

RESEARCH ARTICLE

Open Access

# The genome of the Tiger Milk mushroom, *Lignosus rhinocerotis*, provides insights into the genetic basis of its medicinal properties

Hui-Yeng Y Yap<sup>1\*</sup>, Yit-Heng Chooi<sup>2</sup>, Mohd Firdaus-Raih<sup>3</sup>, Shin-Yee Fung<sup>1</sup>, Szu-Ting Ng<sup>4</sup>, Chon-Seng Tan<sup>5</sup> and Nget-Hong Tan<sup>1</sup>

## Abstract

**Background:** The sclerotium of *Lignosus rhinocerotis* (Cooke) Ryvarden or Tiger milk mushroom (Polyporales, Basidiomycota) is a valuable folk medicine for indigenous peoples in Southeast Asia. Despite the increasing interest in this ethnobotanical mushroom, very little is known about the molecular and genetic basis of its medicinal and nutraceutical properties.

**Results:** The *de novo* assembled 34.3 Mb *L. rhinocerotis* genome encodes 10,742 putative genes with 84.30% of them having detectable sequence similarities to others available in public databases. Phylogenetic analysis revealed a close evolutionary relationship of *L. rhinocerotis* to *Ganoderma lucidum*, *Dichomitus squalens*, and *Trametes versicolor* in the core polyporoid clade. The *L. rhinocerotis* genome encodes a repertoire of enzymes engaged in carbohydrate and glycoconjugate metabolism, along with cytochrome P450s, putative bioactive proteins (lectins and fungal immunomodulatory proteins) and laccases. Other genes annotated include those encoding key enzymes for secondary metabolite biosynthesis, including those from polyketide, nonribosomal peptide, and triterpenoid pathways. Among them, the *L. rhinocerotis* genome is particularly enriched with sesquiterpenoid biosynthesis genes.

**Conclusions:** The genome content of *L. rhinocerotis* provides insights into the genetic basis of its reported medicinal properties as well as serving as a platform to further characterize putative bioactive proteins and secondary metabolite pathway enzymes and as a reference for comparative genomics of polyporoid fungi.

**Keywords:** *Lignosus rhinocerotis*, Genome, Phylogeny, Secondary metabolism, Carbohydrate-active enzymes, Cytochrome P450 superfamily

## Background

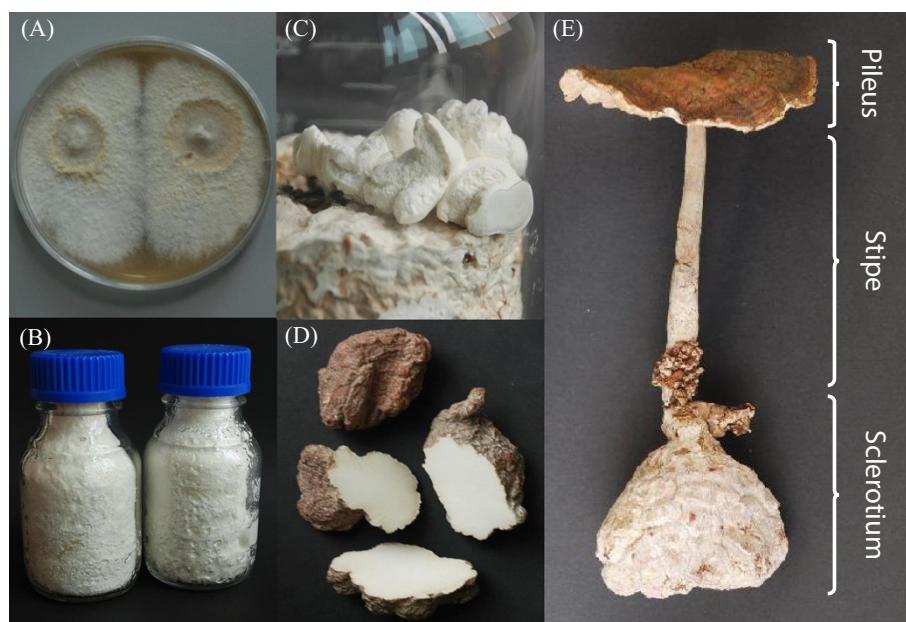
*Lignosus rhinocerotis* (Cooke) Ryvarden, which belongs to the family of Polyporaceae, is characterized by a centrally stipitate pilei arising from its distinct tuber-like sclerotium. This mushroom is widely used by natives of Southeast Asia as a general health tonic for immune enhancement, or as a treatment regime for numerous ailments including cancer, asthma, and bronchitis. It is also used to treat discomfort caused by fright, fever, coughing, vomiting, and cuts [1]. The sclerotium is the part of *L. rhinocerotis* with medicinal value. It is a compact

mass of hardened fungal mycelium and represents one of the stages in the fungal life cycle. This structure is a morphologically variable, nutrient-rich, multihyphal aggregate that serves as a food reserve and can remain dormant until favourable growth conditions arise [2]. Different developmental stages of *L. rhinocerotis* are shown in Figure 1. A two weeks culture of mycelial growth is shown in Figure 1A. Expansion of the mycelium by repeated branching of the germ tube (short, initial hypha) eventually develops into a circular form known as the “Tiger-Eyes”. Cross-linking of the radiating hyphae facilitates nutrient uptake and mobilization around the growing mycelium. After four to six months, the vigorous mycelial growth promotes the development of the sclerotium (Figure 1C). This is often spherical in shape with a dark and tough

\* Correspondence: [yeon\\_ny\\_nie@yahoo.com](mailto:yeon_ny_nie@yahoo.com)

<sup>1</sup>Department of Molecular Medicine, Faculty of Medicine, University of Malaya, 50603 Kuala Lumpur, Malaysia

Full list of author information is available at the end of the article



**Figure 1** Various stages of *L. rhinocerotis* development. (A) Culture of *L. rhinocerotis* mycelium on nutrified agar, also known as the “Tiger-Eyes” (2 weeks culture). (B) Mycelial cultures of *L. rhinocerotis* on solid medium (1 to 2 months cultures). (C) Newly formed sclerotia on the surface of culture medium (4 to 6 months culture). (D) *L. rhinocerotis* sclerotia, the part with medicinal value. (E) Fruiting body of *L. rhinocerotis* with pileus (cap) and stipe (stalk) attached to the sclerotium.

outer skin that keeps the internal compacted hyphal mass from drying out (Figure 1D). The stipe of the mushroom was formed after 12 months culturing, which was preceded by formation of the pileus (Figure 1E). In Malaysia, isolates of *L. rhinocerotis* have been found in Penang Island, Cameron Highlands, Hulu Langat, and Gerik. All of these isolates showed high nucleotide sequence identity (approximately 98%) in their internal transcribed spacer (ITS) gene regions [3,4].

This mushroom is rich in carbohydrates and dietary fiber with moderate amounts of protein while being low in fat [5]. Previous research reported the medical benefits of *L. rhinocerotis* against hypertension, cancer cell cytotoxicity along with enhancement of immunomodulatory activity and antioxidant properties [5-8]. The non-digestible carbohydrates of *Polyporus rhinocerus*, a synonym for *L. rhinocerotis*, was also reported as a potential novel prebiotic for gastrointestinal health [9]. The recent interests in the nutrition and biopharmacology of *L. rhinocerotis* signalled for an immediate need to decipher its biochemical functions, at the genetic level and the identification of its bioactive components.

Rapid advancements in technology has led to the sequencing of numerous fungal genomes with the fungal kingdom becoming one of the most sequenced amongst the eukaryotes [10]. This is not unexpected due to their importance in industry, agriculture, medical, and health. However, the publicly available genome sequences of macrofungi, especially medicinal mushrooms, are still

relatively scarce compared to the plant pathogenic and wood-degrading basidiomycetes or to ascomyceteous microfungi. A recent example is the genome sequence of the medicinal mushroom *Ganoderma lucidum* (lingzhi) by Chen et al. and Liu et al. where the genes in the triterpene biosynthesis and wood degradation pathways were described [11,12]. Other genomes of edible mushrooms include *Volvariella volvacea* (straw mushroom) by Bao et al. [13] and *Agaricus bisporus* (button mushroom) by Foulongne-Oriol et al. [14].

In this paper, we present the *de novo* draft genome sequence of *L. rhinocerotis* TM02 sclerotium. The recent availability of several genome sequences of polyporaceous fungi, especially from the JGI CSP Saprotrophic Agaricomycotina Project [15], has allowed us to gain insights into the *L. rhinocerotis* genome through comparative analyses. We have also surveyed its secondary metabolite production capabilities and identified putative genes that may be involved in the biosynthesis of bioactive proteins and polysaccharides. To our knowledge, this is the first detailed description of the genomic features of *L. rhinocerotis*, an ethnobotanical mushroom of Southeast Asia.

