



Published in final edited form as:

Pediatr Diabetes. 2014 August ; 15(5): 355–362.

Improving Prediction of Type 1 Diabetes by testing Non-HLA Genetic Variants in addition to HLA Markers

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Abstract

Objective—The purpose of this study was to explore whether non-HLA genetic markers can improve type 1 diabetes (T1D) prediction in a prospective cohort with high-risk HLA-DR,DQ genotypes.

Methods—The Diabetes Autoimmunity Study in the Young (DAISY) follows prospectively for development of T1D and islet autoimmunity (IA) children at increased genetic risk. A total of 1709 non-Hispanic White DAISY participants have been genotyped for 27 non-HLA single nucleotide polymorphisms and one microsatellite.

Results—In multivariate analyses adjusting for family history and HLA-DR3/4 genotype, *PTPN22* (rs2476601) and two *UBASH3A* (rs11203203 and rs9976767) SNPs were associated with development of IA (HR=1.87, 1.55 and 1.54 respectively, all $p < 0.003$), while *GLIS3* and *IL2RA* showed borderline association with development of IA. *INS*, *UBASH3A* and *IFIH1* were significantly associated with progression from IA to diabetes (HR=1.65, 1.44 and 1.47 respectively, all $p < 0.04$), while *PTPN22* and *IL27* showed borderline association with progression from IA to diabetes. In survival analysis, 45% of general population DAISY children with *PTPN22* rs2476601 TT or HLA-DR3/4 and *UBASH3A* rs11203203 AA developed diabetes by age 15, compared to 3% of children with all other genotypes ($p < 0.0001$). Addition of non-HLA markers to HLA-DR3/4,DQ8 did not improve diabetes prediction in first-degree relatives.

Conclusion—Addition of *PTPN22* and *UBASH3A* SNPs to HLA-DR,DQ genotyping can improve T1D risk prediction.

Keywords

Type 1 diabetes; islet autoimmunity; non-HLA genetic markers; prediction

Introduction

The HLA region on chromosome 6p21 is considered the major susceptibility locus for type 1 diabetes (odds ratio > 6) with an estimated 30-50% of the genetic risk for diabetes attributed to this region ¹. With the advent of genome wide association studies (GWAS), more than 50 non-HLA susceptibility gene markers have been associated with type 1 diabetes ²⁻⁵. A majority of these loci appear to have effects in the immune system. *INS* ⁶ and *PTPN22* ⁷ show the strongest association (odds ratio ~2), notably weaker compared to the HLA region.

Class II HLA genotypes in combination with islet autoantibodies can predict diabetes risk in first-degree relatives (FDR) of persons with type 1 diabetes. We have previously published on the association of *INS*, *PTPN22* and *UBASH3A* with islet autoimmunity (IA) and type 1 diabetes in the Diabetes Autoimmunity Study in the Young (DAISY) ^{8, 9}. In an article published in *Pediatric Diabetes* in 2012 ⁹, we reported on the association of *UBASH3A* with both IA and type 1 diabetes with a cumulative risk for diabetes of 22% by age 10 for those general population DAISY children having *UBASH3A* AA genotype with HLA-DR3/4,DQB1*0302. In this study, we have genotyped an additional 8 non-HLA single nucleotide polymorphisms (SNPs) in 7 genes (*ERBB3* (rs2292239), *CLEC16A* (rs12708716), *IL27* (rs4788084), *CTRB* (rs7202877), *C14orf* (rs4900384), *GSDM* (rs2290400), *HORMAD2* (rs5753037), *UBASH3A* (rs9976767)), and further explored the independent predictive value of novel non-HLA markers on the risk of IA and progression from IA to diabetes, controlling for the effects of HLA-DR,DQ genotypes. We have also developed a genetic risk model, adding non-HLA markers (*UBASH3A* AA, *PTPN22* TT) to high risk HLA-DR3/4 in order to refine diabetes risk prediction, and report a risk of diabetes by age 15 years of 45% for those DAISY general population children in the high risk genetic stratum. Finally, we tested a set of SNPs, previously found to significantly discriminate diabetes in the BABYDIAB cohort ¹⁰, in the DAISY study.

