

Published in final edited form as:

FEBS Lett. 2008 January 9; 582(1): 132–141. doi:10.1016/j.febslet.2007.11.063.

## Neuronal control of energy homeostasis

Qian Gao<sup>1</sup> and Tamas L. Horvath<sup>1,2,3</sup>

<sup>1</sup>Section of Comparative Medicine, Yale University School of Medicine, New Haven CT 06520

<sup>2</sup>Department of Neurobiology, Yale University School of Medicine, New Haven CT 06520

<sup>3</sup>Department of Obstetrics, Gynecology & Reproductive Sciences, Yale University School of Medicine, New Haven CT 06520

### Abstract

Neuronal control of body energy homeostasis is the key mechanism by which animals and humans regulate their long-term energy balance. Various hypothalamic neuronal circuits (which include the hypothalamic melanocortin, midbrain dopamine reward and caudal brainstem autonomic feeding systems) control energy intake and expenditure to maintain body weight within a narrow range for long periods of a life span. Numerous peripheral metabolic hormones and nutrients target these structures providing feedback signals that modify the default “settings” of neuronal activity to accomplish this balance. A number of molecular genetic tools for manipulating individual components of brain energy homeostatic machineries, in combination with anatomical, electrophysiological, pharmacological and behavioral techniques, have been developed, which provide a means for elucidating the complex molecular and cellular mechanisms of feeding behavior and metabolism. This review will highlight some of these advancements and focus on the neuronal circuitries of energy homeostasis.

### Keywords

Leptin; Ghrelin; Arcuate nucleus; NPY; POMC; GABA; glutamate

## 1. Introduction

Feeding and energy metabolism is a basic and vital life process essential for individual and, consequently, species survival. It could be argued that the emergence and evolution of the central nervous system is to promote the most effective means of dealing with feeding and metabolism in support of survival. Accumulating experimental evidence is in agreement with this basic assertion. Thus, it is not unreasonable to claim that even higher brain regions, such as the archio- and neocortex, emerged under environmental pressures to support

---

© 2007 Federation of European Biochemical Societies. Published by Elsevier B.V. All rights reserved.

tamas.horvath@yale.edu, qian.gao@yale.edu

**Publisher's Disclaimer:** This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

behaviors that more effectively deal with available energy resources. The pursuit to understand the role that the brain plays in the control of body energy balance has continued for more than a century, when patients with pituitary adenomas were initially observed to rapidly gain body weight (1, 2). Increasingly sophisticated technologies and methods have since been brought to bear on the topic and a vast number of observations have been made, particularly within the last decade due to the cloning of the product of the *obesity (ob)* gene, leptin, the mutation of this gene causes obesity in rodents (3), which brought about a wave of discovery of other metabolic hormones, neuropeptides and signaling pathways, and established their relationship to key neuronal structures in the hypothalamus (4, 5, 6, 7). Based upon these fundamental findings, the studies of more complex signal intergrations and neuronal adaptations that actually initiate changes in ingestive behavior and metabolism have become possible. This review will follow this line of research history and highlight some important new advances.

## 2. Discovery of the brain regions that control energy balance

The initial observations of Mohr (1840) and Frohlich (1901) regarding pituitary adenomas led to the hypothesis that adenoma-related obesity is caused by damage to the hypothalamus (8). This important idea was supported by the observation that obesity developed in an experimental dog given that underwent a hypothalamic lesion (9). The actual identification of specific hypothalamic structures that are directly involved in energy homeostasis, however, was accomplished about three decades later, when a set of systematic lesion experiments was performed in rats, in which various hypothalamic nuclei that include the hypothalamic ventromedial (VMH), paraventricular (PVH) and dorsomedial (DMH) nuclei were destroyed. The lesions of these nuclei induced hyperphagia and obesity (10, 11, 12); whereas, lesions in the lateral hypothalamus (LH) led to hypophagia (13). This study, which stands as a milestone in linking brain function to body energy balance, led to the proposal of a “dual center model” that identified the hypothalamic VMN as the “satiety centre,” and the LH as the “hunger centre” (14). More studies, including chemical lesions as well as pharmacological applications of various hormones and drugs, which either stimulate or inhibit the neuron populations locally, have since been done and all supported the initial observations that lead to the proposed “dual center model.” In retrospect to our current knowledge of specific hypothalamic neuronal populations, these lesion studies are strikingly precise in distinguishing between subregions that contain circuits that either promote or suppress feeding. These findings have since been used as a road map for studies concerning the neurobiology of feeding and metabolism.

## 3. Linking the periphery to the brain

The brain regions that control energy homeostasis, by definition, need to “know” the amount of pre-existing body energy stores in order to “wisely” determine the levels of energy intake and expenditure. The nature of the signal(s) that conveys such information to the brain was the subject of a couple of hypotheses. The “glucostatic hypothesis,” (15) suggested that small changes in plasma glucose levels trigger meal initiation or termination. This “depletion-repletion” model of energy intake correlated well with the initiation or termination of feeding per se, however, it did not provide information about or a mechanism

by which to ensure that food consumption with respect to existing energy stores would be appropriate and it correlated poorly with energy expenditure (16). The “lipostatic model,” on the other hand, hypothesized that signal(s) proportional to the amount of fat in the body modulates the amount of food eaten at each meal to maintain overall energy balance (17). Later discoveries have supported this hypothesis that food intake is controlled within a lipostatic system to maintain energy homeostasis.

The findings from the initial parabiosis studies on hypothalamic lesioned rats appear to be essential to the conclusion of the existence of humoral signal(s) in relation to an animal’s lipostatics (18). In these studies, two live animals, one with a lesion in the VMH, were joined by suturing, which allowed humoral factors to pass from one animal to the other. As obesity developed in the lesioned rat, its partner became hypohagic and lost weight, suggesting that a signal, in proportion to the amount of fat mass, is highly potent in inhibiting food intake. Additional parabiosis studies on genetically obese mutant mice, *ob/ob* and *db/db* (19, 20), concluded that *ob/ob* mice lack this lipostatic signal, whereas *db/db* mice are insensitive to it. These hypotheses were later confirmed by the discoveries of the *ob* gene that encodes leptin (3) and the *db* gene that encodes leptin receptor.

The detection of leptin and its receptor soon led to the recognition that all of the hypothalamic nuclei associated with energy regulation, i.e., the VMH, DMH, PVN and LH, are regions where leptin receptors are highly expressed. This promoted a new flourish of gene discoveries, which identified novel neuropeptides, their receptors and transcription factors that mediate leptin function in the hypothalamus and subsequently, led to the discovery of the hypothalamic melanocortin system, the key neuronal system in the control of energy balance by leptin signaling. Other metabolic signals, such as insulin, ghrelin, estrogen, prolactin, glucocorticoids, resistin and interleukins, etc., as well as nutrients such as glucose and free fatty acids (21, 22, 23), which, to a certain extent, modify energy balance and body weight, also target the hypothalamic melanocortin system (16, 5 24, 25, 26), presumably in coordination with leptin function.

#### 4. The importance of the melanocortin system in energy homeostasis

The melanocortin model has held the most significance in explaining the neuronal control of energy balance. In this model, the hypothalamic arcuate nucleus (ARC) is considered a critical region for various reasons. First, the neurons within the ARC are anatomically placed in close proximity to fenestrated capillaries at the base of the hypothalamus, giving them access to humoral signals that are restricted from other regions of the brain (27, 28). Indeed, they respond rapidly to fluctuations in nutrients (7, 29) and metabolic hormones. These neurons are also innervated by axons containing all major neurotransmitters and express receptors for most metabolic hormones (5, 27, 30, 31), implying that an extensive neuronal control exists. Finally, they project broadly to the brain and periphery both directly as well as indirectly.

Two sets of neurons, with opposite effects on feeding, have been identified in the ARC. The neurons that express proopiomelanocortin (POMC) and cocaine-amphetamine regulated transcript (CART) are anorexigenic (32, 33, 34), due to the release of the cleavage products

of POMC precursor,  $\alpha$ -, and  $\beta$ -melanocyte stimulating hormones ( $\alpha$ - and  $\beta$ -MSH) (34). These, in turn, reduce food intake and body weight as well as increase energy expenditure in animals and humans (35, 36, 37) by acting on melanocortin receptor subtypes 3 and 4 (MC3 and 4R) (38), found to be abundant in the ARC, PVN, LH and DMH (39). In contrast, cells containing neuropeptide Y (NPY) and the agouti gene related transcript (AgRP) are orexigenic (40). NPY potently stimulates food intake and reduces energy expenditure (41). In genetically obese animals, i.e., *ob/ob* and *db/db* mice, as well as those in a negative energy state, i.e., fasted or lactating animals, the ARC NPY mRNA and protein content are elevated (42, 43, 44, 45). AgRP acts as a natural antagonist of MC3R and MC4R, thereby, reducing the anorectic effect of  $\alpha$ -MSH (40, 46, 47, 48). An ectopic expression of Agouti, an AgRP like peptide, results in an obese phenotype of the *yA* agouti mouse (40).

The selective expression of fluorescent reporter genes in specific neurons allows for the visualization of hypothalamic neuronal subpopulations so that they can be studied in the context of feeding circuits while the cells are still alive. Through the use of this method, substantial neuroanatomical and electrophysiological data have been generated, which demonstrate that NPY neurons contact nearby POMC cells and inhibit them through the release of GABA (49). This unidirectional NPY to POMC interaction (50) may represent a wiring blueprint that favors the tonic inhibition of satiety signals to not only promote feeding, but also overfeeding when food is available in excess (51).

