

Altered adipocyte metabolism leads to a loss of fat mass and adipose atrophy in cancer cachexia. Metabolically, adipose tissue wasting is characterized by suppressed lipogenesis at increased lipolysis. A high rate of lipolysis appears to be a key factor underlying fat loss, while inhibition of adipocyte development and lipid deposition may also contribute to the problem (83). This metabolic pattern is partly explained reduced substrate supply (in anorectic patients) and the increase in inflammatory markers (like cytokines, TNF $\alpha$ , IL6) as part of a tumor-adipose tissue interaction. These cytokines are also generated locally in macrophages reflecting a state of chronic inflammation. Zinc- $\alpha$ 2-glycoprotein (ZAG), a 43-kDa protein, is overexpressed in certain human malignant tumors and acts as a lipid-mobilizing factor to stimulate lipolysis in adipocytes leading to cachexia (86,87). ZAG mRNA-levels in white adipose tissue (WAT) are positively correlated with weight loss and the rate of lipolysis. By contrast, leptin mRNA levels are reduced in WAT of cachectic cancer patients and showed a negative correlation with weight loss. However, plasma leptin levels were shown to be decreased or even increased in cachectic cancer patients.

Fat–muscle crosstalks might be a critical regulator in the development of cachexia. This is brought about through free fatty acids, myokines, or adipokines. Data derived from a model of cachexia and lipolysis in tumor-bearing mice with wild-type adipose tissue triglyceride lipase (Atgl +/+) or Atgl–/– (i.e. the gene has been ablated) suggested that at normal plasma free fatty acid concentrations (due to the lack of triglyceride lipase and thus failing to increase lipolysis) fat tissue mass was conserved and muscle wasting and, thus, cancer cachexia did not occur (83). The mechanism through which skeletal-muscle mass is maintained in the presence of the systemic mediators is unknown but may involve muscle–adipose cross-talk through free fatty acids. Up to now there has been little effort to manipulate the integrative physiology of adipose tissue and muscle tissue for therapeutic weight gain in tumor patients.

## PERSPECTIVES

### Rethinking why obesity develops and why it is harmful

Thorkild I.A. Sørensen

The prevailing conventional ideas about obesity are that it is a passive reflection of a positive energy balance due to an ‘obesogenic’ environment favoring excess food intake and physical inactivity, eventually leading to an excessive amount of stored fat that by itself threatens the health. In other words, obesity results from a passive storage of surplus energy which may give rise to inflammation and thus co-morbidities. This is also reflected by the definition of obesity and its association with diabetes, cardio- and cerebro-vascular diseases and certain cancers. A more positive view of fat mass is that body fat is an energy reserve available for the future. Then, obesity may be seen as a healthy and active response to an expected future lack of energy (88,89). This challenges conventional ideas about obesity.

The ability to store and mobilize triglycerides in adipocytes is a very efficient and biologically inert way of securing the energy supplies in the future when needs for energy cannot be met by available food. Fat storage is without health problems unless triglyceride stores can be expanded and have not reached their limits. Excessive storage of fat (i.e. obesity) may be considered as a consequence of a spurious unopposed signal of expected future needs of energy that leads to the harmful metabolic effects when the limits of storage capacity are reached. Weight gain and overweight happen if the body senses the risk of possible future shortage of food, although today this nearly never happens to people living in Western societies. This unifying theory requires profound revision of the approach to obesity, which may lead to new modalities of prevention and treatment of obesity.

One may ask how and when this sensing occurs, how it translates into obesity and, finally, how comorbidity results. Evidence for sensing future lack of energy as a cause of overweight comes from experimental data, as for example groups of mice develop greater fat stores with a reduced food supply (90). Sensing may be based on social conditions and, thus, social insecurity is considered as a driver. Thus besides other consequences any social challenge also would imply food insecurity in the future. This is independent of the present food supply. Evidence for this idea comes from the observation that when compared with middle and high socioeconomic state, obesity is highly prevalent in low socioeconomic

states and social insecurity. Faced with the developments of modern societies a considerable segment of populations, even in rich countries, live in financial and social insecurity. Sensing social insecurity will vary between different age groups and is unlikely to occur in children where obesity rates also have increased during the last decades. It is tempting to speculate that in children the sensing of insecurity is transferred from their parents, perhaps by the way they treat them. Another possibility is that the parental sensing of insecurity is transferred to the next generation by epigenetic mechanisms either before or during pregnancy or direct effects on the fetus. Obviously, there may well be individual genetic differences in the sensing and response to the insecurity

From a biological point of view, it is presently unclear how sensing of future food shortages is translated into deposition of triglycerides and, thus, an increase in fat mass. It is likely that central-peripheral psycho-neurobiological mechanisms influence this process which is finally characterized by an expansion and renewal of adipocytes (91). Both the number of fat cells and their size (i) have an upper limit and (ii) determine the upper limit of weight gain. Since adaptation of adipocytes and increased storage may reflect different and at least partly independent processes, this may explain the development of different forms of obesity resulting (i) from simple overfeeding or (ii) true obesity as a disease.

In terms of survival during food shortages, triglyceride synthesis in adipocytes is beneficial as long as limits of storage capacity have been not reached. By contrast, when storage exceeds its' limits this will result in ectopic fat deposition, insulin resistance and inflammation. Then negative effects on metabolism and health become evident. Thus, the association between storage of triglycerides and metabolic disturbances are not directly due to the triglyceride accumulation but are explained by factors co-occurring with it (89). This idea is supported by the finding that many obese individuals do not have adverse health effects and survive until old age (92). As a consequence, the arbitrary definition of obesity simply based on BMI cut offs as well as comparing fat mass between different individuals in clinical practice becomes questionable unless the capacity of fat storage is taken into account. In conclusion, there is need of a re-definition of fat accumulation and obesity faced with its benefits and harms.