Methods

Study population

Since 1993, DAISY has followed two cohorts of young children at increased risk of type 1 diabetes: FDR of type 1 diabetes patients and general population children found through a newborn screening to carry high-risk HLA-DR,DQ genotypes. The details of screening and follow-up have been previously published ¹¹. Briefly, 31,881 newborns from the general population of Denver, Colorado have been screened for HLA-DR,DQ genotypes that carry susceptibility to type 1 diabetes. All children with DR3/4,DQB1*0302, DR3/3 and DR4/4,DQB1*0302 and a sample of those with DR4/DRx, DQB1*0302 or DR3/DRx (where DRx = DR3 or DR4) were invited to participate in DAISY. Although general population children were included in DAISY only if they had the above susceptibility HLA genotypes, non-diabetic offspring and siblings of patients with type 1 diabetes were invited to participate regardless of their HLA genotype. A total of 1709 non-Hispanic White (NHW) participants (858 general population children and 851 FDR children, including 477 multiple siblings) were genotyped for 27 non-HLA single nucleotide polymorphisms and one

microsatellite. Of those, 116 developed persistent IA and 66 of these progressed to diabetes during the 10-year mean prospective follow-up. Informed consent was obtained from the parents of each study subject. The Colorado Multiple Institutional Review Board approved all study protocols.

Islet Autoantibodies

Measurement of islet autoantibodies to insulin, GAD65, IA-2 and ZnT8 was performed in the Clinical Immunology Laboratory at the Barbara Davis Center using previously described radio-immunoassays¹². IA was defined as presence of one or more of the autoantibodies to insulin, GAD65, IA-2 or ZnT8 on at least 2 consecutive visits 3-12 months apart, and still positive at last visit.

Genotyping

INS-23Hph1 (rs689), *CTLA-4* T17A (rs231775), and *PTPN22* R620W (rs2476601) polymorphisms were genotyped using a linear array (immobilized probe) method essentially as described in Mirel et al.¹³. The following SNPs were genotyped in the laboratory of Dr. Cisca Wijmenga using Illumina GoldenGate Beadexpress assays (veracode 48-plex): *IL2RA* (rs12251307), *SH2B3* (rs3184504), *PTPN2* (rs1893217), *C10orf59* (rs10509540), *IL18RAP* (rs917997), *BACH2* (rs11755527) and *TAGAP* (rs1738074).

Taqman SNP genotyping assays (Applied Biosystems, CA USA) were utilized to obtain genotype information on the following SNPs as described previously⁸: *CD69* (rs4763879), *GAB3* (rs2664170), *GLIS3* (rs7020673), *IL10* (rs3024496), *SIRPG* (rs2281808), *PRKD2* (rs425105), *UBASH3A* (rs11203203), *IFIH1* (rs1990760) and *SLC30A8* (rs13266634).

The following SNPs were genotyped by utilizing the Taqman SNP genotype based OpenArray platform [Applied Biosystems, CA USA]: *ERBB3* (rs2292239), *CLEC16A* (rs12708716), *IL27* (rs4788084), *CTRB* (rs7202877), *C14orf* (rs4900384), *GSDM* (rs2290400), *HORMAD2* (rs5753037), *UBASH3A* (rs9976767). Custom designed arrays were loaded using the OpenArray AccuFill system and cycling was performed on a GeneAmp 9700 PCR system, all gDNA template load and run parameters according to manufacturer protocol. Genotypes were analyzed using the OpenArray SNP genotyping analysis software v.1.0.3 and Taqman Genotyper Software 2.0.

CCR5 genotypes were determined using a fluorescent-based method. PCR fragments were generated using primers that differentiate between the wild type genotype (CCR5/CCR5) at 225bp and the homozygous mutant (32/32) at 193bp. Reactions (25 µl) were assembled using FailSafe PCR PreMix J, 2.5 U MasterAmp Taq polymerase (Epicentre), 10 nmol each primer and 100 ng of genomic template. The PCR product was amplified via 35 PCR cycles of 94°C for 30 sec, 57°C for 35 sec, 72°C for 1 min, and a final extension of 72°C for 45 min. Products were diluted 1:60 and separated by capillary electrophoresis on an ABI3100-Avant Genetic Analyzer (Applied Biosystems). Alleles were identified using GeneMapper v3.5 (Applied Biosystems).