The ARC POMC and NPY neurons project to various parts of the brain, e.g., the PVN, LH and perifornical hypothalamic region, all of which contain substantial numbers of MC3 and 4 receptors (39, 52). The projection from the ARC to the PVN is important for the regulation of neurons that produce corticotropin and thyrotropin releasing hormones (CRH and TRH, respectively) and for the modulation of sympathetic activity, both of which are significant mechanisms in energy metabolism (53, Fig. 2). A recent study using a *cre/loxP* genetic approach showed that mice with mutations to the MC4R gene (*loxTB mc4*) ate and weighed less if the MC4R gene was restored selectively in the PVN (54). However, it was not sufficient to bring energy expenditure levels to those of controls, suggesting that energy expenditure is regulated by melanocortins elsewhere in the hypothalamus and/or through receptors other than MC4R (54).

Recent studies showed that intact melanocortin neuronal circuitry is important for acute regulation of feeding (55, 56). These studies stemmed from the considerable frustration encountered due to the fact that neither NPY nor AgRP single gene knockout mice (57) nor NPY/AgRP double knockout mice (58) exhibited the expected hypophagia. To examine whether the NPY and POMC neurons themselves are crucial in adult energy regulation, whereas the loss of NPY/AgRP or POMC genes may be compensated for during development, both groups had used the cell-targeted expression of diphtheria toxin receptors that do not exist in mice, making these animals “immune” to diphtheria toxin-induced necrosis. When the otherwise normally developing diphtheria toxin receptor expressing mice were injected with two subsequent doses of diphtheria toxin, the targeted hypothalamic neurons rapidly died resulting in hypophagia in NPY neuron-ablated mice (55, 56) and hyperphagia in POMC neuron-ablated animals (55). Interestingly, the ablation of NPY neurons during the early postnatal period did not result in an overt metabolic phenotype,

indicating that functional effects of degenerated neurons in critical developmental periods may be overcome by re-organization of the circuits.

## 5. The function of satiety center, VMH: does it modulate the melanocortin circuits?

The VMH was directly implicated in feeding when Hetherington and Ranson showed that electrolytic lesions to this region in rats resulted in rapid development of obesity (10, 59, 11, 12). Chemical lesions as well as pharmacological studies have since supported the notion that the VMH inhibits feeding and increases metabolism and by doing so, restricts the amount of body fat (60, 61). The VMH was found to highly express the long form leptin receptor, LRb, and, therefore, is the region that mediates leptin's effect on homeostasis (62, 63). Despite the early discovery of the anorexic effect of the VMH, little is known regarding the cellular mechanisms by which VMH neurons contribute to homeostasis under the control of leptin signals.

Recent studies, however, show that the VMH is equally important and functions alongside the ARC in energy regulation in response to leptin (64). These studies examined the behavioral and metabolic phenotype of animals with the leptin receptor selectively deleted in POMC neurons (65). Surprisingly, these animals were at best only mildly obese and ate similar amounts of food as controls. In contrast, a selective deletion of the leptin receptor gene in the neurons that express steroid factor-1 (SF-1) in the VMH results in mice that are not only obese but also hyperphagic (66, 67). SF-1 is a transcription factor necessary for the development of the VMH (68, 69, 70). Mice with mutations to leptin receptors in both the ARC POMC and the VMH SF1 neurons are more obese than those with mutations of leptin receptors in either set of neurons alone (67).

How do VMH neurons act to deliver the anorectic effect in the context of hypothalamic feeding circuits? A recent study showed that the VMH may directly increase the activity of POMC neurons via microcircuits (71) that were previously difficult to detect by conventional tracing techniques (72, 73, 74). Using laser scanning photo-stimulation (LSPS) in combination with slice electrophysiology, these authors showed that the inputs from the VMH neurons are mostly excitatory, thus increasing the activity of POMC cells; during a fast, these inputs decrease (71). Whether or not these VMH neurons receive leptin signaling requires further investigation.

Conversely, the ARC may also modulate the activity of the VMH. Although afferent projections from the ARC to the VMH are sparser than those to other nuclei (73), the VMH contains MC4R as well as NPY Y1, Y2 and Y5 receptors (75, 76, 77, 78, 79, 80), suggesting that both POMC and NPY neurons project to the VMH. Infusions of NPY into the VMH increase feeding; fasting is associated with elevated levels of NPY in this region (75, 5), whereas electrophysiological responses of neurons in the VMH to  $\alpha$ -MSH are decreased in animals that are fasted and/or treated with AgRP prior to sacrifice than in animals that have free access to food (80).

A notable characteristic of the VMH is that it highly and specifically expresses the brain-derived neurotrophic factor (BDNF) that has been shown to affect the metabolic functions regulated by the VMH. Genetic deficiencies of BDNF or its TrkB receptor resulted in obesity in both humans and mice (81, 82). Interestingly, leptin increases BDNF transcripts (83, 84, 85, 86), whereas fasting decreases them selectively in the VMH, suggesting that the BDNF is a regulatory component of leptin signaling. Since BDNF in the brain is known to promote synaptic morphology and function (81), this raises the possibility that synaptic plasticity in the hypothalamus may be a part of the regulatory mechanism of energy regulation. Notably, the anorectic effects of BDNF are not directly mediated by the melanocortin system since BDNF reduces the body weight and food intake of mice that lack the MC4R (82), yet neurons in the VMH form excitatory microcircuits with POMC neurons in the ARC (71), suggesting that the VMH may influence POMC neuroactivity by other means, such as classic neurotransmitters and/or plasticity. Indeed, leptin and other hormones increase the number of excitatory synapses on POMC cells (87, 88).

## 6. The lateral hypothalamic arousal-feeding system

In the lateral hypothalamus (LH), two sets of neurons that contain either hypocretin (orexin), a peptide implicated in arousal and feeding, or melanin-concentrating hormone (MCH), another potent stimulator of food intake, have been identified. These neurons are innervated by both POMC and NPY terminals, suggesting that their function in motivating food intake is also within the confines of the ARC melanocortin system (89, 90, 91, 92, 93, 48, 94, 95, 96, 97, 98, 99). Both MCH and hypocretin neurons have a wide projection field and modulate a variety of behavioral responses related to learning, memory, emotion, motivation and motor responses in association with changes in the energy state (100, 92, 101, 102, 97). Although the projections of MCH and hypocretin neurons exhibit significant overlap, their overall effects and actual targets are quite different (100, 96). Like that of the NPY and POMC neurons, the activity of MCH and hypocretin neurons is regulated by numerous hormones that include leptin and ghrelin, as well as by practically every neurotransmitter system (91, 103, 104, 105, 106). Within the LH, MCH and hypocretin neurons have reciprocal connections with each other, and with nearby neurons (99). Electrophysiological studies in brain slices or isolated neurons indicated that hypocretin, in general, has a stimulative effect on LH neurons including MCH neurons (107, 108), whereas MCH depresses the synaptic activity of glutamate and GABA neurons from the rat LH (109). It is not clear whether such electrical interactions between hypocretin and MCH neurons, at the intensities and dynamics observed, are more relevant to feeding or arousal behavior.

MCH and hypocretin neurons are able to integrate metabolic signals to modulate energy balance, more or less, independent of each other, despite their proximate location and physiological interaction in the LH. The mRNA levels of hypocretin in the LH are up-regulated upon fasting (93). Hypocretin neurons are rapidly activated by fasting in rodents and non-human primates (110) and exhibit leptin-dependent synaptic plasticity during fasting (111). Moreover, all of the hypothalamic neurons that are activated by fasting receive strong hypocretin input. These observations, together with the earlier demonstration of a massive hypocretin input to the ARC, particularly to the NPY neurons (112), as well as the dependence of hypocretin function on NPY signaling (in particular on the Y1 and Y5

receptors) (113, 114), and a synergistic action between NPY and hypocretin (at low concentrations) to induce feeding (115) argue that hypocretin neurons may be upstream to the NPY system with regard to feeding and metabolism. Mice with deletions of the hypocretin gene are hypophagic, but maintain normal growth curves, suggesting that their metabolic rates are reduced (116). This, in association with the critical role of hypocretin neurons to promote arousal (117) through direct projections to the brain stem, including the locus coeruleus (112) make the hypocretin neurons a likely candidate to provide the link between obesity and insomnia.

The orexigenic peptide MCH, on the other hand, shows little or no interaction with NPY or hypocretin in inducing food intake when injected together into the third ventricle of the rat (115). This observation provides a physiological component concomitant to the pre-existing morphological data that shows that the strength of MCH projections to the ARC is limited compared to that of the hypocretin neurons. Thus, the action of MCH on feeding is likely independent of NPY and hypocretin. However, like NPY, MCH does exhibit characteristics of orexigenic genes: its mRNA levels are increased in obese mutant animals and fasting further increases its expression in both normal and obese animals (89); and it has a potent orexigenic effect. Targeted deletion of the MCH gene resulted in a phenotype of hypophagia and leanness with an inappropriately high metabolic rate (118), suggesting that MCH is a typical “thrifty gene” that increases energy intake and reduces energy expenditure simultaneously. Consistent with this idea, MCH suppresses thyroid stimulating hormone (TSH) release (119).

