

Cellular trafficking and anticancer activity of *Garcinia mangostana* extract-encapsulated polymeric nanoparticles

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Abstract: *Garcinia mangostana* Linn extract (GME) is a natural product that has received considerable attention in cancer therapy, and has the potential to reduce side effects of chemotherapeutics and improve efficacy. We formulated GME-encapsulated ethyl cellulose (GME-EC) and a polymer blend of ethyl cellulose and methyl cellulose (GME-EC/MC) nanoparticles. We achieved high drug-loading and encapsulation efficiency using a solvent-displacement method with particle sizes around 250 nm. Cellular uptake and accumulation of GME was higher for GME-encapsulated nanoparticles compared to free GME. In vitro cytotoxicity analysis showed effective anticancer activity of GME-EC and GME-EC/MC nanoparticles in HeLa cells in a dose-dependent manner. GME-EC/MC nanoparticles showed approximately twofold-higher anticancer activity compared to GME-EC nanoparticles, likely due to their enhanced bioavailability. GME-encapsulated nanoparticles primarily entered HeLa cells by clathrin-mediated endocytosis and trafficked through the endolysosomal pathway. As far as we know, this is the first report on the cellular uptake and intracellular trafficking mechanism of drug-loaded cellulose-based nanoparticles. In summary, encapsulation of GME using cellulose-derivative nanoparticles – GME-EC and GME-EC/MC nanoparticles – successfully improved the bioavailability of GME in aqueous solution, enhanced cellular uptake, and displayed effective anticancer activity.

Keywords: phytotherapy, cancer, nanoparticles, cellulose, intracellular trafficking

Introduction

Leading chemotherapeutic agents, such as paclitaxel, cisplatin, and doxorubicin, are widely used in the clinic for cancer treatment; however, systemic side effects and the development of drug resistance have limited their therapeutic efficiency. To overcome these potential problems with chemotherapeutics, a number of phytochemical and herb extracts that show anticancer activities are being widely explored for cancer therapy.^{1–5} Mangosteen (*Garcinia mangostana* Linn) is a well-known tropical fruit in Southeast Asia; xanthenes from mangosteen pericarp extracts such as α -mangostin, β -mangostin, γ -mangostin, garcinone E, and gartanin, have received considerable interest in cancer prevention and cancer therapeutics.^{6,7} Several studies have reported anticancer activities of xanthenes isolated from *G. mangostana* Linn extract (GME) in various human cancer cell lines, including those from the liver, breast, and lungs, and gastric, colorectal, and cervical cancers.^{8–13} These anticancer activities involve cell-cycle arrest, inhibition of cancer cell proliferation, induction of apoptosis, and inhibition of adhesion, invasion, and metastasis of tumors.^{3,6} However, GME's poor aqueous solubility and low bioavailability are major problems that have limited its therapeutic efficiency and pharmaceutical application.¹⁴

Polymeric particles are one of the most promising strategies to improve the solubility and bioavailability of poorly soluble drugs.^{15–17} Drug-encapsulated polymeric nanoparticles are being widely developed for cancer therapy, due to their high drug-encapsulation efficiency, passive tumor targeting, excellent endocytosis efficiency, and ability to deliver a wide range of therapeutic agents.^{15–17} Although pharmacological evidence of the anticancer effects of GME is growing, surprisingly there are few reported studies on the development of GME-encapsulated polymeric nanoparticles. Recently, α -mangostin and xanthenes were encapsulated in polyvinylpyrrolidone micelles and polyethylene glycol-b-poly(*N*-(2-hydroxypropyl)methacrylamide-dilactate) micelles, respectively, and showed effective antitumor activity.^{18,19} Cellulose derivatives, such as ethyl cellulose (EC) and methyl cellulose (MC), are among the most commonly used polymers in the pharmaceutical industry, because they are biocompatible, safe, and inexpensive.²⁰ For example, EC is a hydrophobic polymer that can encapsulate a number of hydrophobic drugs and natural substances. EC can also assemble into nanoparticles when MC, a hydrophilic polymer, is used as a blend polymer (EC/MC) to achieve controlled release of drugs.^{20,21}

Recently, understanding the mechanism of cellular entry and intracellular trafficking of nanoparticles has attracted a great deal of interest.^{22–24} Nanoparticles can enter cells via utilization of different endocytic pathways, including clathrin-mediated endocytosis, caveolae-mediated endocytosis, macropinocytosis, and other non-clathrin- and non-caveolae-mediated endocytosis.^{22–25} It has been reported that the cellular uptake mechanism of nanoparticles depends on the physicochemical properties of the particles, including their shape, size, surface chemistry, and particle compositions, as well as different cell types.^{24–32} Despite their wide use in the pharmaceutical industries, the cellular uptake and intracellular trafficking mechanism of therapeutic cellulose-based nanoparticles have not been previously reported.

In this study, we developed two GME-encapsulated cellulose derivative nanoparticles, GME-EC and GME-EC/MC, with high drug-loading and encapsulation efficiency (EE). The physicochemical properties of GME-encapsulated nanoparticles were analyzed by transmission electron microscopy (TEM), hydrodynamic particle size, and zeta-potential measurements. In vitro cytotoxicity of GME-EC and GME-EC/MC nanoparticles against a cancer cell line (HeLa cells) was evaluated. Finally, we report cellular uptake and intracellular trafficking mechanisms of GME-encapsulated nanoparticles in HeLa cells.