### **Intrauterine growth retardation, fat - FFM relation and metabolic risks in later life**

Angelo Pietrobelli

Low birth weight (LBW) and rapid postnatal growth have been positively associated with obesity in adulthood (for a detailed description of that concept see 93–95). Maternal factors affecting intra-uterine development and growth include age, weight, height, nutrition, stress, smoking, drugs, infections, endocrine disorders, vascular diseases and exposure to environmental contaminants. Normal growth biology involves relationship between hormonal levels, body composition, growth patterns with gender and time sensitive maturation programs. LBW and infants born preterm reflect a stressful intrauterine milieu and demonstrate a constellation of aberrant developmental trajectories, which are apparent by term equivalent age and persist into adult life. In fact, when the fetus is exposed to malnutrition, the organism diverts the limited nutrient supply for favoring survival of vital organs such as brain at the expense growth and other organs such as pancreas. Fetal

malnutrition or, in general, a suboptimal uterine environment adds to permanent anatomical and functional changes in various tissues and organs, ultimately leading to increased risk of metabolic and cardiovascular disease. Adverse long-term effects may reflect a mismatch between early (fetal and neonatal) conditions and environmental and nutritional effects in later life (93,94).

The exposure to a different, and sometimes opposite, environment in intrauterine life affects the expression of multiple genes involved in different metabolic pathways. This is partly brought about by epigenetic changes affecting programming in multiple organs including the liver (95). Epigenetic alterations have been proposed as one mechanism to mediate the influence of early life exposures and gene-environment interactions. These changes alter the ability to coordinate fat and carbohydrate metabolism, favoring a shift to a preferential use of fatty acids as an energy source in order to adapt the organism to the *in utero*-reduced nutrient supply. Altogether, these changes induce adaptations in hypothalamic-lipid sensing mechanisms, ultimately affecting food intake and endogenous glucose production in postnatal life (94–96).

Intrauterine adverse cues have been related obesity, type 2 diabetes mellitus, and hypertension, disturbances in lipid metabolism, nonalcoholic fatty liver, asthma, food allergy, depression and neurobehavioral impairment (96–97). In high school girls, there is a significant and inverse correlation between birth weights and either blood pressure or blood levels of triglycerides and insulin (98). In addition, early postnatal nutrition may affect adult health by altering gut microbiota with long lasting effects on later immunity and overall health status (96). In this regard probiotics, which have the potential to restore the intestinal microbiota balance, may add to prevent the development of chronic immune-mediated diseases (99) If so, this may be explained by epigenetic mechanisms elicited by probiotics through the production of short chain fatty acids (95).

Analyzing the effects of LBW related to childhood family, neighborhood, and socio-demographic conditions on disease onset in adulthood clearly showed that LBW increased the risk of asthma, hypertension, type 2 diabetes mellitus and cardiovascular disease stroke, heart attack with odds ratios by age 50 between 1.51 to 2.16 (100). When compared to normal birth weight, LBW has long-lasting effects on chronic diseases in children; at age 14 there were no differences between age 8 and age 14 but the rate of obesity increased (99). After birth, maternal-child feeding patterns may also promote overweight and eating disorders. However, feeding strategies were associated with reduced body weight rather than child overweight (101).

In light of these findings it is fundamental to monitor early life growth of children and the associated changes in their body composition. Fat and FFM increase rapidly during gestation and infancy, but the majority of studies has relied upon serial measurements of weight, length, and head circumference to assess growth with very little information regarding the composition and the quality of the fat and FFM compartments of the body. Because absolute weight change has limited utility in the identification of infants at risk of later adverse health outcomes that stem from elevated adiposity, and/or reduced FFM, better accurate and

precise body composition assessment techniques are needed to evaluate the quality of the body mass during this key period of life (102).

At birth and normal weights (i.e., about 3500 g for boys and 3300 g for girls), normal fat mass is around 480 to 500 g or 14 to 15% of body weight (103). Concomitantly, the hydration of FFM is about 80% with a protein content of 15% (103,104). The neonate's later risk of obesity is then determined by the presence of gestational diabetes in the mother, parental obesity, ethnic background, high birth weight as well as rapid early growth which deserve a special consideration (105).

### **Age-related changes in fat and muscle: relationship with bone and fracture risk**

Marjolein Visser

Aging is associated with metabolic, physiologic, and functional impairments, in part through age-related changes in body composition. Across the lifespan, body weight generally increases until age 80 after which a decline is observed (106). Even when body weight remains stable in old age, changes in the composition of soft tissues can be detected. Changes in body composition with aging relate to body fat and individual fat depots, skeletal muscle mass and bone mineral content. Body weight, fat and muscle mass are related to bone mineral content and fracture risk. Longitudinal data show mean weight gains in groups up to age 60 years, whereas older age groups are more frequently characterized by weight losses (107). The rate of weight gain decreases from age 20 to 60 years, whereas the rate of weight loss increases from about 2kg/11 years to more than 5kg/11 years in subjects above age 80 years (107). Weight loss, but not weight gain, was associated with increased mortality amongst older men and women (108). People who lost weight had a higher mortality rate compared with those who were weight stable, with similar associations found for cardiovascular and non-cardiovascular mortality (108).

Measurements have been performed in large, prospective cohort studies conducted in older adults that have used dual-energy x-ray absorptiometry and/or computed tomography or magnetic resonance imaging to assess age-related changes in fat mass, muscle fat infiltration and skeletal muscle mass. The total amount of body fat increases until age 75 to 80 years, after which a decline is observed which seems to parallel the decline in body weight in very old age (109). Skeletal muscle and bone masses decreased, and detailed analyses showed a decrease in skeletal muscle mass but an increase in muscle fat infiltration (or intramuscular adipose tissue, IMAT) and visceral adipose tissue (VAT; 110). Even though total body fat tends to decline in very old age, the infiltration of fat into the muscles seems to continue with relative changes between 15 and 30% (in women and men, respectively; 111). With age, fat also accumulates in vertebral bone marrow (112) and bone marrow fat content is related to lower bone mineral content at spine, hip and femoral neck and, thus, osteoporosis and prevalent vertebral fractures (113,114).