The orexigenic effect of MCH appears to compete with the action of  $\alpha$ -MSH (a mechanism conserved from skin color regulation in fish to hypothalamic control of energy balance in mammals), such that MCH administration increases feeding, while  $\alpha$ -MSH acts to decrease it. When the peptides are administered together, depending on the relative ratios, one antagonizes the action of the other (92). Recently, a mouse model of MCH neuron ablation was generated by expressing a toxin gene, ataxin-3, targeted at MCH neurons (120). Mice that express this gene have a chronic loss of MCH neurons. Interestingly, the phenotype of these mice highly resembles that of mice lacking only the MCH gene, exhibiting reduced food intake and increased energy expenditure. Moreover, the ablation of MCH neurons in mice with an *ob/ob* mutant background improved obesity and glucose tolerance. These results suggest that the function of MCH cells in energy regulation is limited to the MCH system itself, but not to other aspects of the cells such as their classic neurotransmitter function and/or synaptic plasticity, which are distinct from NPY cells.

## 7. Other hypothalamic regions that regulate food intake and body weight

Receptors for leptin and ghrelin are also found in other hypothalamic nuclei including the DMH and the suprachiasmatic nucleus (SCN) (103, 121). The DMH has been implicated in the regulation of energy balance, however, its actual role remained obscure until recently (122, 123, 124). The DMH is involved in a variety of regulatory mechanisms that include the modulation of glucocorticoid secretion, body temperature, arousal, and circadian rhythms of locomotor activity (123). The DMH receives inputs from cells in the ARC and from brain stem centers that are also implicated in feeding (122). Lesions restricted to the

DMH typically result in hypophagia, although animals can still maintain their body composition (122). Recently, the DMH was shown to be critical for the entrainment of circadian rhythms to feeding schedules (124). The DMH of animals with restricted access to food (4 hours/day) had increased expression of c-Fos, indicating an increased cellular activation at a time when the food was regularly presented compared to animals that had free access to food throughout the day. Ibotenic lesions of the DMH resulted in reduced levels of locomotor activity and decreased food intake. Furthermore, when lesioned rats were placed in the restricted feeding schedule, they showed less preprandial increases in food anticipatory locomotor activity than those of sham operated animals. DMH lesions also blocked the rise in body temperature that is entrained to the timing of food presentation (124). The phenotype of cells within this region remains obscure. A number of these cells produce glutamate as a neurotransmitter and project to the PVN as well as to the preoptic area, both thought to be involved in the circadian regulation of corticosteroid secretion and body temperature (125, 122). Projections from the DMH to the LH and to the ventrolateral preoptic area have been implicated in sleep and arousal and could presumably relate to the enhanced activity of animals in restricted feeding schedules (126).

The role of the SCN in energy balance has been overshadowed by the critical role of this region as a master clock that mediates circadian patterns of biological function (127). Among these, the SCN controls the circadian secretion of metabolic hormones and is thought to regulate seasonal adipose tissue content and maintain patterns of glucose levels (128, 129). The SCN projects to most hypothalamic nuclei, with a strong projection that terminates in the PVN and DMH (127). Recent evidence suggests that the ARC projects to the SCN as well, indicating that the melanocortin system can influence its activity (130). Importantly, mice with mutations of the *clock* gene, a transcript expressed in the SCN and critical for the generation of circadian rhythms, exhibit an obese phenotype (131). Hypothalamic *clock* gene expression is highest in the SCN, thus directly implicating this region in obesity caused by this mutation (132, 133).

## 8. Midbrain dopamine system-the motivation or reward to eat

The midbrain dopamine system is known to be involved in the regulation of arousal, locomotor activity, mood and reward (134). Dopamine deficiency in mice, generated by selective inactivation of tyrosine hydroxylase, markedly suppresses food intake in a manner similar to that of lesions of the LH (135, 136). These mice fail to eat in response to acute glucose deprivation (137), as well as to PYY administration or leptin deficiency (138), suggesting that dopamine signaling is absolutely required for promoting feeding and acts downstream of the melanocortin system. Among the various functions of the dopamine system, the reward pathways have received particular attention in explaining feeding behavior given the universality of food as a natural reinforcer. Dopaminergic neurons within the midbrain ventral tegmental area (VTA) that innervate the nucleus accumbens (the ventral striatum) have been implicated in the rewarding aspects of food, sex, and drugs of abuse (139). In support of a role for the mesolimbic reward circuitry in feeding regulation, it was recently found that interfering with ghrelin signaling specifically in the VTA diminished ghrelin-induced feeding (140). Surprisingly, however, the restoration of dopamine production within the dorsal striatum restores feeding on normal chow, whereas



restoration of dopamine in the nucleus accumbens does not. While these findings may suggest a fundamental difference between feeding for nourishment and food as a rewarding substance (141), a similar study in human subjects argues in the same vein: by using [(11)C]raclopride positron emission tomography (PET) scanning, feeding was found to be associated with dopamine release in the dorsal, but not the ventral striatum, and yet the amount of dopamine released correlated with the degree of pleasure experienced (142). Thus, further investigations are needed to discern the role of VTA versus nigral dopamine neurons in the regulation of feeding and energy homeostasis in general (143, 144).

Nevertheless, feeding is associated with motivational mechanisms important for the behavioral responses necessary for seeking food (145). Hypothalamic peptides like NPY,  $\alpha$ -MSH, AgRP, hypocretin and MCH, on the other hand, modulate the activity of dopaminergic neurons that target the nucleus accumbens (146). The ARC funnels metabolic information from signals like leptin to modulate the activity of the mesolimbic dopaminergic system via direct projections to the nucleus accumbens, or indirectly through the activation of hypocretin or MCH neurons that also project to both the VTA and nucleus accumbens (146). Emerging evidence, however, supports the idea that at least the VTA is sensitive to leptin, insulin and ghrelin, and that the activity of dopaminergic neurons within the VTA can be modulated by these signals (147, 148, 121). The implications of these observations remain controversial. Further research may reveal that, in contrast to the funnel hypothesis, metabolic signals could act directly on reward systems, including the dorsal striatum, to modulate motivational aspects of feeding in tandem with homeostatic systems (149, 148).

## 9. The caudal brainstem: autonomic control of eating

The caudal brainstem contains neurons and circuits that involve autonomic control of ingestion, digestion, and absorption of food (150), and do so without forebrain influence (151), just as for respiration and circulation, the functions essential for survival. Thus, the regulation of nutrient supply is to a large extent autonomically organized within the brainstem that generates most of the parasympathetic support during ingestive and digestive processes through the vagus nerve, as well as sympathetic responses related to severe energy depletion and energy expenditure. It has immediate access to the locomotor and oromotor apparatuses that enable approach of food and orchestrate ingestion of food and fluid placed into the oral cavity. Furthermore, the brainstem can terminate ingestion when the taste is aversive or when visceral sensors detect noxious stimuli. Thus, complete ingestive behavior and energy balance cannot occur without the circuits of the brainstem (152).

Interestingly, the caudal brainstem is involved in the control of meal size by signals arising from the mouth and gastrointestinal tract based upon the chemical or mechanical properties of food, which are relayed to the nucleus of the solitary tract (NST) via the vagus and other cranial nerves (153). In a chronic decerebrate rat preparation, the taste concentrations, gastric preloads and cholecystokinin (CCK; a peptide released from intestinal endocrine cells during feeding) all affected the meal size in a manner similar to that of intact rats. Thus, the brainstem isolated from its forebrain connection exhibits the basic behavior of satiety (154). This challenges the classic view that hypothalamic integration of visceral signals is necessary for the behavioral output that determines meal size. The specific neural

pathways necessary for the “satiety” response have not yet been identified. CCK appears required for the “satiety” response (155), which also involves neurons in the NTS as well as the dorsomotor nucleus of the vagus (DMV), and is blocked by vagal deafferentation (156). In addition to meal size control, decerebrate rats also show a fully formed sympathoadrenal response to systemic 2DG administration (157), indicating that the brainstem houses a complete system responsive to glucoprivation. However, decerebrate rats are not able to increase meal size appropriately in response to food deprivation (158), and, therefore, have an inadequate response to a long-term homeostatic challenge, which requires interactions between hypothalamic and caudal brainstem nuclei (152).

Consistent with these findings, the peripheral hormones leptin, ghrelin and insulin, neuropeptides  $\alpha$ -MSH and CRH as well as nutrients such as glucose all yield potent effects on food intake and body weight similar to their effects in the hypothalamus (152, 151). LRbs are found throughout the dorsal vagal complex (DVC, area postrema, nucleus of the solitary tract, dorsal motor nucleus of the vagus nerve) and in various other structures in the caudal brainstem (159). Localized stimulation of these receptors reduces food intake and body weight (160); Ghrelin receptors, GHS-R are expressed in the area postrema and nucleus tractus solitarius of the brainstem. Stimulation of GHS-R in the caudal brainstem led to a hyperphagic response (161, 162). Ghrelin delivered to the fourth ventricles significantly increased cumulative food intake, with maximal response approximately 3 h after injection. In a separate experiment, ghrelin microinjected unilaterally into the dorsal vagal complex (DVC) significantly increased food intake measured 1.5 and 3 h after treatment (163). Insulin receptors are also found in the NST (164). Furthermore, unilateral DVC injection of both MC3/4R and CRH1/2R agonists resulted in suppression of food intake, while a MC3/4R antagonist induced hyperphagia (165, 166, 167). The innervation of the brainstem by the perifornical and far-lateral hypothalamic MCH neurons has recently been identified (168). Lastly, it is noteworthy that the brainstem also houses glucose-sensitive neurons that are involved in the sympathoadrenal and ingestive responses to glucopenia in rodents (169).