Materials and methods

Materials

GME with 56% α -mangostin (content by weight) was a generous gift from Tipco (Bangkok, Thailand). EC (viscosity 250–300 cP, ethoxy content 48%), MC, (viscosity 400 cP, 1.60%–1.90% degree of methoxy substitution), dialysis tubing cellulose membrane (molecular weight [MW] 12,400 Da), coumarin 6, chlorpromazine, genistein, amiloride, methyl- β -cyclodextrin, lovastatin, and all other chemicals were purchased from Sigma-Aldrich (St Louis, MO, USA). Lab-Tek™ eight-well glass-bottom tissue-culture plates, minimum essential medium, fetal bovine serum, penicillin/streptomycin (pen/strep), ProLong® Gold antifade reagent, and LysoTracker® Red were from Thermo Fisher Scientific (Waltham, MA, USA).

Preparation and characterization of GME-encapsulated nanoparticles

GME-encapsulated nanoparticles (EC and blended EC/MC) were prepared by the solvent-displacement method as previously described, with a slight modification.^{21,33} GME and EC were dissolved in ethanol (5 mg/mL), while GME and 1:1 (w:w) EC/MC mixture was dissolved in 80% (v/v) ethanol/water (1 mg/mL). Both mixtures (10 mL) were placed into a dialysis bag (cellulose membrane, MW cutoff [MWCO] 12,400 Da) and dialyzed against deionized (DI) water for five exchanges (5×1,000 mL). The resulting aqueous suspension was collected, and particle morphology was characterized by TEM (H7600; Hitachi, Tokyo, Japan). Hydrodynamic particle size and zeta potential of nanoparticles were evaluated using a Zetasizer Nano ZS (Malvern Instruments, Malvern, UK).

The amount of encapsulated GME in the nanoparticles was determined by using an ultraviolet-visible (UV-vis) spectrophotometer (BioMate 3; Thermo Fisher Scientific). Briefly, 5 mL of each GME-encapsulated nanoparticle combination – GME-EC and GME-EC/MC – was concentrated by centrifugation at 3,000× *g* for 20 minutes using an Amicon Ultra-15 membrane (MWCO 100,000; EMD Millipore, Billerica, MA, USA). After centrifugation, the resulting nanoparticles in the membrane were soaked in 5 mL of ethanol for 3 hours to extract GME from the nanoparticles. The amount of GME in the ethanol extract (absorbance at 317 nm) was determined using a calibration curve. Encapsulation efficiency (EE) and loading capacity were calculated as follows:

$$EE (\%) = \frac{\text{Weight of encapsulated GME}}{\text{Weight of GME used}} \times 100 \quad (1)$$

$$\text{Loading (\%)} = \frac{\text{Weight of encapsulated GME}}{\text{Weight of GME loading nanoparticles}} \times 100 \quad (2)$$

In vitro GME release

The release of GME from GBM-EC and GME-EC/MC nanoparticles was measured at 37°C in 1× phosphate-buffered saline (PBS) pH 7.4. The particle suspensions were spray-dried using a Büchi 190 mini spray-dryer (Büchi Labortechnik, Flawil, Switzerland). Eight milligrams of dried GME-EC and GME-EC/MC were resuspended in 20 mL of release medium (1× PBS) and then placed in a dialysis bag (MW 12,000 Da), with continuous stirring at 37°C. A standard curve was generated from the known GME solutions prepared in the same release medium. A small aliquot of the release medium was withdrawn to analyze the amount of released GME outside the dialysis bag at 0, 1, 2, 4, 6, 8, and 24 hours, using the BioMate 3 UV-vis spectrophotometer at 320 nm. The percentage of GME release was calculated as follows:

$$\text{Release (\%)} = \frac{\text{Weight of released GME}}{\text{Weight of GME used}} \times 100 \quad (3)$$

Cell culture

Human cervical carcinoma HeLa cells were obtained from the American Type Culture Collection (Manassas, VA, USA). The cells were cultured at 37°C in 5% CO₂ in minimum essential medium supplemented with 10% fetal bovine serum and 1% pen/strep. For confocal microscopy, cells were seeded between 2.0×10³ and 2.5×10³ cells per plate onto Lab-Tek eight-well glass-bottom culture plates and incubated overnight at 37°C. After incubation, the cell-culture medium was replaced with fresh media before nanoparticles were added.

In vitro cytotoxicity study

The cytotoxicity of free GME and GME-encapsulated nanoparticles was determined using a standard 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay. HeLa cells were plated onto 96-well plates at a seeding density of 1.0×10⁴ cells per well in 200 mL media and incubated overnight. Our test samples were free GME dissolved in dimethyl sulfoxide (DMSO), free GME dispersed in DI water, blank EC nanoparticles, blank EC/MC nanoparticles, GME-EC, and GME-ECMC nanoparticles. The samples were diluted to 1–100 µg/mL with the culture

medium and then added into each well and incubated for 48 hours. The equivalent amounts of DMSO, sterile DI water, and untreated cells were used as controls. After incubation, the medium was replaced with a mixture of 160 µL of fresh medium and 40 µL of 2 mg/mL MTT reagent and incubated at 37°C for 4 hours in the dark. Finally, the MTT solution was carefully removed, and 200 µL of DMSO was added in each well and incubated for 30 minutes to completely dissolve the MTT formazan crystals. The absorbance was measured at 570 nm using the plate reader (BioMate 3). Cell viability (%) and half-maximal inhibitory concentration (IC₅₀) were then calculated for each sample.

