

Original Article

Effects of bone marrow-derived mesenchymal stem cells engraftment on vascular endothelial cell growth factor in lung tissue and plasma at early stage of smoke inhalation injury

Feng Zhu, Guang-hua Guo, Wen Chen, Nian-yun Wang

Department of Critical Care Medicine (Zhu F), Department of Burns (Guo GH, Chen W, Wang NY), First Affiliated Hospital of Nanchang University, Nanchang 330006, China

Corresponding Author: Feng Zhu, Email: 13907919783@139.com

BACKGROUND: This study was undertaken to determine the effect of mesenchymal stem cells (MSCs) engraftment on vascular endothelial cell growth factor (VEGF) in lung tissue, plasma and extravascular lung water at early stage of smoke inhalation injury.

METHODS: A rabbit smoke inhalation injury model was established using a home-made smoke inhalation injury generator, and rabbits were divided into two groups randomly: a control group (S group, $n=32$) and a MSCs treatment group (M group, $n=32$). 10 ml PBS was injected via the ear marginal vein immediately at injury into the S group. Third generation MSCs with a concentration of $1 \times 10^7/10$ ml PBS were injected via the ear marginal vein immediately at injury into the M group. VEGF in peripheral blood and lung tissue were measured at 0 (baseline), 2, 4 and 6 hours after injection respectively and analyzed. The right lungs of rabbits were taken to measure lung water mass fraction.

RESULTS: In the lung tissue, VEGF decreased gradually in the S group ($P<0.05$) and significantly decreased in the M group ($P<0.05$), but it increased more significantly than the values at the corresponding time points ($P<0.05$). In peripheral blood, VEGF increased gradually in the S group ($P<0.05$) and markedly increased in the M group ($P<0.05$), but it decreased more significantly than the values at corresponding time points ($P<0.05$).

CONCLUSION: MSCs engraftment to smoke inhalation injury could increase VEGF in lung tissue, decrease VEGF in plasma and reduce extravascular lung water, indicating its protective effect on smoke inhalation injury.

KEY WORDS: Mesenchymal stem cells; Smoke inhalation injury; Vascular endothelial cell growth factor; Extravascular lung water; Rabbit

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INTRODUCTION

Pulmonary injury from smoke inhalation is common in burn victims. It significantly contributes to the morbidity and mortality of patients with fire-related injuries, and as a chief contributor to the pathophysiology of smoke inhalation injury, it has been extensively studied.^[1] Vascular endothelial cell growth factor (VEGF) plays an important role in smoke inhalation injury.^[2]

Mesenchymal stem cells (MSCs) are multipotential nonhematopoietic progenitor cells capable of differentiating into multiple lineages of the mesenchyme. MSCs have emerged as a promising therapeutic modality for tissue regeneration and repair.^[3,4] But there is few report about the effects of MSCs engraftment on VEGF at early stage of somke inhalation injury. We hypothesized that, like other inflammatory cytokines

in lung injury, VEGF could increase local and systematic inflammatory response in smoke inhalation injury, while MSCs transplantation could decrease VEGF, contributing to smoke inhalation injury. In this study, we transplanted engraft MSCs into rabbits with smoke inhalation injury, and observed the change of VEGF in lung tissue and plasma.

METHODS

Preparation of MSCs

A total of 64 young healthy New Zealand white rabbits, weighing 2 kg, provided by the Animal Experiment Center of Nanchang University, were enrolled in this study. Three percent pentobarbital sodium (1 ml/kg) was used for anesthesia via the ear marginal vein. Bone marrow (4 ml) was obtained from bilateral tibial nodules and bilateral iliac crest, and was mixed with the equal amount of PBS for washing. After washing twice and centrifuged at 1500 r/min for 10 minutes, precipitate was discarded and adhered directly for primary culture. The precipitate was mixed with 10% of bovine serum containing DMEM-F12 medium (Hyclone Company) and put into an incubator at 37 °C with 5% CO₂ and saturated humidity. After the first 7-8 days when the cells reached 80% fusion, serial subcultivation started according to the proportion of 1:2, and third generation MSCs were digested by trypsin for further use. The cell-specific antigens were detected using flow cytometry and morphological observation to identify MSCs. Lee et al^[5] reported that there is no test that can be performed on a single cell to determine whether that cell is an MSC, so in this study we detected CD105, CD44, CD34 and CD45 for identification of MSCs.