## 10. Concluding remarks: questions on which to focus

Despite the complex and extensive involvement of the brain in energy homeostasis, a basic picture of the neuronal control behind this process is emerging. In this representation, the melanocortin system, broadly defined to include the hypothalamic ARC, VMH, DMH, PVN and LH, is positioned at the center, with the other brain regions, such as the midbrain dopamine system (controlling the motivation to eat or reward of eating) and the caudal brainstem (autonomic control of eating) positioned downstream. The key molecule that connects the entire network is the adipokine leptin, which conveys information from the periphery regarding pre-existing energy stores to the brain. These neuronal structures are all targeted, differentially, by leptin to modify their default settings, which otherwise tend to increase energy intake and reduce energy expenditure. Although close interactions between individual nuclei have been found, these nuclei and neuronal subpopulations act more or less in parallel but are not hierarchical.

This relatively simplified representation of the neuronal control of energy regulation highlights several issues that still need to be addressed. These include: 1) an intricate and

detailed labeling of all of the LRB expressing neurons in the brain, with particular attention given to the hypothalamus, midbrain and brainstem, in order to identify novel neuronal subpopulations, specifically in the VMH, DMH and LH. In fact, this work is already quickly progressing (personal communication, Myers); 2) focusing on neuronal adaptations, e.g., electrophysiological and plastic, directly induced by the metabolic hormones, leptin and ghrelin, that actually alter feeding behavior and/or metabolism; 3) discerning the particular effect of each nucleus and/or neuronal subpopulation in relation to homeostasis; identifying the niche, through which different neuronal circuits interact, and, more importantly, addressing the logic behind the parallel arrangement of brain structures/systems that control long-term energy balance.

## Acknowledgments

The work conducted for this review was supported by NIH grants DK-074386, DK-060711 and AG022880 to T.L.H..

## References

- [1]. Mohr B. Hypertrophie der Hypophysis cerebri und adurch bedingter Druck auf die Hirngrundflache, ins besondere auf die Sehnerven, das Chiasma derselben und den linkseitigen Hirnschinkel. *Wschr des Heilk.* 1840; 6:565–571.
- [2]. Frohlich A. Ein Fall von Tumor der Hypophysis cerebri ohne Akromegalie. *Wien Klin Rundschau.* 1901; 15:883–886. 906–908.
- [3]. Zhang Y, Proenca R, Maffei M, Barone M, Leopold L, Friedman JM. Positional cloning of the mouse obese gene and its human homologue. *Nature.* 1994; 372:425–432. [PubMed: 7984236]
- [4]. Friedman JM, Halaas JL. Leptin and the regulation of body weight in mammals. *Nature.* 1998; 395:763–770. [PubMed: 9796811]
- [5]. Kalra SP, Dube MG, Pu S, Xu B, Horvath TL, Kalra PS. Interacting appetite-regulating pathways in the hypothalamic regulation of body weight. *Endocr Rev.* 1999; 20:68–100. [PubMed: 10047974]
- [6]. Elmquist JK, Elias CF, Saper CB. From lesions to leptin: hypothalamic control of food intake and body weight. *Neuron.* 1999; 22:221–232. [PubMed: 10069329]
- [7]. Schwartz MW, Porte D Jr. Diabetes, obesity, and the brain. *Science.* 2005; 307:375–379. [PubMed: 15662002]
- [8]. Erdheim J. Uber Hypophysengangsgeschwulste und Hirncholesteatome. *Sitzungsd d k Akad d Wissensch. Math-naturw cl, Wien.* 1904; 113:537–726.
- [9]. Aschner B. Uber die Funktion der Hypothyse. *Arch f d ges Physiol.* 1912; 146:1–146.
- [10]. Hetherington AW, Ranson SW. Hypothalamic lesions and adiposity in the rat. *Anatomical Record.* 1940; 78:149.
- [11]. Brobeck JR, Tepperman J, Long CNH. Experimental hypothalamic hyperphagia in the albino rat. *Yale Journal of Biology and Medicine.* 1943; 15:831–853. [PubMed: 21434115]
- [12]. Brobeck JR. Mechanisms of the development of obesity in animals with hypothalamic lesions. *Physiological Reviews.* 1946; 26:541–559. [PubMed: 21002972]
- [13]. Anand BK, Brobeck JR. Localization of a feeding center in the hypothalamus of the rat. *Proceedings for the Society of Experimental Biology and Medicine.* 1951; 77:323–324.
- [14]. Stellar E. The physiology of motivation. *Psychol. Rev.* 1954; 61:5–22. [PubMed: 13134413]
- [15]. Mayer J. Regulation of energy intake and the body weight: the glucostatic theory and the lipostatic hypothesis. *Ann N Y Acad Sci.* 1955; 63(1):15–43. [PubMed: 13249313]
- [16]. Woods SC, Seeley RJ, Porte D Jr, Schwartz MW. Signals that regulate food intake and energy homeostasis. *Science.* 1998; 280:1378–1383. [PubMed: 9603721]

- [17]. Kennedy GC. The role of depot fat in the hypothalamic control of food intake in the rat. *Proc. R. Soc. Lond. B.* 1953; 140:579–592.
- [18]. Hervey GR. The effects of lesions in the hypothalamus in parabiotic rats. *J. Physiol.* 1959; 145:336–78. [PubMed: 13642304]
- [19]. Coleman DL, Hummel KP. Effects of parabiosis of normal with genetically diabetic mice. *Am. J. Physiol.* 1969; 217:1298–78. [PubMed: 5346292]
- [20]. Coleman DL. Obese and diabetes: two mutant genes causing diabetes-obesity syndromes in mice. *Diabetologia.* 1978; 14(3):141–8. [PubMed: 350680]
- [21]. Lam TK, Schwartz GJ, Rossetti L. (a) Hypothalamic sensing of fatty acids. *Nat Neurosci.* 2005; 8:579–584. [PubMed: 15856066]
- [22]. Lam TK, Gutierrez-Juarez R, Pocai A, Rossetti L. (b) Regulation of blood glucose by hypothalamic pyruvate metabolism. *Science.* 2005; 309:943–947. [PubMed: 16081739]
- [23]. Seeley RJ, York DA. Fuel sensing and the central nervous system (CNS): implications for the regulation of energy balance and the treatment for obesity. *Obes Rev.* 2005; 6:259–265. [PubMed: 16045641]
- [24]. Kojima M, Hosoda H, Date Y, Nakazato M, Matsuo H, Kangawa K. Ghrelin is a growth-hormone-releasing acylated peptide from stomach. *Nature.* 1999; 402:656–660. [PubMed: 10604470]
- [25]. Bray GA. Afferent signals regulating food intake. *Proc Nutr Soc.* 2000; 59:373–384. [PubMed: 10997653]
- [26]. Woods SC, Schwartz MW, Baskin DG, Seeley RJ. Food intake and the regulation of body weight. *Annu Rev Psychol.* 2000; 51:255–277. [PubMed: 10751972]
- [27]. Benoit S, Schwartz M, Baskin D, Woods SC, Seeley RJ. CNS melanocortin system involvement in the regulation of food intake. *Horm Behav.* 2000; 37:299–305. [PubMed: 10860674]
- [28]. Cone RD, Cowley MA, Butler AA, Fan W, Marks DL, Low MJ. The arcuate nucleus as a conduit for diverse signals relevant to energy homeostasis. *Int J Obes Relat Metab Disord.* 2001; 5(25 Suppl.):S63–67. [PubMed: 11840218]
- [29]. Obici S, Rossetti L. Minireview: nutrient sensing and the regulation of insulin action and energy balance. *Endocrinology.* 2003; 144:5172–5178. [PubMed: 12970158]
- [30]. van den Pol AN. Weighing the role of hypothalamic feeding neurotransmitters. *Neuron.* 2003; 40:1059–1061. [PubMed: 14687541]
- [31]. Seeley RJ, Drazen DL, Clegg DJ. The critical role of the melanocortin system in the control of energy balance. *Annu Rev Nutr.* 2004; 24:133–149. [PubMed: 15189116]
- [32]. Boston BA, Blaydon KM, Varnerin J, Cone RD. Independent and additive effects of central POMC and leptin pathways on murine obesity. *Science.* 1997; 278:1641–1644. [PubMed: 9374468]
- [33]. Ellacott KL, Cone RD. The central melanocortin system and the integration of short- and long-term regulators of energy homeostasis. *Recent Prog Horm Res.* 2004; 59:395–408. [PubMed: 14749511]
- [34]. Cone RD. Anatomy and regulation of the central melanocortin system. *Nat Neurosci.* 2005; 8:571–578. [PubMed: 15856065]
- [35]. Fan W, Boston BA, Kesterson RA, Hruby VJ, Cone RD. Role of melanocortinergic neurons in feeding and the agouti obesity syndrome. *Nature.* 1997; 385:165–168. [PubMed: 8990120]
- [36]. Biebermann H, Castaneda TR, van Landeghem F, von Deimling A, Escher F, Brabant G, Hebebrand J, Hinney A, Tschop MH, Gruters A, Krude H. A role for beta-melanocyte-stimulating hormone in human body-weight regulation. *Cell Metab.* 2006; 3:141–146. [PubMed: 16459315]
- [37]. Lee YS, Challis BG, Thompson DA, Yeo GS, Keogh JM, Madonna ME, Wraight V, Sims M, Vatin V, Meye D, Shield J, Burren C, Ibrahim Z, Cheetham T, Swift P, Blackwood A, Hung CC, Wareham NJ, Froguel P, Millhauser GL, O’Rahilly S, Farooqi IS. A POMC variant implicates beta-melanocyte-stimulating hormone in the control of human energy balance. *Cell Metab.* 2006; 3:135–140. [PubMed: 16459314]
- [38]. Adan RA, Cone RD, Burbach JP, Gispen WH. Differential effects of melanocortin peptides on neural melanocortin receptors. *Mol Pharmacol.* 1994; 46:1182–1190. [PubMed: 7808440]