Drug uptake and accumulation in cells

The cellular uptake and accumulation of free GME dissolved in DMSO, free GME dispersed in DI water, and GME-encapsulated nanoparticles were investigated by flow cytometry. HeLa cells were seeded onto 12-well plates at a density of 2.0×10⁵ cells and incubated overnight at 37°C, 5% CO₂. The old media was replaced with 1 mL serum-free media with different samples at a final concentration of 5 µg/mL. After 1 hour of incubation, the media was removed and the cells washed three times with 1× PBS to remove extracellular GME and/or nanoparticles. Then, cells were trypsinized (0.05% trypsin–ethylenediaminetetraacetic acid [EDTA]) and resuspended in cold 1× PBS. Cell-associated fluorescence (10,000 cells per sample) was detected using a FACSCalibur™ flow cytometer (BD, Franklin Lakes, NJ, USA) to determine the mean fluorescence intensity originating from the fluorescence property of GME (488 nm excitation, 530±15 nm emission). Data from 10,000 events were gated using forward- and side-scatter parameters to exclude cell debris. Standard deviations were calculated from six replicates.

Confocal microscopy

An LSM 510 meta-confocal microscope (Carl Zeiss Meditec, Jena, Germany) was used to capture images. For multicolor microscopy, samples were excited with 405, 488, 543, and 633 nm laser lines, and images were captured by multitracking to avoid bleed-through between fluorophores. All samples were maintained at 37°C and 5% CO₂, and observed under a 63× Plan Apo 1.4 NA oil-immersion lens (Nikon, Tokyo, Japan).

Cellular trafficking of nanoparticles

To study the cellular uptake and intracellular trafficking of nanoparticles, we encapsulated coumarin 6, a

hydrophobic fluorescent dye, into EC/MC (C-EC/MC) nanoparticles (2% w/w) instead of GME. This is because the fluorescence intensity of GME within nanoparticles was not strong enough to image them with high resolution by confocal microscopy. For the colocalization study, HeLa cells were seeded between 2.0×10^3 and 2.5×10^3 cells per plate onto Lab-Tek eight-well glass-bottom culture plates and incubated overnight at 37°C . After overnight incubation, C-EC/MC nanoparticles were added to cells and incubated for 1, 2, and 4 hours. Cells were stained with LysoTracker[®] (100 nM; Thermo Fisher Scientific) for 30 minutes before imaging. After staining, cells were washed three times with $1 \times$ PBS and fixed with 2% formaldehyde and 0.2% glutaraldehyde in $1 \times$ PBS for 15 minutes. Finally, the ProLong Gold reagent with 4',6-diamidino-2-phenylindole solution was added to the sample and imaged with the LSM 510 Meta confocal microscope. The percentage of colocalization between nanoparticles and the LysoTracker was analyzed with MetaMorph[®] software (Molecular Devices, Sunnyvale, CA, USA) on a per-pixel basis.^{24,34,35}

Drug inhibition of endocytic pathway

To determine the mechanism of cellular entry for GME-EC/MC nanoparticles, HeLa cells were plated at a seeding density of 2.0×10^5 cells/well in 12-well plates in serum-free media. Prior to GME-EC/MC nanoparticle addition, serum-free media with different endocytic pathway inhibitors, chlorpromazine (10 $\mu\text{g}/\text{mL}$), sucrose (0.45 M), and genistein (200 μM) were added to the cells and incubated for 30 minutes, followed by the addition of nanoparticles. For methyl- β -cyclodextrin (10 mM) and lovastatin (1 $\mu\text{g}/\text{mL}$), cells were treated for 15 minutes, and then nanoparticles were added and incubated for 30 minutes. For 4°C samples, they were chilled for 30 minutes before the addition of nanoparticles and maintained at 4°C during the incubation period (1 hour). After incubation, all samples were washed three times with $1 \times$ PBS to remove extracellular nanoparticles. Cells were harvested with 0.05% trypsin-EDTA and resuspended with cold $1 \times$ PBS. Cell-associated fluorescence (10,000 cells per sample) was detected using the FACSCalibur flow cytometer to determine the mean fluorescence intensity originating from the fluorescence property of GME (488 nm excitation, 530 ± 15 nm emission). Data from 10,000 events were gated using forward- and side-scatter parameters to exclude cell debris. Standard deviations were calculated from three replicates.

Statistical analysis

Statistical analysis of data was performed using a two-tailed Student's *t*-test or one-way analysis of variance with Tukey's honest significant difference test using SPSS 18.0 (SPSS Inc., Chicago, IL, USA). Differences were considered to be statistically significant at a level of $P < 0.05$.