Statistical analysis

Analyses were performed in SAS version 9.2 and PRISM software. Cox proportional hazard models were used to test the effect of each genetic polymorphism on time to development of IA and progression from IA to diabetes. Multivariate model with Weibull distribution (outcome: IA) and Cox PH model (outcome: diabetes) included family history of diabetes (yes/no) and the presence of the HLA-DR3/4-DQB1*0302 genotype (yes/no); independently significant non-HLA polymorphisms were identified by backward selection at a critical level of 0.05. Each SNP in the model was defined according to the number of risk allele present (0, 1 or 2) and was treated as a continuous variable in the model. In the Cox regression model, we checked proportionality by including time-dependent covariates and all *p* values were non-significant. Since our analyses were based on a priori hypotheses, *P* values were not corrected for multiple testing. We performed survival analysis of progression to IA and diabetes with PRISM software, using the log-rank test and an alpha level for significance set at 0.05. Only one of the two *UBASH3A* SNPs (rs11203203) was included in the model since both *UBASH3A* SNPs are in linkage disequilibrium ($D'=1.0$, $r^2=0.63$). Based on initial significant results, survival analyses were stratified by high and low genetic risk groups. High risk genetic group included all subjects with *UBASH3A* AA in addition to HLA-DR3/4 as well as all subjects with *PTPN22* TT (whether they were HLA-DR3/4 or not, since this group is small), while low risk genetic group included all other genotypes. Finally, survival analyses including nine out of the 12 genes recently tested in a model by Winkler et al ¹⁰ were performed. Receiver operator curve (ROC) analysis was performed with area under the curve (AUC) calculated for the 9 gene variants. The SNPs were in Hardy-Weinberg equilibrium except for *C10orf59*, *PRKD2* and *GAB3*, which were therefore excluded from the multivariate and survival analyses. To determine whether inclusion of multiple siblings per family in this cohort affected our findings, we performed analyses accounting for the clustering of patients within a family (using the robust sandwich estimate for statistical inference) and found similar results (data not shown).

Results

In multivariate analyses adjusting for family history and HLA-DR3/4 high-risk genotype, *PTPN22* (rs2476601) and two *UBASH3A* (rs11203203 and rs9976767) SNPs were associated with development of IA (HR=1.87, 1.55 and 1.54 respectively, all *p* 0.003), while *GLIS3* and *IL2RA* showed borderline association with development of IA (Table 1). On the other hand, *INS*, *UBASH3A* and *IFIH1* were significantly associated with progression from IA to diabetes (HR=1.65, 1.44 and 1.47 respectively, all *p* 0.04), while *PTPN22* and *IL27* showed borderline association with progression from IA to diabetes. Some of these SNPs might be close to reach statistical significance due to smaller numbers, especially in the group looking at progression to diabetes. The other non-HLA markers tested did not predict development of IA or diabetes. There were no significant interactions between any of the SNPs and HLA-DR3/4-DQB1*0302.

Backward multivariate regression analyses including all SNPs were performed for both development of IA and progression from IA to type 1 diabetes (Table 2). SNPs that remained significantly associated with IA, adjusting for family history and HLA-DR3/4-

DQB1*0302, included *PTPN22*, *UBASH3A*, *INS* and *GLIS*, while the final model for progression from IA to diabetes only included *UBASH3A*.

