

Biochem Biophys Res Commun. Author manuscript; available in PMC 2014 August 12.

Published in final edited form as:

Biochem Biophys Res Commun. 2010 July 30; 398(3): 513–518. doi:10.1016/j.bbrc.2010.06.110.

Establishment of sandwich ELISA for soluble alpha-Klotho measurement: Age-dependent change of soluble alpha-Klotho levels in healthy subjects

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Abstract

Background— α -Klotho (α Kl) regulates mineral metabolism such as calcium ion (Ca^{2+}) and inorganic phosphate (Pi) in circulation. Defects in mice result in clinical features resembling disorders found in human aging. Although the importance of transmembrane-type α Kl has been

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demonstrated, less is known regarding the physiological importance of soluble-type αKl (s αKl) in circulation.

Objectives—The aims of this study were: 1) to establish a sandwich ELISA system enabling detection of circulating serum $s\alpha Kl$, and 2) to determine reference values for $s\alpha Kl$ serum levels and relationship to indices of renal function, mineral metabolism, age and sex in healthy subjects.

Results—We successively developed an ELISA to measure serum $s\alpha Kl$ in healthy volunteers (n=142, males 66) of ages (61.1 \pm 18.5 yr). The levels (mean \pm SD) in these healthy control adults were as follows: total calcium (Ca; 9.46 \pm 0.41 mg/dL), Pi (3.63 \pm 0.51 mg/dL), Blood urea nitrogen (BUN; 15.7 \pm 4.3 mg/dL), creatinine (Cre; 0.69 \pm 0.14 mg/dL), 1,25 dihydroxyvitamin D (1,25(OH)₂D; 54.8 \pm 17.7 pg/mL), intact parathyroid hormone (iPTH; 49.2 \pm 20.6 pg/mL), calcitonin (26.0 \pm 12.3 pg/mL) and intact Fibroblast growth factor (FGF23; 43.8 \pm 17.6 pg/mL).

Serum levels of $s\alpha Kl$ ranged from 239 to 1266 pg/mL (mean \pm SD; 562 ± 146 pg/mL) in normal adults. Although $s\alpha Kl$ levels were not modified by gender or indices of mineral metabolism, $s\alpha Kl$ levels were inversely related to Cre and age. However, $s\alpha Kl$ levels in normal children (n=39, males 23, mean \pm SD; 7.1 ± 4.8 years) were significantly higher (mean \pm SD; 952 ± 282 pg/mL) than those in adults (mean \pm SD; 562 ± 146 , P<0.001). A multivariate linear regression analysis including children and adults in this study demonstrated that $s\alpha Kl$ correlated negatively with age and Ca, and positively with Pi. Finally, we measured a serum $s\alpha Kl$ from a patient with severe tumoral calcinosis derived from a homozygous missense mutation of α -klotho gene. In this patient, $s\alpha Kl$ level was notably lower than those of age matched controls.

Conclusion—We established a detection system to measure human serum $s\alpha Kl$ for the first time. Age, Ca and Pi seem to influence serum $s\alpha Kl$ levels in a normal population. This detection system should be an excellent tool for investigating $s\alpha Kl$ functions in mineral metabolism.

Keywords

alpha-Klotho; ELISA; calcium; phosphate; tumoral calcinosis

Introduction

 α -kl gene encodes a type I membrane protein with expression restricted to parathyroid glands, the choroid plexus and the kidney [1-4]. α Kl binds to Na⁺,K⁺-ATPase to regulate PTH secretion and is involved in transepitherial calcium concentration. In response to altered extra-cellular calcium concentrations, α Kl is rapidly translocated from endosomal organella to the plasma membrane together with Na⁺,K⁺-ATPase and simultaneously the extracellular domain of α Kl is cleaved and secreted into the blood circulation and cerebrospinal fluid (CSF) [5,6]. The increased Na⁺ gradient produced by elevated Na⁺,K⁺-ATPase activity drives PTH secretion in parathyroid glands and transepithelial transport of calcium in the kidney and choroid plexus [5]. Accordingly, it is assumed that α Kl levels in the serum and CSF mirror the molecular actions of the cellular form of α Kl in these tissues.

