GASTRIC CANCER



Apoptosis induced by preoperative oral 5'-DFUR administration in gastric adenocarcinoma and its mechanism of action

Wen-He Zhao, Shi-Fu Wang, Wei Ding, Jian-Ming Sheng, Zhi-Min Ma, Li-Song Teng, Min Wang, Fu-Sheng Wu, Bing Luo

Wen-He Zhao, Wei Ding, Jian-Ming Sheng, Zhi-Min Ma, Li-Song Teng, Min Wang, Fu-Sheng Wu, Department of Oncological Surgery, The First Affiliated Hospital of Medical College, Zhejiang University, Hangzhou 310003, Zhejiang Province, China

Telephone: +86-751-87236880

Abstract

AIM: To study the apoptosis induced by preoperative oral 5'-DFUR administration in gastric adenocarcinoma and its mechanism of action.

METHODS: Sixty gastric cancer patients were divided randomly into three groups (20 each group) before operation: group one: 5'-DFUR oral administration at the dose of 800-1200mg/d for 3 - 5 d, group two: 500mg 5-FU + 200 mg/d CF by venous drip for 3 - 5 d, group three (control group). One or two days after chemotherapy, the patients were operated. Fas/FasL, PD-ECGF and PCNA were examined by immunohistochemistry and apoptotic tumor cells were detected by *in situ* TUNEL method. Fifty-four patients received gastrectomy, including 12 palliative resections and 42 radical resections. Six patients were excluded. Finally 18 cases in 5'-DFUR group, 16 cases in CF + 5-FU group, and 20 cases in control group were analyzed.

RESULTS: There was no significant difference in patient mean age, gender, white blood cell count, haematoglobin (HB), thromboplastin, perioperative complication incidence, radical or palliation resection, invasion depth (T), lymphonode involvement (N), metastasis (M) and TNM staging among the three groups. However, the PCNA index (PI) in 5'-DFUR group (40.51 ± 12.62) and 5-FU+CF group (41.12 ± 15.26) was significantly lower than that in control group (58.33 ± 15.69) (F = 9.083, P = 0.000). The apoptotic index (AI) in 5'-DFUR group (14.39 ± 9.49) and 5-FU+CF group (14.11 ± 9.68) was significantly

higher than that in control group (6.88 ± 7.37) (F = 4.409, P = 0.017). The expression rates of Fas and FasL in group one and group three were 66.7% (12/18) and 50% (9/18), 43.8% (7/16) and 81.3% (13/16), 45.0% (9/20) and 85% (17/20), respectively. The expression rate of FasL in 5'-DFUR group was significantly lower than that in the other two groups (χ^2 =6.708, *P*=0.035). Meanwhile, the expression rate of PD-ECGF was significantly lower in 5'-DFUR group (4/18, 28.6%) than in CF+5-FU group (9/16, 56.3%) and control group (13/20, 65.0%) ($\chi^2 = 7.542, P = 0.023$). The frequency of Fas expression was significantly correlated with palliative or radical resection (χ^2 = 7.651, P = 0.006), invasion depth $(\chi^2 = 8.927, P = 0.003)$, lymphatic spread $(\chi^2 = 4.488,$ P = 0.034) and UICC stages ($\chi^2 = 8.063$, P = 0.045) respectively. By the end of March 2005, 45 patients were followed up. The 0.5-, 1-, 2-, 3-year survival rates were 96%, 73%, 60%, 48%, respectively, which were related with T, N, M and Fas expression, but not with PD-ECGF and FasL expression.

CONCLUSION: Preoperative oral 5'-DFUR administration may induce apoptosis of gastric carcinoma cells and decrease tumor cell proliferation index, but cannot improve the prognosis of patients with gastric cancer. Downregulation of FasL and PD-ECGF expression mediated by 5'-DFUR may be one of its anti-cancer mechanisms. Fas expression correlates with the progression of gastric carcinoma and may be an effective prognostic factor.

© 2006 The WJG Press. All rights reserved.

Key words: Apoptosis; Preoperative chemotherapy; 5' -DFUR; Gastric carcinoma

Zhao WH, Wang SF, Ding W, Sheng JM, Ma ZM, Teng LS, Wang M, Wu FS, Luo B. Apoptosis induced by preoperative oral 5'-DFUR administration in gastric adenocarcinoma and its mechanism of action. *World J Gastroenterol* 2006; 12(9): 1356-1361

http://www.wjgnet.com/1007-9327/12/1356.asp

INTRODUCTION

Cell apoptosis and proliferation regulate homeostasis.