Results and discussion

Characterization of nanoparticles

We encapsulated GME into cellulose-derivative polymeric nanoparticles – GME-EC and GME-EC/MC – using a solvent-displacement method. We formulated GME-EC/MC nanoparticles at a lower polymer concentration compared to GME-EC nanoparticles, since the MC chains were highly water-soluble and contributed to the swelling of EC/MC nanoparticles.^{21,33} Both GME-EC and GME-EC/MC nanoparticles achieved high drug-loading efficiency and EE. We achieved an EE of $86.2\% \pm 3.9\%$ and $87.4\% \pm 4.1\%$ at GME loading of $46.7\% \pm 2.2\%$ and $46.7\% \pm 2.2\%$ for GME-EC and GME-EC/MC nanoparticles, respectively (Table 1). GME is a hydrophobic molecule that is suitable for efficient encapsulation into cellulose-derivative nanoparticles.^{18,19} The morphology of GME-encapsulated nanoparticles was confirmed by TEM, which showed spherical shapes for both types of nanoparticles (Figure 1).

Dynamic light-scattering analysis showed that the hydrodynamic diameters of the GME-EC and GME-EC/MC nanoparticles were 253.3 ± 8.6 nm and 249.2 ± 13.1 nm, respectively (Table 1). The zeta potential of the GME-EC and GME-EC/MC nanoparticles was -30.9 ± 0.4 mV and -11.7 ± 0.5 mV, respectively (Table 1). As ethanol was slowly displaced by water, the polymeric chains in EC likely self-assembled into spherical nanoparticles with the hydroxyl groups of the sugar units arranged outside the nanoparticles, while the ethoxy portions oriented away from the surrounding water. This arrangement aided nanoparticles to suspend in water, and at the same time encapsulated hydrophobic GME within the core of nanoparticles.^{21,33} For EC/MC nanoparticles, the MC chains likely entangled with the EC chains as the nanoparticles were formed. Therefore, the shell of EC/MC nanoparticles would have been composed of an EC/MC polymer blend. We concluded that GME-encapsulated nanoparticles improved the bioavailability of GME.

In vitro cytotoxicity to cancer cells

The anticancer activity of GME-encapsulated nanoparticles, free GME dissolved in DMSO or in water, and blank EC and

Table 1 Physicochemical properties, drug-encapsulation efficiency (EE), and drug-loading capacity of GME-encapsulated nanoparticles

Formulation	Diameter (nm) ^a	PDI ^b	ζ-potential (mV) ^c	EE (%)	Loading (%)
GME-EC	253.3±8.6	0.05±0.01	-30.9±0.4	86.2±3.9	43.1±1.9
GME-EC/MC	249.2±13.1	0.07±0.04	-11.7±0.5	87.5±4.1	43.5±2.1
C-EC	239.9±16.9	0.12±0.04	-21.7±1.9	-	-
C-EC/MC	233.8±8.1	0.12±0.02	-18.8±2.2	-	-

Notes: ^aDiameter (number mean) measured by dynamic light scattering; data represent the average of three independent experiments ± SD; ^bthe polydispersity index (PDI), measured by dynamic light scattering, represents relative variance in particle-size distribution; a PDI of 1 indicates a large distribution of particle size, whereas a PDI of 0 indicates a monodisperse size distribution; data represent the average of three independent experiments ± SD; ^cmeasured at pH 7.1; data represent the average of three independent experiments ± SD.

Abbreviations: GME, *Garcinia mangostana* Linn extract; EC, ethyl cellulose; MC, methyl cellulose; SD, standard deviation; C, coumarin 6.

EC/MC nanoparticles were tested using the MTT assay. Both blank EC and blank EC/MC nanoparticles did not show any cytotoxic activities at the concentrations that we tested (Figure 2 and Table 2). This showed the biocompatible and non-toxic nature of our cellulose-based nanoparticles. Free GME dissolved in DMSO or in water and GME-encapsulated nanoparticles (GME-EC and GME-EC/MC nanoparticles) showed a dose-dependent cytotoxicity in HeLa cells (Figure 2A). The order of cytotoxic activities (IC_{50}) was: free GME dissolved in DMSO (IC_{50} 7.33 μ g/mL) > GME-EC/MC nanoparticles (IC_{50} 7.43) > GME-EC nanoparticles (IC_{50} 16.17) > free GME dissolved in water (IC_{50} 23.09) (Table 2). Low cytotoxicity observed from free GME dissolved in water was not surprising, since GME has poor solubility in water, which limits its activity. In contrast, both GME-encapsulated nanoparticles that were dispersed in water – GME-EC and GME-EC/MC nanoparticles – showed significantly higher cytotoxic activities compared to free GME dissolved in water. These results indicate that the encapsulation of GME into a polymeric nanoparticle platform improved the bioavailability of GME.

GME-EC/MC nanoparticles showed approximately twofold-higher cytotoxicity in HeLa cells compared to

GME-EC nanoparticles. This result is in good agreement with previous reports of Suwannateep et al where encapsulation of curcumin in EC/MC blend nanoparticles showed higher cytotoxicity compared to curcumin-encapsulated EC nanoparticles.^{21,33} The in vitro release rate of GME from GBM-EC and GME-EC/MC nanoparticles was comparable, with no statistically significant differences (Figure 3). These results indicated that the difference in IC_{50} between GME-EC and GME-EC/MC nanoparticles was not due to the difference in their GME release rates after 24 hours. Our results show that different types of polymer compositions of nanoparticles can significantly affect the in vitro cytotoxicity of GME. Interestingly, at the highest GME concentration that we tested, 100 μ g/mL, both GME-EC and GME-EC/MC nanoparticles showed higher cytotoxicity than free GME dissolved in DMSO (Figures 2B and S1).