αKl also binds to fibroblast growth factor 23 (FGF23), which was discovered in studies of autosomal dominant hypophosphatemic rickets (ADHR) [7] and later tumor-induced osteomalacia (TIO) [8, 9]. FGF23 (i) is produced and secreted from bone in response to

serum levels of phosphorus and $1,25(OH)_2D$ [10-12], (ii) binds to FGF receptor1 (FGFR1), both suppressing 1α -hydroxylase (CYP27B1) expression and stimulating 24-hydroxylase (CYP24A1) expression in kidney [13,14], and (iii) downregulates protein amounts of Na⁺ dependent-phosphate transporter (NaPi) IIa/c to the brush border membrane of proximal tubules thus decreasing phosphate reabsorption [11].

 αKl contributes to integrate mineral homeostasis. Consequently, disturbances of αKl expression impair mineral metabolism via multiple mechanisms involving FGF23 signaling [13, 14], PTH secretion and transepithelial calcium transport. Recently, a patient with autosomal recessive hyperphosphatemic tumoral calcinosis shed new light on the impact of αKl [15]. Mutation analysis revealed a missense mutation in αKl , and in vitro studies indicated that αKl translocation to the plasma membrane was impaired [16]. Therefore, analysis of serum αKl levels may lead to greater understanding of disorders of mineral homeostasis.

In the present study, we developed an ELISA system to measure circulating $s\alpha Kl$ concentrations in serum from human subjects for the first time. We further analyzed and compared $s\alpha Kl$ levels of both healthy volunteers and a case with the α -klotho gene mutation [16]. Finally, we discuss the potential utility in measuring serum αKl in clinical disorders.

Materials and methods

Plasmid construction

Human full length α -Klotho (fl- α Kl; 1012 amino acids(a.a), RefSeq ID: NP_004786)-cDNA and cDNA encoding extracellular domain of α -Klotho (s α Kl; 1-979a.a.) were amplified from total human kidney cDNAs by PCR and consequently cloned into pLP-CMVneo and pLP-IRESneo, respectively, by In fusion PCR kit (Clonetech).

Cell culture

pLP-CMVneo-fl- α Kl was transfected into HEK293 cells by the calcium-phosphate method. pLP-IRESneo-s α Kl was transfected into CHO cells, by the Lipofectamine method (Invitrogen). Then, cells stably expressing either fl- α Kl or s α Kl were selected by G418 and cloned by limiting dilution. HEK293 cells expressing fl- α Kl were grown in DMEM supplemented with 10% Fetal Calf Serum (FCS) and 1mg/mL of G418. CHO cells expressing s α Kl were grown in MEM α supplemented with 10% FCS and 2mg/mL of G418.

saKI protein purification

Recombinant saKl protein was purified from serum-free conditioned media (SFMII, Gibco) of CHO cells stably expressing saKl by a series of chromatographic steps employing Q Sepharose XL, Butyl Sepharose FF, Heparin Sepharose HP, Q Sepharose HP, SP Sepharose HP, DEAE Sepharose FF, Phenyl Sepharose HP, Q Sepharose HP, Lentil Lectin sepharose 4B, and DEAE Sepharose FF (All sepharoses were purchased from GE healthcare). The concentration of purified saKl protein was determined by absorbance at 280 nm.

Animals

All mice were purchased from Charlesriver Inc. Japan and were maintained in SPF conditions according to the institutional guidelines of Kyowa Hakko Kirin Co., Ltd.