## Results and discussion

### Genome features

Genome sequencing of *L. rhinocerotis* sclerotium with more than 100× coverage produced a total of 6,187 Mb clean data which was further assembled into a 34.3 Mb

draft genome (Table 1). This consisted of 1,338 scaffolds with N50 of 90,329 bp and 53.71% G + C content. Using K-mer (15-mer) analysis with an average insert size of 700 bp, these scaffolds were estimated to cover 70.58% of the whole genome, which has an expected size of 48.6 Mb (Additional file 1). The lower genome coverage can be attributed to the high repeat rate encountered in the assembly. The repeat rate of the number of contigs with lengths shorter than 100 bp and longer than 100 bp is 27.26% and 7.89%, respectively (26.44% of total number of contigs) (Additional file 1). Heterozygosity is unlikely to be the major contribution to the lower genome coverage based on the 15-mer depth analysis (Additional file 1) [16]. The use of fosmid libraries for longer paired end reads (e.g. 10, 20, and 40 kbp), beyond the 5000 bp inserts we have used, may be required to overcome the high repeat rate. Alternatively, the genome assembly can be improved in the future by construction of whole genome physical maps using an optical mapping system, as reported for *G. lucidum* [11]. Nonetheless, this draft genome allows a detailed analysis into the gene content, phylogeny, and metabolic pathways of *L. rhinocerotis*.

#### Repeat elements

Repeat elements of 74 diverse families make up about 4.01% or 1,374,638 bp of the assembled genome of *L. rhinocerotis*, where 1.97% of them are tandem repeat sequences and 2.03% are transposable elements (TEs). The tandem repeats (<4 kbp), not clearly associated with transposons, vary in copy number from 1.8 to 362.5. While among the retrotransposons, long terminal repeats (LTRs) and non-LTR retrotransposons (long and short interspersed

nuclear elements) make up 1.26% and 0.55% of the genome, respectively. DNA transposons (Class II) comprised 0.24% of the genome. The DNA transposons elements were mainly categorized into three classes: Enhancer (En/spm), Tigger (TcMar), and Activator (hAT).

#### Gene prediction

The total 10,742 predicted genes, 216 tRNAs, 17 snRNAs and a single rRNA together comprise 44.33% of the assembled genome. Gene density and the average size of protein coding genes are 5.42 genes/10 kb and 1,414.33 bp, respectively. Among the tRNAs, 13 are possible pseudogenes, four with undetermined anticodons and the remaining 199 anti-codon tRNAs correspond to the 20 common amino acid codons. About half of the tRNAs (109) are predicted to not contain an intron. The genome size of *L. rhinocerotis*, its average gene length, the proportion of repeat sequences, and the average number of exons and introns were comparable to the recently sequenced polyporaceous *G. lucidum* genome [12].

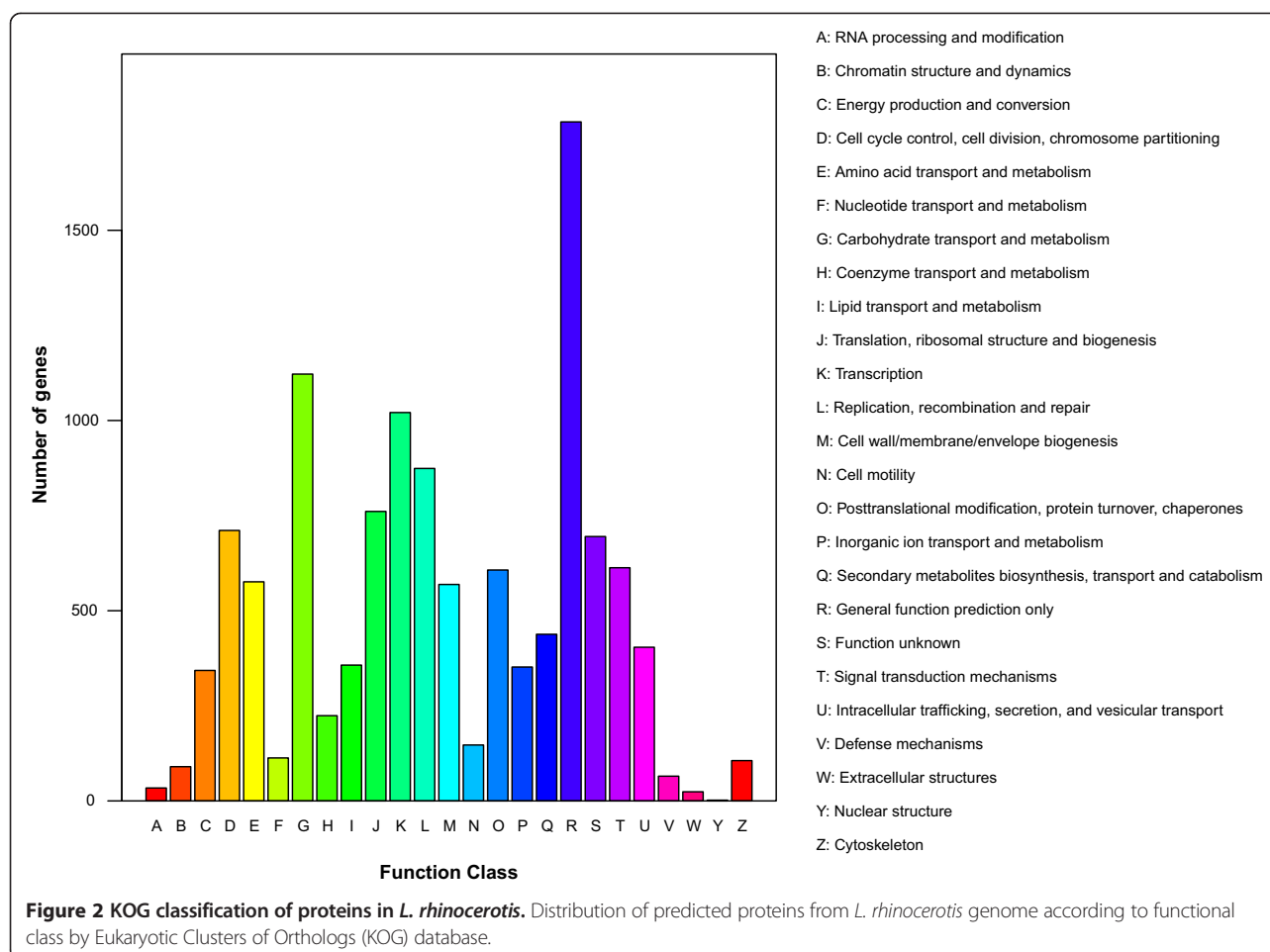
The *L. rhinocerotis* genome revealed a total of 1,686 predicted genes that encode for hypothetical proteins with no apparent homologs to currently available sequences. This is indicative of the uniqueness of *L. rhinocerotis*. On the other hand, up to 8986, 5883, 6669 and 8997 genes are homologous to known proteins in the NCBI nr, SwissProt, InterPro and TrEMBL databases, respectively. These homologous proteins represent 84.30% of the assembled genome.

We further mapped the genome to the Eukaryotic Clusters of Orthologs (KOG), Gene Ontology (GO), and the Kyoto Encyclopedia of Genes and Genomes (KEGG) databases to further characterize the predicted proteins. Based on the phylogenetic classification of the KOGnitor, 4,948 proteins (46.06%) were assigned to KOG (Figure 2). The R category for “General functional prediction only” (typical prediction of biochemical activity), which has the most number of genes (1,785), indicates that these proteins were not unambiguously assigned to a certain group. This is followed by “Carbohydrate transport and metabolism”, “Transcription”, “Replication, recombination and repair”, and “Translation, ribosomal structure and biogenesis” as the top five most abundant gene categories in the KOG grouping. These findings suggest the presence of an enriched and varied array of carbohydrate uptake and metabolism mechanisms that can better facilitate the terrestrial substrate of *L. rhinocerotis* because such a lifestyle results in exposure to more diverse carbohydrate forms and sources as opposed to the wood saprophytes.

The hits from InterPro were used to map to the “Cellular location”, “Molecular function”, and “Biological process” categories of GO (Figure 3A). GO classification revealed that 49.28% (5,294) of the predicted proteins were mainly assigned to “Binding”, “Physiological process”, “Catalytic

**Table 1 Features of *L. rhinocerotis***

<b>Scaffold features</b>	
Total number	1,338
Total length (bp)	34,316,739
N50 (bp)	90,329
N90 (bp)	14,636
Max length (bp)	596,193
Min length (bp)	1,002
Sequence GC content (%)	53.71
<b>Genome features</b>	
Genome assembly (Mb)	34.3
Number of protein-coding genes	10,742
Coding sequences/genome (%)	44.31
Average gene length (bp)	1843.48
Average coding sequence length (bp)	1414.33
Average exon length (nt)	234.30
Average intron length (nt)	85.21
Average number of exons per gene	6.04



activity”, and “Cellular process”. To provide a better understanding of the gene functions in *L. rhinocerotis*, we successfully assigned 4,980 (46.36%) putative proteins to their orthologs in the KEGG database (Figure 3B). The top five categories in KEGG pathway classification, with the highest numbers of annotated genes in *L. rhinocerotis*, are “Genetic information processing - Replication and repair”, “Metabolism - Xenobiotics biodegradation and metabolism”, “Genetic information processing - Folding, sorting, and degradation”, “Metabolism - Carbohydrate metabolism”, and “Metabolism - Amino acid metabolism”.