Based on the results of backward multivariate regression analyses, we performed survival analyses with those significant variables (HLA-DR3/4-DQB1*0302, *PTPN22*, *UBASH3A*, *INS* and *GLIS*) in order to refine diabetes risk prediction. There was no further improvement in prediction by including *INS* or *GLIS*, so the final high-risk stratum includes all subjects with *UBASH3A* AA in addition to HLA-DR3/4 as well as all subjects with *PTPN22* TT (whether they were HLA-DR3/4 or not, since this group is small), while the low risk group has all other genotypes. Cumulative incidence of development of IA showed a higher risk of IA by age 15 years for the high (26%) compared to the low risk group (5%) in the general population (N=843) (Figure 1A). Risk of diabetes by age 15 years was also higher in subjects with high (45%) compared to those with low risk (3%) ($p<0.0001$) (Figure 1B). In comparison, survival analysis stratified by HLA-DR3/4,DQB1*0302 showed a risk of persistent IA and diabetes by age 15 years for HLA-DR3/4 of “only” 12% and 15% respectively (Figure 1C and 1D). In the DAISY general population, the positive predictive value for diabetes is slightly better with this genetic risk stratum than HLA-DR3/4 alone (17.4 vs. 6.4) for similar negative predictive value (98.6 vs. 99.2), while sensitivity was lower (42 vs. 75%) and specificity was better (95 vs. 74%) compared to HLA-DR3/4 alone.

Addition of non-HLA markers to HLA-DR3/4,DQ8 did not improve diabetes prediction in DAISY FDR (Figure 2). The cumulative risk of IA among FDR reached 51% in the high risk group by age 15 (Figure 2A), while the cumulative risk for diabetes was 29% (Figure 2B). Cumulative incidence of IA and diabetes by age 15 years showed similar risk for FDR with HLA-DR3/4 only (39% and 35% respectively) (Figure 2C and 2D).

Nine (*ERBB3*, *PTPN2*, *IFIH1*, *PTPN22*, *KIAA0350/CLEC16A*, *CTLA4*, *SH2B3*, *IL18RAP*, *IL10*) out of the 12 genes recently tested in a model by Winkler et al.¹⁰ have been genotyped in DAISY. For all 9 gene SNPs, a score of 2 was given if the child was homozygous for the susceptible allele, 1 if heterozygous and 0 if homozygous for the non-susceptible allele. The sum of the scores for the 9 genes was assigned as the combined risk score for each child. Although the distribution of the combined risk scores did not reach statistical significance between children who developed diabetes compared to autoantibody negative children, threshold points could be observed at SNP-risk allele score of <5 and >9 , which were used to define low (<5), intermediate (6-9) and high (>9) risk categories (Figure 3). These thresholds showed a similar stratification of diabetes risk in DAISY than in BABYDIAB, although the survival analyses only trended towards significance in the general population ($p=0.06$), likely due to smaller numbers (Figure 4). Receiver operator curve (ROC) analysis was performed, but area under the curve (AUC) was not statistically significant (AUC 0.55, 95% CI 0.45-0.65).

Discussion

High-density SNP analysis, GWAS and follow-up meta-analyses have added to the list of non-HLA loci associated with type 1 diabetes (more than 50 to date)^{2-4, 14}. Still the strongest signals by far are associated with the HLA region ($OR>6$) with the next highest

non-HLA signals in *INS* and *PTPN22*¹⁵. The prospective DAISY study has now genotyped 28 previously confirmed non-HLA loci to test the robustness of these associations with the advantage of evaluating the effect of candidate SNPs on the prospectively observed development of diabetes phenotypes (development of IA and progression from IA to diabetes). In addition to the non-HLA genes most strongly associated with type 1 diabetes in previous studies (*PTPN22* and *INS*)^{16,17}, we recently found a robust association of IA and diabetes for *UBASH3A*⁹. While *PTPN22*, *INS* and *UBASH3A* seem to be the main non-HLA risk factors in the DAISY cohort, some SNPs might not reach statistical significance due to smaller numbers, especially in the group looking at progression to diabetes.