Antibodies

Monoclonal anti-human α -Klotho antibodies were generated by immunizing B6D2F1 mice once a week for 5 weeks with pLP-CMVneo-fl- α Kl vector by TransIT in vivo gene delivery system (Takara Bio). Hybridomas were obtained by fusing spleen cells and Sp2/o mouse myeloma cells (ATCC ID CRL1581) with polyethylene glycol. Then, hybridomas producing anti- α -Klotho antibody were selected by cell ELISA and flow cytometry analysis by using HEK293 cells stably expressing fl- α Kl. Finally, nine monoclonal antibodies were purified with Protein G Sepharose 4FF (GE healthcare) and we next tested all pairs of antibodies with high affinity against recombinant s α KL for sandwich ELISA by using conjugated antibodies with biotin as detection antibodies. A pair of antibodies that have the highest reactivity to s α Kl protein were named 67G3 (IgG1, kappa) as a capture antibody and 91F1 (IgG1, kappa) as a detection antibody, respectively. Rabbit anti-human α -Klotho polyclonal antibodies (N116 and C939) were generated by immunization of two different human α -Klotho partial polypeptides from N-terminal region (PLQPATGDVASDSYNNVFRDT, 116-136a.a.) and C-terminal region (ASMKHYRKIIDSNGFPGPETLERFC, 939-963a.a.), respectively.

Subjects

Healthy Asian volunteers (n=181, 89 males) between 0.1 and 88 years of age participated in our study. Subjects had no known medical conditions. A patient with a missense mutation of the *a-klotho* gene was described in a previous paper [16]. The study protocol was approved by the ethics committee on human research at Kyoto University Graduate School of Medicine, Tokyo Metropolitan Children's Medical Center, Osaka University Hospital, Nara Medical University Hospital and Indiana University. Written informed consent was obtained from subjects prior to enrollment. All subjects had normal serum Ca, Pi, 1,25(OH)₂D, iPTH, calcitonin, FGF23 levels and normal renal function as assessed by BUN and serum Cre levels.

Measurement of serum parameters

Blood samples were drawn from a forearm vein in the morning after overnight fasting. Sera were obtained by centrifugation and immediately stored at -30° C. Serum FGF23 concentrations were measured by a FGF23 kit (Kainos) which can only measure the intact FGF23 [9]. Ca, Pi, BUN, Cre and Alb were measured at SRL Inc. Japan using standard clinical methods. Serum levels of iPTH, calcitonin and $1,25(OH)_2D$ were measured only in adults (>20 years) at SRL Inc. because of limited serum quantity in children. The mean values of age and serum biochemical tests from each group are shown in Table 1.

Immunoprecipitation and Western blot

500μL of sera from two healthy subjects and purified sαKl protein were incubated with resin (NHS-activated Sepharose 4 FF; GE healthcare) conjugated with 67G3, 91F1 or isotype-

matched control antibody (mouse IgG1) at 4°C for 2h. The specifically retained proteins on the resin were solubilized by boiling in SDS-PAGE sample buffer with the reducing agent and separated by SDS-PAGE. Following electrophoresis, proteins were transferred to PVDF membrane and bound with the biotin labeled N116 or C939 antibodies followed by HRP-conjugated streptavidin (DAKO). Protein bands were visualized by the ECL system (GE healthcare)

ELISA procedure

Wells of microtiter plates were coated with 1 μ g/mL of 67G3 monoclonal antibody in 100 μ L of carbonate buffer (pH9.5) for 18h at 4 °C and then blocked with 1% BSA in PBS for 1h at 37°C. Samples (50 μ L) were premixed with dilution buffer (50 μ L; PBS, pH7.4, containing 0.05% Tween20, 1% BSA, and 50 μ g /mL purified normal mouse IgG). The mixtures were loaded and incubated for 1h at room temperature (RT), followed by the addition of 100 μ L 91F1-Fab' conjugated with HRP by using N-(6-Maleimidocaproyloxy)succinimide (EMCS, Dojindo) in PBS for 30 min at RT, and the reaction was visualized by the addition of 100 μ L of chromogenic substrate (TMB+, Dako) for 30 min at RT. The reaction was stopped with 100 μ L of 1N H₂SO₄ and absorbance at 450nm was measured with a reduction at 570 nm using ELISA plate reader. Plates were washed 6 times with washing buffer (PBS, pH7.4, containing 0.1% (v/v) Tween 20) after each step. A standard curve was established by a serial dilution of recombinant sαKl.