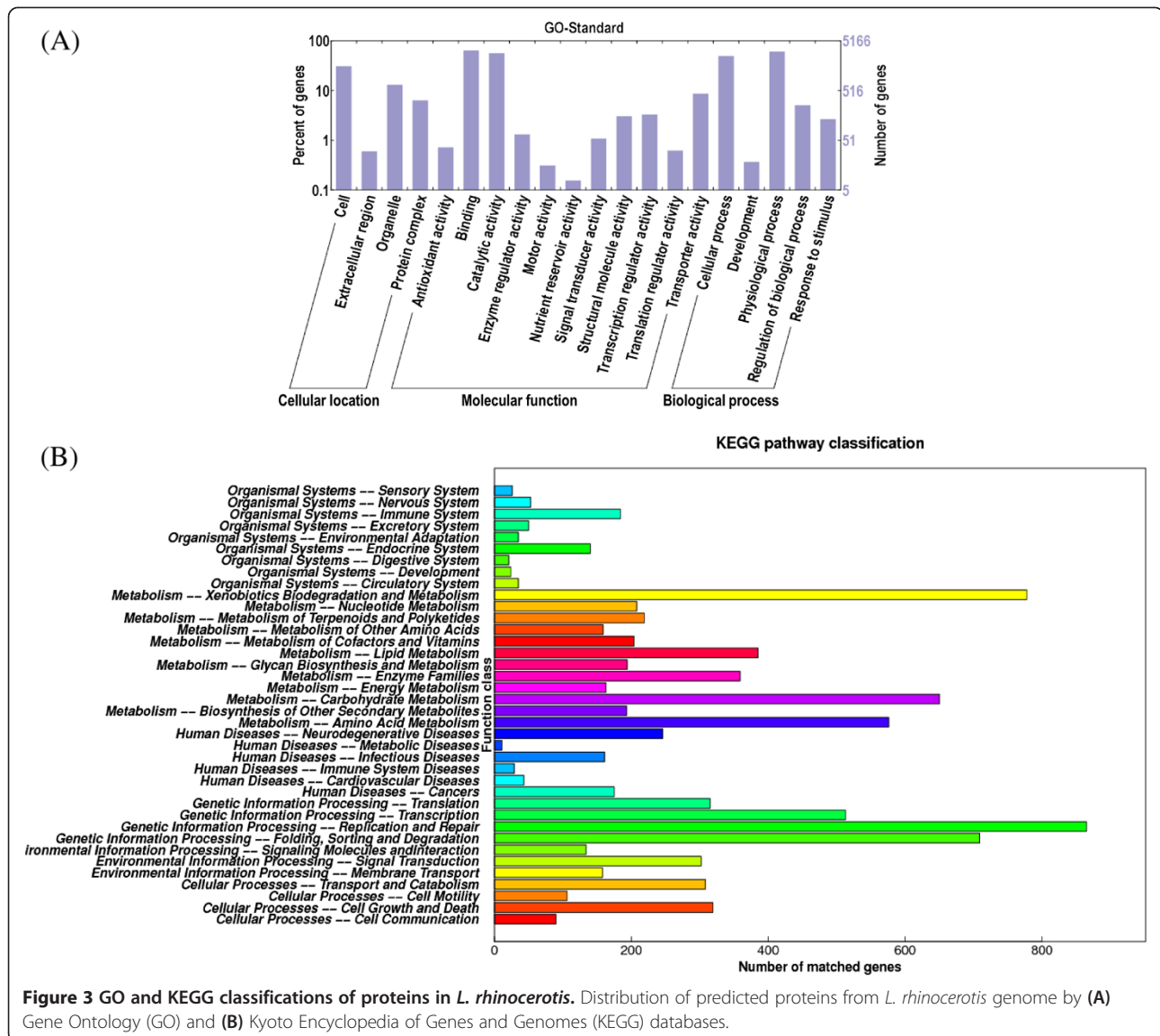
#### KEGG-based comparative genomics analysis

KEGG pathway annotations for seven medicinal Basidiomycota fungi, consisting of five agaricomycetes (*G. lucidum*, *Trametes versicolor*, *Wolfiporia cocos*, *A. bisporus*, *Pleurotus ostreatus*), a basidiomycete (*S. commune*) and tremellomycete (*Tremella mesenterica*), were selected for comprehensive comparison. *L. rhinocerotis* is relatively enriched with genes for catabolism and biosynthesis of secondary metabolites (Table 2, Additional file 2: Table S1) such as “Limonene and pinene degradation”, “Indole alkaloid biosynthesis”, “Penicillin and cephalosporin biosynthesis”,

and “Stilbenoid, diarylheptanoid and gingerol biosynthesis”. However, *L. rhinocerotis* was predicted to have a relatively low number of genes associated with the biosynthesis of siderophore group nonribosomal peptides. The metabolism and biosynthesis of secondary metabolites in *L. rhinocerotis* are further discussed in the subsection “Secondary metabolism”.

The Enzyme Commission (EC) number classification was used to link the respective enzyme genes to their repertoire of metabolic pathways [17]. In the fourth layer of the reference KEGG pathway, *L. rhinocerotis* was found to have 26 putative enzymes that are two-fold greater than the other fungi compared (Additional file 2: Table S2, Additional file 2: Table S3). Among them, 15 were mapped to “Metabolism”, two for “Genetic information processing” and one for “Organismal systems” while two remain “Unclassified” according to KO terms. There were another six enzymes that mapped to multiple sections in the first layer of KEGG.

Interestingly, a total of 535 enzymes are exclusive to *L. rhinocerotis* and not present in any other Basidiomycota fungi compared (Additional file 2: Table S4). Some of the exclusive enzymes participate in multiple KO classes.



**Figure 3** GO and KEGG classifications of proteins in *L. rhinocerotis*. Distribution of predicted proteins from *L. rhinocerotis* genome by (A) Gene Ontology (GO) and (B) Kyoto Encyclopedia of Genes and Genomes (KEGG) databases.

Among them, 79.25% are predicted to involve in “Metabolism” where 20.56% and 18.69% play major roles in “Amino acid metabolism” and “Carbohydrate metabolism”, respectively. This is followed by “Genetic information processing” (8.79%) and “Cellular processes” (5.61%). About 14.21% remain “Unclassified” without mapping to any specific pathway.

### Phylogeny of *L. rhinocerotis*

In this study, we selected eight common KOGs shared by *L. rhinocerotis* and 14 Basidiomycota fungi plus three from Ascomycota fungi for rooting the phylogenetic trees. The four phylogenetic methods according to Kura-mae et al. were employed [18]. The resulting single consensus tree gives highly supported internal branches of 99 to 100%, thus signifying the robustness of our dataset

(Figure 4). Support value of 99% was only observed in the branch of *Puccinia graminis* to *Ustilago maydis* while the others are 100%. Phylogenetic analysis of the 144 concatenated proteins present in the 18 genomes compared revealed a closer evolutionary relationship of *L. rhinocerotis* to *G. lucidum*, *Dichomitus squalens*, and *T. versicolor*, all members of the Polyporaceae family. The Polyporales have been informally divided into several major clades, which include the antrodia, core polyporoid, and phlebioid clades that are well supported by a recent phylogenetic analysis [19]. *L. rhinocerotis* falls into the core polyporoid clade along with *G. lucidum*, *D. squalens*, and *T. versicolor* while *W. cocos* and *Fomitopsis pinicola* belongs to a sister clade known as the antrodia clade.

