



Published in final edited form as:

Expert Opin Ther Targets. 2014 June ; 18(6): 703–714. doi:10.1517/14728222.2014.902938.

Protein-based Therapies for Acute Lung Injury: Targeting Neutrophil Extracellular Traps

Markus Bosmann, M.D.^{1,2} and Peter A. Ward, M.D.³

¹Center for Thrombosis and Hemostasis, University Medical Center, Mainz, Germany

²Department of Hematology, Oncology and Pneumology, University Medical Center, Mainz, Germany

³Department of Pathology, University of Michigan Medical School, Ann Arbor, MI 48109, USA

Abstract

Introduction—Acute lung injury (ALI) and acute respiratory distress syndrome (ARDS) are the acute onset of non-cardiac respiratory insufficiency associated with bilateral lung infiltrations. During the past decade, mechanical ventilation strategies using low tidal volumes have reduced the mortality of ALI/ARDS to around 20-40%. However, ALI/ARDS continues to be a major factor in global burden of diseases, with no pharmacologic agents currently available.

Areas covered—In this review we discuss several inflammatory proteins involved in the molecular pathogenesis of ALI/ARDS. The complement cleavage product, C5a, is a peptide acting as a potent anaphylatoxin. C5a may trigger the formation of neutrophil extracellular traps (NETs) and release of histone proteins to the extracellular compartment during ALI/ARDS. NETs may activate platelets to release TGF β which is involved in tissue remodeling during the later phases of ALI/ARDS. Interception of C5a signaling or blockade of extracellular histones has recently shown promising beneficial effects in small animal models of ALI/ARDS.

Expert opinion—Novel protein-based strategies for the treatment of ALI/ARDS may inspire the hopes of scientists, clinicians and patients. While neutralization of extracellular histones / NETs, C5a and TGF β is effective in experimental models of ALI/ARDS, controlled clinical trials will be necessary for further evaluation in future.

Keywords

Neutrophil extracellular traps; extracellular histones; inflammation; antibody; C5a; TGF β

Corresponding Author: Markus Bosmann, M.D. Center for Thrombosis and Hemostasis Department of Hematology, Oncology and Pneumology University Medical Center Mainz Langenbeckstrasse 1 Mainz, 55131, Germany Phone: +49 (6131) 17-8277 FAX: +49 (6131) 17-6238 markus.bosmann@unimedizin-mainz.de.

Financial and competing interests disclosure The authors have no other relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript apart from those disclosed.

1. Introducing the problem

Acute lung injury (ALI) and acute respiratory distress syndrome (ARDS) are defined by the following diagnostic criteria: acute onset, bilateral lung infiltrates on chest X-rays and respiratory failure with exclusion of cardiac etiologies. Respiratory failure is due to pulmonary edema, resulting in diminished gas diffusion capacity. This is detectable by reduced ratios of arterial oxygen tension (p_aO_2) and fraction of inspired oxygen (FiO_2). In this nomenclature, ALI ($p_aO_2/FiO_2 < 300$ mmHg) is defined as the less severe form of respiratory insufficiency as compared to ARDS ($p_aO_2/FiO_2 < 200$ mmHg). In 2012, the novel '*Berlin definition*' of ARDS was introduced, which classifies three categories of ARDS depending on the degree of hypoxemia: mild ($p_aO_2/FiO_2 < 300$ mmHg), moderate ($p_aO_2/FiO_2 < 200$ mmHg), and severe ($p_aO_2/FiO_2 < 100$ mmHg) ¹.

It is estimated that 200,000 cases of ALI/ARDS occur annually in the United States ^{2, 3}. The overall survival rates of ALI/ARDS are around 20-40% ⁴⁻⁷. The prognosis of survival is better for mild ARDS as compared to severe ARDS with an intermediate risk for moderate ARDS (according to the '*Berlin definition*') ¹. During the past two decades improved survival rates of ALI/ARDS have been accomplished ⁸. Most likely this is attributable to advances in critical care medicine. In particular, novel mechanical ventilation schemes using low tidal volumes (e.g. 6 ml/kg body weight) have reduced the risk of ventilation-induced pulmonary barotrauma and significantly increased survival rates and ventilator-free days in patients with ALI/ARDS ⁴.

A long list of predisposing factors may raise the risk for the development of ALI/ARDS. Clinical conditions that frequently precipitate ALI/ARDS include pneumonia, sepsis, fluids aspiration, severe trauma, massive transfusions and inhalation burns.

The pathophysiology of ALI/ARDS is characterized by initial injury (infection, chemical, mechanical) of the small airways (alveolar epithelium, lung endothelium) ⁸. Lung tissue injury initiates acute inflammation of the alveoli and bronchioles ⁹. This includes activation of Toll-like receptors, the complement system, plasmatic coagulation pathways as well as activation of platelets, alveolar macrophages, adjacent endothelial cells and type II alveolar epithelial cells ^{10, 11}. The activation of coagulation pathways creates intra-alveolar fibrin depositions ('hyaline membranes'). Chemotactic migration of innate immune cells to these sites results in the accumulation of polymorphonuclear neutrophils (PMNs) in the alveolar spaces within a few hours, while adaptive immune cells such as T lymphocytes mainly participate during the later phases of ALI/ARDS ^{12, 13}. PMNs are phagocytic cells and clear infectious microorganisms and debris from the alveolar spaces. Furthermore, PMNs release cytotoxic granules and reactive oxygen species ^{14, 15}.

Immune cells (PMNs, macrophages, T cells) and resident lung cells (epithelium, endothelium, fibroblasts) communicate by the release of a plethora of cytokines / chemokines. IL- 1β , TNF α , Interferon- γ and IL-6 orchestrate and perpetuate lung inflammation during ALI/ARDS ¹⁶⁻¹⁸. These inflammatory cytokines also promote plasmatic coagulation in the small airways, for example via up-regulation of tissue factor in alveolar epithelial cells ¹⁹.

While these inflammatory responses are designed to provide pathogen clearance and restore homeostasis of the lung, they may also cause additional injury in the setting of ALI/ARDS. Loss of barrier function of the epithelial-endothelial interface in alveoli is considered a hallmark of ALI/ARDS. Apoptosis, necrosis and necroptosis of endothelial cells and type I alveolar epithelial cells results in disintegration of the alveolar lining (e.g. disruption of vascular-endothelial cadherin bonds), which is followed by influx of alveolar edema fluid and hemorrhage^{11, 20, 21}. Subsequently, pulmonary gas diffusion capacity is compromised leading to hypoxemia and respiratory failure. The later phases (>7 days) of ALI/ARDS may be dominated by proliferative and fibrotic responses. The mechanisms that distinguish healing and full recovery from ALI/ARDS versus progression to interstitial inflammation and fibrotic lung disease are currently not completely known, but may involve mesenchymal stem cells^{22, 23}.

To date clinical trials using pharmacologic approaches for the treatment of ALI/ARDS have largely failed to demonstrate beneficial effects. The primary endpoints of most studies have been survival at 28 days (or longer) and reduction of days of mechanical ventilation (ventilator-free days) in ALI/ARDS patients. Protein-based strategies have tested the administration of recombinant human proteins such as activated protein C (drotrecogin α), granulocyte macrophage colony stimulating factor (GM-CSF) and surfactant protein C based agents to ALI/ARDS patients²⁴⁻²⁶. Additional trials have used methylprednisolone, nitric oxide, β 2-adrenergic receptor agonists (albuterol, salbutamol), and antioxidants (N-acetylcysteine)^{7, 27-30}. All of these pharmacologic interventions were unable to demonstrate significant clinical efficacy (Table 1). Therefore, there remains a desperate need for better understanding of the pathophysiology of ALI/ARDS. Such knowledge will be helpful to direct future clinical trials testing novel strategies to ameliorate the adverse outcomes of ALI/ARDS. In this review, we will discuss recent progress in identifying potential target proteins, which may be key factors during the unfavorable course of ALI/ARDS.

2. Potential protein targets in Acute Lung Injury

2.1. Blocking of complement component C5a in acute lung injury

The complement system is an ancient part of the innate immune system, which is found in all vertebrates and many invertebrate species³¹. In analogy to the coagulation system, complement proteins constitute a cascade of proteases with the capacity for specific cleavage of its factors during activation. The classical, alternative, MB-lectin and ficolin pathways may all initiate activation of complement during ALI/ARDS³². The major effector functions of complement are direct lysis (membrane-attack complex) and opsonization (C3b) to facilitate phagocytosis of targeted cells (e.g. microbes). In addition, an abundant generation of chemotactic complement anaphylatoxins (C3a, C4a, C5a) occurs, which subsequently activate innate and adaptive immunity. C5a and C3a activate PMNs, macrophages and promote chemotactic migration of those cells to the alveolar spaces during ALI/ARDS^{17, 33}. C5a and C3a are substrates for rapid enzymatic cleavage of the C-terminal arginine residue by plasma carboxypeptidase R and carboxypeptidase N³⁴. The cleavage product, C5a_{desArg}, displays a 10-fold lower biological activity as compared to C5a, while C3a_{desArg} is regarded as completely inactive³⁵. C5a has the strongest biological potency as

compared to C3a and C4a. C5a binds with high affinity to its two receptors, which have been termed C5aR (CD88; C5aR1) and C5L2 (GPR77; C5aR2)^{36, 37}. Both receptors, C5aR and C5L2, are encoded in vicinity on chromosome 19 in humans (mouse: chromosome 7), contain the structural motifs of seven-transmembrane spanning domains and display an amino acid homology of around 38%³⁸.