Shi-Fu Wang, Bing Luo, The Third People's Hospital of Wenling, Taizhou 317523, Zhejiang Province, China

Supported by Natural Science Foundation of Zhejiang Province, No. 20010536

Correspondence to: Professor Wen-He Zhao, Department of Oncological Surgery, the First Affiliated Hospital of Medical College, Zhejiang University, Hangzhou 310003, Zhejiang Province, China. zhaowh@mail.hz.zj.cn

Received: 2005-08-31 Accepted: 2005-10-10

Disordered balance results in tumorigenesis. Cell apoptosis can hinder tumor growth^[1-3]. Therefore, induction of tumor apoptosis is a good anti-cancer therapy. At present, the relationship between chemotherapy and cancer cell apoptosis has drawn more and more attention. However, studies on induction of apoptosis in gastric carcinoma are performed in vitro. Inducers reported include y-ray, betaionone, biological response modifiers, chemotherapeutics, ett^[4-8]. Chemotherapeutics-induced apoptosis is one of its anti-cancer mechanisms. Arsenic trioxide^[9], hydroxycamptothecin^[10], cisplatin^[11], paclitaxel^[12], fluorouracil and its derivant^[13,14], oxaliplatin^[15] can induce apoptosis in human gastric carcinoma cells. 5'-deoxy-5-flurouridine (doxifluridine or 5'-DFUR (known as Furtulon) is a selective anticancer medicine, which can be converted into 5-FU by thymidine phosphorylase (the same substance as traversing platelet-derived endothelial cell growth factor, PD-EC-GF)^[14,19,20]. Accordingly, the concentration of 5-FU is high in tumor tissue^[19, 21]. The therapeutic index of doxifluridine is ten times that of 5-FU^[19]. Doxifluridine has been widely used in treatment of breast cancer, colorectal carcinoma, ovarian adenocarcinoma, bladder cancer and gastric carcinoma^[22-26].

In this study, we used doxifluridine as an apoptosis inducer to study the change of apoptosis and expression of proliferative cell nuclear antigen (PCNA), Fas and Fas ligand (FasL), and PD-ECGF in gastric adenocarcinoma and its mechanism of action.

MATERIALS AND METHODS

Patients

Patients who were diagnosed as malignant gastric neoplasm (age ≤ 70 years, Karnofsky's scale >90) and could endure chemotherapy and operation, were enrolled in this study. The patients were divided into three groups (20 each): group 1: 800 mg - 1 200 mg/d 5'-DFUR for 3-5 days, group 2: 500 mg 5-FU + 200 mg/d CF by venous drip for 3-5 d and group 3 (control group). One or two days after chemotherapy, the patients underwent surgery. From Oct. 2001 to Oct. 2003, 60 gastric cancer patients (20 in each group) were enrolled (37 males and 17 female, mean age 57.5 years, range 32-70 years) .

Groups

Of the 60 patients, 54 underwent gastrectomy (including 12 palliative resections and 42 radical resections, and 6 were excluded. No perioperative mortality occurred during the first 30 days after surgery. Complications were recorded in four of 54 patients resected, one gastric perforation on the fourth day after 5'-DFUR oral chemotherapy, one preoperative gastrorrhagia and one postoperative cerebral infarction in CF+5-FU group, one venous thrombosis of lower extremities in control group complicated by cerebral hemorrhage during thrombolysis treatment. The patients with gastric perforation and gastrorrhagia required emergency laparotomy.

Immunohistochemistry

PCNA, Fas/FasL and PD-ECGF expression was determined

by En Vision immunohistochemistry^[27]. The kit was purchased from DAKO Co., USA. Apoptotic cells were detected by terminal deoxynucleotidyl transferase (TdT)mediated dUTP nick end labeling (TUNEL) method. The kit was purchased from Bochringer Mannheim Co., Germany. The proliferation index (PI) and apoptotic index (AI) of gastric carcinoma cells were evaluated by PCNA immunohistochemical staining and *in situ* TUNEL.

Result assessment

The stained cells had Fas/FasL or PD-ECGF positive expression. Brown-stained nuclei were considered as positive cells of PCNA and apoptosis, brown cellular membrane and cytoplasm were considered as Fas or FasL positive cells (Figures 1A and 1B), brown or yellow staining in cytoplasm and/or cell nuclei was considered as PD-ECGF positive cells (Figure 1C). PCNA index (PI) and apoptotic index (AI) of positive cells in 1000 tumor cells were calculated under high power field (*400) of microscope.

