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# LPA rs10455872 polymorphism is associated with coronary lesions in Brazilian patients submitted to coronary angiography

Paulo CJL Santos<sup>1\*</sup>, Carolina T Bueno<sup>1</sup>, Pedro A Lemos<sup>2</sup>, José E Krieger<sup>1</sup> and Alexandre C Pereira<sup>1\*</sup>

## Abstract

**Background:** Polymorphisms in the *LPA* gene were associated with coronary artery disease (CAD). However, there are differences in the allelic frequencies, Lp(a) levels, and significant association with CAD according to ethnic groups. In this scenario, the main aim of this study was to assess the influence of the *LPA* polymorphisms on coronary lesions in Brazilian patients.

**Methods:** 1,394 consecutive patients submitted to coronary angiography to study suggestive CAD and twenty coronary segments were scored. Genotyping for the *LPA* rs10455872 and rs3798220 polymorphisms were performed by high resolution melting analysis.

**Results:** The frequencies of the rs10455872 G and rs3798220 C variant alleles were 6.4% and 6.2%, respectively. *LPA* rs10455872 G variant allele was associated with higher odds ratio of having coronary lesions in an adjusted model (OR = 2.02, 95% CI = 1.10-3.72, p = 0.02). Scores of coronary lesions (extension, severity, and Gensini scores) were significantly different among rs10455872 genotype groups. Coronary lesions was not associated with *LPA* rs3798220 (OR = 1.09, 95% CI = 0.67-1.76, p = 0.73) and scores of coronary lesions were not different among rs3798220 genotypes.

**Conclusions:** We confirmed the association of the *LPA* rs10455872 with CAD in a large sample of Brazilian patients. For the *LPA* rs3798220, our finding is consistent with studies which showed the lack of this genetic association.

**Keywords:** *LPA* gene, rs10455872, rs3798220, Coronary lesions, Coronary artery disease

## Introduction

Lipoprotein(a) [Lp(a)] is a plasma lipoprotein synthesized by the liver that is composed of a low-density lipoprotein (LDL) molecule, a high molecular weight glycoprotein apolipoprotein(a), and a single molecule of apolipoprotein (B). Physiological and pathogenic roles of Lp(a) remain partially unknown. Studies have suggested that Lp(a) provides a link between the cholesterol transport and the fibrinolytic system acting as a modulator of the blood clotting and fibrinolysis systems [1,2].

The increased concentration of Lp(a) has been associated with incidence and severity of cardiovascular disease (CVD), coronary artery disease (CAD), peripheral

artery disease, and stroke [3-7]. A meta-analysis of published data from 31 prospective studies reported a relative risk for coronary heart disease of 1.60 (95% CI = 1.38-1.85) associated with Lp(a) levels [8,9].

More than 90% of the variance of Lp(a) concentration is explained by genetic variation [1]. A genome-wide association study showed that there is a group of genes strongly associated with CAD, such as solute carrier family 22 member 3 (*SLC22A3*), lipoprotein(a)-like 2 (*LPAL2*), and lipoprotein(a) (*LPA*), but investigators did not identify the functional variants at these loci [10-12].

Two polymorphisms in the *LPA* gene (rs3798220 and rs10455872) were associated with risk for CAD. However, there are differences in the allelic frequencies, Lp(a) levels, and degree of association with CAD according to ethnic groups [13-18]. In this scenario, the main aim of this study was to assess the influence of the *LPA*

\* Correspondence: pacaleb@usp.br; alexandre.pereira@incor.usp.br

<sup>1</sup>Laboratory of Genetics and Molecular Cardiology, Heart Institute (InCor), University of Sao Paulo Medical School, Av. Dr. Enéas de Carvalho Aguiar, 44 Cerqueira César, São Paulo, SP CEP 05403-000, Brazil

Full list of author information is available at the end of the article

polymorphisms on coronary lesions in Brazilian patients submitted to coronary angiography.

## Patients and methods

### Patients submitted to coronary angiography

One thousand three hundred and ninety-four consecutive patients submitted to coronary angiography to study suggestive CAD were selected at the Laboratory of Hemodynamic, Heart Institute (InCor), Sao Paulo, Brazil. All patients had a clinical diagnosis of angina pectoris and stable angina. No patient enrolled in this study was currently experiencing an acute coronary syndrome. Patients with heart failure classes III–IV, hepatic dysfunction, familiar hypercholesterolemia, previous heart or kidney transplantation, and in antiviral treatment were excluded [19,20]. All patients signed an informed consent form and the protocol was approved by the ethics committee from Hospital das Clínicas from São Paulo University (CAPPesq 0398/04).

### Demographic data and laboratory tests

Data regarding general characteristic, weight, height, race/color, main cardiovascular risk factors (hypertension, diabetes, obesity, dyslipidemia, smoking, and current medical treatment) were obtained by interview. Race/color was classified as White, Brown (*Pardo* in Portuguese; person with admixture between White and Black), Black or Asiatic [21].

Triglycerides, total cholesterol (TC), high-density lipoprotein cholesterol, LDL cholesterol, and glucose were evaluated by standard techniques in 12-h fasting blood samples. Diabetes mellitus was diagnosed by the presence of fasting glucose  $\geq 126$  mg/dL or the use of antidiabetic drugs [22]. Hyperlipidemia was defined as TC  $\geq 240$  mg/dL, LDL-C  $\geq 160$  mg/dL, and/or use of lipid-lowering drugs [23].