Cellular uptake and endocytic pathway of nanoparticles

Since GME-EC/MC nanoparticles showed the most promising anticancer activity, we investigated cellular uptake and intracellular trafficking using this nanoparticle formulation.

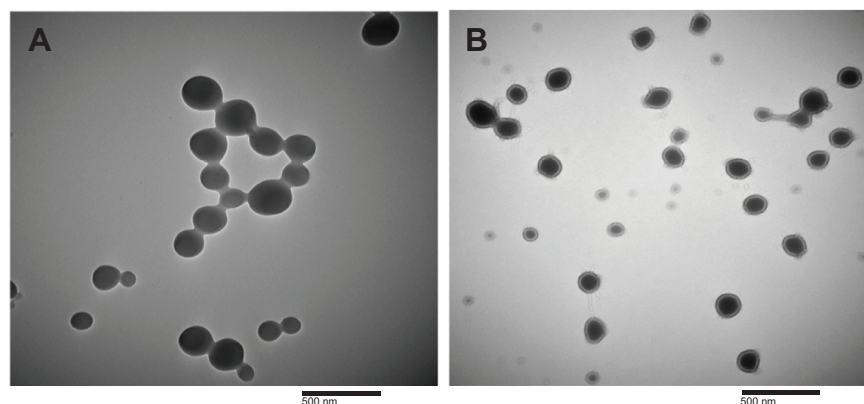


Figure 1 Transmission electron microscopy images of (A) GME-encapsulated EC nanoparticles and (B) GME-encapsulated EC/MC nanoparticles at 50,000 \times magnification.

Note: The scale bar denotes 500 nm.

Abbreviations: GME, *Garcinia mangostana* Linn extract; EC, ethyl cellulose; MC, methyl cellulose.

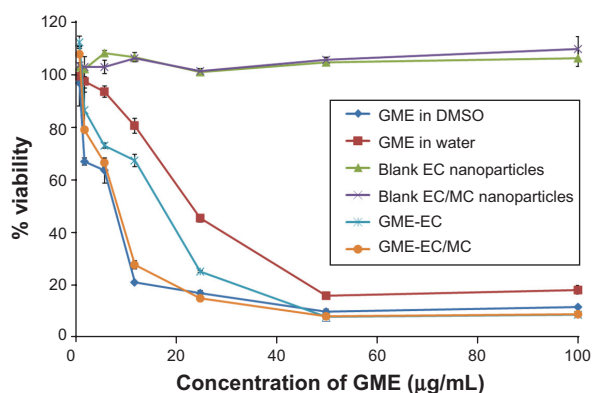


Figure 2 Cytotoxic activity in HeLa cells using different formulations of free GME and GME-encapsulated nanoparticles.

Notes: Cell viability was evaluated by MTT assay. Data represent means \pm standard deviation of three separate experiments ($n=3$).

Abbreviations: GME, *Garcinia mangostana* Linn extract; DMSO, dimethyl sulfoxide; EC, ethyl cellulose; MC, methyl cellulose; MTT, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide.

GME-EC/MC nanoparticles showed significantly higher cellular uptake in HeLa cells compared to free GME dissolved in DI water or in DMSO (Figure 4). These results suggest that the encapsulation of GME in cellulose-derivative nanoparticles is an effective strategy to enhance the cellular uptake and accumulation of GME. As discussed earlier, the cytotoxicity of GME-EC/MC nanoparticles was comparable to free GME dissolved in DMSO, which was likely due to slow and sustained release of encapsulated GME from GME-EC/MC nanoparticles. Therefore, combined with enhanced cellular uptake and accumulation, GME-encapsulated nanoparticles may provide a better alternative for GME cancer therapy.

To determine the endocytic pathway of GME-encapsulated nanoparticles, we measured the uptake of GME-EC/MC nanoparticles in HeLa cells treated with different endocytic pathway inhibitors, using flow cytometry. Compared to the untreated control cells, the uptake of GME-EC/MC nanoparticles was significantly reduced ($P<0.05$) following treatment of cells with chlorpromazine, which blocks clathrin-mediated endocytosis by causing clathrin to accumulate in late endosomes (Figure 5A). Similarly, hypertonic sucrose

Table 2 IC_{50} values for free GME and GME-encapsulated nanoparticles

Formulation	IC_{50} ($\mu\text{g/mL}$)
GME in DMSO	7.33
GME in water	23.09
GME-EC	16.17
GME-EC/MC	7.43

Abbreviations: GME, *Garcinia mangostana* Linn extract; DMSO, dimethyl sulfoxide; EC, ethyl cellulose; MC, methyl cellulose; IC_{50} , half-maximal inhibitory concentration.