AI = TUNEL mark cells/tumor cells (more than one thousand)*100%

PI = PCNA mark cells/tumor cells (more than one thousand)*100%

Statistical analysis

All data were analyzed by ANOVA and chi square test. P < 0.05 was considered statistically significant.

RESULTS

Patients and groups

Fifty-four patients underwent surgical resection, including 18 cases in 5'-DFUR group, 16 cases in CF+5-FU group, and 20 cases in control group. There was no significant differences in patient mean age, gender, white blood cell count, haematoglobin (HB), thromboplastin, perioperative complication incidence, radical or palliation resection, invasion depth(T), lymphonode involvement(N), metastasis (M) and TNM staging among the three groups.

AI and expression of PCNA, Fas/ FasS-L, PD-ECGF in gastric carcinoma

The expression of PI, AI, FasL and PD-ECGF was not statistically significant as compared to the operation procedure (radical or palliation resection), early or advanced tumor, T, N, M and UICC stages. In contrast, positive staining of Fas was closely related to radical resection but not with serosal invasion, lymphnode metastasis and early UICC stages (Table 1, Figure 1).

Influence of preoperative chemotherapy on expression of AI, PI, Fas/FasL and PD-ECGF

Either 5'-DFUR or CF+5-FU preoperative chemotherapy could significantly inhibit cell proliferation and induce apoptosis as compared to control group. The frequency of Fas expression had no significant difference among three groups. However, the frequency of FasL expression was significantly lower in 5'-DFUR group (50%) than those in CF+5-FU group (81.3%) and control group (85%). There was no significant difference between groups two and Table 1 Expression of PI, AI, Fas/ FasL compared to operation procedure, early or advanced tumor, T, N, M and UICC stages

	PI (9()	AI	Fas	FasL	PD-ECGF	
	(%)	(%)	+ -	+ -	+ -	
Way of resection						
radical	46.31±15.30	12.13±9.65	26 16	29 13	19 23	
Palliation	50.74±21.27	9.40±8.35	2 10	10 2	7 5	
P/χ^2 Value P Value	0.654	0.791	7.651 0.006	0.949	0.641	
	0.422	0.378	0.006	0.33	0.423	
Early cancer Yes	45.29±12.34	14.54±12.38	7 2	6 3	3 6	
No	45.29±12.34 47.70±17.51	14.54 ± 12.58 10.92 ± 8.70	21 24	33 12	23 22	
F/χ^2 Value	47.70±17.51 0.154	1.123	2.908	0.166	0.949	
P Value	0.697	0.294	0.088	0.684	0.33	
Invasion depth	0.097	0.294	0.000	0.004	0.55	
T1-2	46.65±15.76	11.96±9.81	20 8	19 9	12 16	
T3-4	47.99±17.92	11.06±9.04	8 18	20 6	14 12	
F/χ^2 Value	0.086	0.121	8.927	0.552	0.652	
P Value	0.770	0.729	0.003	0.457	0.419	
Lymphonode						
N0	50.37±16.34	10.51±9.89	13 5	14 4	9 9	
N1-4	45.76±16.87	12.03±9.20	15 21	25 11	17 19	
F/χ^2 value	0.917	0.312	4.488	0.415	0.037	
P value	0.343	0.579	0.034	0.519	0.847	
Metastasis						
M0	46.73±15.79	11.51±9.10	26 19	33 12	22 23	
M1	50.10±21.5	11.60±11.24	7 2	6 3	4 5	
F/χ^2 value	0.301	0.001	3.798	0.166	0.059	
P value	0.585	0.980	0.051	0.684	0.808	
UICC stage						
Ι	50.42±16.84	10.98±9.99	13 4	13 4	9 8	
II	42.69±13.24	11.78±10.65	5 4	6 3	2 7	
III	46.03±16.2	12.73±10.10	7 9	11 5	7 9	
IV	48.01±20.18	10.52±7.25	3 9	9 3	8 4	
F/χ^2 value	0.452	0.148	8.063	0.434	4.352	
P value	0.717	0.931	0.045	0.933	0.226	

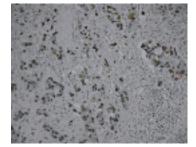


Figure 1 The expression of PCNA (*200).

three. The PD-ECGF expression was detected in 28.6% of 5'-DFUR group, 56.3% of CF+5-FU group and 65.0% of control group (Table 2).