The first evidence that C5a is involved in the development and progression of ALI/ARDS was uncovered several decades ago^{39, 40}. Administration of cobra venom factor (CVF), which activates complement in circulation, resulted in PMN-dependent ALI⁴¹. CVF is a three chain protein and functional analog of C3b, which is purified from cobra venom or recombinantly expressed⁴². In plasma, CVF assembles with other complement proteins to form a C3/C5 convertase complex resulting in uncontrolled complement activation (including generation of C3a and C5a) and, ultimately, consumption of complement components.

The effects of C5a are partly mediated by the production of reactive oxygen species in PMNs (respiratory burst)⁴¹. Administration of blocking anti-C5a antibody protects non-human primates from sepsis-induced ALI and mortality following live *E. coli* challenge⁴³. The complement degradation products, C5a_{desArg} and C3a_{desArg}, are detectable in blood of human patients with ALI/ARDS and sepsis, indicating complement activation⁴⁴. Gene-targeted disruption of the C5aR receptor reduces the severity of immune-complex-induced ALI in mice⁴⁵. Vitamin D binding protein deficient mice are protected against C5a-induced ALI and influx of inflammatory cells⁴⁶, while serum Vitamin D concentrations appear not to influence the severity of LPS-ALI⁴⁷.

C5a-signaling in alveolar macrophages is modulated by urokinase-type plasminogen activator and its receptor⁴⁸. C5 may play a profibrotic role in chronic stages of ALI, as studied in the bleomycin-induced lung injury model using C5-deficient mice⁴⁹. While in the acute phase of ALI the C5aR receptor clearly promotes inflammation^{17, 45}, the function of the C5L2 receptor is still controversial. We have recently reported that the C5L2 receptor is involved in promoting inflammation and disruption of the alveolar/epithelial barrier during ALI, while an earlier report has described an anti-inflammatory role of C5L2 during immune-complex ALI^{17, 50}.

In summary, C5a (via C5aR and C5L2) recruits PMNs and other inflammatory cells to the alveolar spaces during ALI³². The intracellular effects of C5a are related to the activation of Ca²⁺-currents as well as activation of PI3K/Akt and MAPK signaling pathways^{32, 51}. Only recently, the down-stream effects of C5a-induced tissue injury have been associated with the appearance of neutrophil extracellular traps (NETs)¹⁷. These findings may suggest a close pathophysiological link of C5a and NETs in the setting of ALI.

2.2 Targeting neutrophil extracellular traps (NETs) in acute lung injury

Accumulating evidence suggests the biological relevance of neutrophil extracellular traps (NETs) for the pathogenesis of inflammatory diseases. NETs are structures of nuclear chromatin, which contain DNA, histone proteins and other microbicidal / nuclear proteins. NETs are an effector defense mechanism of innate immunity^{52, 53}. PMNs may undergo

programmed cell death and actively release their nucleus to create NETs⁵⁴. The process of formation of NETs may occur either in blood circulation or following chemotactic migration of PMNs to the local site of inflammation. NETs and their major component, extracellular histones, ensnare and kill bacteria in septic blood⁵⁵. Similarly, NETs are capable of killing *Candida species*⁵⁶. PMNs invading tissues form NETs following encountering *Aspergillus fumigatus* in vitro and during lung infection^{57, 58}. Furthermore, certain lung pathogens such as *Streptococcus pneumoniae* have evolved counter strategies such as genes encoding for endonucleases to escape killing by NETs⁵⁹. This demonstrates that NETs are a part of the complex host-pathogen interactions, which have formed during evolution. While NETs may have been evolved to clear infectious pathogens, NETs may also cause adverse tissue injury to the host. Extracellular histones (the major components of NETs) are highly toxic and induce respiratory failure when infused intravenously into healthy research animals (Figure 1)⁶⁰. The cytotoxic activity of extracellular histones / NETs is in line with the fact, that several other intra-cellular proteins (e.g. HMGB1, hemoglobin) have detrimental effects following release to the extracellular compartment^{61, 62}.

Many factors typically present during ALI/ARDS have the potential do induce NET-formation. For instance, live bacteria, LPS, IL-8 or reactive oxygen species (ROS) may all trigger the appearance of NETs⁵⁴. The generation of NETs is an active process and requires an intra-cellular signaling program. Engagement of Raf-MEKERK kinase pathways occurs during NET formation⁶³. In addition, mammalian target of rapamycin (MTOR) and hypoxia inducible factor 1 (HIF-1) regulate the formation of NETs⁶⁴. The down-stream events of the aforementioned signaling pathways include chromatin decondensation, which is a prerequisite for NET generation. This is accomplished by enzymatic hypercitrullination of core histone proteins^{65, 66}.

When NETs derived from activated human PMNs are incubated with mouse or human cell lines of lung epithelial cells, NETs induce cell death of such epithelial cells⁶⁷. NETs are also cytotoxic for lung endothelial cells^{60, 67}. Components of NETs (MPO/DNA/histones) are detectable in broncho-alveolar lavage fluids (BALF) and lung sections by immunofluorescence microscopy of mice following LPS-induced ALI⁶⁷.

The major studies which have investigated the role of NETs (DNA/histones) during ALI/ARDS are summarized in Table 2. In experimental transfusion-related acute lung injury (TRALI), NETs are detectable in the lung microcirculation by immunofluorescence microscopy⁶⁸. In this study, NET formation in TRALI lungs was prevented by inhibition of platelet aggregation using acetylsalicylic acid⁶⁸. The blockade of NETs by administration of neutralizing antibodies directed against extracellular histones reduced lung vascular permeability and the volume of extravascular lung water during TRALI⁶⁸. Similarly, in vivo degradation of NET-derived DNA structures using DNase I reduced the severity of lung injury and mortality in the murine TRALI model⁶⁸. Moreover, myeloperoxidase (MPO)/DNA aggregates as makers of NETs were elevated in plasma samples of human patients with TRALI as compared to healthy controls⁶⁸. In addition, extracellular histones were co-localized with MPO and DNA in lung tissue sections of TRALI patients⁶⁸.

Trauma-associated ALI/ARDS has been recently associated with extracellular histones in blood circulation⁶⁹. In patients with severe nonthoracic blunt trauma, levels of circulating nucleosomes and extracellular histones increase with injury severity scores⁶⁹. When purified histones are infused intravenously in mice, the lung is the most susceptible organ, showing histological signs of inflammation and microvascular thrombi^{17, 69}. This observation may be explained by the fact, that the lungs contain the first capillary bed following intravenous administration of any substance. In response to severe trauma, extracellular histones may be released by dying cells other than PMNs. Circulating histones may be transported via the venous blood flow to the lung endothelium, resulting in extensive lung injury. In fact, it has been described, that extracellular histones derived from dying parenchymal cells rather than NETs can aggravate organ dysfunction⁷⁰. Neutralizing anti-histone single chain variable fragments (scFv) have been used to suppress histone-induced toxicity and reverse coagulation activation⁶⁹.

Treatment of C57BL/6J mice with antibodies targeting histone H4 and H2A is protective in a model of C5a-induced ALI (Figure 2)¹⁷. Extracellular histones are released in BALF during ALI, when induced by LPS, recombinant C5a or IgG immune-complexes¹⁷. In all three models, the appearance of extracellular histones requires the presence of the C5a receptors, C5aR and C5L2¹⁷. Extracellular histones and nucleosomes are also detectable in BALF of around 50% of human patients suffering von ALI/ARDS¹⁷. Extracellular histones mediate the intracellular influx of Ca²⁺ in type II alveolar epithelial cell lines, most likely related to provoking cell death¹⁷. In rats, the direct administration of purified histones to the airways results in severe respiratory acidosis, compromised respiratory excursions, pulmonary edema and acute lung inflammation¹⁷. Histones induce the release of a wide spectrum of inflammatory mediators such as IL-1 β , TNF α , IL-6, Eotaxin, G-CSF, KC, MCP-1, MIP-1 α , MIP-1 β and RANTES^{17, 69}. Extracellular histones may directly or indirectly recruit the activity of TLR2 and TLR4 receptors for promoting inflammation⁷¹. In influenza A virus mediated ALI, the accumulation of PMNs, NETs and lung epithelial injury was reduced by a potent arthropod-derived C5-binding inhibitor of complement activation (OmCI)⁷².