With regard to the lean soft tissue, appendicular skeletal muscle mass decreases with aging and this decline seems to accelerate in very old age (110,111,114). Concomitantly, IMAT increases but subcutaneous fat area in mid-thigh decreases with weight loss (115). Age-related changes in muscle mass impact muscle function and the loss of muscle mass is



considered to be a major determinant of strength loss in aging. An annualized 1% decline in muscle mass with aging is paralleled by a 3% fall in muscle strength (116). Thus, the strength decline is much more rapid than the concomitant loss of muscle mass. The loss of lean mass, higher baseline strength, lower baseline leg lean mass, and older age, are independently associated with strength decline in both men and women. However, maintaining or gaining muscle mass does not prevent aging-associated declines in muscle strength suggesting body composition independent processes in age-related functional impairments (116). All these data give evidence for an age-related remodeling of body composition with reductions in SM and corresponding increases in VAT and IMAT. These changes impact body function.

In white and black women, lean soft tissue and fat mass are both associated with bone mineral density (BMD), whereas in men lean tissue is the only determinant of BMD (117). Weight loss and a low body weight increase the risk of hip, spine and wrist fractures; by contrast, a high BMI decreases the risk (118–121). However, weight-related fracture risk may differ between locations with ankle fractures increasing with increasing BMI but pelvic and rib fractures had an U-shaped association with an increased risk at low as well as at high BMI (118,120). In addition to mass, muscle density also relates the age related incidence of fractures: low thigh muscle attenuation increased the 7-year hip fracture incidence (122).

Sarcopenia parallels osteoporosis. With age above 45 years, the prevalence of osteopenia, osteoporosis and sarcopenia II began to increase with prevalence's of 56%, 12% and 34%, respectively, at ages above 70 years (123). There are numerous connections between bone and skeletal muscle, as for example (i) they share mesodermal origin, (ii) have overlapping signaling pathways during development, (iii) their masses peak around the same time, (iv) have a shared genetic background and (v) both respond to similar anabolic stimuli as well as mechanical forces (123,124). At the cellular level sarcopenia is characterized by a loss in myocyte number, a decrease in myofibrillar protein content and mitochondrial dysfunction.

Inflammation and reduced anabolic stimuli are among other factors to explain sarcopenia. Abnormalities in growth hormone-IGF1-secretion and -signaling may add to both, sarcopenia and osteoporosis (125). In muscle, these abnormalities result in increased myocyte apoptosis, decreased myofiber cross-sectional area and increased expression of proteolytic genes. Concomitantly, low activity of the growth hormone-IGF1 axis adds to delayed mineralization, impaired osteoblast differentiation and proliferation as well as increased apoptosis in bone cells. Low mechanical forces due to bed rest, sedentary behavior and low activity impact gravitational loading as well as muscle contractions (124).

There are correlations between muscle mass and either bone mass and bone quality (123,126,127). As to muscle-bone cross-talks, muscle secretes myokines, e.g. myostatin, leukemia-inhibitory factor, interleukins 6 and 7, IGF1, fibroblast growth factor (FGF 23). Besides its effects on the muscle itself, myostatin, a strong regulator of muscle mass in response exercise and muscle damage also has an effect on bone mass (128). In humans, myostatin gene polymorphisms are associated with peak bone mass. In addition in mice, overexpression of the myostatin pro-peptide increases bone mineral density. Epidemiological data also showed an increased risk of falls and fractures with decreasing

muscle mass (128,129). However, when compared with muscle mass, muscle strength seems to be the more important determinant of falls and fractures (130).

## CONCLUSIONS

The presentations given at the occasion of the 12<sup>th</sup> IASO Stock Conference in 2013 in Hamburg, Germany, supported Michael Stocks' view that integration of research ideas about similarities and extremes in obesity and cachexia benefits future understanding. Body composition data are interpreted within broader contexts of intermediary metabolism and inflammation by taking into account the relationships between organs and tissues rather than their individual masses per se only. Communication is brought about by numerous cross-talks between organs and tissues, which are related to their masses as well as their functions. It is tempting to speculate that changes in body composition with weight changes (i.e., either weight gain resulting in overweight and obesity or voluntary weight loss in response to diet or involuntary weight loss in cancer patients leading to cachexia) go together with and partly result from altered or even disturbed cross-talks between organs and tissues. Since body composition is not fixed throughout life, a functional body composition approach including (i) the masses of organs and tissues, (ii) between organ/tissue masses relationships and (iii) between organ and between tissue cross-talks should become part of a lifetime approach in humans. This will provide a better understanding of trajectories of organ-/tissue masses and their related functions and, thus, add to future modeling as reference of normal integrative physiology throughout life. This will again serve as a background for understanding extremes like obesity and cancer cachexia. Then, understanding obesity and cachexia is more about their differences to models and references rather than the issue itself.

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## REFERENCES

1. Shen, W.; St-Onge, M-P.; Wang, Z.; Heymsfield, SB. Study of Body Composition: An Overview. In: Heymsfield, SB.; Lohman, TG.; Wang, Z.; Going, SB., editors. Human Body Composition. 2nd ed.. 2005. p. 3-14. Human Kinetics Champaign, IL.
2. Müller MJ. From BMI to functional body composition. Eur J Clin Nutr. 2013; 67:1119–1121. [PubMed: 24193256]
3. Ahima R, Lazar MA. Physiology. The health risk of obesity--better metrics imperative. Science. 2013; 341:856–858. [PubMed: 23970691]
4. Benedict, FG. A study of prolonged fasting. Vol. 203. Washington, DC: Carnegie Institute of publication; 1915.