Results

Establishment of anti-aKl mouse monoclonal antibodies for sandwich ELISA

To establish mouse monoclonal antibodies with strong affinity for human αKl protein, we immunized mice with a human full-length αKl expressing vector and screened the resulting hybridomas by measuring the binding affinities to human αKl expressing cells. Nine antibodies which showed high and specific affinities to an extracellular domain of human αKl protein were tested for compatibility as a capture antibody or detection antibody in sandwich ELISA to detect saKl protein. We finally selected a pair of antibodies (67G3 for a capture antibody and 91F1 for a detection antibody). While these antibodies are not useful for Western blot assays, both antibodies have a high affinity in immunoprecipitation to saKl protein as well as in Cell ELISA and Flow cytometry to a transmembrane type of αKl (data not shown), indicating that both antibodies specifically recognize a tertiary protein structure of an extracellular domain of αKl .

Sandwich ELISA for the detection of saKI in human sera

A typical standard curve is shown in Fig.1A of the purified recombinant human $s\alpha Kl$ protein concentration between 94 pg/mL and 6000 pg/mL. The intra- and inter-assay coefficients of variation (CV) were calculated by measuring three different doses with purified human $s\alpha Kl$ protein. As shown in Table S1A and S1B, both CVs ranged from 2.7 to 9.8%. The accuracy of this assay in the presence of human serum and plasma was determined by serially diluting recombinant human $s\alpha Kl$ in 50% human serum and plasma. The measured $s\alpha Kl$ levels were then compared with theoretical $s\alpha Kl$ levels contributed by the addition of recombinant $s\alpha Kl$ and the endogenous $s\alpha Kl$ in serum and plasma. As shown in Table S1C, measured $s\alpha Kl$

levels were 92.6% (in serum) and 94.1% (in plasma) of expected values. Furthermore, the serum dose-response curves were parallel to the purified recombinant saKl dose-response curves (data not shown). Moreover, addition of recombinant human purified FGF23 (1µg/mL) did not affect a measurement (data not shown). Serum samples subjected to quick freezing at -80° C and thawing at RT did not show a significant decrease during three cycles (data not shown). These results indicate that saKl can be measured with acceptable accuracy in the presence of 50% human serum or plasma.

Immunoprecipitation of plasma saKI protein from healthy subjects

To address whether both antibodies bind to the endogenous saKl in human plasma as well as the recombinant extracellular (soluble) α Kl, immunoprecipitation was performed. Both 67G3 and 91F1 antibodies immunoprecipitated saKl from plasma from healthy adult subjects (Fig.1B) as well as the recombinant human saKl (rhsaKl). Furthermore, the densities of the immunopositive bands for precipitated saKl (about 130kDa) precisely corresponded to the concentrations of saKl represented by ELISA (rhsaKl: 5000 pg/mL, plasma 1: 575 pg/mL and plasma 2: 1272 pg/mL). These data suggest that both capture and detection antibodies can bind to endogenous saKl in blood, indicating that our sandwich ELISA system can specifically detect and accurately measure the circulating saKl.

Circulating saKl concentrations in healthy adults

In order to determine the reference range of serum $s\alpha Kl$ and to address the correlation of several mineral parameters with $s\alpha Kl$ in healthy adults, we next analyzed serum levels of $s\alpha Kl$ and indices of mineral metabolism in 142 healthy adults (age 20 years old or over) (Table 1). The serum $s\alpha Kl$ concentrations of healthy adults ranged from 239 to 1266 pg/mL (mean \pm SD; 562 ± 146 pg/mL). There was no apparent correlation of $s\alpha Kl$ levels with gender, iPTH, 1,25(OH)₂D, calcitonin, Ca, Pi, BUN, Cre or FGF23 by simple regression analyses. However, $s\alpha Kl$ levels were slightly, but significantly correlated with age (r=-0.199, P=0.017) and Cre (r=-0.183, P=0.030) by simple regression analyses.