This is the first study to describe a genetic risk stratum for diabetes in a prospective cohort following general population children screened at birth for high-risk HLA-DR,DQ genotypes. The risk definition includes HLA class II, *PTPN22* and *UBASH3A*. This genetic risk stratum significantly improves prediction of type 1 diabetes in DAISY general population children with a risk of diabetes by age 15 years of 45% for those subjects in the high risk compared to 3% for those in the low risk stratum. If confirmed in another population, these prediction models could be used for screening high-risk general population children into potential clinical research trials. We have previously published on two SNPs (rs2040410 and rs7454108) that are 98.6% sensitive and 99.7% specific for HLA DR3/4-DQ8¹⁸. A new genetic stratum including a total of 4 SNPs (rs2476601, rs11203203, rs2040410 and rs7454108) could potentially be applicable for screening of type 1 diabetes risk, followed by diabetes antibody testing in those subjects found at high genetic risk. Although the negative predictive value is good, the positive predictive value remains low. These results should be confirmed in additional cohorts with long-term follow-up such as The Environmental Determinants of Diabetes in the Young (TEDDY) and ideally in a general population without HLA susceptibility genotypes for type 1 diabetes.

Winkler et al recently published on improved prediction of diabetes by including 12 non-HLA risk genes in children with high-risk HLA genotypes¹⁰. Stratified survival analyses showed risk ranging from 0% by age 14 years for children in the low-risk category to 7.1% for children in the high-risk category. Interestingly, our risk score distribution (Figure 3) seems to be shifted to the left compared to the paper by Winkler et al., which might be due to population stratification (although we limited our study to non-Hispanic White subjects) or to higher population frequencies of the three SNPs (*CD25* rs11594656, *IL2* rs4505848, *COBL* rs4948088) not included in our study. One of the strengths and similarities for these two studies is that they are prospective studies in which timing of IA and diabetes onset are closely monitored and time to event analyses are possible. However, there are several important differences between the BABYDIAB article and our study. First, BABYDIAB population only includes first-degree relatives. Second, the number and type of SNPs included were different and HLA high-risk genotypes were defined according to the TEDDY criteria in BABYDIAB¹⁹, while for the DAISY study, only high-risk HLA DR3/4-DQ8 genotype was considered as a categorical variable.

Another study looked at the joint effects of HLA, *INS*, *PTPN22* and *CTLA4* genes and found that multiple susceptibility loci confer a very high risk of diabetes, but only a small proportion of the population carries all high risk alleles²⁰. When assessing the predictive

utility of these genetic risk markers by ROC curve, multiple susceptibility genotypes seemed to improve disease prediction only marginally compared to HLA genotype alone²⁰. ROC analyses did not improve disease prediction in this DAISY study. Limitations of ROC analysis include the fact that it does not take into account time to event, which is actually one of the strength of this prospective DAISY cohort study.

So far, prediction of type 1 diabetes has mainly been based on family history, age of onset of proband, autoantibody number and levels, and genetic susceptibility markers such as *INS* and HLA-DR3/4-DQB1*0302²¹⁻²⁵. Addition of *PTPN22* and *UBASH3A* SNPs to HLA-DR,DQ genotyping can help improve prediction of type 1 diabetes.

Acknowledgments

This research was supported by NIH grants R37 DK32493, RO1 DK32083, DK050979, N01 AI15416. A.K.S. was supported by the JDRF Grant 11-2010-206 Early Career Patient-oriented Diabetes Award.

The authors declare that there is no duality of interest associated with this manuscript.

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Abbreviations

GWAS	genome wide association studies
FDR	first-degree relatives
DAISY	Diabetes Autoimmunity Study in the Young
IA	islet autoimmunity
NHW	non-Hispanic White
SNPs	single nucleotide polymorphisms
TEDDY	The Environmental Determinants of Diabetes in the Young