5. Keys, A.; Brozek, J.; Henschel, A.; Mickelsen, O.; Taylor, HL. *The Biology of Human Starvation*. Minneapolis, London: The University of Minnesota Press; 1950.
6. Dulloo AG, Jacquet J. The control of partitioning between protein and fat during human starvation: its internal determinants and biological significance. *Br J Nutr*. 1999; 82:339–356. [PubMed: 10673906]
7. Forbes G. Lean body mass-body fat interrelationships in humans. *Nutr Rev*. 1987; 45:225–231. [PubMed: 3306482]
8. Hall KG. Modeling metabolic adaptations and energy regulation in humans. *Ann Rev Nutr*. 2012; 32:35–44. [PubMed: 22540251]
9. Müller MJ, Wang Z, Heymsfield SB, Schautz B, Bosy-Westphal A. Advances in the understanding of specific metabolic rates of major organs and tissues in humans. *Curr Opin Clin Nutr Metab Care*. 2013; 16:501–508. [PubMed: 23924948]
10. Wikipedia. 2013 Sep 4. [http://en.wikipedia.org/wiki/Crosstalk\\_\(electronics\)](http://en.wikipedia.org/wiki/Crosstalk_(electronics)).
11. Stock, MJ. Obesity and Cachexia: Possibilities of an Integrated Approach.. In: Rothwell, NJ.; Stock, MJ., editors. *Obesity and Cachexia. Physiological mechanisms and new approaches to pharmacological control*. Chichester: John Wiley&Sons; 1991. p. 1-12.
12. Müller, MJ.; Danforth, E.; Burger, AG.; Siedentopp, U., editors. *Hormones and Nutrition in Obesity and Cachexia*. Heidelberg: Springer; 1990.
13. Leibel RL, Rosenbaum M, Hirsch J. Changes in energy expenditure resulting from altered body weight. *N Engl J Med*. 1995; 332:621–628. [PubMed: 7632212]
14. Heilbronn LK, de Jonge L, Frisard MI, et al. Pennington CALERIE Team. Effect of 6-month calorie restriction on biomarkers of longevity, metabolic adaptation, and oxidative stress in overweight individuals: a randomized controlled trial. *JAMA*. 2006; 295:1539–1548. [PubMed: 16595757]
15. Hall KD. Computational model of in vivo human energy metabolism during semistarvation and refeeding. *Am J Physiol Endocrinol Metab*. 2006; 291:E23–E37. [PubMed: 16449298]
16. Hall KD. Predicting metabolic adaptation, body weight change, and energy intake in humans. *Am J Physiol Endocrinol Metab*. 2010; 298:E449–E466. [PubMed: 19934407]
17. Hall KD, Sacks G, Chandramohan D, Chow CC, Wang YC, Gortmaker SL, Swinburn BA. Quantification of the effect of energy imbalance on body weight. *Lancet*. 2011; 378:826–837. [PubMed: 21872751]
18. Hall KD. Diet versus exercise in "the biggest loser" weight loss competition. *Obesity (Silver Spring)*. 2013 May; 21(5):957–959. [PubMed: 23404767]
19. Hallgreen CE, Hall KD. Allometric relationship between changes of visceral fat and total fat mass. *Int J Obes*. 2008; 32:845–852.
20. Liefvers JR, Mourtzakis M, Hall KD, McCargar LJ, Prado CM, Baracos VE. A viscerally driven cachexia syndrome in patients with advanced colorectal cancer: contributions of organ and tumor mass to whole-body energy demands. *Am J Clin Nutr*. 2009 Apr; 89(4):1173–1179. [PubMed: 19244378]
21. Dulloo AG, Jacquet J, Montani JP, Schutz Y. Adaptive thermogenesis in human body weight regulation: more of a concept than a measurable entity? *Obes Rev*. 2012; 13:105–121. [PubMed: 23107264]
22. Dulloo AG, Jacquet J, Montani JP. How dieting makes some fatter: from a perspective of human body composition autoregulation. *Proc Nutr Soc*. 2012; 71:379–389. [PubMed: 22475574]
23. Dulloo AG, Jacquet J, Girardier L. Poststarvation hyperphagia and body fat overshooting in humans: a role for feedback signals from lean and fat tissues. *Am J Clin Nutr*. 1997; 65:717–723. [PubMed: 9062520]
24. Dulloo AG, Jacquet J. Adaptive reduction in basal metabolic rate in response to food deprivation in humans: a role for feedback signals from fat stores. *Am J Clin Nutr*. 1998; 68:599–606. [PubMed: 9734736]
25. Montani JP, Viecelli AK, Prévot A, Dulloo AG. Weight cycling during growth and beyond as a risk factor for later cardiovascular diseases: the 'repeated overshoot' theory. *Int J Obes*. 2006; 30(Suppl 4):S58–S66. (Lond).