- [39]. Mountjoy KG, Mortrud MT, Low MJ, Simerly RB, Cone RD. Localization of the melanocortin-4 receptor (MC4-R) in neuroendocrine and autonomic control circuits in the brain. *Mol Endocrinol.* 1994; 8:1298–1308. [PubMed: 7854347]
- [40]. Ollmann MM, Wilson BD, Yang YK, Kerns JA, Chen Y, Gantz I, Barsh GS. Antagonism of central melanocortin receptors in vitro and in vivo by agouti-related protein. *Science.* 1997; 278:135–138. [PubMed: 9311920]
- [41]. Stanley BG, Leibowitz SF. Neuropeptide Y: stimulation of feeding and drinking by injection into the paraventricular nucleus. *Life Sci.* 1984; 35:2635–2642. [PubMed: 6549039]
- [42]. Morley JE, Hernandez EN, Flood JF. Neuropeptide Y increases food intake in mice. *Am J Physiol.* 1987; 253:R516–522. [PubMed: 3631310]
- [43]. Sahu A, Kalra PS, Kalra SP. Food deprivation and ingestion induce reciprocal changes in neuropeptide Y concentrations in the paraventricular nucleus. *Peptides.* 1988; 9:83–86. [PubMed: 3362745]
- [44]. Sanacora G, Kershaw M, Finkelstein JA, White JD. Increased hypothalamic content of preproneuropeptide Y messenger ribonucleic acid in genetically obese Zucker rats and its regulation by food deprivation. *Endocrinology.* 1990; 127:730–737. [PubMed: 2373052]
- [45]. Wilding JP, Gilbey SG, Bailey CJ, Batt RA, Williams G, Ghatei MA, Bloom SR. Increased neuropeptide-Y messenger ribonucleic acid (mRNA) and decreased neurotensin mRNA in the hypothalamus of the obese (ob/ob) mouse. *Endocrinology.* 1993; 132:1939–1944. [PubMed: 7682936]
- [46]. Quillan JM, Sadee W, Wei ET, Jimenez C, Ji L, Chang JK. A synthetic human Agouti-related protein-(83-132)-NH2 fragment is a potent inhibitor of melanocortin receptor function. *FEBS Lett.* 1998; 428:59–62. [PubMed: 9645475]
- [47]. Rossi M, Kim MS, Morgan DG, Small CJ, Edwards CM, Sunter D, Abusnana S, Goldstone AP, Russell SH, Stanley SA, Smith DM, Yagaloff K, Ghatei MA, Bloom SR. A C-terminal fragment of Agouti-related protein increases feeding and antagonizes the effect of alpha-melanocyte stimulating hormone in vivo. *Endocrinology.* 1998; 139:4428–4431. [PubMed: 9751529]
- [48]. Tritos NA, Elmquist JK, Mastaitis JW, Flier JS, Maratos-Flier E. Characterization of expression of hypothalamic appetite-regulating peptides in obese hyperleptinemic brown adipose tissue-deficient (uncoupling protein-promoter-driven diphtheria toxin A) mice. *Endocrinology.* 1998; 139:4634–4641. [PubMed: 9794475]
- [49]. Cowley MA, Smart JL, Rubinstein M, Cerdan MG, Diano S, Horvath TL, Cone RD, Low MJ. Leptin activates anorexigenic POMC neurons through a neural network in the arcuate nucleus. *Nature.* 2001; 411:480–484. [PubMed: 11373681]
- [50]. Horvath TL, Naftolin F, Kalra SP, Leranath C. Neuropeptide-Y innervation of beta-endorphin-containing cells in the rat mediobasal hypothalamus: a light and electron microscopic double immunostaining analysis. *Endocrinology.* 1992; 131:2461–2467. [PubMed: 1425443]
- [51]. Bates SH, Myers MG Jr. The role of leptin receptor signaling in feeding and neuroendocrine function. *Trends Endocrinol Metab.* 2003; 14:447–452. [PubMed: 14643059]
- [52]. Sahm UG, Qarawi MA, Olivier GW, Ahmed AR, Branch SK, Moss SH, Pouton CW. The melanocortin (MC3) receptor from rat hypothalamus: photoaffinity labelling and binding of alanine-substituted alpha-MSH analogues. *FEBS Lett.* 1994; 350:29–32. [PubMed: 8062918]
- [53]. Cowley MA, Pronchuk N, Fan W, Dinulescu DM, Colmers WF, Cone RD. Integration of NPY, AGRP, and melanocortin signals in the hypothalamic paraventricular nucleus: evidence of a cellular basis for the adipostat. *Neuron.* 1999; 24:155–163. [PubMed: 10677034]
- [54]. Balthasar N, Dalgaard LT, Lee CE, Yu J, Funahashi H, Williams T, Ferreira M, Tang V, McGovern RA, Kenny CD, Christiansen LM, Edelstein E, Choi B, Boss O, Aschkenasi C, Zhang CY, Mountjoy K, Kishi T, Elmquist JK, Lowell BB. Divergence of melanocortin pathways in the control of food intake and energy expenditure. *Cell.* 2005; 123:493–505. [PubMed: 16269339]
- [55]. Gropp E, Shanabrough M, Borok E, Xu AW, Janoshek R, Buch T, Plum L, Balthasar N, Hampel B, Waisman A, Barsh GS, Horvath TL, Bruning J. Agouti-related peptide-expressing neurons are mandatory for feeding. *Nature Neuroscience.* 2005; 8:1289–91.
- [56]. Luquet S, Perez FA, Hnasko TS, Palmiter RD. NPY/AgRP neurons are essential for feeding in adult mice but can be ablated in neonates. *Science.* 2005; 310:683–5. [PubMed: 16254186]

- [57]. Erickson JC, Clegg KE, Palmiter RD. Sensitivity to leptin and susceptibility to seizures of mice lacking neuropeptide Y. *Nature*. 1996; 381:415–21. [PubMed: 8632796]
- [58]. Qian S, Chen H, Weingarth D, Trumbauer ME, Novi DE, Guan X, Yu H, Shen Z, Feng Y, Frazier E, Chen A, Camacho RE, Shearman LP, Gopal-Truter S, MacNeil DJ, Van der Ploeg LH, Marsh DJ. Neither agouti-related protein nor neuropeptide Y is critically required for the regulation of energy homeostasis in mice. *Mol Cell Biol*. 2002; 22:5027–35. [PubMed: 12077332]
- [59]. Hetherington AW, Ranson SW. The relation of various hypothalamic lesions to adiposity in the rat. *Journal of Comparative Neurology*. 1942; 76:475–499.
- [60]. Tejwani GA, Richard CW 3rd. Effect of electrolytic and chemical ventromedial hypothalamic lesions on food intake, body weight, analgesia and the CNS opioid peptides in rats and mice. *NIDA Res. Monogr*. 1986; 75:497–78. [PubMed: 2893279]
- [61]. Bagnasco M, Dube MG, Kalra PS, Kalra SP. Evidence for the existence of distinct central appetite, energy expenditure, and ghrelin stimulation pathways as revealed by hypothalamic site-specific leptin gene therapy. *Endocrinology*. 2002; 143:4409–78. [PubMed: 12399438]
- [62]. Mercer JG, Hoggard N, Williams LM, Lawrence CB, Hannah LT, Trayhurn P. Localization of leptin receptor mRNA and the long form splice variant (Ob-Rb) in mouse hypothalamus and adjacent brain regions by in situ hybridization. *FEBS Lett*. 1996; 387:113–116. [PubMed: 8674530]
- [63]. Fei H, Okano HJ, Li C, Lee GH, Zhao C, Darnell R, Friedman JM. Anatomic localization of alternatively spliced leptin receptors (Ob-R) in mouse brain and other tissues. *Proc Natl Acad Sci U S A*. 1997; 94(13):7001–5. [PubMed: 9192681]
- [64]. King BM. The rise, fall, and resurrection of the ventromedial hypothalamus in the regulation of feeding behavior and body weight. *Physiol Behav*. 2006; 87:221–244. [PubMed: 16412483]
- [65]. Balthasar N, Coppari R, McMinn J, Liu SM, Lee CE, Tang V, Kenny CD, McGovern RA, Chua SC Jr, Elmquist JK, Lowell BB. Leptin receptor signaling in POMC neurons is required for normal body weight homeostasis. *Neuron*. 2004; 42:983–991. [PubMed: 15207242]
- [66]. Majdic G, Young M, Gomez-Sanchez E, Anderson P, Szczepaniak LS, Dobbins RL, McGarry JD, Parker KL. Knockout mice lacking steroidogenic factor 1 are a novel genetic model of hypothalamic obesity. *Endocrinology*. 2002; 143:607–614. [PubMed: 11796516]
- [67]. Dhillon H, Zigman JM, Ye C, Lee CE, McGovern RA, Tang V, Kenny CD, Christiansen LM, White RD, Edelstein EA, Coppari R, Balthasar N, Cowley MA, Chua S Jr, Elmquist JK, Lowell BB. Leptin directly activates SF1 neurons in the VMH, and this action by leptin is required for normal body-weight homeostasis. *Neuron*. 2006; 49:191–203. [PubMed: 16423694]
- [68]. Parker KL, Rice DA, Lala DS, Ikeda Y, Luo X, Wong M, Bakke M, Zhao L, Frigeri C, Hanley NA, Stallings N, Schimmer BP. Steroidogenic factor 1: an essential mediator of endocrine development. *Recent Prog Horm Res*. 2002; 57:19–36. [PubMed: 12017543]
- [69]. Davis AM, Seney ML, Stallings NR, Zhao L, Parker KL, Tobet SA. Loss of steroidogenic factor 1 alters cellular topography in the mouse ventromedial nucleus of the hypothalamus. *J Neurobiol*. 2004; 60:424–436. [PubMed: 15307147]
- [70]. Segal JP, Stallings NR, Lee CE, Zhao L, Socci N, Viale A, Harris TM, Soares MB, Childs G, Elmquist JK, Parker KL, Friedman JM. Use of laser-capture microdissection for the identification of marker genes for the ventromedial hypothalamic nucleus. *J Neurosci*. 2005; 25:4181–4188. [PubMed: 15843621]
- [71]. Sternson SM, Shepherd GM, Friedman JM. Topographic mapping of VMH --> arcuate nucleus microcircuits and their reorganization by fasting. *Nat Neurosci*. 2005; 8:1356–1363. [PubMed: 16172601]
- [72]. Saper CB, Swanson LW, Cowan WM. The efferent connections of the ventromedial nucleus of the hypothalamus of the rat. *J Comp Neurol*. 1976; 169:409–442. [PubMed: 61975]
- [73]. Zaborszky L, Makara GB. Intrahypothalamic connections: an electron microscopic study in the rat. *Exp Brain Res*. 1979; 34:201–215. [PubMed: 105917]
- [74]. Canteras NS, Simerly RB, Swanson LW. Organization of projections from the ventromedial nucleus of the hypothalamus: a Phaseolus vulgaris-leucoagglutinin study in the rat. *J Comp Neurol*. 1994; 348:41–79. [PubMed: 7814684]