Intra-alveolar hemorrhage is a typical histologic finding of ALI/ARDS. A growing list of evidence suggests the interactions of NETs / extracellular histones with the coagulation system. The major components of NETs, extracellular histones and DNA, are detectable in venous thrombus formations⁷³. In detail, NETs can activate the intrinsic plasmatic coagulation pathway by direct interaction with coagulation factor XII, resulting in generation of the fibrin clot⁷⁴. During severe sepsis, platelets are activated via their TLR4 receptor and bind to PMNs, which triggers the formation of NETs in pulmonary capillaries⁵⁵. On the other hand, platelets can bind to preformed NETs. In fact, NETs provide a scaffold for platelet aggregation under shear stress, thereby promoting thrombosis⁷³. Extracellular DNA and histones are detectable during acute thrombotic microangiopathies and may potentially precipitate the formation of microthrombi⁷⁵. In co-incubation studies of human PMNs with platelets, the formation of NETs was induced, when platelets were pre-activated using proteinase-activated receptor (PAR-1) agonist or thrombin receptor-activating peptide (TRAP)⁶⁸.

Platelets are a major source of TGF β in plasma. NET-induced platelet activation would be expected to promote the release of TGF β from the α -granules of platelets. As highlighted below, TGF β is considered a critical factor especially during the later stages of ALI that determine tissue repair and remodeling.

2.3 Manipulation of transforming growth factor-beta in acute lung injury

Transforming growth factor-beta (TGF β) is a highly conserved cytokine with several existing isoforms (TGF β 1, TGF β 2, TGF β 3) ⁷⁶. TGF β typically forms homodimers.

The release of TGF β occurs as a large latent protein complex (LLC) consisting of TGF β , latency-associated peptide (LAP) and latent TGF β -binding protein (LTBP) ⁷⁷. The LLC binds to components of the extracellular matrix. Latent TGF can also bind to glycoprotein A repetitions predominant (GARP) expressed on regulatory T cells ⁷⁸.

The multiple molecular events transforming latent TGF β into active TGF β are not completely understood. One mechanism of activation is dependent on integrins such as α V β 6 integrin, which is expressed by lung epithelial cells ⁷⁹. In addition, integrin-independent activation steps may include reactive oxygen species, pH reduction (acidification), metallo-proteases (e.g. MMP-2, MMP-9, MMP-14) and thrombospondin-1 ⁸⁰⁻⁸².

TGF β initiates intracellular signaling by binding to the tetrameric TGF β receptor complex. This complex is composed of two homodimers of TGF β receptor I (TGF β RI) and two homodimers of TGF β receptor II (TGF β RII) ⁸³. Initially, TGF β binds to TGF β RII and subsequently TGF β RI is recruited. Both receptors have intracellular domains with serine/threonine kinase activity. In addition, a TGF β RIII is required for optimal signaling of the TGF β 2 isoform ⁸⁴. For intracellular signaling, TGF β recruits activin receptor-like kinases (ALK1, ALK5), that subsequently phosphorylate SMAD proteins ^{85, 86}. In addition, non-Smad-dependent signaling pathways such as p38 MAPK and DAXX are activated ^{87, 88}.

Mice with targeted genetic disruption of TGF β 1 die prematurely from spontaneous generalized inflammation ⁸⁹. TGF β is a central player of acute lung injury ^{79, 90}. A soluble chimeric TGF β type II receptor has the activity of reducing pulmonary edema fluid following LPS-induced ALI ⁷⁹. In monolayers of type II alveolar epithelial cells, recombinant TGF β decreases the transepithelial resistance via depletion of the intracellular amounts of the reduced tripeptide, glutathione ⁷⁹. A genetic defect in TGF β activation, as present in mice with deficiency of α V β 6 integrin, results in reduced pulmonary edema and lung epithelial permeability during ALI ⁷⁹. Furthermore, TGF β 1 directly regulates the transport of ions and water in type II alveolar epithelial cells by MAPK-dependent and ERK1/2-dependent suppression of ion transporter proteins such as the epithelial sodium channel alpha subunit (alphaENaC) ⁹¹. In addition, wounded epithelial cell monolayers release HMGB1, that appears to accelerates healing of the alveolar lung epithelium via α V β 6-dependent activation of TGF β 1 ⁹².

TGF β is a prototypic cytokine for promoting pulmonary fibrosis ^{93, 94}. TGF β facilitates the proliferation and chemotaxis of fibroblasts ⁹⁵. The local transformation of fibroblasts or lung

epithelial cells into myofibroblasts is promoted, while the apoptosis of myofibroblasts is limited by TGF β ⁹⁶⁻⁹⁸. Myofibroblasts are important for the production of extracellular matrix components and produce additional TGF β in a positive feedback loop ⁹⁹. TGF β reduces the activity of matrix metallo-proteases and other matrix-degrading proteases ¹⁰⁰. In addition, TGF β facilitates the production of several other profibrotic cytokines such as platelet-derived growth factor (PDGF) in vascular endothelial cells ¹⁰¹.

In summary, TGF β may be especially important during the later profibrotic stages of ALI/ARDS, although there is also the view that TGF β might be active early in ALI ¹⁰². Blockade of TGF β activation (e.g. anti- α v β 6 integrin antibody) or modulation of TGF β signaling (e.g. ALK5 inhibitors) are currently evaluated as therapeutic strategies to prevent adverse tissue remodeling and pulmonary fibrosis ^{103, 104}.

3. Conclusion

In this review, we have discussed three potential protein targets for future therapies of ALI/ARDS (Figure 3). The complement fragment, C5a, NETs and TGF β orchestrate the development and progression of ALI/ARDS at different stages of disease. C5a is considered an early proinflammatory mediator of the innate immune system. Accumulating evidence suggest, that NETs are an integral effector mechanism of immunity but may mediate adverse outcomes of ALI/ARDS. Extracellular histones and DNA are major components of NETs, but may also be passively released by lung epithelial or endothelial cells. Finally, the later profibrotic phases of ALI/ARDS may involve TGF β , resulting in healing (restoration of homeostasis) or progression to fibrosis with impaired organ function. Therapeutic modulation of the activity of C5a, NETs or TGF β (e.g. by specific neutralizing antibodies) may be helpful in future for patients with ALI/ARDS. However, the clinical efficacy and feasibility of such strategies remains to be seen.

4. Expert opinion

The improvement in survival rates of ALI/ARDS which has been observed during the past 1-2 decades are mainly explained by advances in supportive critical care medicine ⁸. The optimization of mechanical ventilation strategies has been clearly beneficial. On the other hand, even with best supportive care, death occurs in at least 20% of patients with ALI/ARDS. To date all clinical trials investigating novel or traditional pharmacologic substances have failed to show an advantage in the treatment of lung injury. Despite these sobering results, emerging experimental studies have recently raised new enthusiasm to achieve a better pathophysiologic understanding of ALI/ARDS in future.

The role of NETs and extracellular histones during the development and perpetuation of ALI/ARDS may turn out to be an important aspect for this disease. However, it should also be mentioned that the reports on NETs have been faced with some degree of skepticism. ¹⁰⁵ Some scientists have questioned that the presence of extracellular DNA and histones on microscopic slides of activated PMNs may represent laboratory artefacts due to mechanical disruption of cells during isolation and staining procedures rather than a phenomenon of biological significance. On the other hand, several studies have successfully employed intra-

vital imaging of NETs or strategies to antagonize NETs in vivo (e.g. treatment with DNase or anti-histone antibodies). The controversy extends to the level of importance that NET-formation may play for innate host defense. While it is recognized that certain bacteria encode DNases for degradation of NETs as a strategy of immune evasion¹⁰⁶, it is not entirely clear how essential NETs are to achieve pathogen clearance.

In analogy to NETs and appearance of histones, the release of other intracellular proteins such as myoglobin (e.g. during rhabdomyolysis) or hemoglobin (e.g. during destruction of red blood cells) has long been recognized as a risk factor for tissue damage and organ dysfunction¹⁰⁷. In the area following the discovery of penicillin many substances had been evaluated for their bactericidal activity. Indeed, it was already in the 1940s when extracts of purified histone proteins were demonstrated to effectively kill bacteria in culture media¹⁰⁸. It is somewhat tragic, that it took modern science until the 21st century to appreciate that the endogenous release of histone/DNA in the form of NETs may constitute an innate defense mechanism⁵². As true for several immune effector mechanisms, NETs may not only be beneficial for clearance of infectious pathogens, but may rather cause collateral damage to endothelial and epithelial cells during ALI/ARDS. In our opinion, extracellular histones / NETs clearly display adverse effects in the setting of ALI/ARDS. The bactericidal effects of extracellular histones / NETs may not be needed in patients with ALI/ARDS, as long as control of pathogen growth can effectively be achieved by the administration of antibiotics. Hence, selective suppression of certain host immune defense systems (NETs, C5a), which cause tissue injury, may be save to exploit as a pharmacologic principle. It was a key finding that C5a via C5aR and C5L2 promotes the appearance of extracellular histones, which can subsequently mediate cell death of epithelial cells and production of inflammatory mediators in the alveolar compartment during ALI/ARDS¹⁷.