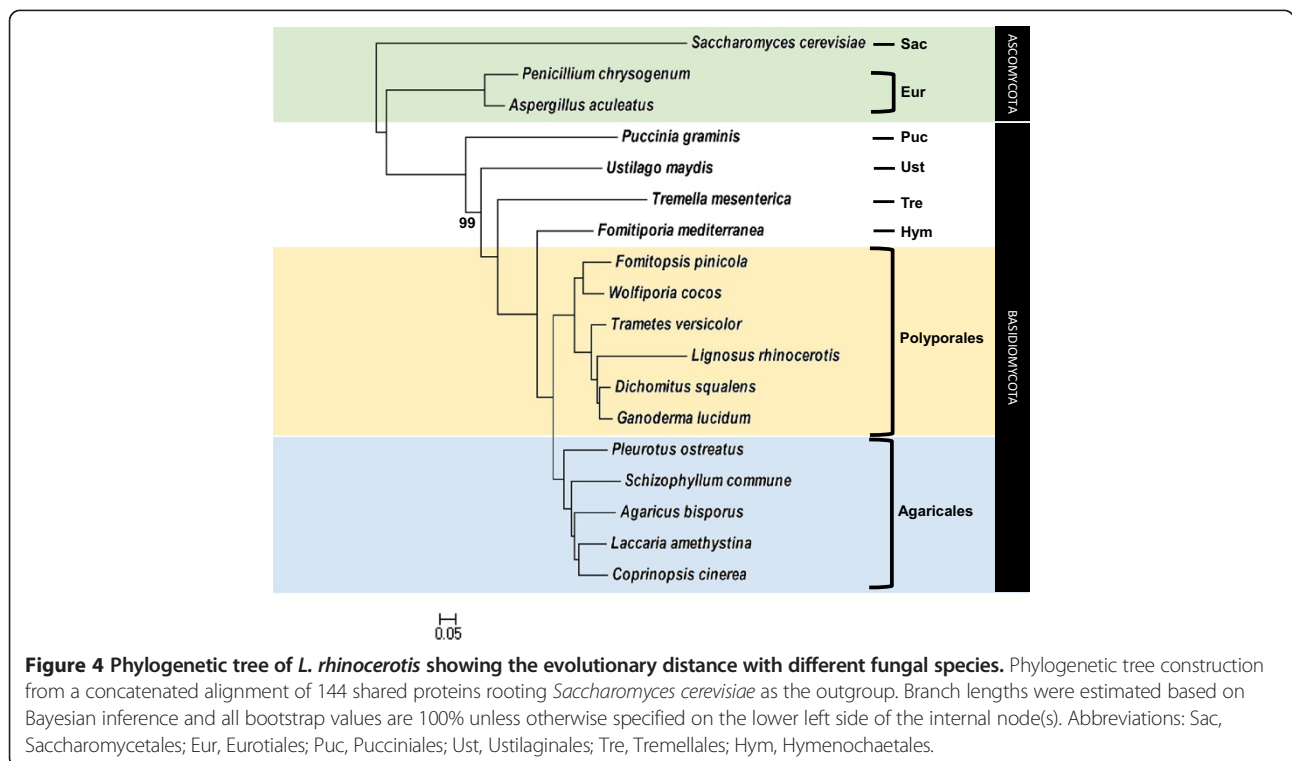
**Table 2 Fungal genes distribution in P450 family and the third layer of KEGG pathways**

	L.rhi	G.luc	T.ver	W.coc	T.mes	S.com	P.ost	A.bis
<b>P450</b>	136	262	248	240	33	159	204	152
<b>KEGG pathway</b>								
00281: Geraniol degradation	17*	13	13	14	6	13	11	10
00900: Terpenoid backbone biosynthesis	13	14	14	17	14	20	14	18
00903: Limonene and pinene degradation	144*	79	72	75	29	53	64	56
01053: Biosynthesis of siderophore group nonribosomal peptides	2	110	119	82	97	141	64	53
00311: Penicillin and cephalosporin biosynthesis	7*	0	0	0	0	0	3	0
00312: beta-Lactam resistance	3*	0	0	0	0	0	0	0
00901: Indole alkaloid biosynthesis	4*	1	1	1	0	1	1	1
00945: Stilbenoid, diarylheptanoid and gingerol biosynthesis	83*	80	65	73	39	53	57	47
00960: Tropane, piperidine and pyridine alkaloid biosynthesis	32	26	46	19	8	20	24	18
00980: Metabolism of xenobiotics by cytochrome P450	33	113	146	119	12	95	91	90
00982: Drug metabolism - cytochrome P450	36	116	142	129	14	93	94	90

\*represents *L. rhinocerotis* having relatively more genes than the average of seven Basidiomycota fungi. Abbreviations: L.rhi, *L. rhinocerotis*; G.luc, *G. lucidum*; T.ver, *T. versicolor*; W.coc, *W. cocos*; T.mes, *T. mesenterica*; S.com, *S. commune*; P.ost, *P. ostreatus*; A.bis, *A. bisporus*.

It should be noted that, although *L. rhinocerotis* falls into the same clade with *G. lucidum*, *D. squalens*, and *T. versicolor*, it is relatively distant from them and shows distinct morphological features. Unlike the other white-rot members from the core polyporoid clade that grow on wood, *L. rhinocerotis* has a terrestrial growth habit similar to the brown-rot *W. cocos* with the development of an underground sclerotium [20]. The sclerotium of *L. rhinocerotis* is oblong to irregular shape and its fruiting

body (basidiocarp) is centrally stipitate with an isodiametric cap. On the other hand, the cap of *G. lucidum* and *T. versicolor* is offset and sometimes indistinct with either a bare stipe for the former or lacking one for both fungi. On the contrary, *D. squalens* has a basidiocarp with poroid hymenophore and lacks a stipe. Although *L. rhinocerotis* shows similar growth habit to *W. cocos* with the presence of a sclerotium, the latter has resupinate fruiting body and spherical sclerotium. Therefore, *L.*



*rhinocerotis* is relatively unique among the sequenced Basidiomycota mushrooms.

### The CAZymes family

As *L. rhinocerotis* is known to thrive on cellulosic substrates, its genome was mapped to the CAZy database to identify the presence of carbohydrate-active enzymes (CAZymes) and auxiliary proteins [20]. A total of 332 non-overlapping CAZyme-coding gene homologs were identified. This includes 178 glycoside hydrolases (GH), 77 glycosyl transferases (GT), three polysaccharide lyases (PL), 102 carbohydrate esterases (CE), 205 carbohydrate-binding module (CBM), and 37 with auxiliary activities (AA) distributed among 39, 26, 1, 13, 32, and 6 coinciding EC activities respectively (Additional file 3). The mapped EC activities may not be directly associated with the family but simply a result of similarity to adjacent modules due to the modular nature of CAZymes. The number of CAZyme candidates identified was almost similar to the average reported in several studies for Basidiomycota fungi [12,13]. The high number of putative GH and GT genes suggests their plausible roles in the degradation of plant cell wall polysaccharides. These polysaccharides consist mainly of cellulose, hemicellulose (including xylan, xyloglucan, glucogalactomannan, galactan, and respective side chains), and pectin (composed of galacturonan, rhamnogalacturonan, and respective side chains).

### The CYPs family

The cytochrome P450 (CYP) superfamily is a diverse group of enzymes involved in various physiological processes, including detoxification, degradation of xenobiotics and the biosynthesis of secondary metabolites [21]. Although not substantial, when compared to most other fungi, it is noted that *L. rhinocerotis* has 33 genes engaged in “Metabolism of xenobiotics by cytochrome P450” and 36 in “Drug metabolism - cytochrome P450” KEGG sub-pathways (Table 2), respectively.

For further comparison, the P450 genes in *L. rhinocerotis* and seven other basidiomycetes were retrieved using BLAST searches against the P450 database (Table 2). *G. lucidum* had the most number of putative P450 genes of 262 followed by *T. versicolor* (246 functional genes and two known pseudogenes) and *W. cocos* (238 functional genes and two known pseudogenes). On the other hand, *T. mesenterica*, a tremellomycete, formed the smallest group among the eight fungi compared with 32 functional genes and a known pseudogene. *L. rhinocerotis* had a total of 136 CYP sequences (135 functional genes and a known pseudogene), which can be classified into 37 families according to Nelson’s nomenclature (Table 2, Additional file 2: Table S5) [22]. The CYP5144 family was found to have the most number of genes (35 genes), followed by CYP5150 (20 genes) and CYP5037 (14 genes) families

**Table 3 P450 genes and subfamilies in *L. rhinocerotis***

Family	Subfamily	Corresponding gene number	Total gene number
CYP5144	A; C; D; F	6; 23; 1; 5	35
CYP5150	A; B; C	17; 1; 2	20
CYP5037	A; B	2; 12	14
CYP504	E	6	6
CYP512	A	6	6
CYP5035	A; B; D	3; 2; 1	6
CYP63	A	5	5
CYP505	D; G	3; 2	5
CYP620	E	3	3
CYP5136	A	3	3
CYP5139	A	3	3
CYP66	A; B	1; 1	2
CYP5046	A	2	2
CYP5065	A	2	2
CYP5151	A; B	1; 1	2
Others	—	—	22

Others are minority CYP families with single gene number which are not shown here.

(Table 3). The CYP5144 family may play a role in triterpenoid biosynthesis (see subsection “Secondary metabolism”) while genes from the CYP5037 and CYP5065 families were found to cluster with terpene synthases (Additional file 2: Table S6). *L. rhinocerotis* also harbours five genes from the CYP63 family, which has been implicated in xenobiotic degradation in *Phanerochaete chrysosporium* [23]. However, the exact roles of these CYPs remain to be determined.