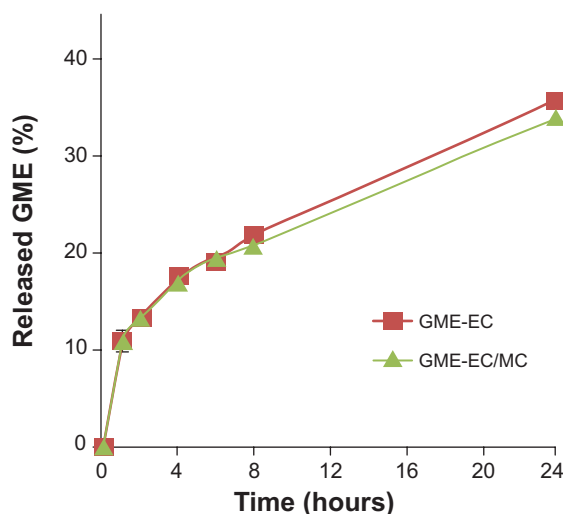


Figure 3 Cumulative in vitro release of GME from GME-EC and GME-EC/MC nanoparticles in 1x PBS (pH 7.4).

Note: Data represent means \pm standard deviation of three separate experiments ($n=3$).
Abbreviations: GME, *Garcinia mangostana* Linn extract; EC, ethyl cellulose; MC, methyl cellulose; PBS, phosphate-buffered saline.

(0.45 M), which inhibits clathrin-mediated endocytosis, also reduced the cellular uptake of GME-EC/MC nanoparticles (~23% reduction). In contrast, the cellular uptake of GME-EC/MC nanoparticles was not affected by genistein or amiloride, inhibitors of caveolae-mediated uptake and macropinocytosis, respectively (Figure 5, B and D). In addition, the uptake of GME-EC/MC nanoparticles was significantly inhibited when the cells were incubated at 4°C, confirming that they entered HeLa cells via energy-dependent endocytosis and not via direct penetration of plasma membrane (Figure 5C). The cellular uptake of GME-EC/MC nanoparticles was significantly reduced ($P<0.05$) by cholesterol depletion, using methyl- β -cyclodextrin (extracts

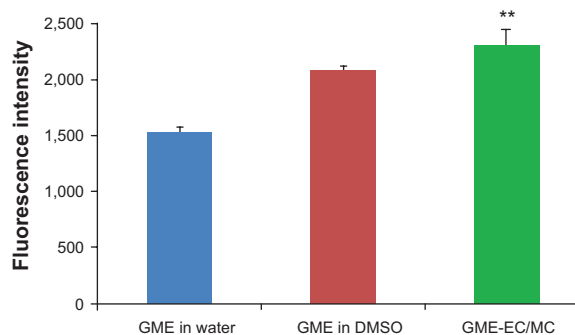


Figure 4 Cellular uptake and accumulation of free GME and GME-encapsulated nanoparticles in HeLa cells.

Notes: Data represent means \pm standard deviation of three separate experiments ($n=3$). Statistical analysis was performed using analysis of variance with Tukey's test. $**P<0.05$

Abbreviations: GME, *Garcinia mangostana* Linn extract; DMSO, dimethyl sulfoxide; EC, ethyl cellulose; MC, methyl cellulose.