Establishment of models with smoke inhalation injury

The procedures were conducted according to the principles and procedures outlined in the NIH Guide for the Care and Use of Laboratory Animals, and the protocols were approved by the Animal Care and Use Committee of Medical College of Nanchang University. Models were established by using a home-made smoke inhalation injury generator. The desiccated pine wood chip and kerosene were used to produce smoke. Rabbits inhaled smoke for 10 minutes and repeated at a 2-minute interval. The success of establishing severe smoke inhalation injury was evidenced by clinical manifestations, blood gas analysis and lung histopathological observation.^[6]

Sample collection

After establishment of the models, the 64 rabbits were randomly divided into two groups: an injury group (S group, *n*=32) and a treatment group (M group, *n*=32). The rabbits in the S group were injected with 10 ml PBS via the ear marginal vein immediately after injury, and served as a control; the rabbits in the M group were injected with PBS containing 1×10⁷/10 ml of third generation MSCs via the ear marginal vein immediately after injury. The lung tissue was collected for preparation of homogenate at pre-injury (baseline) and at 2, 4, and 6 hours post-injury respectively. VEGF concentration in the lung homogenate supernatant was detected by the enzyme-linked immunosorbent assay (ELISA). The test was performed in accordance with the kit instructions (VEGF quantitative enzyme-linked immunosorbent assay kit Carson offered by Shanghai Senxiong Science and Technology Industrial Co., Ltd.).

Detection of water content in the lung

At 6 hours after experiment, 0.5 g right lung tissue was collected and put into an incubator at 80 °C and dried for 72 hours to a constant weight.

Lung water content = (lung wet quality - lung dry quality) / lung wet quality × 100%.

Statistical analysis

Data were expressed as (mean± SD) and SPSS 15.0 for Windows was used for statistical analysis. Student's *t* test was used for the comparison between the two groups. *P*<0.05 was considered statistically significant.

RESULTS

Isolation, culture and identification of MSCs

Direct adhesion was used to separate MSCs in rabbits. MSCs grew well after passage. Under an invert microscope, MSCs were found to be spindle-shaped, numerous, and closely arranged. Cluster arrangement appeared at the early stage, and MSCs were equally distributed like whirlpool (Figure 1). CD34(-), CD45(-) and CD44(+), and CD105(+) were detected by flow cytometry, and this indicated that the cells were MSCs cultured (Figure 2).

Changes of VEGF in the lung tissue of rabbits

Compared to the baseline, VEGF in the S group and M group significantly decreased at 2, 4 and 6 hours after injury (*P*<0.05). VEGF concentration at every time point in the M group increased more significantly than in the S group (*P*<0.05) (Table 1).

Changes of VEGF of peripheral blood in rabbits

Compared to the baseline, VEGF in the S group and M group increased significantly at 2, 4 and 6 hours after injury ($P<0.05$). VEGF concentration at every time point in the M group decreased more significantly than in the S group ($P<0.05$) (Table 2).

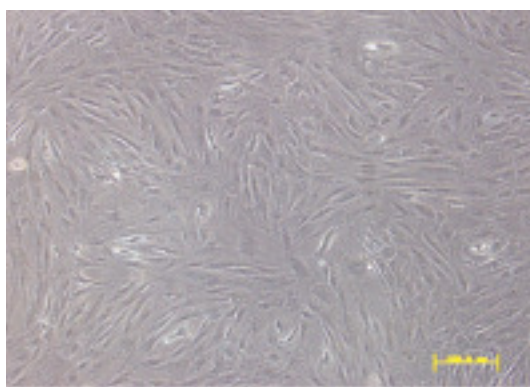
Changes of VEGF in the lung tissue of rabbits

Compared to the baseline, VEGF in the S group and M group significantly decreased at 2, 4 and 6 hours after