- [75]. Bouali SM, Fournier A, St-Pierre S, Jolicoeur FB. Effects of NPY and NPY2-36 on body temperature and food intake following administration into hypothalamic nuclei. *Brain Res Bull.* 1995; 36:131–135. [PubMed: 7895090]
- [76]. Lopez-Valpuesta FJ, Nyce JW, Griffin-Biggs TA, Ice JC, Myers RD. Antisense to NPY-Y1 demonstrates that Y1 receptors in the hypothalamus underlie NPY hypothermia and feeding in rats. *Proc Biol Sci.* 1996; 263:881–886. [PubMed: 8760491]
- [77]. Harrold JA, Widdowson PS, Williams G. Altered energy balance causes selective changes in melanocortin-4(MC4-R), but not melanocortin-3 (MC3-R), receptors in specific hypothalamic regions: further evidence that activation of MC4-R is a physiological inhibitor of feeding. *Diabetes.* 1999; 48:267–271. [PubMed: 10334300]
- [78]. Wisialowski T, Parker R, Preston E, Sainsbury A, Kraegen E, Herzog H, Cooney G. Adrenalectomy reduces neuropeptide Y-induced insulin release and NPY receptor expression in the rat ventromedial hypothalamus. *J Clin Invest.* 2000; 105:1253–1259. [PubMed: 10792000]
- [79]. Kishi T, Aschkenasi CJ, Lee CE, Mountjoy KG, Saper CB, Elmquist JK. Expression of melanocortin 4 receptor mRNA in the central nervous system of the rat. *J Comp Neurol.* 2003; 457:213–235. [PubMed: 12541307]
- [80]. Li YZ, Davidowa H. Food deprivation decreases responsiveness of ventromedial hypothalamic neurons to melanocortins. *J Neurosci Res.* 2004; 77:596–602. [PubMed: 15264229]
- [81]. Rios M, Fan G, Fekete C, Kelly J, Bates B, Kuehn R, Lechan RM, Jaenisch R. Conditional deletion of brain-derived neurotrophic factor in the postnatal brain leads to obesity and hyperactivity. *Mol Endocrinol.* 2001; 15(10):1748–57. [PubMed: 11579207]
- [82]. Xu B, Goulding EH, Zang K, Cepoi D, Cone RD, Jones KR, Tecott LH, Reichardt LF. Brain-derived neurotrophic factor regulates energy balance downstream of melanocortin-4 receptor. *Nat Neurosci.* 2003; 6:736–742. [PubMed: 12796784]
- [83]. Nakagawa T, Ono-Kishino M, Sugaru E, Yamanaka M, Taiji M, Noguchi H. Brain-derived neurotrophic factor (BDNF) regulates glucose and energy metabolism in diabetic mice. *Diabetes Metab Res Rev.* 2002; 18:185–191. [PubMed: 12112936]
- [84]. Tsuchida A, Nonomura T, Nakagawa T, Itakura Y, Ono-Kishino M, Yamanaka M, Sugaru E, Taiji M, Noguchi H. Brain-derived neurotrophic factor ameliorates lipid metabolism in diabetic mice. *Diabetes Obes Metab.* 2002; 4:262–269. [PubMed: 12099975]
- [85]. Nakagawa T, Ogawa Y, Ebihara K, Yamanaka M, Tsuchida A, Taiji M, Noguchi H, Nakao K. Anti-obesity and anti-diabetic effects of brain-derived neurotrophic factor in rodent models of leptin resistance. *Int J Obes Relat Metab Disord.* 2003; 27:557–565. [PubMed: 12704399]
- [86]. Komori T, Morikawa Y, Nanjo K, Senba E. Induction of brain-derived neurotrophic factor by leptin in the ventromedial hypothalamus. *Neuroscience.* 2006
- [87]. Pinto S, Roseberry AG, Liu H, Diano S, Shanabrough M, et al. Rapid rewiring of arcuate nucleus feeding circuits by leptin. *Science.* 2004; 304:110–78. [PubMed: 15064421]
- [88]. Gao Q, Mezei G, Nie Y, Rao Y, Choi CS, et al. Anorectic estrogen mimics leptin's effect on the rewiring of melanocortin cells and Stat3 signaling in obese animals. *Nat. Med.* 2007; 13:89–78. [PubMed: 17195839]
- [89]. Qu D, Ludwig DS, Gammeltoft S, Piper M, Pellemounter MA, Cullen MJ, Mathes WF, Przypek R, Kanarek R, Maratos-Flier E. A role for melanin-concentrating hormone in the central regulation of feeding behaviour. *Nature.* 1996; 380:243–247. [PubMed: 8637571]
- [90]. de Lecea L, Kilduff TS, Peyron C, Gao X, Foye PE, Danielson PE, Fukuhara C, Battenberg EL, Gautvik VT, Bartlett FS 2nd, Frankel WN, van den Pol AN, Bloom FE, Gautvik KM, Sutcliffe JG. The hypocretins: hypothalamus-specific peptides with neuroexcitatory activity. *Proc Natl Acad Sci U S A.* 1998; 95:322–327. [PubMed: 9419374]
- [91]. Elias CF, Saper CB, Maratos-Flier E, Tritos NA, Lee C, Kelly J, Tatro JB, Hoffman GE, Ollmann MM, Barsh GS, Sakurai T, Yanagisawa M, Elmquist JK. Chemically defined projections linking the mediobasal hypothalamus and the lateral hypothalamic area. *J Comp Neurol.* 1998; 402:442–459. [PubMed: 9862320]
- [92]. Ludwig DS, Mountjoy KG, Tatro JB, Gillette JA, Frederich RC, Flier JS, Maratos-Flier E. Melanin-concentrating hormone: a functional melanocortin antagonist in the hypothalamus. *Am J Physiol.* 1998; 274:E627–633. [PubMed: 9575823]