Prognosis

By the end of March 2005, 45 patients were followed up. During the follow-up, 22 cases died, 23 remained alive (including one patient with cerebral hemorrhage after operation). The 0.5-, 1-, 2-, 3-year survival rates were 96%, 73%, 60%, 48% respectively, which were related with T (χ^2 =30.32, P=0.0000), N (χ^2 =22.10, P=0.0000), M (χ^2 =17.04, P=0.0000) and Fas expression (χ^2 = 12.24, P=0.0005, Figure 2), but not with PD-ECGF (χ^2 = 0.78, P=0.3775) and FasL expression (χ^2 =0.7, P=0.7967).

DISCUSSION

Gastric carcinoma is one of the most common malignant tumors and remains a leading cause of cancer-related death worldwide ^[15, 16, 28-30]. In China, it still ranks the first^[31, 32]. Operative resection is the most effective treatment nowadays. However, the 5-year survival rate was 25%~60% even after radical resection^[33-36], micrometastasis is the major reason for recurrence^[29,37,38]. Preoperation chemotherapy can inhibit micrometastasis, but strong chemotherapy may potentially increase operation compli-cations in resectable patients^[30,35,36]. It was reported that surgical resection may serve as a stimulus for the growth of residual tumor^[30]. Inada *et al*^{13]} have reported the effectiveness of preoperative 5-Fu venous chemotherapy, which can induce cancer cell apoptosis in vivo. Our study showed that preoperative venous CF+5-Fu chemotherapy and 5' -DFUR oral administration could induce gastric carcinoma cell apoptosis and inhibit cancer cell proliferation. Wang et al^{22} reported that preoperative 5'-DFUR chemotherapy is able to partially inhibit the expression of extracellular signal-regulated kinase (ERK) which is closely related with cell proliferation in breast cancer. PCNA index decrease in our studies might be related with inhibition of ERK-1 and ERK-2 expression. Liang et al^[39] studied apoptosis in ovarian cancer and found that apoptosis induced by che-

Table 2 Influence of preoperative chemotherapy on expression of Fas/FasL, PD-ECGF and AI, PI												
Group	N	Fas		FasL		PI	AI	PD-ECGF				
		+	-	+	-	(%)	(%)	+	-			
5'-DFUR	18	12	6	9	9	40.51 ± 12.62	14.39 ± 9.49	4	14			
CF+5-FU	16	7	9	13	3	41.12 ± 15.26	14.11 ± 9.68	9	7			
Control	20	9	11	17	3	58.33 ± 15.69	6.88 ± 7.37	13	7			
X2/F value	e	2.3	79	6.7	08	9.083	4.409	7.5	542			
P value		0.3	0.035		0	0.017	0.023					

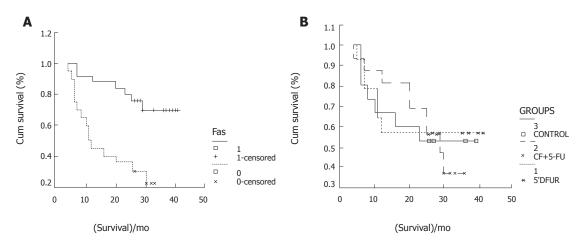


Figure 2 Fas expression (A) and survival rate (B) in three groups.

motherapy is decreased, suggesting that apoptosis induced by chemotherapy has a time limit. Therefore, preoperative chemotherapy-induced tumor cell apoptosis can inhibit malignant behavior in some degree.

Fas(CD95/APO-1)/Fas ligand (FasL) system is one of the major apoptotic pathways and plays an important role in maintenance of cell colony, elimination of malignant transformation cells and regulation of immune system^[1-3, 40]. In general, Fas/FasL system plays an important role in prognosis and immune escape. Fas of activated T lymphocytes and tumor-infiltrating lymphocytes(TIL) can increase apoptosis^[1-3,40-45]. Tumor cells combined with activated T lymphocytes and TIL can kill surrounding normal infiltrating lymphocytes, escape immune system^[1,2,22,23,43,44]. Our investigation showed that preoperative 5'-DFUR oral administration chemotherapy could down regulate FasL expression, which may be one of its mechanisms underlying tumor cell apoptosis. But CF+5-FU venous chemotherapy had no influence on Fas/FasL expression, suggesting that 5'-DFUR and 5-FU have a different mechanism of action. Induction of 5-FU is relevant with activation of apoptosis gene bax^[45], and expression of bcl-2 oncogene^[13].