### Hemodynamic and angiographic data

Blood pressure was measured in the sitting position with the use of a standard mercury sphygmomanometer on the left arm after 5 min rest. The first and fifth phases of Korotkoff sounds were used for systolic blood pressure (SBP) and diastolic blood pressure (DBP), respectively. The SBP and DBP were calculated from two readings with a minimal interval of 10 min apart. Hypertension was defined as mean SBP  $\geq 140$  mm Hg and/or DBP  $\geq 90$  mm Hg and/or antihypertensive drug use [24].

Twenty coronary segments were scored: each vessel was divided into three segments (proximal, medial, and distal), except for the secondary branches of the right coronary artery (posterior ventricular and posterior descending), which were divided into proximal and distal segments. Stenosis higher than 50% in any coronary segment was graded 1 point and the sum of points for all 20 segments constituted the Extension Score. Lesion severity was calculated as follows: none and irregularities, 0

points; <50%, 0.3 points; 50–70%, 0.6 points; >70–90%, 0.8 points; and >90–100%, 0.95 points. The Severity Score was calculated through the sum of points for all 20 coronary segments [25].

### Genotyping

Genomic DNA from subjects was extracted from peripheral blood following standard salting-out procedure. Additional file 1: Figure S1 shows genotyping detected by polymerase chain reaction (PCR) followed by high resolution melting (HRM) analysis with the Rotor Gene 6000<sup>®</sup> instrument (Qiagen, Courtaboeuf, France). The QIAgility<sup>®</sup> (Qiagen, Courtaboeuf, France), an automated instrument, was used according to instructions to optimize the sample preparation step [26].

Amplification of the fragment for the *LPA* rs10455872 (A > G, intron 25) polymorphism was performed using the primer sense 5'- ATGGGCTGGCAACACATAG - 3' and antisense 5'- CACTTCTCTCTAACCTGTAA -3' (78 pairs base). Amplification of the fragment for the *LPA* rs3798220 (T > C, p.Ile1891Met) polymorphism was performed using the primer sense 5'- GGCTCCAAGAACAGCCTAGA -3' and antisense 5'- TCCTCAAGGCCTTCATCCTA -3' (104 pairs base). A 40-cycle PCR was carried out with the following conditions: denaturation of the template DNA for first cycle of 94°C for 120 s, denaturation of 94°C for 20 s, annealing of 56.7°C for 20 s, and extension of 72°C for 22 s. PCR was performed with addition of fluorescent DNA-intercalating SYTO9<sup>®</sup> (1.5  $\mu$ M; Invitrogen, Carlsbad, USA). In the HRM phase, the Rotor Gene 6000<sup>®</sup> measured the fluorescence in each 0.1°C temperature increase in the range of 72–81°C. Melting curves were generated by the decrease in fluorescence with the increase in the temperature; and in analysis, nucleotide changes result in three different curve patterns (Additional file 1: Figure S1). Samples of the three observed curves were analyzed using bidirectional sequencing as a validation procedure (ABI Terminator Sequencing Kit<sup>®</sup> and ABI 3500XL Sequencer<sup>®</sup> - Applied Biosystems, Foster City, CA, USA) [27]. The two methods gave identical results in all tests. The wild-type, heterozygous and mutant homozygous genotypes could be easily discernible by HRM analysis. In addition, 4% of the samples were randomly selected and reanalyzed as quality controls and gave identical results.

### Statistical analysis

Categorical variables are presented as percentage while continuous variables are presented as mean  $\pm$  standard deviation. Chi-square test was performed for comparative analysis of general characteristics and coronary lesion frequency according to *LPA* polymorphisms. Dominant models (AG + GG) for the rs10455872 or (TC + CC) for the rs3798220 were performed because the frequencies of

the GG and CC homozygous genotypes are low. Student's t-test was performed for comparing the age, body mass index (BMI), biochemical data, blood pressures, and angiographic data means according to *LPA* polymorphisms. Biochemical data, blood pressures, and angiographic data were adjusted for age, gender, and race/color. Logistic regression univariate and multivariate analyses were performed to evaluate OR (odds ratio) for coronary lesions. Two adjusted models were performed: one using age, gender, and race/color and another with the additional covariates BMI, hyperlipidemia, statin use, and smoking. Linkage disequilibrium, Hardy-Weinberg equilibrium, and haplotype analyses were conducted with Haploview 4.0. All statistical analyses were carried out using the SPSS software (v. 16.0), with the level of significance set at  $p \leq 0.05$ .

## Results

### General characteristics and *LPA* polymorphisms

Of the 1,394 patients, mean age of  $59.7 \pm 10.4$ , 1,126 (80.8%) had coronary lesions indicated by coronary angiography. The frequency of the rs10455872 G variant allele was 6.4% and the distribution of the genotypes was 0.6% ( $n = 8$ ) for variant homozygous, 11.7% ( $n = 163$ ) for heterozygous and 87.7% ( $n = 1223$ ) for wild-type. The frequency of the rs3798220 C variant allele was 6.2% and the distribution of the genotypes was 0.4% ( $n = 6$ ) for variant homozygous, 11.6% ( $n = 165$ ) for heterozygous and 88.0% ( $n = 1248$ ) for wild-type. The genotypic distribution for the *LPA* rs10455872 and rs3798220 polymorphisms were in accordance with the Hardy-Weinberg equilibrium ( $X^2 = 1.01$ ,  $p = 0.31$  and  $X^2 = 0.05$ ,  $p = 0.82$ , respectively). Linkage disequilibrium analysis shows that the *LPA* rs3798220 and rs10455872 variant alleles had no strong disequilibrium ( $LD = 73$ ) (Additional file 2: Figure S2).