Circulating sa KI concentrations in healthy children

As mineral metabolism dramatically changes in childhood, we compared concentrations of $s\alpha Kl$ between adults and children (<17 years) (Table 1). As expected, all factors measured in this study (Ca, Pi, Cre, BUN and FGF23) were significantly different between adults and children. The serum $s\alpha Kl$ concentrations of normal children were significantly higher than those of adults (mean \pm SD; 952 \pm 282 pg/mL in children vs 562 ± 146 pg/mL in adults, student's t-test, P<0.001, Fig.2A), indicating that serum $s\alpha Kl$ correlated with age. The extent of correlation of $s\alpha Kl$ with other serum factors examined by simple regression analysis was shown in Fig.2B-F. S αKl levels were found to correlate considerably with Pi (Fig.2C r=0.517, P<0.001), Cre (Fig.2E, r=-0.538, P<0.001) and FGF23 (Fig.2F r=-0.350, P<0.001) concentrations as well as age (Fig2B, r=-0.599, P<0.001). In addition, $s\alpha Kl$ levels also slightly, but significantly, correlated with BUN (Fig.2D, r=-0.203, P=0.006) but $s\alpha Kl$ levels did not significantly correlate with calcium.

Next, factors independently associated with serum $s\alpha Kl$ levels were examined using a multiple regression model. Serum $s\alpha Kl$ exhibited significant and independent associations

with age (P<0.001), Ca (P=0.004) and Pi (P=0.001), but no significant associations with BUN (P=0.411), Cre (P=0.051), or FGF23 (P=0.089) as shown in Table S2. Pi was positively correlated with s α Kl. Interestingly, Ca and age were negatively correlated with s α Kl in this model, while Ca was a not significant factor with s α Kl by simple regression analysis.

Examination of saKl levels in a patient with a aKl mutation

Ichikawa et al. have recently reported a case of hyperphosphatemic tumoral calcinosis with hypervitaminosis D due to missense mutations in human α -klotho gene (a single amino acid substitution from histidine 193 to arginine). The authors concluded that, at least in part, disruption of the architecture of α Kl protein results in a failure to translocate to the plasma membrane [16]. Consequently the subject's s α Kl level was expected to be low. To test this prediction, we measured serum from the patient and found that the serum s α Kl level was 337 pg/mL. Because the patient was 14 years old, we prepared serum from age-matched controls (n=11, mean \pm SD: 13.5 \pm 1.7 years from 11.2 to 16.6 years old). As shown in Fig. 2G, s α Kl levels of age-matched controls ranged from 652 to 1146 pg/mL, and these values all fit within the mean value \pm 2SD. As anticipated, the subject with α -Klotho mutation had low concentrations of s α Kl compared with age-matched controls.

Discussion

We developed an ELISA to successfully measure $s\alpha Kl$ in human subjects and demonstrated positive relationships with phosphorus, and negative relationships with age and Ca^{2+} in multiple linear regression analysis. Since low calcium stimuli induce αKl secretion [5], the negative relationship appears compatible. We have previously proposed a novel negative feedback system involving FGF23 and αKl for maintenance of normal phosphate and 1,25(OH)₂D levels [16,13]. Based on the fact that FGF23 is a negative regulator of phosphate concentration (high FGF23 suppresses phosphate reabsorption in kidney), FGF23 correlates with phosphate in a positive manner (high phosphate induces high FGF23 level). Predictably, $s\alpha Kl$ correlates negatively with FGF23, but positively with phosphate in simple regression analysis. We have proposed a mineral homeostatic system integrated by αKl , where Ca^{2+} and PTH levels are acutely regulated by Na^+, K^+ -ATPase activity [5] and phosphate and 1,25(OH)₂D levels are suppressed by FGF23 signals at late phase [8]. In this system, FGF23 signals lead to the suppression of α -klotho gene expression and the decreased synthesis of 1,25(OH)₂D, an enhancer of α -klotho gene expression. A negative correlation between $s\alpha Kl$ and FGF23 levels could reflect this putative feedback system.

FGF23 concentrations are maintained at a low or normal level regardless of high phosphate concentrations in children (Fig.S1). Since high phosphate has been reported to induce high FGF23, low or normal FGF23 concentrations in children seem to be paradoxical. Although the mechanism is unknown, this is postulated to be related to a higher need for adequate bone mineralization during childhood. In contrast, $s\alpha Kl$ is maintained at a high concentration in children. Based on the preceding statistics that $s\alpha Kl$ correlates positively with phosphate, the high value of $s\alpha Kl$ is not inconsistent with a high phosphate level even at a young age. The balance of mineral metabolism in children is different from that in

adults. It is very likely that a shift of metabolic equilibrium sustains higher phosphate concentration and optimizes Ca^{2+} reabsorption in tubules. Further studies are required to delineate the mechanism.