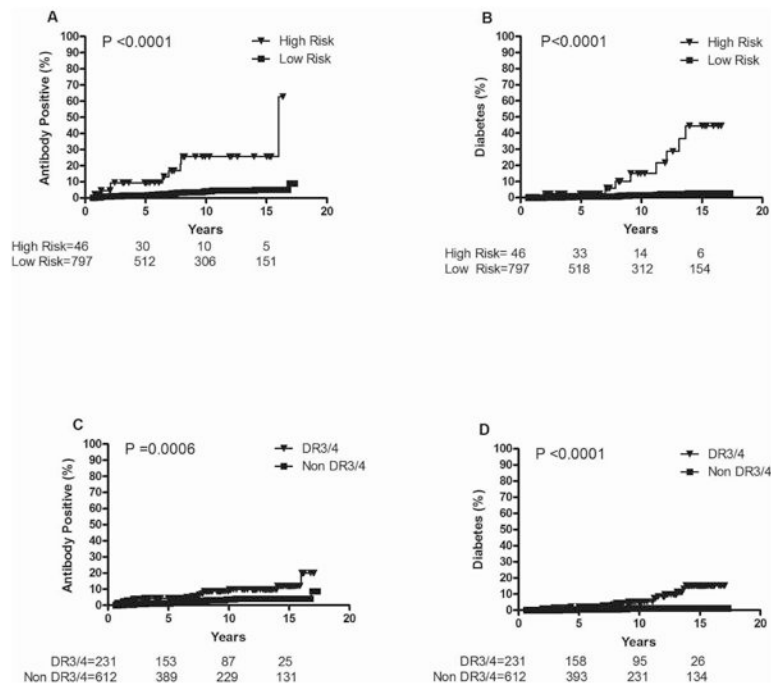


Figure 1. Progression to Islet Autoimmunity (IA) and Diabetes in DAISY NHW general population children (N=843)*: IA by genetic risk strata (1A), Diabetes by genetic risk strata (1B), IA by HLA-DR3/4 (1C) and Diabetes by HLA-DR3/4 (1D)

High risk: all subjects with *UBASH3A* AA in addition to HLA-DR3/4 as well as all subjects with *PTPN22* TT; Low risk: all other genotypes

Follow-up time was defined as the age of the child at the 1st of the 2 consecutive positive visits for affected children and age of the child at the last visit for unaffected children.

Non DR3/4 refers to not having the highest risk HLA DR3/4,DQB1*0302 genotype.

*15 subjects not included due to missing either *UBASH3A* or *PTPN22* genotyping

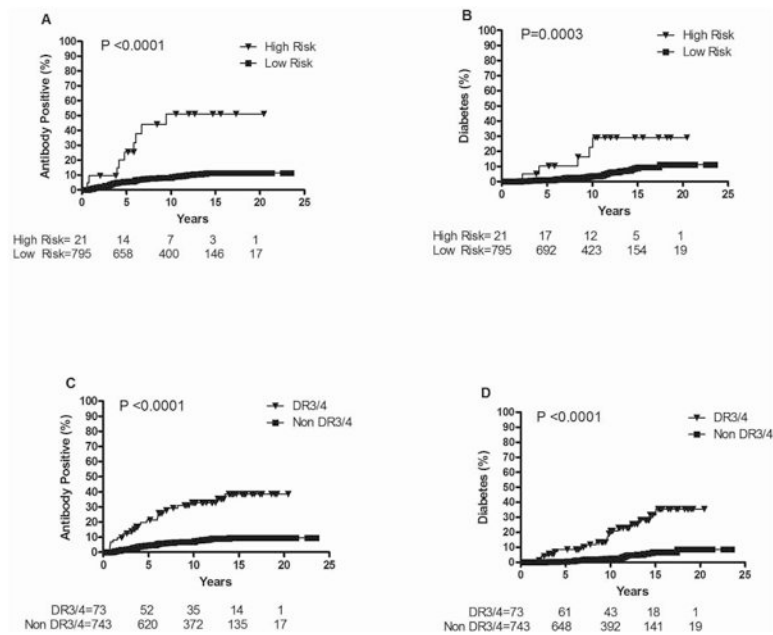


Figure 2. Progression to Islet Autoimmunity (IA) and Diabetes in DAISY NHW first-degree relatives (N=816)*: IA by genetic risk strata (1A), Diabetes by genetic risk strata (1B), IA by HLA-DR3/4 (1C) and Diabetes by HLA-DR3/4 (1D)

High risk: all subjects with *UBASH3A* AA in addition to HLA-DR3/4 as well as all subjects with *PTPN22* TT; Low risk: all other genotypes

Follow-up time was defined as the age of the child at the 1st of the 2 consecutive positive visits for affected children and age of the child at the last visit for unaffected children.

Non DR3/4 refers to not having the highest risk HLA DR3/4,DQB1*0302 genotype.