PD-ECGF, one of the angiogenesis factors, extracted from fresh thrombocytolysis matters is the same substance as thymidine phosphorylase(TP), PyNPase^[20,46-49]. It can promote angiogenesis, cell proliferation and inhibit apoptosis^[46-49]. Konno *et al*^[47] reported that expression of PD-ECGF in gastric carcinoma has a positive correlation with PCNA, and PD-ECGF can also promote tumor growth. Osaki *et al*^[20] found that increased PD-ECGF expression is closely related with decreased apoptosis in gastric carcimoma. The mean apoptotic index in early and advanced gastric cancer is significantly lower in positive PD-ECGF than in negative PD-ECGF. Koizumi et $at^{[21]}$ determined PD-ECGF expression in gastric carcinoma and found that the responsive rate to chemotherapy is 56.8% in positive PD-ECGF group and 0% in negative group, and 82.4% in PD-ECGF over expression group, indicating that chemosensitivity is closely related with PD-ECGF expression. PD-ECGF can increase the activity of 5-FU and other anti-cancer medicines^[50]. Cytokine such as interferon, can up-regulate PD-ECGF and cytokine levels, thus detecting PD-ECGF level in tumor tissue can predict chemotherapy sensitivity and its efficacy^[18,26,51]. PD-ECGF inhibitors can decrease tumor angiogenesis, prevent infiltration and metastasis^[52]. PD-ECGF is the key enzyme for the metabolism of Fortulon. Satoh B et al¹⁴ reported that by oral administration of 5'-DFUR, 1 200 mg per day for seven days significantly decreases PD-ECGF activity and acidic protein. Our results showed that preoperative oral administration of Fortulon for 3 -5 d can down-regulate PD-ECGF expression and induce cancer cell apoptosis in gastric carcinoma. Over expression of PD-ECGF is an important factor of tumor metastasis^[53]. Our results also showed that patients with PD-ECGF overexpression had a higher tendency toward liver metastasis. For this reason, down regulation of PD-ECGF expression by preoperative 5'-DFUR chemotherapy plays a certain role in preventing postoperative recurrence and metastasis of gastric carcinoma. It was reported that PD-ECGF level is closely related with tumor metastasis^[24,25].

Kabayashi and kimur⁸ performed a multicenter clinical trial of preoperative chemotherapy for gastric cancer and

found that oral administration of 5'-DFUR may induce apoptosis of gastric carcinoma and decrease proliferation index in old patients with advanced gastric cancer, but cannot improve their prognosis. However, Fas expression is often presented in early stage of tumors and shows a better prognosis. Thus, Fas expression in gastric carcinoma may be an effective prognostic factor for survival.

In conclusion, 5'-DFUR or CF+5-FU chemotherapy can induce apoptosis and inhibit proliferation of tumor cells. Down regulation of FasL and PD-ECGF induced by 5'-DFUR may be one of its anti-cancer mechanisms. Fas expression is correlated with progression of gastric carcinoma and may be an effective prognostic factor.