26. Braun TP, Zhu X, Szumowski M, Scott GD, et al. Central nervous system inflammation induces muscle atrophy via activation of the hypothalamic-pituitary-adrenal axis. *J Exp Med*. 2011; 208:2449–2463. [PubMed: 22084407]
27. Ravussin E, Bogardus C. Relationship of genetics, age, and physical fitness to daily energy expenditure and fuel utilization. *Am J Clin Nutr*. 1989; 49(Suppl):968–975. [PubMed: 2655422]
28. Tschöp MH, Speakman JR, Arch JR, et al. A guide to analysis of mouse energy metabolism. *Nat Methods*. 2011 Dec 28; 9(1):57–63. [PubMed: 22205519]
29. Heymsfield SB, Thomas D, Bosy-Westphal A, Shen W, Peterson CM, Müller MJ. Evolving concepts on adjusting human resting energy expenditure measurements for body size. *Obes Rev*. 2012 Nov; 13(11):1001–1014. [PubMed: 22863371]
30. Later W, Bosy-Westphal A, Kossel E, Glüer CC, Heller M, Müller MJ. Is the 1975 Reference Man still a suitable reference? *Eur J Clin Nutr*. 2010; 64:1035–1042. [PubMed: 20664617]
31. Bosy-Westphal A, Braun W, Schautz B, Müller MJ. Issues in characterizing resting energy expenditure in obesity and after weight loss. *Front Physiol*. 2013; 4:1–9. article 47. [PubMed: 23372552]
32. Müller MJ, Langemann D, Gehrke I, et al. Effect of constitution on mass of individual organs and their association with metabolic rate in humans—a detailed view on allometric scaling. *PLoS One*. 2011; 6:e22732. [PubMed: 21818376]
33. Prado CM, Sawyer MB, Ghosh S, Liefers JR, Esfandiari N, Antoun S, Baracos VE. Central tenet of cancer cachexia therapy: do patients with advanced cancer have exploitable anabolic potential? *Am J Clin Nutr*. 2013; 98:1012–1019. [PubMed: 23966429]
34. Rosenbaum M, Leibel RL. Adaptive thermogenesis in humans. *Int J Obes (Lond)*. 2010; 34(Suppl 1):S47–S55. [PubMed: 20935667]
35. Müller MJ, Bosy-Westphal A. Adaptive thermogenesis with weight loss in humans. *Obesity (Silver Spring)*. 2013; 21:218–228. [PubMed: 23404923]
36. Larsen TM, Dalskov SM, van Baak M, et al. Diet, Obesity, and Genes (Diogenes) Project. Diets with high or low protein content and glycemic index for weight-loss maintenance. *N Engl J Med*. 2010; 363:2102–2113. [PubMed: 21105792]
37. Rebelos E, Muscelli E, Natali A, et al. RISC Study Investigators. Body weight, not insulin sensitivity or secretion, may predict spontaneous weight changes in nondiabetic and prediabetic subjects: the RISC study. *Diabetes*. 2011; 60:1938–1945. [PubMed: 21617179]
38. Swinburn BA, Nyomba BL, Saad MF, et al. Insulin resistance associated with lower rates of weight gain in Pima Indians. *J Clin Invest*. 1991; 88:168–173. [PubMed: 2056116]
39. Wong MH, Holst C, Astrup A, et al. Caloric restriction induces changes in insulin and body weight measurements that are inversely associated with subsequent weight regain. *PLoS One*. 2012; 7(8):e42858. [PubMed: 22905179]
40. Lagerpusch M, Enderle J, Later W, Eggeling B, Pape D, Müller MJ, Bosy-Westphal A. Impact of glycaemic index and dietary fibre on insulin sensitivity during the refeeding phase of a weight cycle in young healthy men. *Br J Nutr*. 2013; 109:1606–1616. [PubMed: 23191994]
41. Lagerpusch M, Enderle J, Eggeling B, et al. Carbohydrate Quality and Quantity Affect Glucose and Lipid Metabolism during Weight Regain in Healthy Men. *J Nutr*. 2013; 143:1593–1601. [PubMed: 23946346]
42. Baracos V, Caserotti P, Earthman CP, et al. Advances in the science and application of body composition measurement. *J Parenter Enteral Nutr*. 2012 Jan; 36(1):96–107.
43. Müller MJ, Bosy-Westphal A, Lagerpusch M, Heymsfield SB. Use of balance methods for assessment of short-term changes in body composition. *Obesity (Silver Spring)*. 2012; 20:701–707. [PubMed: 21869755]
44. Kelley DE, He J, Menshikova EV, Ritov VB. Dysfunction of mitochondria in human skeletal muscle in type 2 diabetes. *Diabetes*. 2002; 51:2944–2950. [PubMed: 12351431]
45. Raschke S, Eckardt K, Bjørklund Holven K, Jensen J, Eckel J. Identification. *PLoS One*. 2013; 8(4):e62008. [PubMed: 23637948]
46. Hartwig S, Raschke S, Knebel B, et al. Secretome profiling of primary human skeletal muscle cells. *Biochim Biophys Acta*. 2013 Aug 27. doi:pii: S1570-9639(13)00295-1. 10.1016/j.bbapap.2013.08.004. [Epub ahead of print].

47. Boström P, Wu J, Jedrychowski MP, et al. A PGC1- $\alpha$ -dependent myokine that drives brown-fat-like development of white fat and thermogenesis. *Nature*. 2012; 481(7382):463–468. [PubMed: 22237023]
48. Raschke S, Elsen M, Gassenhuber H, et al. Evidence against a Beneficial Effect of Irisin in Humans. *PLoS One*. 2013; 8(9):e73680. [PubMed: 24040023]
49. Raschke S, Eckel J. Adipo-myokines: two sides of the same coin--mediators of inflammation and mediators of exercise. *Mediators Inflamm*. 2013; 2013:320724. Epub 2013 Jun 3. [PubMed: 23861558]
50. Rajala MW, Scherer PE. The adipocyte - at the crossroads of energy homeostasis, inflammation, and atherosclerosis. *Endocrinology*. 2003; 144:3765–3773. [PubMed: 12933646]
51. Trayhurn P, Wood IS. Adipokines: Inflammation and the pleiotropic role of white adipose tissue. *Br J Nutr*. 2004; 92:347–355. [PubMed: 15469638]
52. Trayhurn P. Endocrine and signalling role of adipose tissue: new perspectives on fat. *Acta Physiol Scand*. 2005; 184:285–293. [PubMed: 16026420]
53. Hotamisligil GS. Inflammation and metabolic disorders. *Nature*. 2006; 444:860–867. [PubMed: 17167474]
54. Rosen ED, Spiegelman BM. Adipocytes as regulators of energy balance and glucose homeostasis. *Nature*. 2006; 444:847–853. [PubMed: 17167472]
55. Dahlman I, Elsen M, Tennagels N, Korn M, Brockmann B, Sell H, et al. Functional annotation of the human fat cell secretome. *Archiv Physiol Biochem*. 2012; 118:84–91.
56. Hotamisligil GS. Inflammatory pathways and insulin action. *Int J Obesity*. 2003; 27(Suppl 3):S53–S55.
57. Zhang YY, Proenca R, Maffei M, Barone M, Leopold L, Friedman JM. Positional cloning of the mouse obese gene and its human homolog. *Nature*. 1994; 372:425–432. [PubMed: 7984236]
58. Arita Y, Kihara S, Ouchi N, et al. Paradoxical decrease of an adipose-specific protein, adiponectin, in obesity. *Biochem Biophys Res Commun*. 1999; 257:79–83. [PubMed: 10092513]
59. Hotta K, Funahashi T, Arita Y, et al. Plasma concentrations of a novel, adipose-specific protein, adiponectin, in type 2 diabetic patients. *Arterioscl Thromb Vasc Biol*. 2000; 20:1595–1599. [PubMed: 10845877]
60. Berg AH, Combs TP, Du X, Brownlee M, Scherer PE. The adipocyte-secreted protein Acrp30 enhances hepatic insulin action. *Nature Med*. 2001; 7:947–953. [PubMed: 11479628]
61. Ouchi N, Kihara S, Arita Y, et al. Novel modulator for endothelial adhesion molecules - Adipocyte-derived plasma protein adiponectin. *Circulation*. 1999; 100:2473–2476. [PubMed: 10604883]
62. Yamauchi T, Kamon J, Waki H, et al. The fat-derived hormone adiponectin reverses insulin resistance associated with both lipoatrophy and obesity. *Nature Med*. 2001; 7:941–946. [PubMed: 11479627]
63. Yokota T, Oritani K, Takahashi I, et al. Adiponectin, a new member of the family of soluble defense collagens, negatively regulates the growth of myelomonocytic progenitors and the functions of macrophages. *Blood*. 2000; 96:1723–1732. [PubMed: 10961870]
64. Trayhurn P. Hypoxia and adipose tissue function and dysfunction in obesity. *Physiol Rev*. 2013; 93:1–21. [PubMed: 23303904]
65. Pedersen BK, Steensberg A, Keller P, et al. Muscle-derived interleukin-6: lipolytic, anti-inflammatory and immune regulatory effects. *Pflügers Archiv Eur J Physiol*. 2003; 446:9–16. [PubMed: 12690457]
66. Pedersen BK, Febbraio MA. Muscles, exercise and obesity: skeletal muscle as a secretory organ. *Nat Rev Endocrinol*. 2012; 8:457–465. [PubMed: 22473333]
67. Pedersen BK, Febbraio MA. Muscle as an Endocrine Organ: Focus on Muscle-Derived Interleukin-6. *Physiol Rev*. 2008; 88:1379–1406. [PubMed: 18923185]
68. Haugen F, Norheim F, Lian H, Wensaas A, Dueland S, Berg O, et al. IL-7 is expressed and secreted by human skeletal muscle cells. *Am J Physiol Cell Physiol*. 2010; 298:C807–C816. [PubMed: 20089933]