*35 subjects not included due to missing either *UBASH3A* or *PTPN22* genotyping

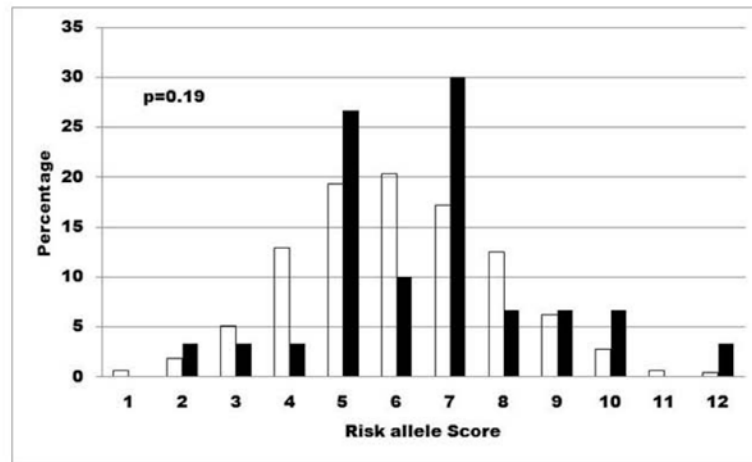


Figure 3. Non-HLA gene SNP-risk allele score distribution in DAISY general population and first-degree relatives

Distribution of risk allele scores derived from 9 (ERBB3, PTPN2, IFIH1, PTPN22, KIAA0350/CLEC16A, CTLA4, SH2B3, IL18RAP, IL10) type 1 diabetes susceptibility genes in nondiabetic autoantibody negative children (unfilled bars) compared to children who progressed to diabetes (filled bars) in DAISY

N=1332 antibody negative children and N=37 children with type 1 diabetes (antibody positive subjects who have not developed diabetes and subjects missing data for one or more SNPs are not included)

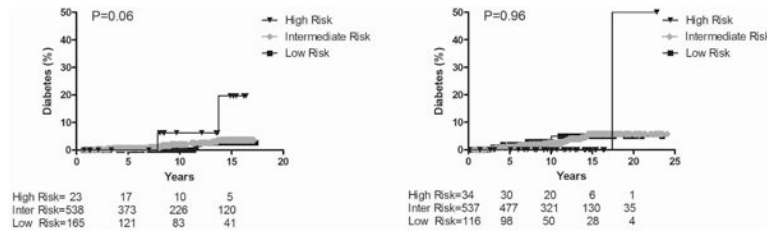


Figure 4. Progression to Diabetes in DAISY general population children (left) and DAISY first-degree relatives (right)

SNP-risk allele score categories are low <5, intermediate 5-9 and high >9

Inter: intermediate

N=726 general population children and N=687 first-degree relatives (subjects missing data for one or more SNPs were not included)

Table 1
Non-HLA gene polymorphisms as potential predictors of islet autoimmunity (IA) and progression from IA to type 1 diabetes (T1D) in DAISY non-Hispanic white population^a

