

Immune cells in pancreatic cancer

Joining the dark side

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Abbreviations: ADM, acinar–ductal metaplasia; GM-CSF, granulocyte macrophage colony stimulating factor; IFN γ , interferon-gamma; IL-6, interleukin 6; IL-17, interleukin 17; IL-17RA, IL-17 receptor A; MDSC, myeloid-derived suppressor cell; PanIN, pancreatic intraepithelial neoplasia; PDA, pancreatic ductal adenocarcinoma; T_H17, T helper 17 cell

Pancreatic tumors are rich in immune cell infiltrates that include CD4⁺ T-cell subsets encompassing both regulatory T cells and T_H17 cells. Rather than protecting the organism by exerting an anticancer effect, these T-cell subsets promote tumor formation. Thus, re-activation of antitumor immunity should be investigated for use in pancreatic cancer prevention and therapy.

Pancreatic ductal adenocarcinoma (PDA), the most common form of pancreatic cancer, is one of the deadliest human malignancies.¹ In the past 40 y, the 5-y survival rate of pancreatic cancer has not markedly improved pointing to a dire need to develop novel therapeutic approaches. In humans, PDA is characterized by the accumulation of an extensive stroma comprising extracellular matrix components, activated fibroblasts, vascular components, and abundant immune cell infiltrates. Targeting the tumor-associated microenvironment is emerging as a new intervention approach for this deadly disease. However, our understanding of how immune infiltrates regulate pancreatic carcinogenesis is currently incomplete.

Pancreatic cancer in humans is preceded by precursor lesions known as pancreatic intraepithelial neoplasia (PanIN). PanIN is driven by expression of an oncogenic form of the *KRAS* gene in the pancreatic epithelium² and is markedly accelerated by the induction of pancreatitis.³ PanIN formation is accompanied by extensive infiltration of immune cells;⁴ however, the majority of the infiltrating

immune cells are immunosuppressive. Among infiltrating T lymphocytes, CD8⁺ T cells are rare whereas CD4⁺ T cells are abundant.⁴

Here, we describe two recent studies that address the functional role of CD4⁺ T cells within the pancreatic cancer microenvironment.^{5,6} Of note, two different genetically engineered mouse models were used: the iKras^{*} mouse model⁷ and the KC^{iMist1} (*Mist1*^{CreERT2/+}; *LSL-Kras*^{G12D}) mouse model. In these models, oncogenic Kras is expressed in a tissue-specific and inducible manner using the Tet or CreER system respectively.

In the first study,⁵ we genetically depleted CD4⁺ T cells by crossing iKras^{*} mice with CD4^{-/-} mice. The resulting iKras^{*};CD4^{-/-} mice were found to be susceptible to caerulein-induced pancreatitis. However, unlike iKras^{*} mice, they do not develop PanIN lesions following the induction of pancreatitis. In fact, the low-grade PanIN lesions found in these animals undergo extensive apoptosis and do not persist over time. This is in sharp contrast to iKras^{*} mice, which rapidly develop tissue-wide PanINs after the induction of pancreatitis and show progression to

higher grade lesions over time. Thus, this study highlighted the requirement of CD4⁺ T cells for pancreatic cancer initiation. Of note, CD4⁺ T cell depletion also led to reduced PanIN formation in the KC^{iMist1} model.⁶

Infiltration of CD8⁺ T cells was increased in iKras^{*};CD4^{-/-} mice and was often observed in close proximity to epithelial cells in areas of acinar ductal metaplasia and low-grade PanINs. More importantly, CD8⁺ T cells extracted from iKras^{*};CD4^{-/-} mice were more active, based on expression of interferon γ (IFN γ) and Granzyme B, and more receptive to further activation upon treatment with anti-CD3 and anti-CD28 antibodies *in vitro*. When freshly sorted PanIN and CD8⁺ T cells were co-cultured, CD8⁺ T cells derived from iKras^{*};CD4^{-/-} mice, but not CD8⁺ T cells derived from iKras^{*} mice, were activated in response to PanINs. Finally, depletion of CD8⁺ T cells in iKras^{*};CD4^{-/-} mice rescued PanIN formation. Taken together, these data show that CD4⁺ T cells promote PanIN formation by blocking the antitumor immune responses mediated by CD8⁺ T cells (Fig. 1).