69. Trayhurn P, Drevon CA, Eckel J. Secreted proteins from adipose tissue and skeletal muscle - adipokines, myokines and adipose/muscle cross-talk. *Arch Physiol Biochem*. 2011; 117:47–56. [PubMed: 21158485]
70. Bortoluzzi S, Scannapieco P, Cestaro A, Danieli G, Schiaffino S. Computational reconstruction of the human skeletal muscle secretome. *Proteins*. 2006; 62:776–792. [PubMed: 16342272]
71. Yoon J, Yea K, Kim J, et al. Comparative proteomic analysis of the insulin-induced L6 myotube secretome. *Proteomics*. 2009; 9:51–60. [PubMed: 19053084]
72. Henningsen J, Rigbolt K, Blagoev B, Pedersen B, Kratchmarova I. Dynamics of the skeletal muscle secretome during myoblast differentiation. *Mol Cell Proteomics*. 2010; 9:2482–2496. [PubMed: 20631206]
73. Bostrom P, Wu J, Jedrychowski MP, Korde A, et al. A PGC1- $\alpha$ -dependent myokine that drives brown-fat-like development of white fat and thermogenesis. *Nature*. 2012; 481:463–468. [PubMed: 22237023]
74. Wu J, Boström P, Sparks Lauren M, et al. Beige Adipocytes are a distinct type of thermogenic fat cell in mouse and human. *Cell*. 2012; 150:366–376. [PubMed: 22796012]
75. Fearon K, Arends J, Baracos V. Understanding the mechanisms and treatment options in cancer cachexia. *Nat Rev Clin Oncol*. 2013
76. Martin L, Birdsell L, Macdonald N, et al. Cancer cachexia in the age of obesity: skeletal muscle depletion is a powerful prognostic factor, independent of body mass index. *J Clin Oncol*. 2013 Apr 20; 31(12):1539–1547. [PubMed: 23530101]
77. Prado CM, Lieffers JR, McCargar LJ, Reiman T, Sawyer MB, Martin L, Baracos VE. Prevalence and clinical implications of sarcopenic obesity in patients with solid tumours of the respiratory and gastrointestinal tracts: a population-based study. *Lancet Oncol*. 2008; 9:629–635. [PubMed: 18539529]
78. Lieffers JR, Bathe OF, Fassbender K, Winget M, Baracos VE. Sarcopenia is associated with postoperative infection and delayed recovery from colorectal cancer resection surgery. *Br J Cancer*. 2012; 107:931–936. [PubMed: 22871883]
79. Parsons HA, Baracos VE, Dhillon N, Hong DS. Kurzrock Body composition, symptoms, and survival in advanced cancer patients referred to a phase I service. *R.PLoS One*. 2012; 7(1):e29330. Epub 2012 Jan 3.
80. Antoun S, Lanoy E, Iacovelli R, Albiges-Sauvin L, et al. Skeletal muscle density predicts prognosis in patients with metastatic renal cell carcinoma treated with targeted therapies. *Cancer*. 2013; 119:3377–3384. [PubMed: 23801109]
81. Bing C, Trayhurn P. New insights into adipose tissue atrophy in cancer cachexia. *Proc Nutr Soc*. 2009; 68:385–392. [PubMed: 19719894]
82. Stephens NA, Skipworth RJ, Macdonald AJ, Greig CA, Ross JA, Fearon KC. Intramyocellular lipid droplets increase with progression of cachexia in cancer patients. *J Cachexia Sarcopenia Muscle*. 2011; 2:111–117. [PubMed: 21766057]
83. Fearon KC. Cancer cachexia and fat-muscle physiology. *N Engl J Med*. . 2011; 365:565–567. [PubMed: 21830971]
84. Johns N, Stephens NA, Fearon KC. Muscle wasting in cancer. *Int J Biochem Cell Biol*. 2013; 45:2215–2229. [PubMed: 23770121]
85. Gallagher IJ, Stephens NA, MacDonald AJ, et al. Suppression of skeletal muscle turnover in cancer cachexia: evidence from the transcriptome in sequential human muscle biopsies. *Clin Cancer Res*. 2012; 18:2817–2827. [PubMed: 22452944]
86. Chen, Bing; Yi, Bao; John, Jenkins, et al. Zinc- $\alpha$ 2-glycoprotein, a lipid mobilizing factor, is expressed in adipocytes and is up-regulated in mice with cancer cachexia. *PNAS*. 2004; 101:2500–2505. [PubMed: 14983038]
87. Mracek T, Stephens NA, Gao D, Bao Y, Ross JA, Rydén M, Arner P, Trayhurn P, Fearon KC, Bing C. Enhanced ZAG production by subcutaneous adipose tissue is linked to weight loss in gastrointestinal cancer patients. *Br J Cancer*. 2011; 104:441–447. [PubMed: 21245862]
88. Sørensen TIA. Is Obesity a Healthy Active Response to an Expected Future Lack of Energy rather than a Passive Storage of Surplus Energy. *Obes Facts*. 2012; 5:431–435. [PubMed: 22797370]