### Secondary metabolism

Secondary metabolite biosynthetic genes are often clustered [24]. The *L. rhinocerotis* genome contains several secondary metabolite gene clusters that suggest the potential for production of certain biologically active compounds (Additional file 2: Table S6). There are ten gene clusters encoding key enzymes, such as terpene synthases (TS), non-ribosomal peptide synthetase (NRPS), and polyketide synthase (PKS), that are crucial for the biosynthesis of terpenes, peptides, and polyketides, respectively. It is noted that, like most basidiomycetes, *L. rhinocerotis* has very few PKS genes and multi-domain NRPS genes compared to ascomycetes. The only PKS gene that can be found in *L. rhinocerotis* is GME5066\_g, which encodes a non-reducing PKS which are often associated with the biosynthesis of aromatic polyketides. This non-reducing PKS appears to be conserved among basidiomycetes and an ortholog of the gene can be found in most of the

sequenced basidiomycetes genomes, including *G. lucidum*, *T. versicolor*, and *A. bisporus*. Interestingly, GME5066\_g shared a head-to-tail homology (38% identity and 55% similarity) and domain architecture with the orsellinic acid synthase from *Coprinopsis cinerea* (CC1G\_05377), the only basidiomycete PKS gene that has been characterized so far [25]. Like CC1G\_05377, GME5066\_g contains a starter unit acyl-carrier protein transacylase (SAT), ketosynthase (KS), acyltransferase (AT), product template (PT), two acyl-carrier proteins (ACPs) and a thioesterase (TE) domain. GME5066\_g is clustered with GME5065\_g, which is a predicted flavin-dependent oxidoreductase. It remains to be determined if the GME5066\_g gene cluster produces orsellinic acid derivatives or related polyketides. The *L. rhinocerotis* genome also harbours a single multi-domain NRPS gene. The NRPS has a single adenylation domain along with three thiolation and condensation domains, and are conserved among several basidiomycetes, including *D. squalens* DICSQDRAFT\_132068 (61% identity) and *T. versicolor* TRAVEDRAFT\_27949 (59% identity), but none are characterized.

Terpenoids (or isoprenoids) is one group of secondary metabolites that are well recognized for their pharmaceutical uses and are known to be one of the major groups of therapeutic compounds in *G. lucidum*. The triterpenoid ganoderic acids, for example, have been reported to have anti-tumor, immuno-regulation, and antioxidative functions [26]. Other notable examples include the diterpenoid antibiotic pleuromutilins from *Clitopilus* mushrooms [27], as well as the sesquiterpenoid anticancer illudins [28]. In the genome of *L. rhinocerotis*, we identified 15 key enzymes involved in the mevalonate (MVA) pathway but not the 2-C-methyl-D-erythritol 4-phosphate/1-deoxy-D-

xylulose 5-phosphate (MEP/DOXP) pathway (Additional file 4). This indicates that the terpenoid backbone biosynthesis in *L. rhinocerotis*, like most fungi, can only proceed via the MVA pathway. All the core enzymes involved in the MVA pathway are listed in Table 4. The enzymes 3-hydroxy-3-methylglutaryl-CoA reductase and solanesyl diphosphate synthase are each encoded by two copies of the genes whereas the remaining nine enzymes are encoded by single copy genes. As in the case for the *G. lucidum* genome reported by Liu et al. [12], we found a fusion gene (GME67\_g) that was partially homologous to cystathionine beta-lyase (metC; K01760; EC4.4.1.8) and mevalonate kinase (MVK; K00869; EC2.7.1.36) at the N- and C-terminal respectively. This metC-MCK gene, which encodes for 918 amino acids, is the only gene that matches the key enzyme mevalonate kinase, implying the role in terpenoid backbone biosynthesis.

We next searched the *L. rhinocerotis* genome for potential triterpenoid biosynthesis genes and found an open reading frame (GME631\_g) that encodes a single copy gene for lanosterol synthase (LSS; K01852; EC5.4.99.7). LSS is a squalene/oxidosqualene cyclase family enzyme that catalyzes the cyclization of the triterpenes squalene or 2-3-oxidosqualene to lanosterol, the precursor of all steroids [29]. This enzyme has been implicated in biosynthesis of ganoderic acids, which are the bioactive triterpenes in *G. lucidum* [11]. Similarly, the LSS in *L. rhinocerotis* can be involved in biosynthesis of bioactive triterpenes. The CYP5144 and CYP512 families of P450 genes have been previously implicated in triterpenoid biosynthesis in *G. lucidum* due to their co-expression with LSS [11]. As mentioned earlier, CYP5144 is the largest P450 family in *L. rhinocerotis* with 35 genes, while

**Table 4 Putative genes involved in terpenoid backbone biosynthesis**

Gene name and definition	EC No.	KO term	Gene ID
E1.1.1.34; 3-hydroxy-3-methylglutaryl-CoA reductase	1.1.1.34	K00021	GME845_g; GME10429_g
metC; cystathionine beta-lyase (homologous to mevalonate kinase at the C-terminal)	4.4.1.8	K01760	GME67_g
atoB; acetyl-CoA C-acetyltransferase	2.3.1.9	K00626	GME4142_g
FDPS; farnesyl diphosphate synthase	2.5.1.1 2.5.1.10	K00787	GME3582_g
GGPS1; geranylgeranyl diphosphate synthase, type III	2.5.1.1 2.5.1.10 2.5.1.29	K00804	GME4885_g
mvaK2; phosphomevalonate kinase	2.7.4.2	K00938	GME148_g
MVD; diphosphomevalonate decarboxylase	4.1.1.33	K01597	GME2603_g
E2.3.3.10; hydroxymethylglutaryl-CoA synthase	2.3.3.10	K01641	GME2144_g
idi; isopentenyl-diphosphate delta-isomerase	5.3.3.2	K01823	GME7794_g
hexPS; hexaprenyl diphosphate synthase	2.5.1.33	K05355	GME1421_g
SPS; solanesyl diphosphate synthase	2.5.1.11	K05356	GME7467_g; GME8769_g
DHDDS; cis-prenyltransferase, dehydrololichyl diphosphate synthase	2.5.1.-	K11778	GME2072_g



six CYP512 family genes are present in its genome as well. This suggests that *L. rhinocerotis* may be a potential triterpenoid source.

Mushrooms are known to be prolific producers of bioactive sesquiterpenes [30]. The *L. rhinocerotis* genome is enriched with sesquiterpenoid biosynthesis genes compared to some of the other seven Basidiomycota genomes (Additional file 2: Table S1). The *L. rhinocerotis* genome encodes up to 12 sesquiterpene cyclase genes. This is comparable to the number of sesquiterpene cyclase genes found in the recently sequenced genome of the Jack O'Lantern mushroom *Omphalotus olearius*, which is the producer of anticancer illudins [31]. Many of these terpene synthase genes are clustered together with various modifying enzymes (Additional file 2: Table S6). Comparatively, the common filamentous ascomycetes have fewer sesquiterpene cyclase genes, for example, both the *Aspergillus nidulans* and *Aspergillus oryzae* have only one sesquiterpene cyclase gene, while the *Aspergillus fumigatus* has none [32]. This suggests that unlike the filamentous ascomycetes that are enriched with the polyketide and nonribosomal peptide biosynthesis genes, the sesquiterpenoids are major secondary metabolites produced by *L. rhinocerotis*.

#### Biosynthesis of potential bioactive proteins and polysaccharides

Polysaccharides are the most extensively investigated mushroom constituents due to their pharmaceutical potential. The water soluble 1,3- $\beta$ - and 1,6- $\beta$ -glucans are some of the most active immunomodulatory compounds reported [33]. Additional file 2: Table S7 lists enzymes that may be involved in the biosynthesis of uridine diphosphate glucose (UDP-glucose, precursor of glucans) and 1,6- $\beta$ -glucans in *L. rhinocerotis*. These enzymes include hexokinase,  $\alpha$ -phosphoglucosyltransferase, UTP-glucose-1-phosphate uridylyltransferase, 1,3- $\beta$ -glucan synthases, and  $\beta$ -glucan biosynthesis-associated proteins.

Mushrooms have also been an important source of bioactive proteins, which include lectins, fungal immunomodulatory proteins (FIP), ribosome inactivating proteins (RIP), antimicrobial proteins, ribonucleases, and laccases [34]. The genome of *L. rhinocerotis* codes for nine putative lectins, two putative fungal immunomodulatory proteins (FIP), and four putative laccases (Additional file 2: Table S7). It is interesting to note that both of the *L. rhinocerotis* FIPs (GME7566\_g and GME10641\_g) are homologous to LZ-8 (64% identities), a member of the FIP family from *G. lucidum* that has been shown to possess immunomodulation and anti-cancer activity [35,36]. Both the putative FIP proteins carry an Fve domain (pfam09259) found in the major fruiting body protein isolated from *Flammulina velutipes* with immunomodulatory activity [37].