- [93]. Sakurai T, Amemiya A, Ishii M, Matsuzaki I, Chemelli RM, Tanaka H, Williams SC, Richardson JA, Kozlowski GP, Wilson S, Arch JR, Buckingham RE, Haynes AC, Carr SA, Annan RS, McNulty DE, Liu WS, Terrett JA, Elshourbagy NA, Bergsma DJ, Yanagisawa M. Orexins and orexin receptors: a family of hypothalamic neuropeptides and G protein-coupled receptors that regulate feeding behavior. *Cell*. 1998; 92:573–585. [PubMed: 9491897]
- [94]. Dube MG, Kalra SP, Kalra PS. Food intake elicited by central administration of orexins/hypocretins: identification of hypothalamic sites of action. *Brain Res*. 1999; 842:473–477. [PubMed: 10526145]
- [95]. Edwards CM, Abusnana S, Sunter D, Murphy KG, Ghatei MA, Bloom SR. The effect of the orexins on food intake: comparison with neuropeptide Y, melanin-concentrating hormone and galanin. *J Endocrinol*. 1999; 160:R7–12. [PubMed: 10077743]
- [96]. Horvath TL, Diano S, van den Pol AN. Synaptic interaction between hypocretin (orexin) and neuropeptide Y cells in the rodent and primate hypothalamus: a novel circuit implicated in metabolic and endocrine regulations. *J Neurosci*. 1999; 19:1072–1087. [PubMed: 9920670]
- [97]. Saito Y, Cheng M, Leslie FM, Civelli O. Expression of the melanin-concentrating hormone (MCH) receptor mRNA in the rat brain. *J Comp Neurol*. 2001; 435:26–40. [PubMed: 11370009]
- [98]. Tritos NA, Mastaitis JW, Kokkotou E, Maratos-Flier E. Characterization of melanin concentrating hormone and preproorexin expression in the murine hypothalamus. *Brain Res*. 2001; 895:160–166. [PubMed: 11259773]
- [99]. Guan JL, Uehara K, Lu S, Wang QP, Funahashi H, Sakurai T, Yanagisawa M, Shioda S. Reciprocal synaptic relationships between orexin- and melanin-concentrating hormone-containing neurons in the rat lateral hypothalamus: a novel circuit implicated in feeding regulation. *Int J Obes Relat Metab Disord*. 2002; 26:1523–1532. [PubMed: 12461668]
- [100]. Broberger C, De Lecea L, Sutcliffe JG, Hokfelt T. Hypocretin/orexin- and melanin-concentrating hormone-expressing cells form distinct populations in the rodent lateral hypothalamus: relationship to the neuropeptide Y and agouti gene-related protein systems. *J Comp Neurol*. 1998; 402:460–474. [PubMed: 9862321]
- [101]. Peyron C, Tighe DK, van den Pol AN, de Lecea L, Heller HC, Sutcliffe JG, Kilduff TS. Neurons containing hypocretin (orexin) project to multiple neuronal systems. *J Neurosci*. 1998; 18:9996–10015. [PubMed: 9822755]
- [102]. Lu XY, Bagnol D, Burke S, Akil H, Watson SJ. Differential distribution and regulation of OX1 and OX2 orexin/hypocretin receptor messenger RNA in the brain upon fasting. *Horm Behav*. 2000; 37:335–344. [PubMed: 10860677]
- [103]. Hakansson ML, Brown H, Ghilardi N, Skoda RC, Meister B. Leptin receptor immunoreactivity in chemically defined target neurons of the hypothalamus. *J Neurosci*. 1998; 18:559–572. [PubMed: 9412531]
- [104]. Torrealba F, Yanagisawa M, Saper CB. Colocalization of orexin and glutamate immunoreactivity in axon terminals in the tuberomammillary nucleus in rats. *Neuroscience*. 2003; 119:1033–1044. [PubMed: 12831862]
- [105]. Toshinai K, Date Y, Murakami N, Shimada M, Mondal MS, Shimbara T, Guan JL, Wang QP, Funahashi H, Sakurai T, Shioda S, Matsukura S, Kangawa K, Nakazato M. Ghrelin-induced food intake is mediated via the orexin pathway. *Endocrinology*. 2003; 144:1506–1512. [PubMed: 12639935]
- [106]. Sakurai T, Nagata R, Yamanaka A, Kawamura H, Tsujino N, Muraki Y, Kageyama H, Kunita S, Takahashi S, Goto K, Koyama Y, Shioda S, Yanagisawa M. Input of orexin/hypocretin neurons revealed by a genetically encoded tracer in mice. *Neuron*. 2005; 46:297–308. [PubMed: 15848807]
- [107]. van den Pol AN, Acuna-Goycolea C, Clark KR, Ghosh PK. Physiological properties of hypothalamic MCH neurons identified with selective expression of reporter gene after recombinant virus infection. *Neuron*. 2004; 42:635–78. [PubMed: 15157424]
- [108]. Li Y, Gao XB, Sakurai T, van den Pol AN. Hypocretin/orexin excites hypocretin neurons via a local glutamate neuron-A potential mechanism for orchestrating the hypothalamic arousal system. *Neuron*. 2002; 36:1169–78. [PubMed: 12495630]



- [109]. Gao XB, van den Pol AN. Melanin concentrating hormone depresses synaptic activity of glutamate and GABA neurons from rat lateral hypothalamus. *J. Physiol.* 2001; 533:237–78. [PubMed: 11351031]
- [110]. Diano S, Horvath B, Urbanski HF, Sotonyi P, Horvath TL. Fasting activates the nonhuman primate hypocretin (orexin) system and its postsynaptic targets. *Endocrinology.* 2003; 144:3774–78. [PubMed: 12933647]
- [111]. Horvath TL, Gao XB. Inpuro organization and plasticityof hypocretin neurons: Possible clues for obesity's association with insomnia. *Cell Metabolism.* 2005; 1:279–286. [PubMed: 16054072]
- [112]. Horvath TL, Peyron C, Diano S, Ivanov A, Aston-Jones G, Kilduff TS, van den Pol AN. Hypocretin (orexin) activation and synaptic innervation of the locus coeruleus noradrenergic system. *J Comp Neurol.* 1999; 415:145–159. [PubMed: 10545156]
- [113]. Jain MR, Horvath TL, Kalra PS, Kalra SP. Evidence that NPY Y1 receptors are involved in stimulation of feeding by orexins (hypocretins) in sated rats. *Regulatory Peptides.* 1999; 87:19–24. [PubMed: 10710284]
- [114]. Yamanaka A, Kunii K, Nambu T, Tsujino N, Sakai A, et al. Orexin-induced food intake involves neuropeptide Y pathway. *Brain Res.* 2000; 859:404–78. [PubMed: 10719096]
- [115]. Sahu A. Interactions of neuropeptide Y, hypocretin-I (orexin A) and melanin-concentrating hormone on feeding in rats. *Brain Res.* 2002; 944:232–8. [PubMed: 12106685]
- [116]. Willie JT, Chemelli RM, Sinton CM, Yanagisawa M. To eat or to sleep? Orexin in the regulation of feeding and wakefulness. *Annu. Rev. Neurosci.* 2001; 24:429–78. [PubMed: 11283317]
- [117]. Chemelli RM, Willie JT, Sinton CM, Elmquist JK, Scammell T, et al. Narcolepsy in orexin knockout mice: molecular genetics of sleep regulation. *Cell.* 1999; 98:437–78. [PubMed: 10481909]
- [118]. Shimada M, Tritos NA, Lowell BB, Flier JS, Maratos-Flier E. Mice lacking melanin-concentrating hormone are hypophagic and lean. *Nature.* 1998; 396:670–78. [PubMed: 9872314]
- [119]. Kennedy AR, Todd JF, Stanley SA, Abbott CR, Small CJ, et al. Melanin-concentrating hormone (MCH) suppresses thyroid stimulating hormone (TSH) release, in vivo and in vitro, via the hypothalamus and the pituitary. *Endocrinology.* 2002; 142:3265–78. [PubMed: 11416052]
- [120]. Alon T, Friedman JM. Late-onset leanness in mice with targeted ablation of melanin concentrating hormone neurons. *J. Neurosci.* 2006; 26:389–78. [PubMed: 16407534]
- [121]. Zigman JM, Jones JE, Lee CE, Saper CB, Elmquist JK. Expression of ghrelin receptor mRNA in the rat and the mouse brain. *J Comp Neurol.* 2006; 494:528–548. [PubMed: 16320257]
- [122]. Bellinger LL, Bernardis LL. The dorsomedial hypothalamic nucleus and its role in ingestive behavior and body weight regulation: lessons learned from lesioning studies. *Physiol Behav.* 2002; 76:431–442. [PubMed: 12117580]
- [123]. Chou TC, Scammell TE, Gooley JJ, Gaus SE, Saper CB, Lu J. Critical role of dorsomedial hypothalamic nucleus in a wide range of behavioral circadian rhythms. *J Neurosci.* 2003; 23:10691–10702. [PubMed: 14627654]
- [124]. Gooley JJ, Schomer A, Saper CB. The dorsomedial hypothalamic nucleus is critical for the expression of food-entrainable circadian rhythms. *Nat Neurosci.* 2006; 9:398–407. [PubMed: 16491082]
- [125]. ter Horst GJ, Luiten PG. The projections of the dorsomedial hypothalamic nucleus in the rat. *Brain Res Bull.* 1986; 16:231–248. [PubMed: 3697791]
- [126]. Chou TC, Bjorkum AA, Gaus SE, Lu J, Scammell TE, Saper CB. Afferents to the ventrolateral preoptic nucleus. *J Neurosci.* 2002; 22:977–990. [PubMed: 11826126]
- [127]. Morin LP, Allen CN. The circadian visual system. *Brain Res Brain Res Rev.* 2005
- [128]. Buijs RM, van Eden CG, Goncharuk VD, Kalsbeek A. The biological clock tunes the organs of the body: timing by hormones and the autonomic nervous system. *J Endocrinol.* 2003; 177:17–26. [PubMed: 12697033]
- [129]. Sumova A, Bendova Z, Sladek M, Kovacicova Z, Illnerova H. Seasonal molecular timekeeping within the rat circadian clock. *Physiol Res.* 2004; 1(53 Suppl.):S167–176. [PubMed: 15119947]