Histone H4 appears to be the histone protein with the highest cytotoxic activity. Blockade of H4 using antibodies reduced the severity of ALI by around 50% in several studies^{17, 68}. An important development in future would be the identification of antibodies with higher avidity for the cytotoxic domains of several core histones in order to fully maintain normal alveolar/endothelial barrier function during ALI/ARDS.

The ultimate goal in the field of research on ALI/ARDS would be the availability of an easy-to-administer recombinant protein / enzyme to efficiently antagonize the unfavorable effects of certain endogenous proteins (NETs/extracellular histones, C5a, TGF β). Protein-based therapies for ALI/ARDS may hold the promise to reduce mortality rates and prevent the need for invasive mechanical ventilation, which can be a traumatizing event for patients and their relatives. Current treatment strategies with supportive critical care treatment result in substantial costs for the national health systems. It is unclear, if a cost benefit of protein-based anti-ALI/ARDS drugs would be accomplished, especially when such agents may require de novo pharmacologic development and extensive clinical testing.

The humanized monoclonal anti-C5 antibody, eculizumab, would have the advantage that it has already been FDA-approved for the treatment of atypical hemolytic uremic syndrome (aHUS) and paroxysmal nocturnal hemoglobinuria (PNH)¹⁰⁹. It is tempting to speculate

that eculizumab may also block C5a-induced release of NETs / extracellular histones in the setting of ALI/ARDS. At least, this could be easily tested in experimental models of ALI.

We may at least expect a good amount of scientific progress in the coming years. In particular, the area of experimental research on the role of NETs / extracellular histones during ALI/ARDS may provide additional insights how NETs are situated in the immunologic networks.

A weakness of the current data on NETs is that it was mainly (but not exclusively) accumulated in animal models of ALI/ARDS. It should be cautioned that a certain degree of disconnect between experimental and clinical studies of ALI/ARDS is likely to exist. Experimental studies typically use rodents (*M. musculus*, *R. norvegicus*) with defined induction of ALI by a single insult. For example, lipopolysaccharide (LPS) is frequently used to induce ALI in rodents. It should be mentioned that rodents are much more resistant against the adverse effects of LPS as compared to humans, which is partly explained by a high level of genetic variability in the extracellular domain of their respective TLR4 receptors¹¹⁰. Experimental TRALI may involve a two hit model starting with LPS injections followed by administration of mouse MHC class I antibodies⁶⁸. This experimental approach in rodents provides a more standardized insult as compared to the complex etiology of TRALI in humans¹¹¹. In most small animal studies measurements of p_aO_2 are not routinely performed and the term ALI is used, therefore, discrimination between ALI and ARDS would be difficult. In fact, the severity of many ALI/ARDS models in rodents is less as compared to ALI/ARDS observed in human patients. More severe experimental models of ALI/ARDS in rodents would require the use of small animal mechanical ventilation, which is not feasible in most laboratories.

Consequently, the relevance of experimental results obtained by studies with small animal models for the situation in human patients with ALI/ARDS is not entirely clear¹¹². For example, the use of β_2 -adrenoceptor agonists or glucocorticoids has shown some efficacy in experimental ALI using rodents^{113, 114}, but such agents have failed to show protective results in clinical trials in humans^{28, 30, 115}. These observations may suggest the existence of considerable discrepancies in the molecular pathophysiology of experimental ALI in small animals as compared to ALI/ARDS in humans. Unlike experimental ALI, most clinical studies with humans have employed the evaluation of 30-60-day mortality and ventilator-free days. These primary endpoints are often supplemented by collection of BALF and plasma at serial time points. Clinical studies typically require high numbers of patients given the heterogeneity of underlying causes of ALI/ARDS. In addition, study populations may vary because of different definitions of ALI/ARDS (e.g. severity of disease according to the traditional definition versus the novel '*Berlin definition*') making the comparison of study results sometimes difficult.

In future, the use of novel experimental models such as 'humanized mice' may have some potential to overcome the limited relevance of current animal models for human diseases. 'Humanized mice' are generated by transplantation of human CD34+ hematopoietic stem cells into immunocompromised mouse strains (e.g. NSG mice) resulting in the co-presence of a human immune system in a mouse lung during experimental ALI/ARDS. Additional

progress may be achieved by the utilization of novel methodologies such as next-generation-sequencing approaches, the use of flow cytometry based high-throughput immunocytofluorescence (e.g. ImageStream^X Mark II, Amnis) and single-cell protein biomarker analysis (e.g. chipcytometry technology, Zellkraftwerk).

In conclusion, subsequent decades may be full of scientific advancement as far as the understanding of the pathophysiology and drug development for the treatment of ALI/ARDS is concerned.

Acknowledgments

Peter Ward was supported by grants from the National Institutes of Health, USA (GM-29507, GM-61656.), Markus Bosmann was supported by the Federal Ministry of Education and Research (BMBF, 01EO1003), Deutsche Forschungsgemeinschaft (BO 3482/3-1), B. Braun Foundation, MAIFOR Program of the University Medical Center Mainz, Marie Curie Career Integration Grant of the European Union (project 334486) and a Clinical Research Fellowship of the European Hematology Association. The authors are responsible for the scientific content of this publication.

References

Papers of special note have been highlighted as either of interest (•) or of considerable interest (••) to readers.

1. Force ADT, Ranieri VM, Rubenfeld GD, et al. Acute respiratory distress syndrome: the Berlin Definition. *JAMA*. Jun 20; 2012 307(23):2526–33. [PubMed: 22797452]
2. Goss CH, Brower RG, Hudson LD, et al. Incidence of acute lung injury in the United States. *Critical care medicine*. 2003; 31(6):1607–11. [PubMed: 12794394]
3. Rubenfeld GD, Caldwell E, Peabody E, et al. Incidence and outcomes of acute lung injury. *N Engl J Med*. Oct 20; 2005 353(16):1685–93. [PubMed: 16236739]
4. Ventilation with lower tidal volumes as compared with traditional tidal volumes for acute lung injury and the acute respiratory distress syndrome. The Acute Respiratory Distress Syndrome Network. *N Engl J Med*. May 4; 2000 342(18):1301–8. NoAuthorsListed. * This clinical study was a milestone to improve the clinical management of ALI/ARDS by simple reduction of tidal volumes.
5. National Heart L; Blood Institute Acute Respiratory Distress Syndrome Clinical Trials N. Wiedemann HP, et al. Comparison of two fluid-management strategies in acute lung injury. *N Engl J Med*. Jun 15; 2006 354(24):2564–75. [PubMed: 16714767]
6. Rice TW, Wheeler AP, Thompson BT, et al. Enteral omega-3 fatty acid, gamma-linolenic acid, and antioxidant supplementation in acute lung injury. *JAMA*. Oct 12; 2011 306(14):1574–81. [PubMed: 21976613]
7. National Heart L; Blood Institute Acute Respiratory Distress Syndrome Clinical Trials N. Matthay MA, et al. Randomized, placebo-controlled clinical trial of an aerosolized beta(2)-agonist for treatment of acute lung injury. *Am J Respir Crit Care Med*. Sep 1; 2011 184(5):561–8. [PubMed: 21562125]
8. Matthay MA, Ware LB, Zimmerman GA. The acute respiratory distress syndrome. *J Clin Invest*. Aug 1; 2012 122(8):2731–40. [PubMed: 22850883]
9. Ward PA. Acute lung injury: how the lung inflammatory response works. *Eur Respir J Suppl*. Sep. 2003 44:22s–23s. [PubMed: 14582896]
10. Gao H, Neff T, Ward PA. Regulation of lung inflammation in the model of IgG immune-complex injury. *Annual review of pathology*. 2006; 1:215–42.
11. Corada M, Mariotti M, Thurston G, et al. Vascular endothelial-cadherin is an important determinant of microvascular integrity in vivo. *Proc Natl Acad Sci U S A*. Aug 17; 1999 96(17): 9815–20. [PubMed: 10449777]