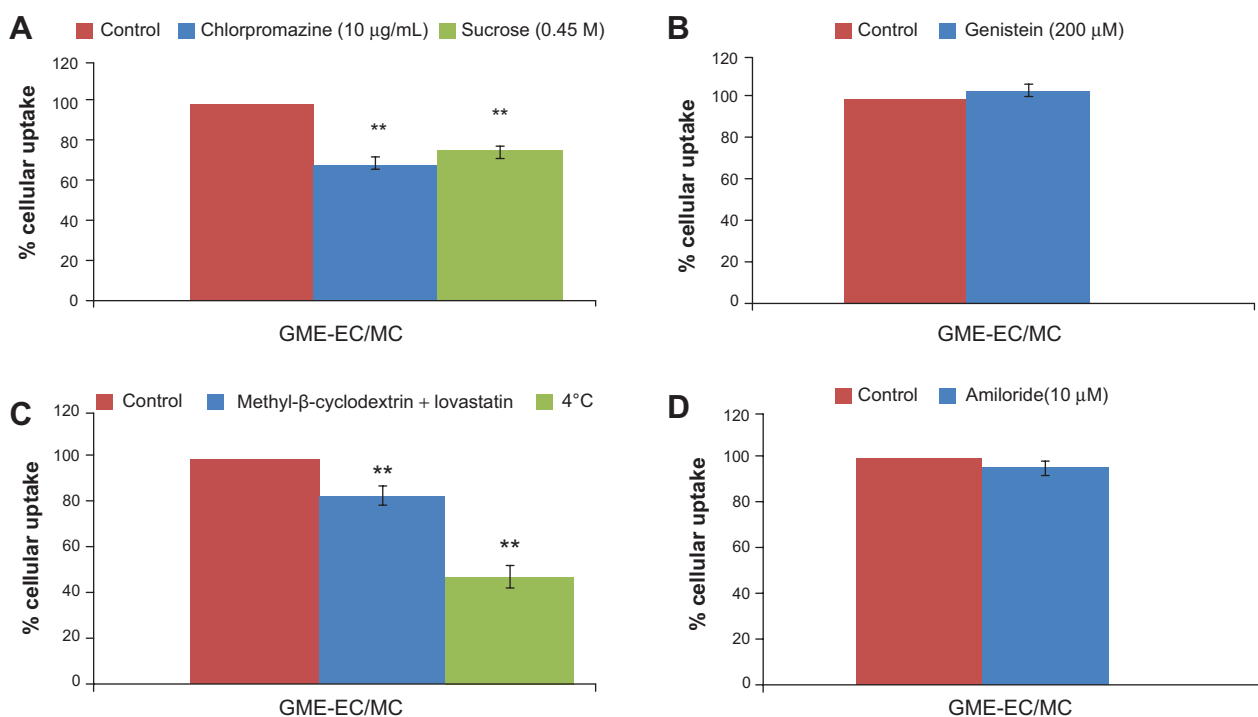


Figure 5 Endocytic mechanism of GME-EC/MC nanoparticles in HeLa cells. Cells were treated with various endocytic inhibitors prior to nanoparticle addition.

Notes: (A) Chlorpromazine and sucrose 0.45 M, inhibitors of clathrin-mediated endocytosis; (B) genistein, an inhibitor of caveolae-mediated endocytosis; (C) methyl-β-cyclodextrin and lovastatin, which depleted cholesterol, and uptake of nanoparticles at 4°C; and (D) amiloride, an inhibitor of macropinocytosis. ** $P < 0.05$ compared to no-treatment control, calculated using two-tailed Student's *t*-test.

Abbreviations: GME, *Garcinia mangostana* Linn extract; EC, ethyl cellulose; MC, methyl cellulose.

cholesterol from plasma membrane) and lovastatin (inhibitor of de novo cholesterol synthesis), in good agreement with the cholesterol-dependent nature of clathrin-mediated endocytosis (Figure 5C).^{36–39} Together, these findings suggest that GME-EC/MC nanoparticles entered HeLa cells primarily via clathrin-mediated endocytosis.

Intracellular trafficking of nanoparticles

The general fate of most nanoparticles following clathrin-mediated endocytosis is the endolysosomal pathway, where early endosomes mature into late endosomes, and finally to lysosomes.⁴⁰ To determine whether EC/MC nanoparticles were trafficked via the endolysosomal pathway, we performed colocalization studies using the LysoTracker, a well-characterized marker for late endosomes and lysosomes. The hydrodynamic size and zeta potential of C-EC/MC nanoparticles were similar to GME-EC/MC nanoparticles (Table 1). Confocal microscopy images showed a high degree of colocalization between C-EC/MC nanoparticles and the LysoTracker within 1 hour (Figure 6), which continued to increase significantly until the 2-hour time point (Figure 6). Interestingly, the degree of colocalization between C-EC/MC nanoparticles and the LysoTracker

decreased at the 4-hour time point (Figure 6), which suggests that C-EC/MC nanoparticles were providing controlled release of drugs once the particles were entrapped in the acidic vesicles. This is in good agreement with previous reports, where curcumin-encapsulated EC/MC nanoparticles showed faster drug release at lower pH, compared to physiological pH.^{21,33} These results indicate that EC/MC nanoparticles traffic through the endolysosomal pathway following their cellular entry via clathrin-mediated endocytosis.

Conclusion

In this study, GME (with 56% α-mangostin content) was successfully encapsulated into cellulose-derivative nanoparticles – GME-EC and GME-EC/MC nanoparticles – with high EE (~87%) and loading capacity (~46%). GME-EC and GME-EC/MC nanoparticles showed spherical morphology and a particle size around 250 nm with surface charges of –30.9 mV and –11.7 mV, respectively. GME-EC and GME-EC/MC nanoparticles showed effective anticancer activity in HeLa cells, and enhanced the cellular uptake of encapsulated GME compared to free GME. GME-encapsulated nanoparticles primarily entered HeLa cells by clathrin-mediated