- [130]. Yi CX, van der Vliet J, Dai J, Yin G, Ru L, Buijs RM. Ventromedial arcuate nucleus communicates peripheral metabolic information to the suprachiasmatic nucleus. *Endocrinology*. 2006; 147:283–294. [PubMed: 16195398]
- [131]. Turek FW, Joshu C, Kohsaka A, Lin E, Ivanova G, McDearmon E, Laposky A, Losee-Olson S, Easton A, Jensen DR, Eckel RH, Takahashi JS, Bass J. Obesity and metabolic syndrome in circadian Clock mutant mice. *Science*. 2005; 308:1043–1045. [PubMed: 15845877]
- [132]. Hastings MH, Reddy AB, Garabette M, King VM, Chahad-Ehlers S, O'Brien J, Maywood ES. Expression of clock gene products in the suprachiasmatic nucleus in relation to circadian behaviour. *Novartis Found Symp*. 2003; 253:203–217. discussion 102-209, 218-222, 281204. [PubMed: 14712923]
- [133]. Maywood ES, O'Brien JA, Hastings MH. Expression of mCLOCK and other circadian clock-relevant proteins in the mouse suprachiasmatic nuclei. *J Neuroendocrinol*. 2003; 15:329–334. [PubMed: 12622829]
- [134]. Wise RA. Brain reward circuitry: insights from unsensed incentives. *Neuron*. 2002; 36:229–240. [PubMed: 12383779]
- [135]. Szczyepka MS, Mandel RJ, Donahue BA, Snyder RO, Leff SE, Palmiter RD. Viral gene delivery selectively restores feeding and prevents lethality of dopamine-deficient mice. *Neuron*. 1999; 22:167–178. [PubMed: 10027299]
- [136]. Szczyepka MS, Rainey MA, Kim DS, Alaynick WA, Marck BT, Matsumoto AM, Palmiter RD. Feeding behavior in dopamine-deficient mice. *Proc Natl Acad Sci U S A*. 1999; 96:12138–12143. [PubMed: 10518589]
- [137]. Hnasko TS, Szczyepka MS, Alaynick WA, Daring MJ, Palmiter RD. A role for dopamine in feeding responses produced by orexigenic agents. *Brain Res*. 2004; 1023:309–78. [PubMed: 15374756]
- [138]. Szczyepka MS, Rainey MA, Palmiter RD. Dopamine is required for hyperphagia in *Lep(ob/ob)* mice. *Nat. Genet*. 2000; 25:102–78. [PubMed: 10802666]
- [139]. Volkow ND, Wise RA. How can drug addiction help us understand obesity? *Nat Neurosci*. 2005; 8:555–560. [PubMed: 15856062]
- [140]. Abizaid A, Liu ZW, Andrews ZB, Shanabrough M, Borok E, et al. Ghrelin modulates the activity and synaptic input organization of midbrain dopamine neurons while promoting appetite. *J. Clin. Invest*. 2006; 116:3229–78. [PubMed: 17060947]
- [141]. Szczyepka MS, Kwok K, Brot MD, Marck BT, Matsumoto AM, et al. Dopamine production in the caudate putamen restores feeding in dopamine-deficient mice. *Neuron*. 2001; 30:819–78. [PubMed: 11430814]
- [142]. Small DM, Jones-Gotman M, Dagher A. Feeding-induced dopamine release in dorsal striatum correlates with meal pleasantness ratings in healthy human volunteers. *Neuroimage*. 2003; 19:1709–78. [PubMed: 12948725]
- [143]. Cannon CM, Palmiter RD. Reward without dopamine. *J. Neurosci*. 2003; 23:10827–78. [PubMed: 14645475]
- [144]. Sotak BN, Hnasko TS, Robinson S, Kremer EJ, Palmiter RD. Dysregulation of dopamine signaling in the dorsal striatum inhibits feeding. *Brain Res*. 2005; 1061:88–78. [PubMed: 16226228]
- [145]. Berridge KC. Food reward: brain substrates of wanting and liking. *Neurosci Biobehav Rev*. 1996; 20:1–25. [PubMed: 8622814]
- [146]. Berthoud HR. Multiple neural systems controlling food intake and body weight. *Neurosci Biobehav Rev*. 2002; 26:393–428. [PubMed: 12204189]
- [147]. Guan XM, Yu H, Palyha OC, McKee KK, Feighner SD, Sirinathsinghji DJ, Smith RG, Van der Ploeg LH, Howard AD. Distribution of mRNA encoding the growth hormone secretagogue receptor in brain and peripheral tissues. *Brain Res Mol Brain Res*. 1997; 48:23–29. [PubMed: 9379845]
- [148]. Figlewicz DP. Adiposity signals and food reward: expanding the CNS roles of insulin and leptin. *Am J Physiol Regul Integr Comp Physiol*. 2003; 284:R882–892. [PubMed: 12626355]
- [149]. Fulton S, Woodside B, Shizgal P. Modulation of brain reward circuitry by leptin. *Science*. 2000; 287:125–128. [PubMed: 10615045]

- [150]. Berthoud, HR. The caudal brainstem and the control of food intake and energy balance. In: Stricker, EM.; Woods, SC., editors. Handbook of behavioral neurobiology. Plenum; New York: 2004.
- [151]. Grill HJ, Kaplan JM. The neuroanatomical axis for control of energy balance. *Front Neuroendocrinol.* 2002; 23:2–40. [PubMed: 11906202]
- [152]. Grill HJ, Kaplan JM. Interoceptive and integrative contributions of forebrain and brainstem to energy balance control. *Int J Obes Relat Metab Disord.* 2001; 5(25 Suppl.):S73–77. [PubMed: 11840220]
- [153]. Travers SP, Norgren R. Coding the sweet taste in the nucleus of the solitary tract: differential roles for anterior tongue and nasoincisor duct gustatory receptors in the rat. *J. Neurophysiol.* 1991; 65:1372–78. [PubMed: 1875246]
- [154]. Grill HJ, Norgren R. Chronically decerebrate rats demonstrate satiation but not bait shyness. *Science.* 1978; 201:267–269. [PubMed: 663655]
- [155]. Silver AJ, Flood JF, Song AM, Morley JE. Evidence for a physiological role for CCK in the regulation of food intake in mice. *Am. J. Physiol.* 1989; 256:646–78.
- [156]. Smith GP, Jerome C, Norgren R. Afferent axons in abdominal vagus mediate satiety effect of cholecystokinin in rats. *Am. J. Physiol.* 1985; 249:638–641.
- [157]. DiRocco RJ, Grill HJ. The forebrain is not essential for sympathoadrenal hyperglycemic response to glucoprivation. *Science.* 1979; 204:1112–1114. [PubMed: 451558]
- [158]. Seeley RJ, Grill HJ, Kaplan JM. Neurological dissociation of gastrointestinal and metabolic contributions to meal size control. *Behav. Neurosci.* 1994; 108:347–352. [PubMed: 8037879]
- [159]. Grill HJ, Schwartz MW, Kaplan JM, Foxhall JS, Breninger J, Baskin DG. Evidence that the caudal brainstem is a target for the inhibitory effect of leptin on food intake. *Endocrinology.* 2002; 143(1):239–46. [PubMed: 11751615]
- [160]. Jacob RJ, Dziura J, Medwick MB, Leone P, Caprio S, During M, Shulman GI, Shvrwin RS. The effect of leptin is enhanced by microinjection into the ventromedial hypothalamus. *Diabetes.* 1997; 46:150–2. [PubMed: 8971096]
- [161]. Bailey AR, Von Englehardt N, Leng G, Smith RG, Dickson SL. Growth hormone secretagogue activation of the arcuate nucleus and brainstem occurs via a non-noradrenergic pathway. *J Neuroendocrinol.* 2000; 12:191–197. [PubMed: 10718914]
- [162]. Lawrence CB, Snape AC, Baudoin FM, Luckman SM. Acute central ghrelin and GH secretagogues induce feeding and activate brain appetite centers. *Endocrinology.* 2002; 143:155–62. [PubMed: 11751604]
- [163]. Faulconbridge LF, Cummings DE, Kaplan JM, Grill HJ. Hyperphagic effects of brainstem ghrelin administration. *Diabetes.* 2003; 52:2260–5. [PubMed: 12941764]
- [164]. Unger JW, Moss AM, Livingston JN. Immunohistochemical localization of insulin receptors and phosphotyrosine in the brainstem of the adult rat. *Neuroscience.* 1991; 42(3):853–61. [PubMed: 1720228]
- [165]. Grill HJ, Markison S, Ginsberg A, Kaplan JM. Long-term effects on feeding and body weight after stimulation of forebrain or hindbrain CRH receptors with urocortin. *Brain Res.* 2000; 867:19–28. [PubMed: 10837794]
- [166]. Williams DL, Kaplan JM, Grill HJ. The role of the dorsal vagal complex and the vagus nerve in feeding effects of melanocortin-3/4 receptor stimulation. *Endocrinology.* 2000; 141:1332–7. [PubMed: 10746636]
- [167]. Grill HJ, Ginsberg AB, Seeley RJ, Kaplan JM. Brainstem application of melanocortin receptor ligands produces long-lasting effects on feeding and body weight. *J Neurosci.* 1998; 18(23):10128–35. [PubMed: 9822766]
- [168]. Zheng H, Patterson LM, Morrison C, Banfield BW, Randall JA, Browning KN, Travagli RA, Berthoud HR. Melanin concentrating hormone innervation of caudal brainstem areas involved in gastrointestinal functions and energy balance. *Neuroscience.* 2005; 135:611–25. [PubMed: 16111819]
- [169]. Ritter RC, Slusser PG, Stone S. Glucoreceptors controlling feeding and blood glucose: location in the hindbrain. *Science.* 1981; 213:451–3. [PubMed: 6264602]