12. Shaw JO, Henson PM. Pulmonary intravascular sequestration of activated neutrophils: failure to induce light-microscopic evidence of lung injury in rabbits. *Am J Pathol.* Jul; 1982 108(1):17–23. [PubMed: 7091301]
13. Garibaldi BT, D'Alessio FR, Mock JR, et al. Regulatory T cells reduce acute lung injury fibroproliferation by decreasing fibrocyte recruitment. *Am J Respir Cell Mol Biol.* Jan; 2013 48(1):35–43. [PubMed: 23002097]
14. Imai Y, Kuba K, Neely GG, et al. Identification of oxidative stress and Toll-like receptor 4 signaling as a key pathway of acute lung injury. *Cell.* Apr 18; 2008 133(2):235–49. [PubMed: 18423196]
15. Chabot F, Mitchell JA, Gutteridge JM, et al. Reactive oxygen species in acute lung injury. *Eur Respir J.* Mar; 1998 11(3):745–57. [PubMed: 9596132]
16. Warren JS, Yabroff KR, Remick DG, et al. Tumor necrosis factor participates in the pathogenesis of acute immune complex alveolitis in the rat. *The Journal of clinical investigation.* Dec; 1989 84(6):1873–82. [PubMed: 2531759]
17. Bosmann M, Grailer JJ, Ruemmler R, et al. Extracellular histones are essential effectors of C5aR- and C5L2-mediated tissue damage and inflammation in acute lung injury. *FASEB J.* Aug 27.2013
* In this report, the intra-tracheal administration of histone extracts to healthy rodents resulted in ALI/ARDS-like symptoms.
18. Chen ES, Greenlee BM, Wills-Karp M, et al. Attenuation of lung inflammation and fibrosis in interferon-gamma-deficient mice after intratracheal bleomycin. *Am J Respir Cell Mol Biol.* May; 2001 24(5):545–55. [PubMed: 11350823]
19. Bastarache JA, Sebag SC, Grove BS, et al. Interferon-gamma and tumor necrosis factor-alpha act synergistically to up-regulate tissue factor in alveolar epithelial cells. *Exp Lung Res.* Oct; 2011 37(8):509–17. [PubMed: 21913843]
20. Bardales RH, Xie SS, Schaefer RF, et al. Apoptosis is a major pathway responsible for the resolution of type II pneumocytes in acute lung injury. *Am J Pathol.* Sep; 1996 149(3):845–52. [PubMed: 8780388]
21. Polunovsky VA, Chen B, Henke C, et al. Role of mesenchymal cell death in lung remodeling after injury. *J Clin Invest.* Jul; 1993 92(1):388–97. [PubMed: 8326006]
22. Curley GF, Hayes M, Ansari B, et al. Mesenchymal stem cells enhance recovery and repair following ventilator-induced lung injury in the rat. *Thorax.* Jun; 2012 67(6):496–501. [PubMed: 22106021]
23. Gotts JE, Matthay MA. Mesenchymal stem cells and acute lung injury. *Crit Care Clin.* Jul; 2011 27(3):719–33. [PubMed: 21742225]
24. Liu KD, Levitt J, Zhuo H, et al. Randomized clinical trial of activated protein C for the treatment of acute lung injury. *Am J Respir Crit Care Med.* Sep 15; 2008 178(6):618–23. [PubMed: 18565951]
25. Paine R 3rd, Standiford TJ, Dechert RE, et al. A randomized trial of recombinant human granulocyte-macrophage colony stimulating factor for patients with acute lung injury. *Crit Care Med.* Jan; 2012 40(1):90–7. [PubMed: 21926600]
26. Spragg RG, Lewis JF, Walmrath HD, et al. Effect of recombinant surfactant protein C-based surfactant on the acute respiratory distress syndrome. *N Engl J Med.* Aug 26; 2004 351(9):884–92. [PubMed: 15329426]
27. Gao Smith F, Perkins GD, Gates S, et al. Effect of intravenous beta-2 agonist treatment on clinical outcomes in acute respiratory distress syndrome (BALTI-2): a multicentre, randomised controlled trial. *Lancet.* Jan 21; 2012 379(9812):229–35. [PubMed: 22166903]
28. Bernard GR, Luce JM, Sprung CL, et al. High-dose corticosteroids in patients with the adult respiratory distress syndrome. *N Engl J Med.* Dec 17; 1987 317(25):1565–70. [PubMed: 3317054]
29. Bernard GR, Wheeler AP, Arons MM, et al. A trial of antioxidants N-acetylcysteine and procysteine in ARDS. The Antioxidant in ARDS Study Group. *Chest.* Jul; 1997 112(1):164–72. [PubMed: 9228372]
30. Steinberg KP, Hudson LD, Goodman RB, et al. Efficacy and safety of corticosteroids for persistent acute respiratory distress syndrome. *N Engl J Med.* Apr 20; 2006 354(16):1671–84. [PubMed: 16625008]

31. Nonaka M, Yoshizaki F. Evolution of the complement system. *Molecular immunology*. 2004; 40(12):897–902. [PubMed: 14698228]
32. Bosmann M, Ward PA. Role of C3, C5 and anaphylatoxin receptors in acute lung injury and in sepsis. *Adv Exp Med Biol*. 2012; 946:147–59. [PubMed: 21948367]
33. Mulligan MS, Schmid E, Beck-Schimmer B, et al. Requirement and role of C5a in acute lung inflammatory injury in rats. *J Clin Invest*. Jul 15; 1996 98(2):503–12. [PubMed: 8755663]
34. Campbell WD, Lazoura E, Okada N, et al. Inactivation of C3a and C5a octapeptides by carboxypeptidase R and carboxypeptidase N. *Microbiol Immunol*. 2002; 46(2):131–4. [PubMed: 11939578]
35. Zwirner J, Gotze O, Sieber A, et al. The human mast cell line HMC-1 binds and responds to C3a but not C3a(desArg). *Scand J Immunol*. Jan; 1998 47(1):19–24. [PubMed: 9467653]
36. Gerard NP, Gerard C. The chemotactic receptor for human C5a anaphylatoxin. *Nature*. 1991; 349(6310):614–7. [PubMed: 1847994]
37. Okinaga S, Slattery D, Humbles A, et al. C5L2, a nonsignaling C5A binding protein. *Biochemistry*. Aug 12; 2003 42(31):9406–15. [PubMed: 12899627]
38. Li R, Coulthard LG, Wu MC, et al. C5L2: a controversial receptor of complement anaphylatoxin, C5a. *FASEB J*. Mar; 2013 27(3):855–64. [PubMed: 23239822]
39. Ward PA. Immune complex injury of the lung. *Am J Pathol*. Oct; 1979 97(1):85–92. [PubMed: 158982]
40. Larsen GL, McCarthy K, Webster RO, et al. A differential effect of C5a and C5a des Arg in the induction of pulmonary inflammation. *Am J Pathol*. Jul; 1980 100(1):179–92. [PubMed: 7395964]
41. Till GO, Johnson KJ, Kunkel R, et al. Intravascular activation of complement and acute lung injury. Dependency on neutrophils and toxic oxygen metabolites. *J Clin Invest*. May; 1982 69(5): 1126–35. [PubMed: 7068850]
42. Vogel CW, Fritzing DC, Hew BE, et al. Recombinant cobra venom factor. *Molecular immunology*. Jun; 2004 41(2-3):191–9. [PubMed: 15159065]
43. Stevens JH, O’Hanley P, Shapiro JM, et al. Effects of anti-C5a antibodies on the adult respiratory distress syndrome in septic primates. *J Clin Invest*. Jun; 1986 77(6):1812–6. [PubMed: 3711336]
44. Weinberg PF, Matthay MA, Webster RO, et al. Biologically active products of complement and acute lung injury in patients with the sepsis syndrome. *Am Rev Respir Dis*. Nov; 1984 130(5): 791–6. [PubMed: 6497161]
45. Bozic CR, Lu B, Hopken UE, et al. Neurogenic amplification of immune complex inflammation. *Science*. Sep 20; 1996 273(5282):1722–5. [PubMed: 8781237]
46. Trujillo G, Habel DM, Ge L, et al. Neutrophil recruitment to the lung in both C5a- and CXCL1-induced alveolitis is impaired in vitamin D-binding protein-deficient mice. *J Immunol*. Jul 15; 2013 191(2):848–56. [PubMed: 23752613]
47. Klaff LS, Gill SE, Wisse BE, et al. Lipopolysaccharide-induced lung injury is independent of serum vitamin D concentration. *PLoS One*. 2012; 7(11):e49076. [PubMed: 23185294]
48. Shushakova N, Eden G, Dangers M, et al. The urokinase/urokinase receptor system mediates the IgG immune complex-induced inflammation in lung. *J Immunol*. Sep 15; 2005 175(6):4060–8. [PubMed: 16148155]
49. Addis-Lieser E, Kohl J, Chiamonte MG. Opposing regulatory roles of complement factor 5 in the development of bleomycin-induced pulmonary fibrosis. *J Immunol*. Aug 1; 2005 175(3):1894–902. [PubMed: 16034133]
50. Gerard NP, Lu B, Liu P, et al. An anti-inflammatory function for the complement anaphylatoxin C5a-binding protein, C5L2. *Journal of Biological Chemistry*. 2005; 280(48):39677–80. [PubMed: 16204243]
51. Bosmann M, Haggadone MD, Hemmila MR, et al. Complement activation product C5a is a selective suppressor of TLR4-induced, but not TLR3-induced, production of IL-27(p28) from macrophages. *J Immunol*. May 15; 2012 188(10):5086–93. [PubMed: 22491257]
52. Brinkmann V, Reichard U, Goosmann C, et al. Neutrophil extracellular traps kill bacteria. *Science*. Mar 5; 2004 303(5663):1532–5. [PubMed: 15001782] ** This is the initial report introducing the concept of neutrophil extracellular traps.