### Biochemical, hemodynamic, and angiographic data according to *LPA* rs10455872 polymorphism

Table 1 shows data from patients submitted to coronary angiography according to *LPA* rs10455872 genotypes. The frequency of the *LPA* rs10455872 AG or GG genotypes was lower in non-Whites compared with Whites ( $p = 0.02$ ). Patient carrying AG or GG genotypes had higher TC mean and proportion of hyperlipidemia. Regarding the angiographic data, higher frequency of coronary lesions ( $p = 0.004$ ) was found in patients with AG or GG genotypes (88.9%) compared to patient with AA genotype (79.6%). Also, scores of coronary lesions (extension, severity, and Gensini scores) were significantly different among genotype groups ( $p < 0.001$ ,  $p < 0.001$ , and  $p = 0.05$ , respectively) (Table 1).

Furthermore, the presence of the *LPA* rs10455872 G variant allele was associated with higher OR of having coronary lesion, which we compared normal coronary arteries versus one-vessel, two-vessel, and three-vessel disease. Table 2

**Table 1 General characteristics, biochemical, hemodynamic, and angiographic data according to *LPA* rs10455872 genotypes in the patients submitted to coronary angiography**

(n=1394, 100%)	Genotypes		p value
	AA (n=1223)	AG+GG (n=171)	
Age (years)	59.9 ± 10.1	59.5 ± 10.8	0.62
Gender, female (%)	40.6	37.4	0.42
Race/color (%)			
White	65.0	74.9	
Intermediate	29.8	23.4	0.02
Black	5.2	1.8	
Hypertension (%)	70.0	68.4	0.68
Diabetes (%)	31.2	27.5	0.33
Hyperlipidemia (%)	57.2	71.3	0.002
Statin use (%)	28.0	30.5	0.61
Smokers (%)	35.6	38.0	0.44
Body mass index (Kg/m <sup>2</sup> )	27.7 ± 4.8	27.6 ± 4.9	0.99
Total cholesterol (mg/dL)	228 ± 49	237 ± 46	0.05
LDL-C (mg/dL)	147 ± 44	155 ± 39	0.11
HDL-C (mg/dL)	42 ± 12	42 ± 10	0.82
Triglycerides (mg/dL)	183 ± 130	179 ± 108	0.75
Systolic blood pressure (mmHg)	149 ± 34	152 ± 39	0.61
Diastolic blood pressure (mmHg)	82 ± 15	84 ± 15	0.29
Ejection fraction (%)	60.6 ± 14.4	56.8 ± 17.7	0.07
Coronary lesions (%)	79.6	88.9	0.004
Extension score	2.1 ± 1.6	2.6 ± 1.7	<0.001
Severity score	1.5 ± 1.2	1.9 ± 1.3	<0.001
Gensini score	19.7 ± 28.2	25.7 ± 32.0	0.05

HDL-C: high density lipoprotein; LDL-C: low density lipoprotein. Biochemical data, blood pressures, and angiographic scores were adjusted for age, gender, and race/color. Coronary lesions frequency was compared between normal coronary arteries versus one-vessel, two-vessel, and three-vessel disease.

shows three models, being the OR of 2.02 (95% CI = 1.10-3.72,  $p = 0.02$ ) in an adjusted model with age, gender, race/color, BMI, hyperlipidemia, statin use, and smoking. Also, Additional file 3: Table S1 shows logistic regression univariate analysis of the OR for coronary lesions. Additional file 4: Table S2 shows significant association for the variables age (as a continuous variable presented in the Table or as categorical variable - median age of 61 years as a cut-off - resulting in an OR of 1.88, 95% CI = 1.43-2.47,  $p < 0.001$ ), gender, BMI, statin use, and hyperlipidemia in a logistic regression multivariate analysis.

### Analysis stratified by race for the *LPA* rs10455872 polymorphism

In the White group ( $n = 901$ ), patients with AG or GG genotypes ( $n = 128$ ) had higher frequency of coronary lesions

**Table 2 Analysis of the coronary lesions odds ratio associated with *LPA* rs10455872 AG or GG genotypes in the patients submitted to coronary angiography**

(n=1394, 100%)	OR	95% CI	p value
<b>Models</b>			
<b>Unadjusted</b>	2.04	1.24-3.36	0.005
<b>Adjusted*</b>	2.05	1.22-3.43	0.006
<b>Adjusted**</b>	2.02	1.10-3.72	0.02

Coronary lesions frequency was compared between normal coronary arteries versus one-vessel, two-vessel, and three-vessel disease.

\*Adjusted for age, gender, and race/color.

\*\*Adjusted for age, gender, race/color, body mass index, hyperlipidemia, statin use, and smoking.

compared to patient with AA genotype (n = 773) (90.6% and 81.0%, respectively, p = 0.008). The presence of the *LPA* rs10455872 G variant allele was associated with higher OR of having coronary lesion in an adjusted model (OR = 2.17, 95% CI = 1.04-4.61, p = 0.03). Also, scores of coronary lesions were significantly different among genotype (extension: 2.7 ± 1.6 and 2.2 ± 1.6; severity: 1.9 ± 1.3 and 1.6 ± 1.2; and Gensini scores: 26.3 ± 30.0 and 19.7 ± 29.8) (p = 0.001, p = 0.01, and p = 0.04, respectively).