#### Conclusions

*L. rhinocerotis* genome sequencing has allowed us to perform comparative genomics and phylogenetic analysis. This has provided valuable insights into the biology of this medicinal mushroom from Southeast Asia. A survey of the secondary metabolite biosynthesis genes in the *L. rhinocerotis* genome shows that it is particularly enriched with sesquiterpenoid biosynthesis genes. Thus, future bioactive molecule discovery efforts should focus on this class of metabolites. Furthermore, *L. rhinocerotis* appears to encode the capabilities to produce 1,3- $\beta$ - and 1,6- $\beta$ -glucans as well as bioactive proteins, such as lectins and FIPs. Our genomic analysis of *L. rhinocerotis* will provide the foundation for future research and exploitation of *L. rhinocerotis* in pharmacological and functional food applications.

#### Methods

##### Fungal material, sequencing, and assembly

Sclerotium of *L. rhinocerotis* was collected from tropical forest at Lata Iskandar, Cameron Highland, Pahang, Malaysia (4.3245°N, 101.3324°E) in 2010. The fungus was deposited at Royal Botanic Gardens, Kew (Richmond, London, England) with the accession number K(M) 177812. Genomic DNA was extracted using a modified cetrimonium bromide (CTAB) procedure [38] from the sclerotium of *L. rhinocerotis* TM02 strain cultivated by Ligno Biotech Sdn. Bhd. (Balakong Jaya, Selangor, Malaysia). Paired-end reads were generated by sequencing of four cloned insert libraries of 200, 700, 2,000, and 5,000 bp using Hiseq2000 system (Illumina Inc., San Diego, CA, USA) at BGI-Shenzhen, China. Reads of low complexity and low quality with adapter and duplication contamination were removed from the raw data to strengthen the accuracy of follow-up analysis. To avoid issues associated with heterozygosity and/or low sequence quality, reads with significant poly-A structure and kmer frequency of 1 were removed. The clean short reads were then assembled using SOAPdenovo based on *de Bruijn* graph theory [39]. The gaps were filled using the GapCloser module from SOAPdenovo. During scaffold construction, contigs with certain distance relationships but without genotypes were connected with wildcards. The GapCloser module then replaced these wildcards using the context and paired-end reads information. The GapCloser assembled sequences iteratively in the gaps to fill large gaps where at each iterative cycle, GapCloser considered only the reads that could be aligned in the current cycle.

##### Gene prediction and annotation

Protein coding genes were predicted using the *ab initio* gene predictors Augustus [40], GeneMark-ES [41], and SNAP [42]. The resulting gene sets were integrated to

get the most comprehensive and non-redundant reference gene. Transposon sequences were predicted by aligning the assembled gene sequences with the transposon Repbase database [43] using RepeatMasker software at <http://www.repeatmasker.org> and RepeatProteinMasker software (transposon protein library). Tandem repeat sequences were predicted using Tandem Repeat Finder (TRF) [44]. rRNA sequences were identified by rRNA pool alignment and rRNAmmer *de novo* prediction software [45]. tRNA genes were predicted by tRNAscan-SE software [46] while others non-coding RNAs (miRNA, sRNA, and snRNA) were predicted by Rfam.

Predicted genes were functionally annotated based on homology to annotated genes in various databases via BLAST. Protein models were aligned to SwissProt, TrEMBL, InterPro and NCBI nr (BLASTP cut-off  $e$ -value  $\leq 1e-5$ ); and further classified according to GO [47], KOG [48], and KEGG pathways [49]. KEGG terms are assigned into four layers. The first layer consists of seven main sections, including “Metabolism”, “Genetic information processing”, “Environmental information processing”, “Cellular processes”, “Organismal systems”, “Human diseases”, and “Drug development”; and is further divided into several small entries, the second layers. The third and fourth layers are the specific pathway map and specific genes regulated in each pathway, respectively.

### Phylogenetic analysis

Together with *L. rhinocerotis*, 14 Basidiomycota of agaricomycetes (*G. lucidum*, *T. versicolor*, *D. squalens*, *W. cocos*, *F. pinicola*, *Fomitiporia mediterranea*, *A. bisporus*, *Coprinus cinereus* (synonym *C. cinerea*), *Laccaria amethystina*, *P. ostreatus*), basidiomycete (*S. commune*), tremellomycete (*T. mesenterica*), pucciniomycete (*P. graminis*), and ustilaginomycete (*U. maydis*) were selected for analysis. Three Ascomycota of eurotiomycetes (*Aspergillus aculeatus*, *Penicillium chrysogenum*) and saccharomycete (*Saccharomyces cerevisiae*) were added to root the phylogenetic trees. All the selected genomic gene models were downloaded from the US Department of Energy Joint Genome Institute website at <http://genome.jgi.doe.gov/> (Additional file 2: Table S8).

Protein sequences of each shared KOG family from the different species were aligned using CLUSTAL X [50]. The multiple sequence alignments were concatenated using DAMBE5 [51] upon the removal of poorly aligned regions by GBlocks server [52]. PROTTEST [53] was used to select the best fit empirical substitution model of protein evolution for the concatenated alignment.

Maximum Parsimony and Neighbor Joining analyses were executed with Phylip [54] using PROTPARS and PROTDIST (JTT model). TREE-PUZZLE [55] was used to construct maximum likelihood

quartet puzzling trees using the VT model [56]. Bayesian inference using Markov chain Monte Carlo phylogenetic analysis was performed with MrBayes 3.2.1 [57] under Rtrev + G + F model. Trees were sampled every 10 generations over a random starting trees of 50,000 generations, resulting in a total of 5,000 sampling trees, of which the first 2,000 were discarded. The remaining 3,000 trees were used to build a single consensus tree with  $\geq 70\%$  majority-rule using MEGA [58].

### CYP and CAZy family classifications

*L. rhinocerotis* gene models were aligned to fungi P450 sequences and the detected CYPs were named according to nomenclature in the P450 database (cut-off  $e$ -value  $\leq 1e-10$ , identity  $> 30\%$ ) at Cytochrome P450 homepage (<http://drnelson.uthsc.edu/CytochromeP450.html>) [22]. Annotation of carbohydrate-active enzymes in *L. rhinocerotis* genome was carried out by BLASTP analysis against CAZy database at <http://www.cazy.org/> [59].

### Secondary metabolites gene clusters annotation

Secondary metabolite gene clusters were determined using Secondary Metabolite Unique Regions Finder (SMURF, <http://jcv.org/smurf/index.php>) [60] based on PFAM and TIGRFAM resources along with the gene's chromosomal position and antibiotics & Secondary Metabolite Analysis Shell (antiSMASH 2.0, <http://antismash.secondarymetabolites.org/>) [61]; a web-based analysis platform.

### Accession number

This Whole Genome Shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession AXZM0000000. The version described in this paper is version AXZM01000000.

### Additional files

**Additional file 1: Heterogeneous sequence (k-mer) and repeat rate analyses of *L. rhinocerotis* genome.**

**Additional file 2: Table S1.** Gene distribution of *L. rhinocerotis* and the other seven compared Basidiomycota fungi in the third layer of KEGG pathway. **Table S2.** Enzyme distribution of *L. rhinocerotis* with not less than 2-fold genes more than the other seven compared Basidiomycota fungi in the fourth layer of KEGG pathway. **Table S3.** The classes of 26 enzymes of *L. rhinocerotis* with not less than 2-fold genes more than the other seven compared Basidiomycota fungi in the fourth layer of KEGG pathway. **Table S4.** The 535 enzymes which are exclusive to only *L. rhinocerotis*. **Table S5.** Cytochrome P450 genes in *L. rhinocerotis*. **Table S6.** The putative backbone genes and clusters involved in *L. rhinocerotis* secondary metabolism. **Table S7.** Putative genes involved in the biosynthesis of bioactive polysaccharides and proteins in *L. rhinocerotis*. **Table S8.** Download portal of additional fungi used in this study.

**Additional file 3: Assignment of *L. rhinocerotis* genes to CAZy families and their respective EC activities.**

**Additional file 4: Potential terpenoids biosynthesis pathway in *L. rhinocerotis*.**

**Competing interests**

S-TN is affiliated with Ligno Biotech Sdn. Bhd. which commercialised the Tiger Milk mushroom.

**Authors' contributions**

H-YYY performed the molecular genetic studies, bioinformatics analysis, sequence alignment, and drafted the manuscript. Y-HC and MF-R participated in the design of the study, bioinformatics analysis and helped to draft the manuscript. N-HT, C-ST, S-YF, and S-TN conceived the study and participated in its design and coordination. N-HT and S-YF helped to draft the manuscript. All authors read and approved the final manuscript.