Gene	SNP	Risk Allele	Islet Autoimmunity (N=1709) ^b		Progression from IA to T1D (N=116) ^c	
			HR ^d (95% CI)	p-value	HR (95% CI)	p-value
ERBB3	rs2292239	T	1.12 (0.85-1.48)	0.43	1.32 (0.81-2.15)	0.26
CLEC16A	rs12708716	G	0.94 (0.70-1.27)	0.69	0.94 (0.63-1.40)	0.77
IL27	rs4788084	T	0.98 (0.74-1.29)	0.87	1.45 (0.98-2.14)	0.06
CTRB	rs7202877	G	1.01 (0.65-1.59)	0.96	0.75 (0.39-1.41)	0.37
C14orf	rs4900384	G	0.77 (0.56-1.06)	0.11	0.64 (0.40-1.05)	0.08
GSDM	rs2290400	T	0.91 (0.70-1.19)	0.51	0.97 (0.66-1.42)	0.86
HORMAD2	rs7575307	T	0.92 (0.69-1.23)	0.59	1.16 (0.79-1.71)	0.44
BACH2	rs11755527	G	1.01 (0.76-1.35)	0.95	0.88 (0.59-1.32)	0.54
C10orf59	rs10509540	G	0.84 (0.69-1.03)	0.10	0.96 (0.72-1.26)	0.74
CD69	rs4763879	A	1.10 (0.84-1.44)	0.49	0.93 (0.65-1.35)	0.72
GAB3	rs2664170	G	0.91 (0.71-1.15)	0.42	0.92 (0.67-1.26)	0.61
GLIS3	rs7020673	C	0.77 (0.60-1.00)	0.05	0.77 (0.55-1.09)	0.14
IFIH1	rs1990760	C	1.07 (0.81-1.40)	0.65	1.47 (1.02-2.12)	0.04
IL10	rs3024496	A	1.27 (0.97-1.65)	0.08	1.21 (0.85-1.73)	0.29
IL18RAP	rs917997	A	1.16 (0.84-1.60)	0.36	0.94 (0.57-1.54)	0.80
IL2RA	rs12251307	A	0.61 (0.37-1.02)	0.06	1.15 (0.58-2.29)	0.69
INS	rs689	A	1.29 (0.93-1.79)	0.12	1.65 (1.05-2.59)	0.03
PRKD2	rs425105	C	0.87 (0.61-1.26)	0.47	0.83 (0.49-1.38)	0.47
PTPN2	rs1893217	G	1.33 (0.95-1.84)	0.09	1.11 (0.70-1.77)	0.65
PTPN22	rs2476601	T	1.87 (1.32-2.66)	0.001	1.59 (0.97-2.61)	0.06
SH2B3	rs3184504	A	0.99 (0.76-1.31)	0.97	0.80 (0.56-1.15)	0.23
SIRPG	rs2281808	T	0.85 (0.64-1.13)	0.25	1.03 (0.72-1.48)	0.87
TAGAP	rs1738074	A	0.90 (0.68-1.19)	0.47	1.20 (0.83-1.73)	0.32
UBASH3A	rs11203203	A	1.55 (1.19-2.02)	0.001	1.44 (1.01-2.04)	0.04

Gene	SNP	Risk Allele	Islet Autoimmunity (N=1709) ^b		Progression from IA to T1D (N=116) ^c	
			HR ^d (95% CI)	p-value	HR (95% CI)	p-value
UBASH3A	rs9976767	G	1.54 (1.16-2.05)	0.003	1.47 (0.97-2.21)	0.07
SLC30A8	rs13266634	T	0.95 (0.70-1.27)	0.71	1.07 (0.70-1.65)	0.75
CTLA4	rs231775	G	1.20 (0.93-1.56)	0.16	0.95 (0.66-1.36)	0.76
CCR5	microsatellite	Del32	0.93 (0.60-1.47)	0.77	1.04 (0.55-1.98)	0.89

^a Multivariate analyses, adjusted for HLA-DR3/4,DQB1*0302 and family history of type 1 diabetes

^b Total N=1709 (116 subjects with IA)

^c Total N=116 (66 subjects with T1D)

^d HR = hazard ratio

Table 2
Predictors of islet autoimmunity and progression to type 1 diabetes ^a

Predictor	Islet Autoimmunity		Progression to diabetes	
	HR ^b (95% CI)	p-value	HR (95% CI)	p-value
HLA-DR3/4, DQB1*0302	3.96 (2.22-7.06)	<0.0001	5.93 (2.83-12.43)	<0.0001
Cohort (FDR) ^c	2.09 (1.20-3.64)	0.009	2.68 (1.21-5.96)	0.02
rs689 (INS)	1.95 (1.20-3.19)	0.008	NA ^d	
rs2476601 (PTPN22)	1.93 (1.16-3.22)	0.01	NA	
rs9976767 (UBASH3A)	1.63 (1.12-2.37)	0.01	2.11 (1.14-3.89)	0.02
rs7020673 (GLIS3)	0.65 (0.45-0.93)	0.02	NA	

^a Multivariate model including all variables with an a level of <0.05

^b HR = hazard ratio

^c FDR = first-degree relatives

^d NA = not significant for progression from IA to diabetes