In the non-White group (n = 460, formed for Black and Brown individuals), frequency of coronary lesions was not statistically different (83.7% for AG or GG genotypes and 77.0% for AA genotype, p = 0.30). However, the presence of the G variant allele was associated with higher OR (OR = 1.95, 95% CI = 1.09-4.39, p = 0.04 – adjusted model) and scores of coronary lesions were different between AG or GG and AA genotypes (extension score: 2.6 ± 1.7 and 2.0 ± 1.6; and severity score: 1.9 ± 1.3 and 1.4 ± 1.2) (p = 0.04, and p = 0.03, respectively). Gensini score was marginally different between genotypes (24.7 ± 29.6 and 21.2 ± 27.5; p = 0.06).

#### Biochemical, hemodynamic, and angiographic data according to *LPA* rs3798220 polymorphism

The proportion of hyperlipidemia was not different among rs3798220 genotypes (p = 0.62). Regarding the angiographic data, the frequency of coronary lesions was not associated with rs3798220 (p = 0.57) and no significant OR was observed in an adjusted model (OR = 1.09, 95% CI = 0.67-1.76, p = 0.73). Also, scores of coronary lesions (extension, severity, and Gensini scores) were not significantly different among genotype groups (p = 0.85, p = 0.56, and p = 0.46, respectively).

#### Discussion

The two *LPA* polymorphisms studied in this study are some of the most important genetic markers for CAD. In this context, our main finding was that the *LPA* rs10455872 polymorphism is associated with coronary lesions in Brazilian patients submitted to coronary angiography. On the other hand, no association for the

*LPA* rs3798220 was observed for any of the tested phenotypes.

Studies have reported higher Lp(a) concentration in sub-Saharan African descent and lower Lp(a) concentration in European descent [13,15,28,29]. Regarding the ethnicity, Brazil has one of the most heterogeneous population of the world, composed by a mixture of different ethnic groups, mainly European descent, African descent and Amerindians. In our data, a stratified analysis by race supported a role for the rs10455872 polymorphism independent of ethnic group.

Corroborating with our study, Anderson et al. found that the rs10455872 polymorphism strongly predicted prevalent CAD (per allele OR = 1.43, 95% CI = 1.07-1.91) [30]. Helgadottir et al. showed that patients with CAD carrying *LPA* risk alleles have increased susceptibility to atherosclerotic manifestations outside of the coronary tree and they are more likely to be diagnosed earlier with CAD than are CAD cases not carrying this variant [31]. Other studies that analyzed the rs10455872 and rs3798220 polymorphisms together reported an increased risk of coronary disease and Lp(a) level that can be explained by these *LPA* polymorphisms [11,32]. *LPA* rs10455872 is an intronic polymorphism associated with short KIV-2 repeat region (kringle IV type 2) which is associated with Lp(a) levels. In the present study, the frequency of the rs10455872 G variant allele was 6.4% for overall, but we observed higher allelic frequency in White compared with non-White groups. The association of the rs10455872 with CAD was significant even in the non-White group which had a small sample size. These data suggest that rs10455872 is also a strong genetic marker for CAD risk in ethnically mixed populations.

For the *LPA* rs3798220 polymorphism, we did not observe significant association with coronary lesions. Data from other studies also did not support a relationship between this *LPA* variant and CAD [33,34]. Furthermore, Anderson et al., studying 1,400 participants with coronary angiography (more than 90% Whites), did not find an association signal between rs3798220 and CAD (OR = 1.47, 95% CI = 0.81-2.67, p = 0.20) [30]. In contrast to our study, the rs3798220 has previously been reported to have an association with the Lp(a) level and the risk of coronary disease [35-37]. *LPA* rs3798220 results in an aminoacid substitution in the protease domain of *LPA*, but it can not provide stronger association than rs10455872 which might be representing a more complex group of genetic variants or repeat structures. A possible hypothesis for the lack of this association in the present study could be the lower value of linkage disequilibrium between rs3798220 and rs10455872 identified in the Brazilian patients compared with some studies with patients predominantly from European descent [11,35-37]. Another hypothesis may be low statistical power, but less likely if the impact of



rs3798220 was approximately equal to the impact of rs10455872. However, the exact reason is unclear and other genetic components differently expressed due to ethnicity might be important modulators.