**Acknowledgements**

This research is supported by High Impact Research Grant UMC/625/1/HIR/MoE/E20040-20001 from the University of Malaya/Ministry of Education, Malaysia. H-YYY is supported by the postgraduate research grant (PPP) PV024/2012A from University of Malaya, Malaysia. Y-HC is a recipient of Australian Research Council Discovery Early Career Researcher Award (ARC DECRA). We are grateful to the team at BGI-Shenzhen for their assistance in genomic sequencing and analysis. Oliver Mead is thanked for proofreading the manuscript.

**Author details**

<sup>1</sup>Department of Molecular Medicine, Faculty of Medicine, University of Malaya, 50603 Kuala Lumpur, Malaysia. <sup>2</sup>Plant Sciences Division, Research School of Biology, The Australian National University, Canberra 0200, Australia. <sup>3</sup>School of Biosciences and Biotechnology, Faculty of Science and Technology and Institute of Systems Biology, Universiti Kebangsaan Malaysia, 43600 UKM Bangi, Malaysia. <sup>4</sup>Ligno Biotech Sdn. Bhd., 43300 Balakong Jaya, Selangor, Malaysia. <sup>5</sup>Malaysian Agricultural Research and Development Institute (MARDI), 43400 Serdang, Selangor, Malaysia.

Received: 17 February 2014 Accepted: 23 July 2014

Published: 29 July 2014

**References**

1. Lee SS, Chang YS: **Ethnomycology**. In *Malaysian Fungal Diversity*. Edited by Jones EBG, Hyde KD, Sabaratnam V. Malaysia: Mushroom Research Centre, University of Malaya and Natural Resources and Environment, Malaysia; 2007:307–317.
2. Willetts HJ, Bullock S: **Developmental biology of sclerotia**. *Mycol Res* 1992, **96**:801–816.
3. Cunningham GH: **Polyporaceae of New Zealand**. *Bull New Zealand Dept Sci Industr Res* 1965, **164**:1–304.
4. Tan CS, Ng ST, Vikineswary S, Lo EP, Tee CS: **Genetic markers for identification of a Malaysian medicinal mushroom *Lignosus rhinocerus* (Cendawan susu rimau)**. *Acta Hort* 2010, **859**:161–167.
5. Yap YHY, Tan NH, Fung SY, Aziz AA, Tan CS, Ng ST: **Nutrient composition, antioxidant properties, and anti-proliferative activity of *Lignosus rhinocerus* Cooke sclerotium**. *J Sci Food Agric* 2013, **9**:2945–2952.
6. Lai CKM, Wong KH, Cheung PCK: **Antiproliferative effects of sclerotial polysaccharides from *Polyporus rhinocerus* Cooke (Aphyllophoromycetideae) on different kinds of leukemic cells**. *Int J Med Mushrooms* 2008, **10**:255–264.
7. Lee ML, Tan NH, Fung SY, Tan CS, Ng ST: **The antiproliferative activity of sclerotia of *Lignosus rhinocerus* (Tiger Milk Mushroom)**. *Evid Based Complement Alternat Med* 2012, **2012**:697603.
8. Wong KH, Lai CKM, Cheung PCK: **Immunomodulatory activities of mushroom sclerotial polysaccharides**. *Food Hydrocoll* 2011, **25**:150–158.
9. Gao S, Lai CKM, Cheung PCK: **Nondigestible carbohydrates isolated from medicinal mushroom sclerotia as novel prebiotics**. *Int J Med Mushrooms* 2009, **11**:1–8.
10. Galagan JE, Henn MR, Ma LJ, Cuomo CA, Birren B: **Genomics of the fungal kingdom: insights into eukaryotic biology**. *Genome Res* 2005, **15**:1620–1631.
11. Chen S, Xu J, Liu C, Zhu Y, Nelson DR, Zhou S, Li C, Wang L, Guo X, Sun Y, Luo H, Li Y, Song J, Henrissat B, Levasseur A, Qian J, Li J, Luo X, Shi L, He L, Xiang L, Xu X, Niu Y, Li Q, Han MV, Yan H, Zhang J, Chen H, Lv A, Wang Z,

- et al: **Genome sequence of the model medicinal mushroom *Ganoderma lucidum***. *Nat Commun* 2012, **3**:913.
12. Liu D, Gong J, Dai W, Kang X, Huang Z, Zhang HM, Liu W, Liu L, Ma J, Xia Z, Chen Y, Chen YW, Wang D, Ni P, Guo AY, Xiong X: **The genome of *Ganoderma lucidum* provides insights into triterpenes biosynthesis and wood degradation**. *PLoS ONE* 2012, **7**:e36146.
  13. Bao D, Gong M, Zheng H, Chen M, Zhang L, Wang H, Jiang J, Wu L, Zhu Y, Zhu G, Zhou Y, Li C, Wang S, Zhao Y, Zhao G, Tan Q: **Sequencing and comparative analysis of the straw mushroom (*Volvariella volvacea*) genome**. *PLoS ONE* 2013, **8**:e58294.
  14. Foulongne-Oriol M, Murat C, Castanera R, Ramirez L, Sonnenberg AS: **Genome-wide survey of repetitive DNA elements in the button mushroom *Agaricus bisporus***. *Fungal Genet Biol* 2013, **55**:6–21.
  15. Floudas D, Binder M, Riley R, Barry K, Blanchette RA, Henrissat B, Martinez AT, Otilar R, Spatafora JW, Yadav JS, Aerts A, Benoit I, Boyd A, Carlson A, Copeland A, Coutinho PM, de Vries RP, Ferreira P, Findley K, Foster B, Gaskell J, Glotzer D, Gorecki P, Heitman J, Hesse C, Hori C, Igarashi K, Jurgens JA, Kallen N, Kersten P, et al: **The Paleozoic origin of enzymatic lignin decomposition reconstructed from 31 fungal genomes**. *Science* 2012, **336**:1715–1719.
  16. Zheng W, Huang L, Huang J, Wang X, Chen X, Zhao J, Guo J, Zhuang H, Qiu C, Liu J, Liu H, Huang X, Pei G, Zhan G, Tang C, Cheng Y, Liu M, Zhang J, Zhao Z, Zhang S, Han Q, Han D, Zhang H, Zhao J, Gao X, Wang J, Ni P, Dong W, Yang L, Yang H, et al: **High genome heterozygosity and endemic genetic recombination in the wheat stripe rust fungus**. *Nat Commun* 2013, **4**:2673.
  17. Kotera M, Okuno Y, Hattori M, Goto S, Kanehisa M: **Computational assignment of the EC numbers for genomic-scale analysis of enzymatic reactions**. *J Am Chem Soc* 2004, **126**:16487–16498.
  18. Kuramae EE, Robert V, Snel B, Weiss M, Boekhout T: **Phylogenomics reveal a robust fungal tree of life**. *FEMS Yeast Res* 2006, **6**:1213–1220.
  19. Binder M, Justo A, Riley R, Salamov A, Lopez-Giraldez F, Sjakovik E, Copeland A, Foster B, Sun H, Larsson E, Larsson KH, Townsend J, Grigoriev IV, Hibbett DS: **Phylogenetic and phylogenomic overview of the Polyporales**. *Mycologia* 2013, **105**:1350–1373.
  20. Wong KH, Cheung PCK: **Sclerotia: Emerging Functional Food Derived from Mushrooms**. In *Mushrooms as Functional Foods*. Edited by Cheung PCK. New Jersey: John Wiley & Sons, Inc; 2009:111–146.
  21. Crešnar B, Petrič S: **Cytochrome P450 enzymes in the fungal kingdom**. *Biochim Biophys Acta* 1814, **2011**:29–35.
  22. Nelson DR: **The cytochrome P450 homepage**. *Hum Genomics* 2009, **4**:59–65.
  23. Doddapaneni H, Yadav JS: **Differential regulation and xenobiotic induction of tandem P450 monooxygenase genes pc-1 (CYP63A1) and pc-2 (CYP63A2) in the white-rot fungus *Phanerochaete chrysosporium***. *Appl Microbiol Biotechnol* 2004, **65**:559–665.
  24. Keller NP, Turner G, Bennett JW: **Fungal secondary metabolism - from biochemistry to genomics**. *Nat Rev Microbiol* 2005, **3**:937–947.
  25. Ishiuchi K, Nakazawa T, Ookuma T, Sugimoto S, Sato M, Tsunematsu Y, Ishikawa N, Noguchi H, Hotta K, Moriya H, Watanabe K: **Establishing a new methodology for genome mining and biosynthesis of polyketides and peptides through yeast molecular genetics**. *ChemBiochem* 2012, **13**:846–854.
  26. Shi L, Ren A, Mu D, Zhao M: **Current progress in the study on biosynthesis and regulation of ganoderic acids**. *Appl Microbiol Biotechnol* 2010, **88**:1243–1251.
  27. Novak R, Shlaes DM: **The pleuromutilin antibiotics: a new class for human use**. *Curr Opin Investig Drugs* 2010, **11**:182–191.
  28. Schobert R, Knauer S, Seibt S, Biersack B: **Anticancer active illudins: recent developments of a potent alkylating compound class**. *Curr Med Chem* 2011, **18**:790–807.
  29. Corey EJ, Matsuda SP, Bartel B: **Isolation of an *Arabidopsis thaliana* gene encoding cycloartenol synthase by functional expression in a yeast mutant lacking lanosterol synthase by the use of a chromatographic screen**. *Proc Natl Acad Sci U S A* 1993, **90**:11628–11632.
  30. Kramer R, Abraham W-R: **Volatile sesquiterpenes from fungi: what are they good for?** *Phytochem Rev* 2012, **11**:15–37.
  31. Wawrzyn GT, Quin MB, Choudhary S, López-Gallego F, Schmidt-Dannert C: **Draft genome of *Omphalotus olearius* provides a predictive framework for sesquiterpenoid natural product biosynthesis in Basidiomycota**. *Chem Biol* 2012, **19**:772–783.
  32. Nierman WC, Pain A, Anderson MJ, Wortman JR, Kim HS, Arroyo J, Berriman M, Abe K, Archer DB, Bermejo C, Bennett J, Bowyer P, Chen D, Collins M, Coulson R, Davies R, Dyer PS, Farman M, Fedorova N, Fedorova N,