REFERENCES

- Pinti M, Troiano L, Nasi M, Moretti L, Monterastelli E, Mazzacani A, Mussi C, Ventura P, Olivieri F, Franceschi C, Salvioli G, Cossarizza A. Genetic polymorphisms of Fas (CD95) and FasL (CD178) in human longevity: studies on centenarians. *Cell Death Differ* 2002; 9: 431-438
- 2 Chang YC, Xu YH. Expression of Bcl-2 inhibited Fas-mediated apoptosis in human hepatocellular carcinoma BEL-7404 cells. *Cell Res* 2000; 10: 233-242
- 3 Eichhorst ST, Muller M, Li-Weber M, Schulze-Bergkamen H, Angel P, Krammer PH. A novel AP-1 element in the CD95 ligand promoter is required for induction of apoptosis in hepatocellular carcinoma cells upon treatment with anticancer drugs. *Mol Cell Biol* 2000; 20: 7826-7837
- 4 Ramp U, Caliskan E, Mahotka C, Krieg A, Heikaus S, Gabbert HE, Gerharz CD. Apoptosis induction in renal cell carcinoma by TRAIL and gamma-radiation is impaired by deficient caspase-9 cleavage. Br J Cancer 2003; 88: 1800-1807
- 5 Yamamoto M, Maehara Y, Sakaguchi Y, Kusumoto T, Ichiyoshi Y, Sugimachi K. Transforming growth factor-beta 1 induces apoptosis in gastric cancer cells through a p53-independent pathway. *Cancer* 1996; 77: 1628-1633
- 6 Timmermann W, Illert B, Vollmers HP, Krenn V, Ruckle-Lanz H, Wilhelm M, Thiede A. Induction of apoptosis by preoperative passive immunotherapy in resectable stomach carcinoma. *Kongressbd Dtsch Ges Chir Kongr* 2002; **119**: 396-397
- 7 Yan J, Xu YH. Tributyrin inhibits human gastric cancer SGC-7901 cell growth by inducing apoptosis and DNA synthesis arrest. *World J Gastroenterol* 2003; 9: 660-664
- 8 Kobayashi T, Kimura T. Long-term outcome of preoperative chemotherapy with 5'-deoxy-5-fluorouridine (5'-DFUR) for gastric cancer. Gan To Kagaku Ryoho 2000; 27: 1521-1526
- 9 Xing M, Zhang EJ, Ye X. Investigation of the pathway of apoptosis induced by arsenic trioxide in cancer cells. *Zhongguo Yaolixue Tongbao* 2002; **18**: 87-90
- 10 Tu SP, Jiang SH, Tan JH, Zhong J, Qiao MM, Jiang XH, Zhang YP, Yuan YZ, Wu YL, Wu YY. The mechanism of apoptosis induced by hydroxycamptothecin in gastric cancer cells. *Zhon-ghua Xiaohua Zazhi* 1999; 19: 19-21
- 11 **Jones NA,** Turner J, McIlwrath AJ, Brown R, Dive C. Cisplatinand paclitaxel-induced apoptosis of ovarian carcinoma cells and the relationship between bax and bak up-regulation and the functional status of p53. *Mol Pharmacol* 1998; **53**: 819-826
- 12 **Zhou HB**, Zhu JR. Paclitaxel induces apoptosis in human gastric carcinoma cells. *World J Gastroenterol* 2003; **9**: 442-445
- 13 Inada T, Ichikawa A, Igarashi S, Kubota T, Ogata Y. Effect of preoperative 5-fluorouracil on apoptosis of advanced gastric cancer. J Surg Oncol 1997; 65: 106-110
- 14 Satoh B, Ohtoshi M, Ishida Y, Hen-mi K, Kaneko I, Soda M, Sugihara J, Shibagaki F, Iwai N, Nakamura T, Yamasaki T, Matsumoto S. Correlation between pyrimidine nucleoside phosphorylase (PyNPase)/thymidine phosphorylase/plateletderived endothelial cell growth factor and histological prognostic factor, and influence of 5'-deoxy-5-fluorouridine (5' -DFUR) administration on PyNPase activities and serum im-

munosuppressive acidic protein levels. A study group of oral anti-cancer drugs in Seiban/Tajima area. *Gan To Kagaku Ryoho* 1998; **25**: 359-364