The exact mechanism by which an increased Lp(a) level increases the CAD risk is not fully understood. Pathways modulated by Lp(a) may involve the LDL-cholesterol transport system, the inhibition of the expression of tissue factor, the inhibition of conversion of plasminogen to plasmin, the carriage of pro-inflammatory oxidized phospholipids, and an atherosclerotic stenotic mechanism [30,38-44]. Some studies reported that the severity of coronary artery disease is associated with Lp(a) levels or LDL concentration [45-48]. Regarding to the Lp(a) level as a risk factor in different ethnic groups, Lp(a) has been associated with risk in European populations [1], but not unequivocally in African Americans [17,18]. However, a recent study identified that the increased risk of CVD was at least as strong in African Americans as in White Americans [49]. Another study investigated differential frequencies of *LPA* polymorphisms in non-Hispanic whites, non-Hispanic blacks, and Mexican Americans [50]. Interestingly, 15 of the 19 polymorphisms tested were strongly associated with Lp(a) levels in at least one subpopulation, six in at least two subpopulations, and none in all three subpopulations. The lack of generalization of associations across ethnicities suggests that specific *LPA* variants may be contributing to the observed Lp(a) between-population variance. Authors also compared the allele frequencies in HapMap, and observed extremely high correlations ( $r \geq 0.99$ ) in allele frequencies between non-Hispanic whites and HapMap CEU (US individuals of northern and western European ancestry) and between non-Hispanic blacks and both HapMap YRI (Yoruba from West Africa) and ASW (individuals with African ancestry from the Southwest USA) [50].

There are some limitations in our study. First, we did not measure Lp(a) levels and we also did not genotype KIV-2 repeats to check their association with both the *LPA* polymorphisms and/or the CAD phenotype. Second, we did not assess ancestry through genetic markers; instead, we used a self-declared classification which is commonly applied in Brazil and correlates with genetic ancestry determination. In addition, in our stratified analysis by race, we observed significant association of the rs10455872 with CAD in the White and non-White patient groups. Third, our plaque burden data are derived from institutional records and represent real-life data, as opposed to core-lab derived hemodynamic data. Thus, and despite the greater external validity of our results, we were not able to determine inter- or intra-observed variability estimates. In addition, our chosen method for establishing atherosclerotic burden in the studied patients has relied upon the Gensini Score, which has been

shown to highly correlate with this end-point. Other scores could also be used, although they are not as well fitted for quantifying plaque burden. One example is the Syntax score, an angiographic tool for grading the complexity of CAD and designed to better anticipate the risks of percutaneous or surgical revascularization. Finally, it is not possible to completely exclude the interaction of the covariates as other genetic markers, use of concomitant drugs, ethnicity, gender and age on our findings [51-54]. Nonetheless, our findings remained after multivariate analysis.

## Conclusions

In conclusion, we confirmed the association of the *LPA* rs10455872 with CAD in a large sample of Brazilian patients. For the *LPA* rs3798220, our finding is consistent with studies which showed the lack of this genetic association.

## Additional files

**Additional file 1: Figure S1.** Graphs of the *LPA* rs10455872 (A > G, intron 25) genotyping. Nucleotide changes results in different curve patterns using high resolution melting analysis. A: Graph of normalized fluorescence by temperature. B: Graph of normalized fluorescence (based on genotype 2) by temperature. 1: wild-type genotype (AA); 2: heterozygous genotype (AG); 3: mutant homozygous genotype (GG).

**Additional file 2: Figure S2.** Linkage disequilibrium and haplotype analyses for the *LPA* rs3798220 and rs10455872 polymorphisms in the patients submitted to coronary angiography.

**Additional file 3: Table S1.** Logistic regression univariate analysis of the coronary lesions odds ratio in the patients submitted to coronary angiography.

**Additional file 4: Table S2.** Logistic regression multivariate analysis of the coronary lesions odds ratio in the patients submitted to coronary angiography.

## Competing interests

The authors declare that they have no competing interests.

## Authors' contributions

PCJLS and CTB carried out molecular genetic analyses, performed the statistical analysis and drafted the manuscript. PCJLS, ACP, PAL, JEK conceived the study, and participated in its design. All authors read and approved the final manuscript.

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## Author details

<sup>1</sup>Laboratory of Genetics and Molecular Cardiology, Heart Institute (InCor), University of Sao Paulo Medical School, Av. Dr. Enéas de Carvalho Aguiar, 44 Cerqueira César, São Paulo, SP CEP 05403-000, Brazil. <sup>2</sup>Hemodynamic Laboratory, Heart Institute (InCor), University of Sao Paulo Medical School, Sao Paulo, Brazil.