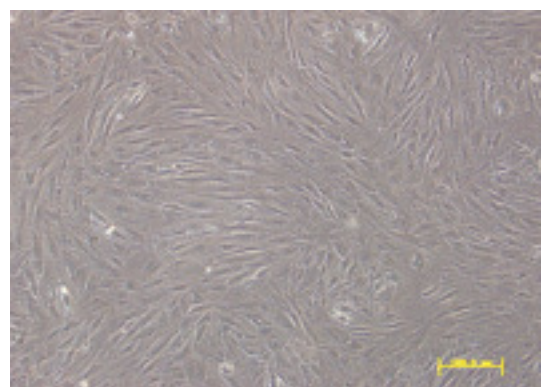
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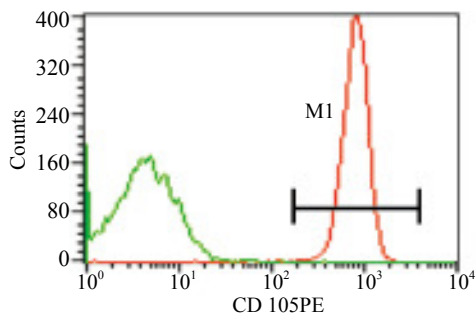


Primary culture

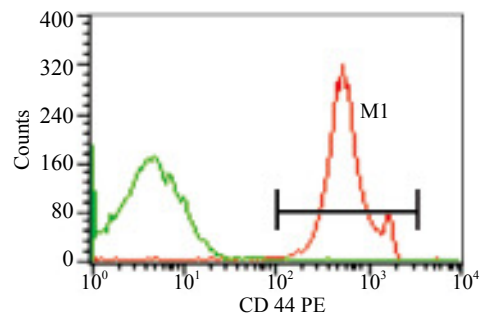


Third subculture

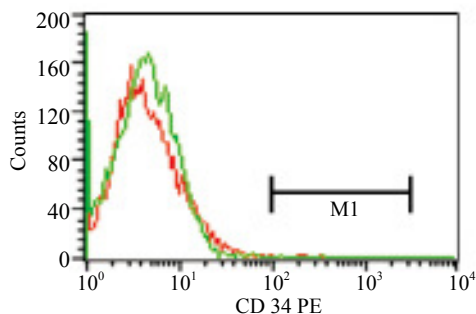
Figure 1. MSCs of the rabbits (original magnification×100)



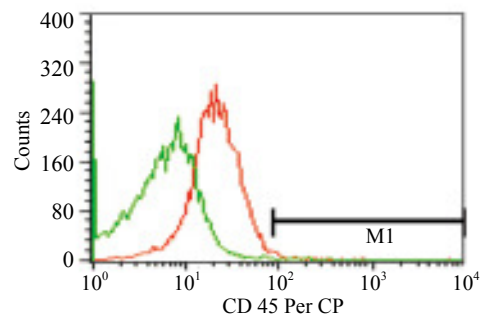
Marker	% Gated
AII	100.00
M1	99.79



Marker	% Gated
AII	100.00
M1	99.39



Marker	% Gated
AII	100.00
M1	0.11



Marker	% Gated
AII	100.00
M1	0.70

Figure 2 MSCs detected by flow cytometry in rabbits

Table 1. Change and comparison of VEGF at every time point in lung tissue (pg/ml, mean \pm SD, n=8)

Group	Baseline	2h post-injury	4h post-injury	6h post-injury
S	95.19 \pm 10.08	55.32 \pm 24.08 [#]	38.33 \pm 7.99 [#]	29.15 \pm 7.19 [#]
M	96.07 \pm 11.65	70.95 \pm 13.85 ^{*#}	50.74 \pm 14.10 ^{*#}	45.19 \pm 12.06 ^{*#}

Compared to values at every time points in the S group, * P <0.05; compared to baselines, [#] P <0.05

Table 2. Change and comparison of VEGF at every time point in plasma (pg/ml, mean \pm SD, n=8)

Group	Baseline	2h post-injury	4h post-injury	6h post-injury
S	25.19 \pm 1.78	70.62 \pm 4.08 [#]	98.39 \pm 3.09 [#]	119.77 \pm 7.49 [#]
M	23.98 \pm 1.65	50.95 \pm 3.85 ^{*#}	60.74 \pm 2.10 ^{*#}	85.19 \pm 2.06 ^{*#}

Compared to values at every time point in the S group, * P <0.05; compared to baselines, [#] P <0.05

Comparison of water content in the lung at 6 hours after injury

Lung water content was lower in the M group than in the S group (59.17% \pm 6.93% vs. 77.43% \pm 6.13%) at 6 hours after injury (P <0.05).