53. Brinkmann V, Zychlinsky A. Beneficial suicide: why neutrophils die to make NETs. *Nat Rev Microbiol.* Aug; 2007 5(8):577–82. [PubMed: 17632569]
54. Fuchs TA, Abed U, Goosmann C, et al. Novel cell death program leads to neutrophil extracellular traps. *J Cell Biol.* Jan 15; 2007 176(2):231–41. [PubMed: 17210947]
55. Clark SR, Ma AC, Tavener SA, et al. Platelet TLR4 activates neutrophil extracellular traps to ensnare bacteria in septic blood. *Nat Med.* Apr; 2007 13(4):463–9. [PubMed: 17384648]
56. Urban CF, Reichard U, Brinkmann V, et al. Neutrophil extracellular traps capture and kill *Candida albicans* yeast and hyphal forms. *Cell Microbiol.* Apr; 2006 8(4):668–76. [PubMed: 16548892]
57. Bruns S, Kniemeyer O, Hasenberg M, et al. Production of extracellular traps against *Aspergillus fumigatus* in vitro and in infected lung tissue is dependent on invading neutrophils and influenced by hydrophobin RodA. *PLoS Pathog.* Apr.2010 6(4):e1000873. [PubMed: 20442864]
58. McCormick A, Heesemann L, Wagener J, et al. NETs formed by human neutrophils inhibit growth of the pathogenic mold *Aspergillus fumigatus*. *Microbes Infect.* Nov; 2010 12(12-13):928–36. [PubMed: 20603224]
59. Beiter K, Wartha F, Albiger B, et al. An endonuclease allows *Streptococcus pneumoniae* to escape from neutrophil extracellular traps. *Curr Biol.* Feb 21; 2006 16(4):401–7. [PubMed: 16488875]
60. Xu J, Zhang X, Pelayo R, et al. Extracellular histones are major mediators of death in sepsis. *Nat Med.* Nov; 2009 15(11):1318–21. [PubMed: 19855397] ** This article provides elegant experimental evidence for the cleavage of extracellular histones by activated protein C in sepsis.
61. Andersson U, Tracey KJ. HMGB1 is a therapeutic target for sterile inflammation and infection. *Annu Rev Immunol.* 2011; 29:139–62. [PubMed: 21219181]
62. Klune JR, Dhupar R, Cardinal J, et al. HMGB1: endogenous danger signaling. *Mol Med.* Jul-Aug; 2008 14(7-8):476–84. [PubMed: 18431461]
63. Hakkim A, Fuchs TA, Martinez NE, et al. Activation of the Raf-MEK-ERK pathway is required for neutrophil extracellular trap formation. *Nat Chem Biol.* Feb; 2011 7(2):75–7. [PubMed: 21170021]
64. McInturff AM, Cody MJ, Elliott EA, et al. Mammalian target of rapamycin regulates neutrophil extracellular trap formation via induction of hypoxia-inducible factor 1 alpha. *Blood.* Oct 11; 2012 120(15):3118–25. [PubMed: 22919032]
65. Li P, Li M, Lindberg MR, et al. PAD4 is essential for antibacterial innate immunity mediated by neutrophil extracellular traps. *J Exp Med.* Aug 30; 2010 207(9):1853–62. [PubMed: 20733033] * This article highlights histone hypercitrullination as a prerequisite for NET-formation.
66. Neeli I, Khan SN, Radic M. Histone deimination as a response to inflammatory stimuli in neutrophils. *J Immunol.* Feb 1; 2008 180(3):1895–902. [PubMed: 18209087]
67. Saffarzadeh M, Juenemann C, Queisser MA, et al. Neutrophil extracellular traps directly induce epithelial and endothelial cell death: a predominant role of histones. *PLoS One.* 2012; 7(2):e32366. [PubMed: 22389696]
68. Caudrillier A, Kessenbrock K, Gilliss BM, et al. Platelets induce neutrophil extracellular traps in transfusion-related acute lung injury. *J Clin Invest.* Jul 2; 2012 122(7):2661–71. [PubMed: 22684106] * This is the first report focusing on the role of NETs during ALI/ARDS.
69. Abrams ST, Zhang N, Manson J, et al. Circulating Histones Are Mediators of Trauma-associated Lung Injury. *Am J Respir Crit Care Med.* Jan 15; 2013 187(2):160–9. [PubMed: 23220920] * This article nicely combines experimental and clinical data suggesting that the lung is the most susceptible organ to trauma-induced release of histones.
70. Allam R, Scherbaum CR, Darisipudi MN, et al. Histones from dying renal cells aggravate kidney injury via TLR2 and TLR4. *J Am Soc Nephrol.* Aug; 2012 23(8):1375–88. [PubMed: 22677551]
71. Xu J, Zhang X, Monestier M, et al. Extracellular histones are mediators of death through TLR2 and TLR4 in mouse fatal liver injury. *J Immunol.* Sep 1; 2011 187(5):2626–31. [PubMed: 21784973]
72. Garcia CC, Weston-Davies W, Russo RC, et al. Complement C5 activation during influenza A infection in mice contributes to neutrophil recruitment and lung injury. *PLoS One.* 2013; 8(5):e64443. [PubMed: 23696894]
73. Fuchs TA, Brill A, Duerschmied D, et al. Extracellular DNA traps promote thrombosis. *Proc Natl Acad Sci U S A.* Sep 7; 2010 107(36):15880–5. [PubMed: 20798043]

74. von Bruhl ML, Stark K, Steinhart A, et al. Monocytes, neutrophils, and platelets cooperate to initiate and propagate venous thrombosis in mice in vivo. *J Exp Med*. Apr 9; 2012 209(4):819–35. [PubMed: 22451716]
75. Fuchs TA, Kremer Hovinga JA, Schatzberg D, et al. Circulating DNA and myeloperoxidase indicate disease activity in patients with thrombotic microangiopathies. *Blood*. Aug 9; 2012 120(6):1157–64. [PubMed: 22611154]
76. Li MO, Wan YY, Sanjabi S, et al. Transforming growth factor-beta regulation of immune responses. *Annu Rev Immunol*. 2006; 24:99–146. [PubMed: 16551245]
77. Rifkin DB. Latent transforming growth factor-beta (TGF-beta) binding proteins: orchestrators of TGF-beta availability. *J Biol Chem*. Mar 4; 2005 280(9):7409–12. [PubMed: 15611103]
78. Stockis J, Colau D, Coulie PG, et al. Membrane protein GARP is a receptor for latent TGF-beta on the surface of activated human Treg. *Eur J Immunol*. Dec; 2009 39(12):3315–22. [PubMed: 19750484]
79. Pittet JF, Griffiths MJ, Geiser T, et al. TGF-beta is a critical mediator of acute lung injury. *J Clin Invest*. Jun; 2001 107(12):1537–44. [PubMed: 11413161] * This article demonstrated that integrin $\alpha\text{v}\beta\text{6}$ activates latent TGF β for promoting ALI.
80. Schultz-Cherry S, Murphy-Ullrich JE. Thrombospondin causes activation of latent transforming growth factor-beta secreted by endothelial cells by a novel mechanism. *J Cell Biol*. Aug; 1993 122(4):923–32. [PubMed: 8349738]
81. Barcellos-Hoff MH, Dix TA. Redox-mediated activation of latent transforming growth factor-beta 1. *Mol Endocrinol*. Sep; 1996 10(9):1077–83. [PubMed: 8885242]
82. Karsdal MA, Larsen L, Engsig MT, et al. Matrix metalloproteinase-dependent activation of latent transforming growth factor-beta controls the conversion of osteoblasts into osteocytes by blocking osteoblast apoptosis. *J Biol Chem*. Nov 15; 2002 277(46):44061–7. [PubMed: 12226090]
83. Wrana JL, Attisano L, Wieser R, et al. Mechanism of activation of the TGF-beta receptor. *Nature*. Aug 4; 1994 370(6488):341–7. [PubMed: 8047140]
84. Wang XF, Lin HY, Ng-Eaton E, et al. Expression cloning and characterization of the TGF-beta type III receptor. *Cell*. Nov 15; 1991 67(4):797–805. [PubMed: 1657407]
85. Heldin CH, Miyazono K, ten Dijke P. TGF-beta signalling from cell membrane to nucleus through SMAD proteins. *Nature*. Dec 4; 1997 390(6659):465–71. [PubMed: 9393997]
86. Birukova AA, Adyshev D, Gorshkov B, et al. ALK5 and Smad4 are involved in TGF-beta1-induced pulmonary endothelial permeability. *FEBS Lett*. Jul 18; 2005 579(18):4031–7. [PubMed: 16004987]
87. Perlman R, Schiemann WP, Brooks MW, et al. TGF-beta-induced apoptosis is mediated by the adapter protein Daxx that facilitates JNK activation. *Nat Cell Biol*. Aug; 2001 3(8):708–14. [PubMed: 11483955]
88. Hannigan M, Zhan L, Ai Y, et al. The role of p38 MAP kinase in TGF-beta1-induced signal transduction in human neutrophils. *Biochem Biophys Res Commun*. May 8; 1998 246(1):55–8. [PubMed: 9600067]
89. Shull MM, Ormsby I, Kier AB, et al. Targeted disruption of the mouse transforming growth factor-beta 1 gene results in multifocal inflammatory disease. *Nature*. Oct 22; 1992 359(6397):693–9. [PubMed: 1436033] * This article showed that endogenous TGF β is required to prevent lethal spontaneous inflammation in mice.
90. Murray LA, Chen Q, Kramer MS, et al. TGF-beta driven lung fibrosis is macrophage dependent and blocked by Serum amyloid P. *Int J Biochem Cell Biol*. Jan; 2011 43(1):154–62. [PubMed: 21044893]
91. Frank J, Roux J, Kawakatsu H, et al. Transforming growth factor-beta1 decreases expression of the epithelial sodium channel αENaC and alveolar epithelial vectorial sodium and fluid transport via an ERK1/2-dependent mechanism. *J Biol Chem*. Nov 7; 2003 278(45):43939–50. [PubMed: 12930837]
92. Pittet JF, Koh H, Fang X, et al. HMGB1 accelerates alveolar epithelial repair via an IL-1beta and $\alpha\text{v}\beta\text{6}$ integrin-dependent activation of TGF-beta1. *PLoS One*. 2013; 8(5):e63907. [PubMed: 23696858]