- Feldblyum TV, Fischer R, Fosker N, Fraser A, García JL, García MJ, Goble A, Goldman GH, Gomi K, Griffith-Jones S, et al: **Genomic sequence of the pathogenic and allergenic filamentous fungus *Aspergillus fumigatus*.** *Nature* 2005, **438**:1151–1156.
33. Xu Z, Chen X, Zhong Z, Chen L, Wang Y: ***Ganoderma lucidum* polysaccharides: immunomodulation and potential anti-tumor activities.** *Am J Chin Med* 2011, **39**:15–27.
34. Xu X, Yan H, Chen J, Zhang X: **Bioactive proteins from mushrooms.** *Biotechnol Adv* 2011, **29**:667–674.
35. Kino K, Yamashita A, Yamaoka K, Watanabe J, Tanaka S, Ko K, Shimizu K, Tsunoo H: **Isolation and characterization of a new immunomodulatory protein, ling zhi-8 (LZ-8), from *Ganoderma lucidum*.** *J Biol Chem* 1989, **264**:472–478.
36. Wu CT, Lin TY, Hsu HY, Sheu F, Ho CM, Chen EI: **Ling Zhi-8 mediates p53-dependent growth arrest of lung cancer cells proliferation via the ribosomal protein S7-MDM2-p53 pathway.** *Carcinogenesis* 2011, **32**:1890–1896.
37. Paaventhan P, Joseph JS, Seow SV, Vaday S, Robinson H, Chua KY, Kolatkar PR: **A 1.7A structure of Fve, a member of the new fungal immunomodulatory protein family.** *J Mol Biol* 2003, **332**:461–470.
38. Cota-Sánchez JH, Remarchuk K, Ubayasena K: **Ready-to-use DNA extracted with a CTAB method adapted for herbarium specimens and mucilaginous plant tissue.** *Plant Mol Biol Rep* 2006, **24**:161–167.
39. Li R, Zhu H, Ruan J, Qian W, Fang X, Shi Z, Li Y, Li S, Shan G, Kristiansen K, Li S, Yang H, Wang J, Wang J: **De novo assembly of human genomes with massively parallel short read sequencing.** *Genome Res* 2010, **20**:265–272.
40. Stanke M, Waack S: **Gene prediction with a hidden Markov model and a new intron submodel.** *Bioinformatics* 2003, **19**(Suppl 2):ii215–ii225.
41. Borodovsky M, Lomsadze A: **Eukaryotic gene prediction using GeneMark.hmm-E and GeneMark-ES.** *Curr Protoc Bioinformatics* 2011, **Chapter 4**:1–10.
42. Korf I: **Gene finding in novel genomes.** *BMC Bioinformatics* 2004, **5**:59.
43. Jurka J, Kapitonov VV, Pavlicek A, Klonowski P, Kohany O, Walichiewicz J: **Repbase update, a database of eukaryotic repetitive elements.** *Cytogenet Genome Res* 2005, **110**:462–467.
44. Benson G: **Tandem repeats finder: a program to analyze DNA sequences.** *Nucleic Acids Res* 1999, **27**:573–580.
45. Lagesen K, Hallin P, Rodland EA, Staerfeldt HH, Rognes T, Ussery DW: **RNAmmr: consistent and rapid annotation of ribosomal RNA genes.** *Nucleic Acids Res* 2007, **35**:3100–3108.
46. Schattner P, Brooks AN, Lowe TM: **The tRNAscan-SE, snoscan and snoGPS web servers for the detection of tRNAs and snoRNAs.** *Nucleic Acids Res* 2005, **33**:W686–W689.
47. Ashburner M, Ball CA, Blake JA, Botstein D, Butler H, Cherry JM, Davis AP, Dolinski K, Dwight SS, Eppig JT, Harris MA, Hill DP, Issel-Tarver L, Kasarskis A, Lewis S, Matese JC, Richardson JE, Ringwald M, Rubin GM, Sherlock G: **Gene ontology: tool for the unification of biology. The Gene Ontology Consortium.** *Nat Genet* 2000, **25**:25–29.
48. Koonin EV, Fedorova ND, Jackson JD, Jacobs AR, Krylov DM, Makarova KS, Mazumder R, Mekhedov SL, Nikolskaya AN, Rao BS, Rogozin IB, Smirnov S, Sorokin AV, Sverdlov AV, Vasudevan S, Wolf YI, Yin JJ, Natale DA: **A comprehensive evolutionary classification of proteins encoded in complete eukaryotic genomes.** *Genome Biol* 2004, **5**:R7.
49. Kanehisa M, Goto S, Kawashima S, Okuno Y, Hattori M: **The KEGG resource for deciphering the genome.** *Nucleic Acids Res* 2004, **32**:D277–D280.
50. Thompson JD, Gibson TJ, Plewniak F, Jeanmougin F, Higgins DG: **The ClustalX windows interface: flexible strategies for multiple sequence alignment aided by quality analysis tools.** *Nucleic Acids Res* 1997, **25**:4876–4882.
51. Xia X: **DAMBES: a comprehensive software package for data analysis in molecular biology and evolution.** *Mol Biol Evol* 2013, **30**:1720–1728.
52. Castresana J: **Selection of conserved blocks from multiple alignments for their use in phylogenetic analysis.** *Mol Biol Evol* 2000, **17**:540–552.
53. Abascal F, Zardoya R, Posada D: **ProtTest: selection of best-fit models of protein evolution.** *Bioinformatics* 2005, **21**:2104–2105.
54. Felsenstein J: **Inferring phylogenies from protein sequences by parsimony, distance, and likelihood methods.** *Methods Enzymol* 1996, **266**:418–427.
55. Schmidt HA, Strimmer K, Vingron M, von Haeseler A: **TREE-PUZZLE: maximum likelihood phylogenetic analysis using quartets and parallel computing.** *Bioinformatics* 2002, **18**:502–504.
56. Müller T, Vingron M: **Modeling amino acid replacement.** *J Comput Biol* 2000, **7**:761–776.
57. Huelsenbeck JP, Ronquist F: **MRBAYES: Bayesian inference of phylogenetic trees.** *Bioinformatics* 2001, **17**:754–755.
58. Kumar S, Tamura K, Nei M: **MEGA3: integrated software for molecular evolutionary genetics analysis and sequence alignment.** *Brief Bioinform* 2004, **5**:150–163.
59. Cantarel BL, Coutinho PM, Rancurel C, Bernard T, Lombard V, Henrissat B: **The carbohydrate-active EnZymes database (CAZy): an expert resource for glycogenomics.** *Nucleic Acids Res* 2009, **37**:D233–D238.
60. Khaldi N, Seifuddin FT, Turner G, Haft D, Nierman WC, Wolfe KH, Fedorova ND: **SMURF: genomic mapping of fungal secondary metabolite clusters.** *Fungal Genet Biol* 2010, **47**:736–741.
61. Blin K, Medema MH, Kazempour D, Fischbach MA, Breitling R, Takano E, Weber T: **antiSMASH 2.0 — a versatile platform for genome mining of secondary metabolite producers.** *Nucleic Acids Res* 2013, **41**:W204–W212.

doi:10.1186/1471-2164-15-635

**Cite this article as:** Yap et al.: The genome of the Tiger Milk mushroom, *Lignosus rhinocerotis*, provides insights into the genetic basis of its medicinal properties. *BMC Genomics* 2014 **15**:635.

**Submit your next manuscript to BioMed Central and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at  
www.biomedcentral.com/submit