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## References

1. Kronenberg F, Utermann G: Lipoprotein(a): Resurrected by genetics. *J Intern Med* 2013, **273**:6–30.
2. Koschinsky ML: Novel insights into lp(a) physiology and pathogenicity: More questions than answers? *Cardiovasc Hematol Disord Drug Targets* 2006, **6**:267–278.
3. Erqou S, Kaptoge S, Perry PL, Di Angelantonio E, Thompson A, White IR, Marcovina SM, Collins R, Thompson SG, Danesh J, Collaboration ERF: Lipoprotein(a) concentration and the risk of coronary heart disease, stroke, and nonvascular mortality. *JAMA* 2009, **302**:412–423.
4. Tsimikas S, Mallat Z, Talmud PJ, Kastelein JJ, Wareham NJ, Sandhu MS, Miller ER, Benessiano J, Tedgui A, Witztum JL, Khaw KT, Boekholdt SM: Oxidation-specific biomarkers, lipoprotein(a), and risk of fatal and nonfatal coronary events. *J Am Coll Cardiol* 2010, **56**:946–955.
5. Danesh J, Collins R, Peto R: Lipoprotein(a) and coronary heart disease. Meta-analysis of prospective studies. *Circulation* 2000, **102**:1082–1085.
6. Klein JH, Hegele RA, Hackam DG, Koschinsky ML, Huff MW, Spence JD: Lipoprotein(a) is associated differentially with carotid stenosis, occlusion, and total plaque area. *Arterioscler Thromb Vasc Biol* 2008, **28**:1851–1856.
7. Ohira T, Schreiner PJ, Morrisett JD, Chambless LE, Rosamond WD, Folsom AR: Lipoprotein(a) and incident ischemic stroke: The atherosclerosis risk in communities (aric) study. *Stroke* 2006, **37**:1407–1412.
8. Bennet A, Di Angelantonio E, Erqou S, Eiriksdottir G, Sigurdsson G, Woodward M, Rumley A, Lowe GD, Danesh J, Gudnason V: Lipoprotein(a) levels and risk of future coronary heart disease: Large-scale prospective data. *Arch Intern Med* 2008, **168**:598–608.
9. Nestel PJ, Barnes EH, Tonkin AM, Simes J, Fournier M, White HD, Colquhoun DM, Blankenberg S, Sullivan DR: Plasma lipoprotein(a) concentration predicts future coronary and cardiovascular events in patients with stable coronary heart disease. *Arterioscler Thromb Vasc Biol* 2013, **33**:2902–2908.
10. Trégouët DA, König IR, Erdmann J, Munteanu A, Braund PS, Hall AS, Grosshennig A, Linsel-Nitschke P, Perret C, DeSuremain M, Meitinger T, Wright BJ, Preuss M, Balmforth AJ, Ball SG, Meisinger C, Germain C, Evans A, Arveiler D, Luc G, Ruidavets JB, Morrison C, van der Harst P, Schreiber S, Neureuther K, Schäfer A, Bugert P, El Mokhtari NE, Schrezenmeir J, Stark K, *et al*: Genome-wide haplotype association study identifies the *slc22a3-lpa2-lpa* gene cluster as a risk locus for coronary artery disease. *Nat Genet* 2009, **41**:283–285.
11. Clarke R, Peden JF, Hopewell JC, Kyriakou T, Goel A, Heath SC, Parish S, Barlera S, Franzosi MG, Rust S, Bennett D, Silveira A, Malarstig A, Green FR, Lathrop M, Gigante B, Leander K, De Faire U, Seedorf U, Hamsten A, Collins R, Watkins H, Farrall M, Consortium P: Genetic variants associated with lp(a) lipoprotein level and coronary disease. *N Engl J Med* 2009, **361**:2518–2528.
12. Hopewell JC, Clarke R, Parish S, Armitage J, Lathrop M, Hager J, Collins R, Group HPSC: Lipoprotein(a) genetic variants associated with coronary and peripheral vascular disease but not with stroke risk in the heart protection study. *Circ Cardiovasc Genet* 2011, **4**:68–73.
13. Chretien JP, Coresh J, Berthier-Schaad Y, Kao WH, Fink NE, Klag MJ, Marcovina SM, Giaculli F, Smith MW: Three single-nucleotide polymorphisms in *lpa* account for most of the increase in lipoprotein(a) level elevation in african americans compared with european americans. *J Med Genet* 2006, **43**:917–923.
14. Lanktree MB, Anand SS, Yusuf S, Hegele RA, Investigators S: Comprehensive analysis of genomic variation in the *lpa* locus and its relationship to plasma lipoprotein(a) in south asians, chinese, and european caucasians. *Circ Cardiovasc Genet* 2010, **3**:39–46.
15. Deo RC, Wilson JG, Xing C, Lawson K, Kao WH, Reich D, Tandon A, Akyzbekova E, Patterson N, Mosley TH, Boerwinkle E, Taylor HA: Single-nucleotide polymorphisms in *lpa* explain most of the ancestry-specific variation in lp(a) levels in african americans. *PLoS One* 2011, **6**:e14581.
16. Mooser V, Scheer D, Marcovina SM, Wang J, Guerra R, Cohen J, Hobbs HH: The apo(a) gene is the major determinant of variation in plasma lp(a) levels in african americans. *Am J Hum Genet* 1997, **61**:402–417.
17. Paultre F, Pearson TA, Weil HF, Tuck CH, Myerson M, Rubin J, Francis CK, Marx HF, Philbin EF, Reed RG, Berglund L: High levels of lp(a) with a small apo(a) isoform are associated with coronary artery disease in african american and white men. *Arterioscler Thromb Vasc Biol* 2000, **20**:2619–2624.
18. Moliterno DJ, Jokinen EV, Miserez AR, Lange RA, Willard JE, Boerwinkle E, Hillis LD, Hobbs HH: No association between plasma lipoprotein(a) concentrations and the presence or absence of coronary atherosclerosis in african-americans. *Arterioscler Thromb Vasc Biol* 1995, **15**:850–855.