93. Bartram U, Speer CP. The role of transforming growth factor beta in lung development and disease. *Chest*. Feb; 2004 125(2):754–65. [PubMed: 14769761]
94. Adamali HI, Maher TM. Current and novel drug therapies for idiopathic pulmonary fibrosis. *Drug Des Devel Ther*. 2012; 6:261–72.
95. Postlethwaite AE, Keski-Oja J, Moses HL, et al. Stimulation of the chemotactic migration of human fibroblasts by transforming growth factor beta. *J Exp Med*. Jan 1; 1987 165(1):251–6. [PubMed: 3491869]
96. Desmouliere A, Geinoz A, Gabbiani F, et al. Transforming growth factor-beta 1 induces alpha-smooth muscle actin expression in granulation tissue myofibroblasts and in quiescent and growing cultured fibroblasts. *J Cell Biol*. Jul; 1993 122(1):103–11. [PubMed: 8314838]
97. Willis BC, Liebler JM, Luby-Phelps K, et al. Induction of epithelial-mesenchymal transition in alveolar epithelial cells by transforming growth factor-beta1: potential role in idiopathic pulmonary fibrosis. *Am J Pathol*. May; 2005 166(5):1321–32. [PubMed: 15855634]
98. Zhang HY, Phan SH. Inhibition of myofibroblast apoptosis by transforming growth factor beta(1). *Am J Respir Cell Mol Biol*. Dec; 1999 21(6):658–65. [PubMed: 10572062]
99. Kramann R, DiRocco DP, Humphreys BD. Understanding the origin, activation and regulation of matrix-producing myofibroblasts for treatment of fibrotic disease. *J Pathol*. Nov; 2013 231(3): 273–89. [PubMed: 24006178]
100. Garcia-Alvarez J, Ramirez R, Checa M, et al. Tissue inhibitor of metalloproteinase-3 is up-regulated by transforming growth factor-beta1 in vitro and expressed in fibroblastic foci in vivo in idiopathic pulmonary fibrosis. *Exp Lung Res*. May; 2006 32(5):201–14. [PubMed: 16908447]
101. Taylor LM, Khachigian LM. Induction of platelet-derived growth factor B-chain expression by transforming growth factor-beta involves transactivation by Smads. *J Biol Chem*. Jun 2; 2000 275(22):16709–16. [PubMed: 10828062]
102. Dhainaut JF, Charpentier J, Chiche JD. Transforming growth factor-beta: a mediator of cell regulation in acute respiratory distress syndrome. *Crit Care Med*. Apr; 2003 31(4 Suppl):S258–64. [PubMed: 12682450]
103. Peng R, Sridhar S, Tyagi G, et al. Bleomycin induces molecular changes directly relevant to idiopathic pulmonary fibrosis: a model for “active” disease. *PLoS One*. 2013; 8(4):e59348. [PubMed: 23565148]
104. STX-100 in Patients With Idiopathic Pulmonary Fibrosis (IPF). <http://clinicaltrials.gov/ct2/show/study/NCT01371305>
105. Nauseef WM. Editorial: Nyet to NETs? A pause for healthy skepticism. *J Leukoc Biol*. Mar; 2012 91(3):353–5. [PubMed: 22379074] * This article provides a refreshing comment on the current field of research on NETs.
106. Buchanan JT, Simpson AJ, Aziz RK, et al. DNase expression allows the pathogen group A *Streptococcus* to escape killing in neutrophil extracellular traps. *Current biology* : CB. Feb 21; 2006 16(4):396–400. [PubMed: 16488874]
107. Bosch X, Poch E, Grau JM. Rhabdomyolysis and acute kidney injury. *N Engl J Med*. Jul 2; 2009 361(1):62–72. [PubMed: 19571284]
108. Hirsch JG. Bactericidal action of histone. *The Journal of experimental medicine*. Dec 1; 1958 108(6):925–44. [PubMed: 13598820]
109. Hillmen P, Young NS, Schubert J, et al. The complement inhibitor eculizumab in paroxysmal nocturnal hemoglobinuria. *N Engl J Med*. Sep 21; 2006 355(12):1233–43. [PubMed: 16990386]
110. Smirnova I, Poltorak A, Chan EK, et al. Phylogenetic variation and polymorphism at the toll-like receptor 4 locus (TLR4). *Genome biology*. 2000; 1(1) RESEARCH002.
111. Looney MR, Gropper MA, Matthay MA. Transfusion-related acute lung injury: a review. *Chest*. Jul; 2004 126(1):249–58. [PubMed: 15249468]
112. Matute-Bello G, Frevert CW, Martin TR. Animal models of acute lung injury. *Am J Physiol Lung Cell Mol Physiol*. Sep; 2008 295(3):L379–99. [PubMed: 18621912]
113. Bosmann M, Grailer JJ, Zhu K, et al. Anti-inflammatory effects of beta2 adrenergic receptor agonists in experimental acute lung injury. *FASEB J*. May; 2012 26(5):2137–44. [PubMed: 22318967]

114. Corbel M, Lagente V, Theret N, et al. Comparative effects of betamethasone, cyclosporin and nedocromil sodium in acute pulmonary inflammation and metalloproteinase activities in bronchoalveolar lavage fluid from mice exposed to lipopolysaccharide. *Pulmonary pharmacology & therapeutics*. 1999; 12(3):165–71. [PubMed: 10419836]
115. Singh B, Tiwari AK, Singh K, et al. beta2 Agonist for the Treatment of Acute Lung Injury: A Systematic Review and Meta-analysis. *Respiratory care*. Feb; 2014 59(2):288–96. [PubMed: 23777655]
116. Taylor RW, Zimmerman JL, Dellinger RP, et al. Low-dose inhaled nitric oxide in patients with acute lung injury: a randomized controlled trial. *JAMA*. Apr 7; 2004 291(13):1603–9. [PubMed: 15069048]
117. Abraham E, Baughman R, Fletcher E, et al. Liposomal prostaglandin E1 (TLC C-53) in acute respiratory distress syndrome: a controlled, randomized, double-blind, multicenter clinical trial. TLC C-53 ARDS Study Group. *Crit Care Med*. Aug; 1999 27(8):1478–85. [PubMed: 10470753]

Article highlights

- Acute lung injury (ALI) is among the leading causes of morbidity and mortality worldwide.
- While advances in critical care medicine (e.g. low-tidal volume ventilation) have somewhat improved the outcomes of ALI/ARDS, no specific pharmacologic therapies are currently available to ameliorate the adverse consequences of ALI.
- Many studies support the idea of an essential role for the complement cleavage product, C5a, during ALI. The knowledge on the precise molecular effector functions of how C5a-modulated inflammation is translated into organ dysfunction have been recently expanded.
- It has recently been uncovered that C5a mediates the appearance of extracellular histones in broncho-alveolar lavage fluids (BALF) during experimental ALI.
- Extracellular histones, which are major components of neutrophil extracellular traps (NETs), are detectable in plasma and BALF of human patients with ALI/ARDS in humans.
- Extracellular histones / NETs compromise lung function and alveolar gas exchange by displaying cytotoxic activity for alveolar epithelial cells / endothelial cells, and promoting acute inflammation and the accumulation of lung edema fluids.
- Neutralization of extracellular histones using anti-H4 antibodies reduces the severity of experimental ALI as induced by C5a or transfusions (TRALI).
- Extracellular histones are a substrate for enzymatic degradation by activated protein C (APC), but this inactivation process may not be sufficient to protect from ALI/ARDS in clinical settings.
- NETs interact with platelets, the latter being major sources of TGF β . TGF β may dominate the fibro-proliferative stages of ALI and may determine the conditional transition to fibrotic conversions in lung architecture.
- Interception of NETs / extracellular histone using neutralizing antibodies or degradation by site specific enzymes may evolve as novel protein-based strategies to cope with ALI/ARDS. Alternatively, interference with up-stream mechanisms (anti-C5a strategies) or down-stream effector proteins (anti-TGF β strategies) could prove helpful for certain patients with ALI/ARDS.