89. Sørensen TIA. Obesity defined as Excess Storage of inert Triglycerides – Do we need a Paradigm Shift? *Obes Facts*. 2011; 4:91–94. [PubMed: 21577014]
90. Li X, Cope BM, Johnson MS, Smith DL, Nagy TR. Mild calorie restriction reduces fat accumulation in female C57BL/6J mice. *Obesity*. 2010; 18:456–462. [PubMed: 19798071]
91. Spalding KL, Arner E, Westermark PO, Bernard S, Buchholz BA, Bergmann O, Blomqvist L, Hoffstedt J, Näslund E, Britton T, Concha H, Hassan M, Rydén M, Frisén J, Arner P. Dynamics of fat cell turnover in humans. *Nature*. 2008; 453:783–787. [PubMed: 18454136]
92. Zimmermann E, Holst C, Sørensen TIA. Lifelong doubling of mortality in men entering adult life as obese. *Int J Obes*. 2011; 35:1193–1199.
93. Barker, DJP. *Mothers, Babies and Health in later life*. 1st. Ed.. London, UK: Churchill Livingstone; 1998.
94. Godfrey KM, Barker DJP. Fetal programming and adult health. *Public Health Nutrition*. 2001; 4:611–624. [PubMed: 11683554]
95. Canani RB, Costanzo MD, Leone L, Bedogni G, Brambilla P, Cianfarani S, Nobili V, Pietrobelli A, Agostoni C. Epigenetic mechanisms elicited by nutrition in early life. *Nutr Res Rev*. 2011; 24:198–205. [PubMed: 22008232]
96. Cianfarani S, Agostoni C, Bedogni G, Berni Canani R, Brambilla P, Nobili V, Pietrobelli A. Effect of intrauterine growth retardation on liver and long-term metabolic risk. *Int J Obes (Lond)*. 2012; 36:1270–1277. [PubMed: 22531091]
97. Mori M, Mori H, Yamori Y, Tsuda K. Low birth weight as cardiometabolic risk in Japanese high school girls. *J Am Coll Nutr*. 2012 Feb; 31(1):39–44. [PubMed: 22661625]
98. Johnson RC, Schoeni RF. Early-life origins of adult disease: national longitudinal population-based study of the United States. *Am J Public Health*. 2011; 101:2317–2324. [PubMed: 22021306]
99. Zampieri N, Pietrobelli A, Biban P, Soffiati M, Dall'agnola A, Camoglio FS. *Lactobacillus paracasei* subsp. *paracasei* F19 in Bell's stage 2 of necrotizing enterocolitis. *Minerva Pediatr*. 2013 Aug; 65(4):353–360. [PubMed: 24051968]
100. Hack M, Schluchter M, Andreias L, Margevicius S, Taylor HG, Drotar D, Cuttler L. Change in prevalence of chronic conditions between childhood and adolescence among extremely low-birth-weight children. *JAMA*. 2011; 306:394–401. [PubMed: 21791688]
101. Faith MS, Heshka S, Keller KL, Sherry B, Matz PE, Pietrobelli A, Allison DB. Maternal-child feeding patterns and child body weight: findings from a population-based sample. *Arch Pediatr Adolesc Med*. 2003; 157:926–932. [PubMed: 12963600]
102. Fomon SJ, Haschke F, Ziegler EE, Nelson SE. Body composition of reference children from birth to age 10 years. *Am J Clin Nutr*. 1982; 35(5 Suppl):1169–1175. [PubMed: 7081099]
103. Cole TJ, Faith MS, Pietrobelli A, Heo M. What is the best measure of adiposity change in growing children: BMI, BMI %, BMI z-score or BMI centile? *Eur J Clin Nutr*. 2005 Mar; 59(3): 419–425. [PubMed: 15674315]
104. Wells, JCK. *Thrift and control*. Cambridge: Cambridge University Press; 2010. *The Evolutionary Biology of Human Body Fatness*.
105. Rudolf M. Predicting babies' risk of obesity. *Arch Dis Child*. 2011; 96:995–997. [PubMed: 21828069]
106. Drøyvold WB, Nilsen TI, Krüger O, Holmen TL, Krokstad S, Midthjell K, Holmen J. Change in height, weight and body mass index: Longitudinal data from the HUNT Study in Norway. *Int J Obes*. 2006; 30:935–939.
107. Drøyvold WB, Lund Nilsen TI, Lydersen S, Midthjell K, Nilsson PM, Nilsson JA, Holmen J. Nord-Trøndelag Health Study. Weight change and mortality: the Nord-Trøndelag Health Study. *J Intern Med*. 2005; 257:338–345. [PubMed: 15788003]
108. Ding J, Kritchevsky SB, Newman AB, et al. Health ABC Study. Effects of birth cohort and age on body composition in a sample of community-based elderly. *Am J Clin Nutr*. 2007; 85:405–410. [PubMed: 17284736]
109. Song MY, Ruts E, Kim J, Janumala I, Heymsfield S, Gallagher D. Sarcopenia and increased adipose tissue infiltration of muscle in elderly African American women. *Am J Clin Nutr*. 2004; 79:874–880. [PubMed: 15113728]