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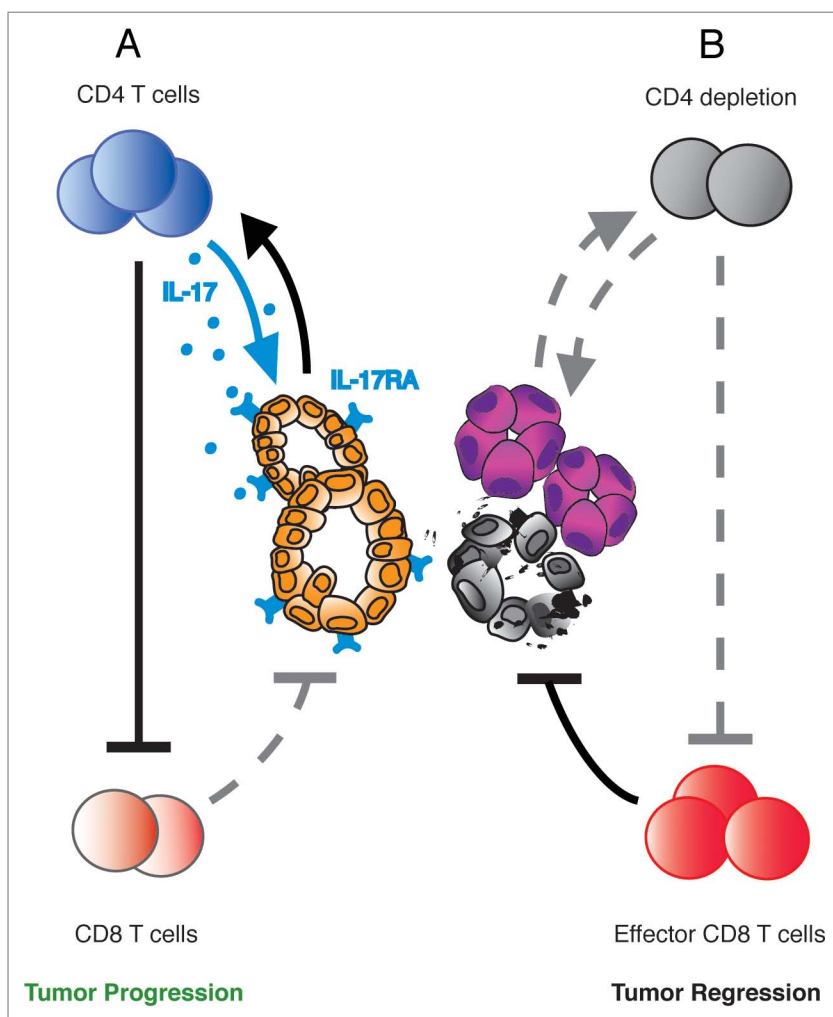


Figure 1. CD4⁺ T lymphocytes and IL-17 signaling are required for oncogenic Kras-driven pancreatic carcinogenesis. (A) Intrapancreatic CD4⁺ T cells suppress the antitumor activity of CD8⁺ T cells during Kras*-driven pancreatic intraepithelial neoplasia (PanIN) formation. T helper 17 (T_H17) cells (as well as $\gamma\delta$ T cells) secrete IL-17A that signals through IL-17RA in acinar-ductal metaplasia and PanINs, thereby inducing tumor initiation and progression. (B) CD4⁺ T cell ablation enables effector CD8⁺ T cell function and induces apoptosis in PanIN cells, thus blocking the onset of pancreatic cancer initiation.

CD4⁺ T cells are a heterogeneous population. High numbers of regulatory T cells and T helper 17 (T_H17) cells were observed in the pancreatic microenvironment in a Kras-dependent manner.⁵ The level of T_H17 cells was also found to be elevated in the pancreatic immune-infiltrates of KC^{iMist1} mice and formed the focus of the second study,⁶ which demonstrated that these cells were required for initiation and progression of pancreatic tumorigenesis.

Using the KC^{iMist1} model, McAllister et al. showed that oncogenic Kras and chronic pancreatitis synergistically recruited T_H17 and interleukin-17 (IL-17)⁺/ $\gamma\delta$ T cells to the pancreatic

microenvironment.⁶ Different and complementary approaches were used to address the function of these cells in pancreatic tumorigenesis. Adenoviral-mediated IL-17 overexpression in the pancreas of KC^{iMist1} mice markedly accelerated PanIN initiation and progression. In contrast, genetic inhibition of IL-17 signaling by transplanting KC^{iMist1} mice with bone marrow from IL-17 deficient mice, or alternatively, by a pharmacological approach in which KC^{iMist1} mice were treated with IL-17 and IL-17RA monoclonal antibodies, prevented PanIN formation (Fig. 1).

Several changes in immune infiltration and inflammatory signaling were

observed in both iKras*;CD4^{-/-} mice and KC^{iMist1} mice upon inactivation of the IL-17 axis. In both studies, a decrease in myeloid-derived suppressor cells (MDSC), typically abundant within the pancreatic cancer microenvironment⁴ and required for pancreatic cancer development, was observed. Moreover, in both investigations, a reduction in the activation of the IL-6/p-Stat3 signaling axis, was detected, a key pathway required for PanIN formation and progression.^{8,9} Expression of granulocyte macrophage colony stimulating factor (GM-CSF, also known as Csf2), another important factor for pancreatic cancer progression,^{10,11} was also reduced, possibly explaining the diminished MDSC recruitment. The interactions between individual immune subsets, as well as the role of other components of the stroma in regulating the inflammatory microenvironment of pancreatic cancer, are areas of active investigation.

Taken together, these studies highlight an important role for CD4⁺ T cells, specifically the Th17 cell subset, during the pancreatic cancer onset and disease progression, and argue for immune modulation as a valid therapeutic approach for this dreaded disease.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

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