This box summarizes key points contained in the article.

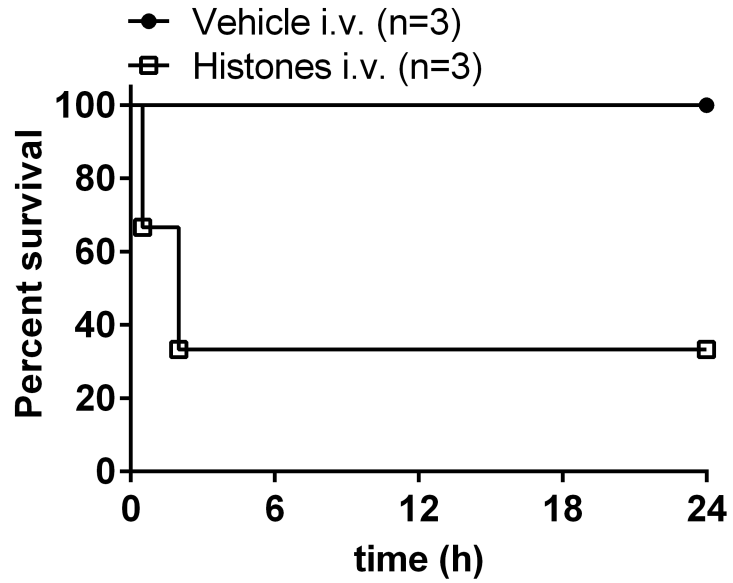


Figure 1. Infusion of extracellular histones (75 mg/kg body weight i.v.) purified from calf thymus mediates lethality in C57BL/6J mice. Death of mice was preceded by clinical signs of respiratory failure. This figure shows data by Bosmann and Ward, which are consistent with reported findings by Xu et al. ⁶⁰.

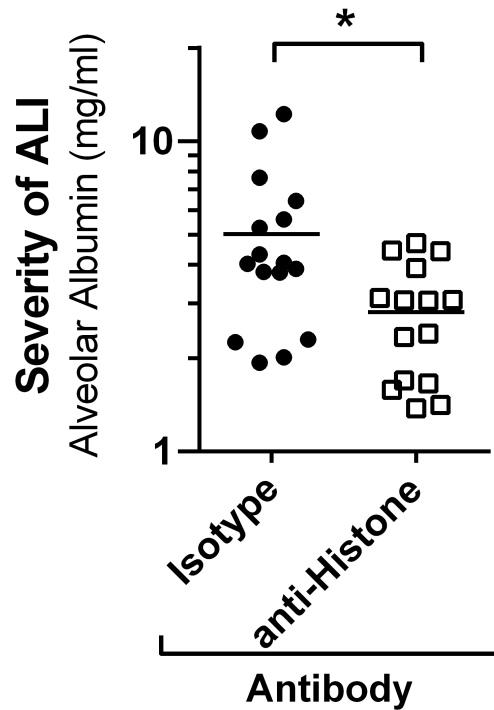


Figure 2.

Neutralization of extracellular histones using monoclonal anti-H4 antibody reduces the severity of ALI in C57BL/6J mice. ALI was induced by intra-tracheal administration of recombinant mouse C5a (500 ng/mouse). Groups of mice were treated with either anti-Histone antibody (300 μ g/mouse) or non-specific matched isotype IgG1 κ antibody (300 μ g/mouse). Severity of ALI was determined by quantification of albumin (ELISA) in broncho-alveolar lavage fluids after 8 h. Data taken from ¹⁷).

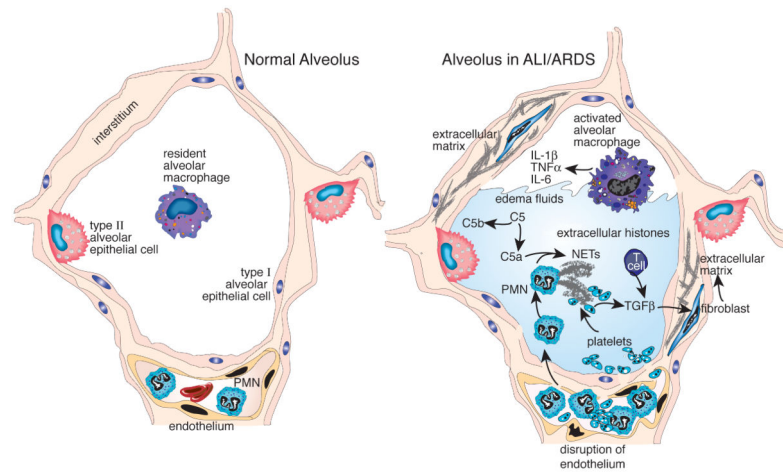


Figure 3.

Current concepts on the pathophysiologic mechanisms of ALI/ARDS involving extracellular histones / NETs, C5a and TGF β . The frame on the left shows the architecture of the alveolus, which is composed of type I and type II alveolar epithelial cells, resident intra-alveolar macrophages and adjacent lung capillaries with intact endothelial lining. The frame on the right displays the injured alveolus in ALI/ARDS: Complement activation products (C5a) and inflammatory mediators released by activated macrophages orchestrate the influx of PMNs, monocytes and adaptive immune cells to the alveolar compartment. C5a promotes release of NETs and extracellular histones, thereby resulting in tissue damage and disruption of the epithelial/endothelial barrier. Intra-alveolar hemorrhage includes the presence of platelets, which interact with NETs and release TGF β . The later phases of ALI/ARDS may include TGF β -mediated fibro-proliferative responses and accumulation of extracellular matrix.

TABLE 1

Selected clinical trials without benefit in patients with ALI/ARDS

References	ARDS patients	Pharmacologic intervention	Result
25	n = 130	Recombinant GM-CSF	No differences in 28-day survival or ventilator-free days
24	n = 75	Recombinant Activated Protein C	No differences in 60-day survival or ventilator-free days
26	n = 448	Recombinant Surfactant Protein C	No differences in 28-day survival or ventilator-free days
7	n = 282	β 2-Agonists (inhalation)	No difference in ventilator-free days or survival before hospital discharge
27	n = 324	β 2-Agonists (intravenous)	No differences in 28-day survival
116	n = 385	Nitric oxide (inhalation)	No differences in survival and ventilator-free days
117	n = 348	Prostaglandin E1 (Liposomes)	No differences in 28-day survival or ventilator-free days
28 / 30	n = 99 / n = 180	Methylprednisolone	No differences in 45-day survival / No differences in 180-day survival
6	n = 272	Omega-3-fatty acids	No differences in 60-day survival
29	n = 46	N-Acetylcysteine / Procysteine	No differences in survival

TABLE 2

Studies on the role of NETs / extracellular histones during ALI

Reference	Species	Disease Model	Major findings
Caudrillier et al. ⁶⁸	<i>M. musculus</i> , <i>H. sapiens</i>	Transfusion-induced ALI (TRALI), clinical samples	-Detection of NETs in lungs and plasma of ALI patients -Blockade of extracellular histones is protective in TRALI -Activated platelets induce NETs during TRALI
Abrams et al. ⁶⁹	<i>H. sapiens</i> , <i>M. musculus</i>	Clinical samples	-Circulating histones in patients with non-thoracic trauma increase the risk for ALI -Histones induce cytokine release -Histones are toxic for endothelial cells
Bosmann et al. ¹⁷	<i>M. musculus</i> , <i>H. sapiens</i>	C5a-induced ALI, clinical samples	-C5a mediates appearance of extracellular histones in lungs during ALI -Blockade of histones is protective in complement-induced ALI -Intra-tracheal administration of purified histones precipitates symptoms of ALI -Detection of extracellular histones in BALF of patients with ALI
Saffarzadeh et al. ⁶⁷	<i>H. sapiens</i> , <i>M. musculus</i>	LPS-induced ALI, cell cultures	-NETs are cytotoxic for human alveolar epithelial cells and endothelial cells