The manifestations seen in the patient with the α -klotho gene mutation resembled features observed in α -klotho ablated mice, although the patient's phenotype was milder than that observed in the mouse model. In an attempt to understand the manifestations seen in this patient, we verified saKl levels using the ELISA system. Since the mutated protein impairs the intracellular trafficking ability in vitro, the saKl level of the patient was expected to be low. Indeed, the saKl value was more than 50% lower than the mean value of healthy agematched subjects. Accordingly, the ELISA may be useful in studying disorders of mineral metabolism.

 $S\alpha Kl$ levels are higher in adolescence and lower in older adults and negatively correlate with Cre and BUN, which are increased in an age-dependent fashion even in healthy humans. Therefore, $s\alpha Kl$ concentrations in serum are expected to be noted as a biomarker for kidney function. In the present study, we are not able to analyze the effect of renal function on $s\alpha Kl$ and vice versa, since our subjects had normal renal function. Notably, in renal failure, α -klotho mRNA expression is decreased [18], though this may be due to death of the cells in the nephron expressing αKl . Further study is necessary to determine the utility of this ELISA in renal failure.

Kurosu et al reported that an increase of saKl levels by genetic manipulation of full-length α -Klotho resulted in a prolonged life span in mice [19]. In that paper, since the levels of normal saKl in mice were measured at >10ng/ml by radioimmunoassay [19], they were estimated to be approximately 20 fold greater than the average normal human saKl concentration in this study. However, as previously reported, murine saKl levels were calculated to be similar to that of humans by immunoprecipitation followed by western blot analysis [6]. These findings indicate that serum saKl concentration should be carefully estimated to evaluate the roles of saKl level in life span elongation. In addition, whereas two to three-fold elevations in saKl level were noted in longevity-elongated mice [19], this magnitude would be within the divergence seen in healthy human subjects. Furthermore, this cross-sectioned study identified that younger subjects have higher saKl, and that older subjects (who have lived longer) have lower saKl. Therefore, careful prospective longitudinal studies would be needed to determine whether future longevity can be predictable from saKl concentrations at one point in time.

Acknowledgments

This work was supported in part by the grants from Ministry of Education, Science and Culture in Japan, 21026017, 21390058 (to A.I.) and 17109004 (to Y.N.) and the Ministry of Health, Labour and Welfare of Japan KH20Q007a-1 (to K.O.). We thank K Sakuma, K Ono and N Yoshii for excellent technical support and Dr. T Sakai for helpful suggestion. We thank the coordinators of Fujiwara-kyo Study [20] from which almost healthy adults were recruited. Notably, subjects from Fujiwara-kyo Study had no history of cardiovascular diseases and diabetes mellitus and no other known medical conditions. Their blood pressures were within the normal range and their serum creatinine levels were less than 1.2mg/dL.

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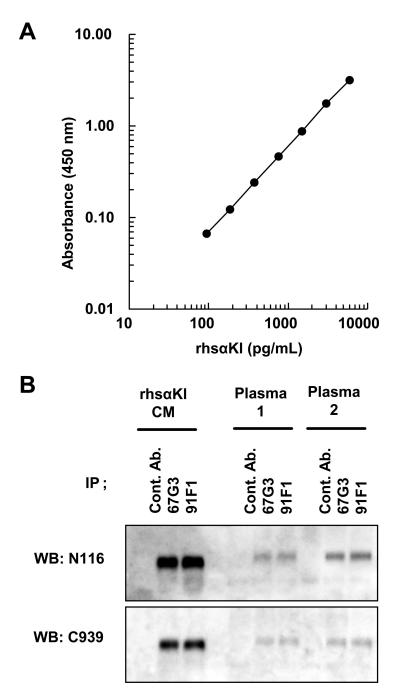
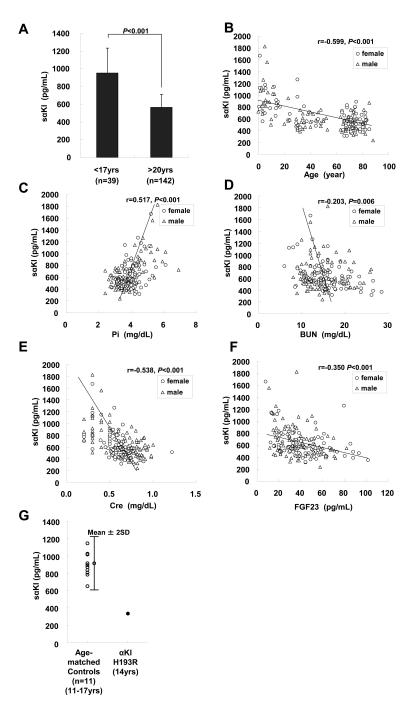