110. Delmonico MJ, Harris TB, Visser M, et al. Health, Aging, and Body. Longitudinal study of muscle strength, quality, and adipose tissue infiltration. *Am J Clin Nutr.* 2009; 90:1579–1585. [PubMed: 19864405]
111. Kugel H, Jung C, Schulte O, Heindel W. Age- and sex-specific differences in the 1H-spectrum of vertebral bone marrow. *J Magn Reson Imaging.* 2001; 13:263–268. [PubMed: 11169833]
112. Lang T, Cauley JA, Tylavsky F, Bauer D, Cummings S, Harris TB. Health ABC Study. Computed tomographic measurements of thigh muscle cross-sectional area and attenuation coefficient predict hip fracture: the health, aging, and body composition study. *J Bone Miner Res.* 2010 Mar; 25(3):513–519. [PubMed: 20422623]
113. Schwartz AV, Sigurdsson S, Hue TF, et al. Vertebral bone marrow fat associated with lower trabecular BMD and prevalent vertebral fracture in older adults. *J Clin Endocrinol Metab.* 2013; 98:2294–2300. [PubMed: 23553860]
114. Frontera WR, Reid KF, Phillips EM, Krivickas LS, Hughes VA, Roubenoff R, Fielding RA. Muscle fiber size and function in elderly humans: a longitudinal study. *J Appl Physiol.* 2008; 105:637–642. [PubMed: 18556434]
115. Goodpaster BH, Park SW, Harris TB, et al. The loss of skeletal muscle strength, mass, and quality in older adults: the health, aging and body composition study. *J Gerontol A Biol Sci Med Sci.* 2006; 61:1059–1064. [PubMed: 17077199]
116. Taaffe DR, Cauley JA, Danielson M, Nevitt MC, Lang TF, Bauer DC, Harris TB. Race and sex effects on the association between muscle strength, soft tissue, and bone mineral density in healthy elders: the Health, Aging, and Body Composition Study. *J Bone Miner Res.* 2001; 16:1343–1352. [PubMed: 11450711]
117. De Laet C, Kanis JA, Odén A, et al. Body mass index as a predictor of fracture risk: a meta-analysis. *Osteoporos Int.* 2005; 16:1330–1338. [PubMed: 15928804]
118. Langlois JA, Visser M, Davidovic LS, Maggi S, Li G, Harris TB. Hip fracture risk in older white men is associated with change in body weight from age 50 years to old age. *Arch Intern Med.* 1998; 158:990–996. [PubMed: 9588432]
119. Miranda EG, Armstrong Benjamin J, et al. Million Women Study Collaborators. Different effects of age adiposity and physical activity on the risk of ankle, wrist and hip fractures in postmenopausal women. *Bone.* 2012; 50:1394–1400. [PubMed: 22465850]
120. Compston JE, Flahive J, Hosmer DW, et al. for the GLOW Investigators. Relationship of weight, height, and body mass index with fracture risk at different sites in postmenopausal women: The global longitudinal study of osteoporosis in women (GLOW). *J Bone Miner Res.* 2013 Jul 22. [Epub ahead of print].
121. Griffith JF, Yeung DK, Antonio GE, et al. Vertebral bone mineral density, marrow perfusion, and fat content in healthy men and men with osteoporosis: dynamic contrast-enhanced MR imaging and MR spectroscopy. *Radiology.* 2005; 236:945–951. [PubMed: 16055699]
122. Walsh MC, Hunter GR, Livingstone MB. Sarcopenia in premenopausal and postmenopausal women with osteopenia, osteoporosis and normal bone mineral density. *Osteoporos Int.* 2006; 17:61–67. [PubMed: 15995793]
123. DiGirolamo DJ, Kiel DP, Esser KA. Bone and skeletal muscle: neighbors with close ties. *J Bone Miner Res.* 2013; 28:1509–1518. [PubMed: 23630111]
124. Kohrt WM, Barry DW, Schwartz RS. Muscle forces or gravity: what predominates mechanical loading on bone? *Med Sci Sports Exerc.* 2009; 41:2050–2055. [PubMed: 19812511]
125. Perrini S, Laviola L, Carreira MC, Cignarelli A, Natalicchio A, Giorgino F. The GH/IGF1 axis and signaling pathways in the muscle and bone: mechanisms underlying age-related skeletal muscle wasting and osteoporosis. *J Endocrinol.* 2010; 205:201–210. [PubMed: 20197302]
126. Szulc P, Beck TJ, Marchand F, Delmas PD. Low skeletal muscle mass is associated with 53 poor structural parameters of bone and impaired balance in elderly men--the MINOS study. *J Bone Miner Res.* 2005; 20:721–729. [PubMed: 15824844]
127. Elkasrawy MN, Hamrick MW. Myostatin (GDF-8) as a key factor linking muscle mass and bone structure. *J Musculoskelet Neuronal Interact.* 2010; 10:56–63. [PubMed: 20190380]

128. Sjöblom S, Suuronen J, Rikkinen T, Honkanen R, Kröger H, Sirola J. Relationship between postmenopausal osteoporosis and the components of clinical sarcopenia. *Maturitas*. 2013; 75:175–180. [PubMed: 23628279]
129. Johannesdottir F, Aspelund T, Siggeirsdottir K, et al. Mid-thigh cortical bone structural parameters, muscle mass and strength, and association with lower limb fractures in older men and women (AGES-Reykjavik Study). *Calcif Tissue Int*. 2012; 90:354–364. [PubMed: 22451219]
130. Schaap LA, Koster A, Visser M. Adiposity, Muscle Mass, and Muscle Strength in Relation to Functional Decline in Older Persons. *Epidemiol Rev*. 2012 Dec 4. Epub ahead of print.