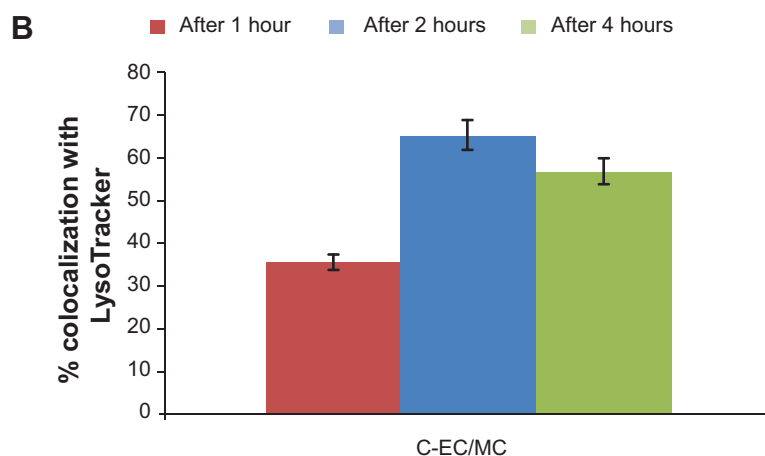
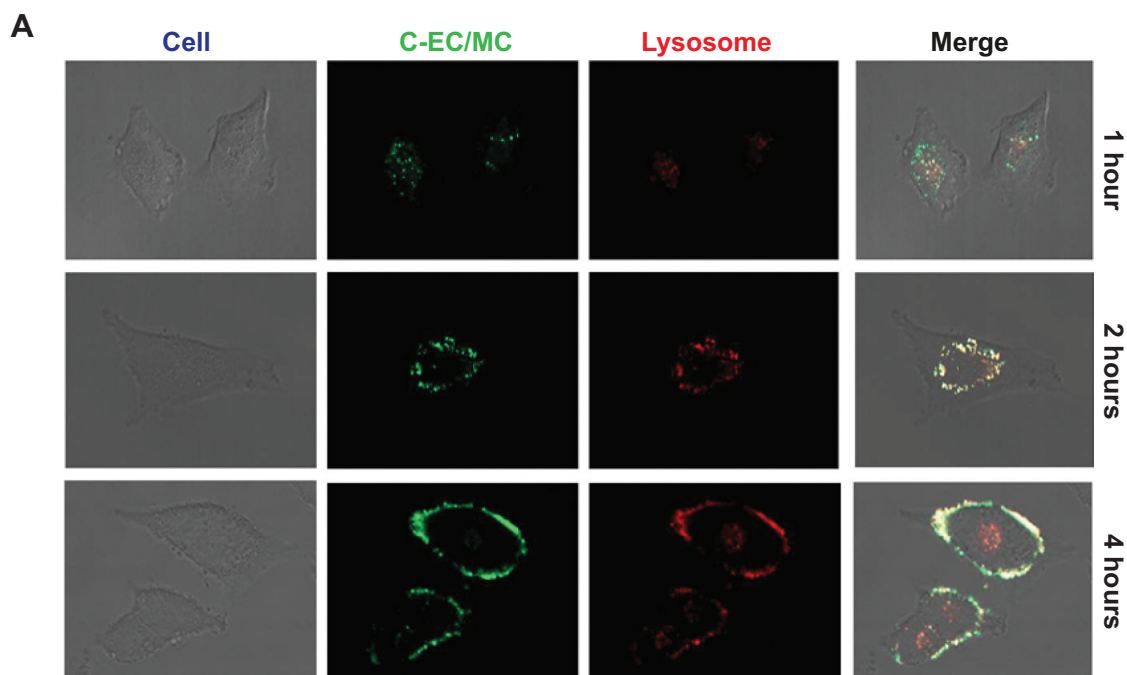


Figure 6 Representative confocal microscopy images of (A) C-EC/MC nanoparticles (green) with LysoTracker (red) in HeLa cells. The colocalization between C-EC/MC nanoparticles and LysoTracker are shown in the merged images (yellow). (B) Quantification of colocalization analysis at different time points.

Abbreviations: EC, ethyl cellulose; MC, methyl cellulose; C, coumarin 6.

endocytosis and were trafficked through the endolysosomal pathway. Our results show that GME-encapsulated cellulose-derivative nanoparticles, especially GME-EC/MC nanoparticles, have the potential to provide a better alternative strategy for cancer therapy.

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Disclosure

The authors report no conflicts of interest in this work.

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Supplementary material

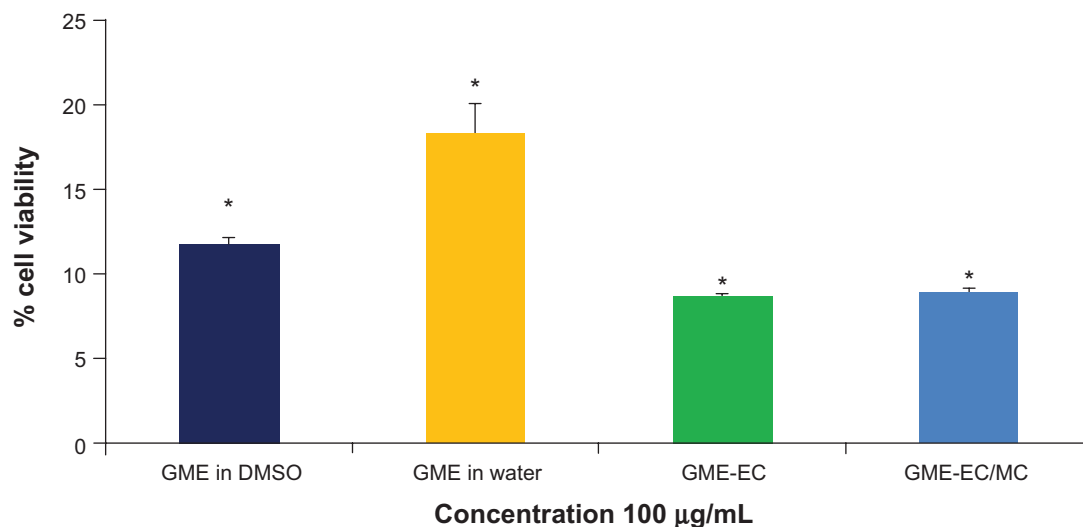


Figure S1 Cytotoxic activity of different formulation of 100 µg/mL free GME and GME-encapsulated nanoparticles in HeLa cells.

Notes: Data represent means \pm standard deviation (n=3). * $P < 0.05$. Statistical analysis was performed by analysis of variance with Tukey's honest significant difference test.

Abbreviations: GME, *Garcinia mangostana* Linn extract; EC, ethyl cellulose; MC, methyl cellulose; DMSO, dimethyl sulfoxide.

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