- 15 Lin WL, Li DG, Chen Q, Lu HM. Clinical and experimental study of oxaliplatin in treating human gastric carcinoma. *World J Gastroenterol* 2004; 10: 2911-2915
- 16 Ji YB, Gao SY, Ji HR, Kong Q, Zhang XJ, Yang BF. Antineoplastic efficacy of Haimiding on gastric carcinoma and its mechanisms. *World J Gastroenterol* 2004; 10: 484-490
- 17 Liu JR, Chen BQ, Yang BF, Dong HW, Sun CH, Wang Q, Song G, Song YQ. Apoptosis of human gastric adenocarcinoma cells induced by beta-ionone. *World J Gastroenterol* 2004; 10: 348-351
- 18 Fukui T, Matsui K, Kato H, Takao H, Sugiyama Y, Kunieda K, Saji S. Significance of apoptosis induced by tumor necrosis factor-alpha and/or interferon-gamma against human gastric cancer cell lines and the role of the p53 gene. *Surg Today* 2003; 33: 847-853
- 19 Sun XN, Yang QC, Hu JB. Pre-operative radiochemotherapy of locally advanced rectal cancer. World J Gastroenterol 2003; 9: 717-720
- 20 Osaki M, Sakatani T, Okamoto E, Goto E, Adachi H, Ito H. Thymidine phosphorylase expression results in a decrease in apoptosis and increase in intratumoral microvessel density in human gastric carcinomas. *Virchows Arch* 2000; **437**: 31-36
- 21 Koizumi W, Saigenji K, Nakamaru N, Okayasu I, Kurihara M. Prediction of response to 5'-deoxy-5-fluorouridine (5'-DFUR) in patients with inoperable advanced gastric cancer by immunostaining of thymidine phosphorylase/platelet-derived endothelial cell growth factor. Oncology 1999; 56: 215-222
- 22 Wang ZZ, Wang S, Zhu FX, Ye YJ, Yu YX, Qiao XM. Expression of extracellular signal regulated kinase and its relationship with clinicopathological characteristics of breast cancer. *Zhonghua Zhongliu Zazhi* 2002; 24: 360-363
- 23 Xu L, Zhan YQ, Li W, Sun XW. Influence of preoperative chemotherapy with 5-fluoro uracilum and citrovorum factor on apoptosis and proliferation of human gastric carcinoma cells. *Zhonghua Weichang Waike Zazhi* 2003; 6: 47-49
- 24 Ueda M, Fujii H, Yoshizawa K, Kumagai K, Ueki K, Terai Y, Yanagihara T, Ueki M. Effects of sex steroids and growth factors on invasive activity and 5'-deoxy-5-fluorouridine sensitivity in ovarian adenocarcinoma OMC-3 cells. *Jpn J Cancer Res* 1998; 89: 1334-1342
- 25 Nishimura G, Izumi R, Matsuki N, Takeyama S, Konishi K, Fujita H, Miyata R, Sasaki T, Kojima Y, Takada M, Takita Y, Kanno M, Ueno K, Mura T, Asano K, Syouji M, Yamazaki H, Ooto T, Ueda H, Takigawa Y, Sawa T, Miyazaki I. Effect on 5' -deoxy-5-fluorouridine (5'-DFUR) of pyrimidine nucleoside phosphorylase (PyNPase), matrix metalloprotease and serum IAP values. Hokuriku Colorectal Cancer Chemotherapy Study Group. Gan To Kagaku Ryoho 1997; 24: 1947-1952
- 26 Li G, Kawakami S, Kageyama Y, Yan C, Saito K, Kihara K. IFN gamma-induced up-regulation of PD-ECGF/TP enhances the cytotoxicity of 5-fluorouracil and 5'-deoxy-5-fluorouridine in bladder cancer cells. *Anticancer Res* 2002; 22: 2607-2612
- 27 Li L, Zhang WY. Expression and clinical significance of p27(kip1), p16 and proliferating cell nuclear antigen in nasopharyngeal carcinoma. *Zhonghua Binglixue Zazhi* 2003; 32: 347-349
- 28 Bani-Hani KE, Yaghan RJ, Heis HA, Shatnawi NJ, Matalka II, Bani-Hani AM, Gharaibeh KA. Gastric malignancies in Northern Jordan with special emphasis on descriptive epidemiology. World J Gastroenterol 2004; 10: 2174-2178
- 29 Chen XM, Chen GY, Wang ZR, Zhu FS, Wang XL, Zhang X. Detection of micrometastasis of gastric carcinoma in peripheral blood circulation. *World J Gastroenterol* 2004; **10**: 804-808
- 30 Fink U, Stein HJ, Schuhmacher C, Wilke HJ. Neoadjuvant chemotherapy for gastric cancer: update. World J Surg 1995; 19: 509-516
- 31 Sun XD, Mu R, Zhou SY, Dai XD, Qiao YL, Zhang SW, Huangpu XM, Sun J, Li LD, Lu FZ. 1990-1992 mortality of stomach cancer in China. *Zhonghua Zhongliu Zazhi* 2002; 24: 4-8
- 32 **Sun XD**, Mu R, Zhou SY, Dai XD, Zhang SW, Huangpu XM, Sun J, Li LD, Lu FZ, Qiao YL. Analysis of mortality rate of

stomach cancer and its trend in twenty years in China. Zhonghua Zhongliu Zazhi 2004; **26**: 4-9