DISCUSSION

The essence of smoke inhalation injury is acute lung injury (ALI), and the main pathological characteristics of the injury include diffuse alveoli-capillary membrane damage, high permeability, pulmonary edema, lower pulmonary compliance, increased intrapulmonary shunt, significantly decreased pulmonary ventilation and exchange function leading to respiratory distress and anoxemia.^[7] As pro-inflammatory cytokines, VEGF, a stronger vascular permeability factor, participates in and mediates inflammatory reaction, formation of pulmonary edema and aggregation of neutrophilic granulocyte in the lung, which play an important role in progress of ALI. Investigations of inhibition and interruption of VEGF showed good prospects for the therapy of ALI.^[8,9] In this study, we found that VEGF in the lung homogenate decreased significantly at 2, 4 and 6 hours, but increased significantly in peripheral blood after smoke inhalation injury. The reasons may be that (1) the production of VEGF decreases significantly in lung homogenate because of destruction of type II alveoli after smoke inhalation injury; (2) the high permeability results in VEGF in lung tissue, flowing into peripheral blood along concentration gradient; and (3) the increased VEGF in plasma is relevant to some other factors, which promote VEGF production in peripheral blood such as activated neutrophilic granulocyte. But the underlying mechanisms are not clear.

Human MSCs avoid allorecognition, interfere with dendritic cells and T-cell function, and generate a local immunosuppressive microenvironment by secreting cytokines.^[10] It has been shown that the immunomodulatory function of human MSCs is enhanced when the cells are exposed to an

inflammatory environment characterized by the presence of elevated local interferon-gamma levels.^[11] Other studies showed contradictory findings, reflecting the highly heterogeneous nature of MSC isolates and the considerable differences between isolates generated by many different methods under development.^[12]

Mesenchymal stem cells can be activated and mobilized if necessary. However, the efficiency is very low. For instance, damage to muscles heals very slowly. However, if there is a method of activating the mesenchymal stem cells, such wounds would heal faster. Direct injection or placement of the cells into a site in need of repair may be the preferred method of treatment, as vascular delivery suffers from a "pulmonary first pass effect" where intravenously injected cells are sequestered in the lungs.^[13] Case reports in orthopedic applications have been published, though the number of patients treated is small and these methods still lack rigorous investigation demonstrating effectiveness. A study^[14] presented a series of nine defects in five knees involving surgical transplantation of mesenchymal stem cells with coverage of the treated chondral defects.

In this study, MSCs significantly increased VEGF in the lung tissue whereas decreased VEGF in peripheral blood and extravascular lung water. This may be due to the anti-inflammation and immunomodulation of MSCs i.e. (1) MSCs can adsorb and neutralize a large number of inflammatory factors and depress inflammatory reaction to alleviate type II alveoli and alveolar capillary membrane damage by expressing a great quantity receptor molecules on its surface,^[15] (2) MSCs can secrete some bioactive molecules such as VEGF,^[16-17] and this on the one hand complements loss of VEGF caused by synthetic inadequacy, on the other hand is against inflammatory factors (for instance TNF- α) and interrupt water fall effect of inflammatory signal in order to create condition for repair and regeneration of type II alveoli and alveolar capillary cells; (3) MSCs inhibit the activation and proliferation of immune cells by secreting inhibitory factor to reduce the production of factors inducing the production of VEGF.^[18,19]

VEGF plays an important role in generation and development of smoke inhalation injury. MSCs engraftment shows protective effect on smoke inhalation injury through regulation of the local and systematic VEGF and reduction of lung water content. But its underlying mechanism needs further clarification.

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Contributors: Zhu F proposed the study, and wrote the first draft. All authors contributed to the design and interpretation of the study and to further drafts.

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