19. Maciel SS, Pereira Ada C, Silva GJ, Rodrigues MV, Mill JG, Krieger JE: Association between glutathione s-transferase polymorphisms and triglycerides and hdl-cholesterol. *Atherosclerosis* 2009, **206**:204–208.
20. Santos PC, Oliveira TG, Lemos PA, Mill JG, Krieger JE, Pereira AC: Mylip p.N342s polymorphism is not associated with lipid profile in the brazilian population. *Lipids Health Dis* 2012, **11**:83.
21. Santos PC, Alvim Rde O, Ferreira NE, De Sa Cunha R, Krieger JE, Mill JG, Pereira AC: Ethnicity and arterial stiffness in brazil. *Am J Hypertens* 2011, **24**:278–284.
22. Executive summary: Standards of medical care in diabetes–2011. *Diabetes Care* 2011, **34**(Suppl 1):S4–S10.
23. Executive summary of the third report of the national cholesterol education program (ncep) expert panel on detection, evaluation, and treatment of high blood cholesterol in adults (adult treatment panel iii). *JAMA* 2001, **285**:2486–2497.
24. The seventh report of the joint national committee on prevention, detection, evaluation, and treatment of high blood pressure. *JAMA* 2003, **2004**.
25. Lanz JR, Pereira AC, Martinez E, Krieger JE: Metabolic syndrome and coronary artery disease: Is there a gender specific effect? *Int J Cardiol* 2006, **107**:317–321.
26. Santos PC, Soares RA, Santos DB, Nascimento RM, Coelho GL, Nicolau JC, Mill JG, Krieger JE, Pereira AC: Cyp2c19 and abcb1 gene polymorphisms are differently distributed according to ethnicity in the brazilian general population. *BMC Med Genet* 2011, **12**:13.
27. Santos PC, Soares RA, Nascimento RM, Machado-Coelho GL, Mill JG, Krieger JE, Pereira AC: Slco1b1 rs4149056 polymorphism associated with statin-induced myopathy is differently distributed according to ethnicity in the brazilian general population: Amerindians as a high risk ethnic group. *BMC Med Genet* 2011, **12**:136.
28. Cobbaert C, Kesteloot H: Serum lipoprotein(a) levels in racially different populations. *Am J Epidemiol* 1992, **136**:441–449.
29. Utermann G: Genetic architecture and evolution of the lipoprotein(a) trait. *Curr Opin Lipidol* 1999, **10**:133–141.
30. Anderson JL, Knight S, May HT, Horne BD, Bair TL, Huntinghouse JA, Rollo JS, Muhlestein JB, Carlquist JF: Validation and quantification of genetic determinants of lipoprotein-a levels and predictive value for angiographic coronary artery disease. *Am J Cardiol* 2013, **112**:799–804.
31. Helgadóttir A, Gretarsdóttir S, Thorleifsson G, Holm H, Patel RS, Gudnason T, Jones GT, van Rij AM, Eapen DJ, Baas AF, Tregouët DA, Morange PE, Emmerich J, Lindblad B, Götsäter A, Kiemeny LA, Lindholt JS, Sakalihan S, Ferrell RE, Carey DJ, Elmore JR, Tsao PS, Grarup N, Jørgensen T, Witte DR, Hansen T, Pedersen O, Pola R, Gaetani E, Magnadóttir HB, *et al*: Apolipoprotein(a) genetic sequence variants associated with systemic atherosclerosis and coronary atherosclerotic burden but not with venous thromboembolism. *J Am Coll Cardiol* 2012, **60**:722–729.
32. Ronald J, Rajagopalan R, Cerrato F, Nord AS, Hatsukami T, Kohler T, Marcovina S, Heagerty P, Jarvik GP: Genetic variation in *lpa2*, *lpa*, and *plg* predicts plasma lipoprotein(a) level and carotid artery disease risk. *Stroke* 2011, **42**:2–9.
33. Li ZG, Li G, Zhou YL, Chen ZJ, Yang JQ, Zhang Y, Sun S, Zhong SL: Lack of association between lipoprotein(a) genetic variants and subsequent cardiovascular events in chinese han patients with coronary artery disease after percutaneous coronary intervention. *Lipids Health Dis* 2013, **12**:127.
34. Lv X, Zhang Y, Rao S, Liu F, Zuo X, Su D, Wang M, Xia M, Guo H, Feng D, Hong C, Li D, Ma W, Quyang P, Li X, Feng X, Yang Y, Ling W, Qiu J: Lack of association between four snps in the *slc22a3-lpa2-lpa* gene cluster and coronary artery disease in a chinese han population: A case control study. *Lipids Health Dis* 2012, **11**:128.
35. Luke MM, Kane JP, Liu DM, Rowland CM, Shiffman D, Cassano J, Catanese JJ, Pullinger CR, Leong DU, Arellano AR, Tong CH, Movsesyan I, Naya-Vigne J, Noordhof C, Feric NT, Malloy MJ, Topol EJ, Koschinsky ML, Devlin JJ, Ellis SG: A polymorphism in the protease-like domain of apolipoprotein(a) is associated with severe coronary artery disease. *Arterioscler Thromb Vasc Biol* 2007, **27**:2030–2036.
36. Chasman DI, Shiffman D, Zee RY, Louie JZ, Luke MM, Rowland CM, Catanese JJ, Buring JE, Devlin JJ, Ridker PM: Polymorphism in the apolipoprotein(a) gene, plasma lipoprotein(a), cardiovascular disease, and low-dose aspirin therapy. *Atherosclerosis* 2009, **203**:371–376.
37. Li Y, Luke MM, Shiffman D, Devlin JJ: Genetic variants in the apolipoprotein (a) gene and coronary heart disease. *Circ Cardiovasc Genet* 2011, **4**:565–573.
38. Kathiresan S, Willer CJ, Peloso GM, Demissie S, Musunuru K, Schadt EE, Kaplan L, Bennett D, Li Y, Tanaka T, Voight BF, Bonycastle LL, Jackson AU,