- 33 Noguchi Y, Imada T, Matsumoto A, Coit DG, Brennan MF. Radical surgery for gastric cancer. A review of the Japanese experience. *Cancer* 1989; 64: 2053-2062
- 34 Nakamura K, Ueyama T, Yao T, Xuan ZX, Ambe K, Adachi Y, Yakeishi Y, Matsukuma A, Enjoji M. Pathology and prognosis of gastric carcinoma. Findings in 10,000 patients who underwent primary gastrectomy. *Cancer* 1992; **70**: 1030-1037
- 35 Ajani JA, Ota DM, Jessup JM, Ames FC, McBride C, Boddie A, Levin B, Jackson DE, Roh M, Hohn D. Resectable gastric carcinoma. An evaluation of preoperative and postoperative chemotherapy. *Cancer* 1991; **68**: 1501-1506
- 36 Ajani JA, Mayer RJ, Ota DM, Steele GD, Evans D, Roh M, Sugarbaker DJ, Dumas P, Gray C, Vena DA. Preoperative and postoperative combination chemotherapy for potentially resectable gastric carcinoma. J Natl Cancer Inst 1993; 85: 1839-1844
- 37 Zhao AL, Li JY, Sun WQ. Detection and significance of lymph node micrometastases in patients with histologically nodenegative gastric carcinoma. *Zhonghua Zhongliu Zazhi* 2000; 22: 222-224
- 38 Zhang XW, Fan P, Yang HY, Yang L, Chen GY. Significance of detecting disseminated tumor cells in peripheral blood of gastric and colorectal cancer patients. *Zhonghua Zhongliu Zazhi* 2003; 25: 66-68
- 39 Liang JF, Liu LY, Cheng SJ. Relation between apoptosis and proliferation or genes expression after chemotherapy in overian epithelial carcinoma. *Zhongguo Aizheng Zazhi* 2002; 12: 26-28
- 40 Ashkenazi A, Dixit VM. Death receptors: signaling and modulation. *Science* 1998; **281:** 1305-1308
- 41 Nagashima H, Mori M, Sadanaga N, Mashino K, Yoshikawa Y, Sugimachi K. Expression of Fas ligand in gastric carcinoma relates to lymph node metastasis. *Int J Oncol* 2001; 18: 1157-1162
- 42 Liu HF, Liu WW, Fang DC, Men RF. Relationship between Fas antigen expression and apoptosis in human gastric carcinoma and adjacent noncancerous tissues. *Huaren Xiaohua Zazhi* 1998; 6: 321-322
- 43 Koyama S, Koike N, Adachi S. Fas receptor counterattack against tumor-infiltrating lymphocytes in vivo as a mechanism of immune escape in gastric carcinoma. J Cancer Res Clin Oncol 2001; 127: 20-26

- 44 Zheng HC, Sun JM, Wei ZL, Yang XF, Zhang YC, Xin Y. Expression of Fas ligand and caspase-3 contributes to formation of immune escape in gastric cancer. World J Gastroenterol 2003; 9: 1415-1420
- 45 Pan C, Feng ZQ, Peng T, Liu NB. Induction of Apoptosis of Gastric Carcinoma Cell Line SGC-7901 by 5-Fluorouracil in Vitro. *Nanjing Yike Daxue Xuebo* 2000; 20: 163-165
- 46 Ogawa K, Konno S, Takebayashi Y, Miura K, Katsube T, Kajiwara T, Aiba M, Aikou T, Akiyama S. Clinicopathological and prognostic significance of thymidine phosphorylase expression in gastric carcinoma. *Anticancer Res* 1999; 19: 4363-4367
- 47 Konno S, Takebayashi Y, Aiba M, Akiyama S, Ogawa K. Clinicopathological and prognostic significance of thymidine phosphorylase and proliferating cell nuclear antigen in gastric carcinoma. *Cancer Lett* 2001; 166: 103-111
- 48 Kimura H, Konishi K, Kaji M, Maeda K, Yabushita K, Miwa A. Correlation between expression levels of thymidine phosphorylase (dThdPase) and clinical features in human gastric carcinoma. *Hepatogastroenterology* 2002; 49: 882-886
- 49 Yoshikawa T, Suzuki K, Kobayashi O, Sairenji M, Motohashi H, Tsuburaya A, Nakamura Y, Shimizu A, Yanoma S, Noguchi Y. Thymidine phosphorylase/platelet-derived endothelial cell growth factor is upregulated in advanced solid types of gastric cancer. Br J Cancer 1999; 79: 1145-1150
- 50 Katayanagi S, Aoki T, Takagi Y, Ito K, Sudo H, Tsuchida A, Koyanagi Y. Measurement of serum thymidine phosphorylase levels by highly sensitive enzyme-linked immunosorbent assay in gastric cancer. Oncol Rep 2003; 10: 115-119
- 51 Hotta T, Taniguchi K, Kobayashi Y, Johata K, Sahara M, Naka T, Watanabe T, Ochiai M, Tanimura H, Tsubota YT. Preoperative endoscopic analysis of thymidine phosphorylase and dihydropyrimidine dehydrogenase in gastrointestinal cancer. Oncol Rep 2004; 11: 1233-1239
- 52 Matsushita S, Nitanda T, Furukawa T, Sumizawa T, Tani A, Nishimoto K, Akiba S, Miyadera K, Fukushima M, Yamada Y, Yoshida H, Kanzaki T, Akiyama S. The effect of a thymidine phosphorylase inhibitor on angiogenesis and apoptosis in tumors. *Cancer Res* 1999; **59**: 1911-1916
- 53 Suda Y, Kuwashima Y, Shioya T, Uchida K, Tanaka Y. The expression of thymidylate synthase and thymidine phosphorylase in the early-stage of gastric cancer. *Gan To Kagaku Ryoho* 1999; 26: 321-327

S-Editor Wang J L-Editor Wang XL E-Editor Ma WH