Fig.1. Enzyme-linked immunosorbent assay (ELISA) and immunoprecipitation analysis by two useful anti-saKl monoclonal antibodies. (A) Typical dose-response curve of recombinant human saKl (rhsaKl). Sandwich ELISA for saKl was performed by 67G3 as a capture antibody and 91F1 as a detection antibody. The results are expressed as the mean \pm standard deviation (SD) of triplicate measurements. (B) Immunoprecipitation of plasma saKl by 67G3 and 91F1 antibodies. Conditioned medium from CHO cells stably expressing FGF23 (rhsaKl) and plasma from healthy volunteers (plasma 1, plasma 2) were immunoprecipitated

anti-Klotho antibodies (67G3 and 91F1) and control mouse IgG1 (cont. Ab.), and analyzed by Western blot with anti-human Klotho rabbit polyclonal antibodies, N116 and C939.



Serum sαKl levels (A) Comparison of serum sαKl levels between healthy adults and children. Serum sαKl levels of healthy adults (n=142, age: >20 years) and children (n=39, age: <17years) were measured. Results represent mean ± standard deviation (SD). Statistical analysis was performed by student's t-test. P values less than 0.05 were taken as statistically significant. Correlation of serum sαKl with (B) age (C) Pi (D) BUN (E) Cre and (F) FGF23 levels in simple regression analysis in healthy subjects. The open circles and triangles represent male and female subjects, respectively. Statistical analysis was performed by

Pearson correlation. P values less than 0.05 were taken as statistically significant. (G) Comparison of serum $s\alpha Kl$ levels between a patient with klotho missense mutation and agematched healthy subjects. Serum $s\alpha Kl$ levels from 11 healthy subjects (left, age: from 11.2 to 16.6 years) and a patient with klotho homozygous gene mutation (right, age: 14 years) were shown. An error bar along the values for healthy controls represents mean \pm 2 fold of standard deviation (2SD) in order to indicate a reference range.

Table 1

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number:	total (male/female)	all 181 (89/92)	children (<17yrs) 39 (23/16)	adults (>20yrs) 142 (66/76)	P children vs adults
Age	year	49.4 ± 27.7	7.1 ± 4.8	61.1 ± 18.5	< 0.001
Ca	mg/dL	9.50 ± 0.44	9.65 ± 0.52	9.46 ± 0.41	0.018
Pi	mg/dL	3.90 ± 0.75	4.88 ± 0.68	3.63 ± 0.51	< 0.001
Ca x Pi	$(mg/dL)^2$	37.1 ± 8.0	47.2 ± 7.9	34.4 ± 5.3	< 0.001
BUN	mg/dL	15.2 ± 4.3	13.3 ± 3.6	15.7 ± 4.3	0.002
Cre	mg/dL	0.62 ± 0.19	0.36 ± 0.12	0.69 ± 0.14	< 0.001
Alb	mg/dL	4315 ± 384	4190 ± 474	4349 ± 349	0.021
$1,25(OH)_2D$	pg/mL	ND	ND	54.8 ± 17.7	ND
iPTH	pg/mL	ND	ND	49.2 ± 20.6	ND
calcitonin	pg/mL	ND	ND	26.0 ± 12.3	ND
FGF23	pg/mL	39.6 ± 18.4	24.2 ± 12.0	43.8 ± 17.6	< 0.001

ND: not determined