- Crawford G, Surti A, Guiducci C, Burt NP, Parish S, Clarke R, Zelenika D, Kubalanza KA, Morken MA, Scott LJ, Stringham HM, Galan P, Swift AJ, Kuusisto J, Bergman RN, Sundvall J, Laakso M, et al: **Common variants at 30 loci contribute to polygenic dyslipidemia.** *Nat Genet* 2009, **41**:56–65.
39. Caplice NM, Panetta C, Peterson TE, Kleppe LS, Mueske CS, Kostner GM, Broze GJ, Simari RD: **Lipoprotein (a) binds and inactivates tissue factor pathway inhibitor: A novel link between lipoproteins and thrombosis.** *Blood* 2001, **98**:2980–2987.
  40. Grainger DJ, Kemp PR, Liu AC, Lawn RM, Metcalfe JC: **Activation of transforming growth factor-beta is inhibited in transgenic apolipoprotein(a) mice.** *Nature* 1994, **370**:460–462.
  41. Tsimikas S, Brilakis ES, Miller ER, McConnell JP, Lennon RJ, Kornman KS, Witztum JL, Berger PB: **Oxidized phospholipids, lp(a) lipoprotein, and coronary artery disease.** *N Engl J Med* 2005, **353**:46–57.
  42. Koschinsky ML, Marcovina SM: **Structure-function relationships in apolipoprotein(a): Insights into lipoprotein(a) assembly and pathogenicity.** *Curr Opin Lipidol* 2004, **15**:167–174.
  43. Miles LA, Plow EF: **Lp(a): An interloper into the fibrinolytic system?** *Thromb Haemost* 1990, **63**:331–335.
  44. Cushing GL, Gaubatz JW, Nava ML, Burdick BJ, Bocan TM, Guyton JR, Weibaecker D, DeBakey ME, Lawrie GM, Morrisett JD: **Quantitation and localization of apolipoproteins [a] and b in coronary artery bypass vein grafts resected at re-operation.** *Arteriosclerosis* 1989, **9**:593–603.
  45. Malek F, Dvorak J, Svitil J, Skalnikova V, Mates M, Kmonicek P, Formanek P, Aschermann O, Kopriva K, Neuzil P: **Correlation of lipoprotein(a) concentration with the extent of coronary artery disease in patients on lipid lowering therapy.** *Neuro Endocrinol Lett* 2012, **33**(Suppl 2):55–59.
  46. Moon JY, Kwon HM, Kwon SW, Yoon SJ, Kim JS, Lee SJ, Park JK, Rhee JH, Yoon YW, Hong BK, Rim SJ, Kim HS: **Lipoprotein(a) and ldl particle size are related to the severity of coronary artery disease.** *Cardiology* 2007, **108**:282–289.
  47. Miller NE, Hammett F, Saltissi S, Rao S, van Zeller H, Coltart J, Lewis B: **Relation of angiographically defined coronary artery disease to plasma lipoprotein subfractions and apolipoproteins.** *Br Med J (Clin Res Ed)* 1981, **282**:1741–1744.
  48. Brilakis ES, McConnell JP, Lennon RJ, Elesber AA, Meyer JG, Berger PB: **Association of lipoprotein-associated phospholipase a2 levels with coronary artery disease risk factors, angiographic coronary artery disease, and major adverse events at follow-up.** *Eur Heart J* 2005, **26**:137–144.
  49. Virani SS, Brautbar A, Davis BC, Nambi V, Hoogeveen RC, Sharrett AR, Coresh J, Mosley TH, Morrisett JD, Catellier DJ, Folsom AR, Boerwinkle E, Ballantyne CM: **Associations between lipoprotein(a) levels and cardiovascular outcomes in black and white subjects: The atherosclerosis risk in communities (aric) study.** *Circulation* 2012, **125**:241–249.
  50. Dumitrescu L, Glenn K, Brown-Gentry K, Shephard C, Wong M, Rieder MJ, Smith JD, Nickerson DA, Crawford DC: **Variation in lpa is associated with lp(a) levels in three populations from the third national health and nutrition examination survey.** *PLoS One* 2011, **6**:e16604.
  51. Ferreira NE, Omae S, Pereira A, Rodrigues MV, Miyakawa AA, Campos LC, Santos PC, Dallan LA, Martinez TL, Santos RD, Mill JG, Krieger JE, Pereira AC: **Thioredoxin interacting protein genetic variation is associated with diabetes and hypertension in the brazilian general population.** *Atherosclerosis* 2012, **221**:131–136.
  52. Alvim RO, Santos PC, Ferreira NE, Mill JG, Krieger JE, Pereira AC: **Thioredoxin interacting protein (txnip) rs7212 polymorphism is associated with arterial stiffness in the brazilian general population.** *J Hum Hypertens* 2012, **26**:340–342.
  53. De Oliveira AR, Santos PC, Musso MM, De Sá CR, Krieger JE, Mill JG, Pereira AC: **Impact of diabetes mellitus on arterial stiffness in a representative sample of an urban brazilian population.** *Diabetol Metab Syndr* 2013, **5**:45.
  54. Santos PC, Morgan AC, Jannes CE, Turolla L, Krieger JE, Santos RD, Pereira AC: **Presence and type of low density lipoprotein receptor (ldlr) mutation influences the lipid profile and response to lipid-lowering therapy in brazilian patients with heterozygous familial hypercholesterolemia.** *Atherosclerosis* 2014, **233**:206–210.

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