

Submit a Manuscript: http://www.wjgnet.com/esps/ Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx DOI: 10.3748/wjg.v20.i30.10238

*World J Gastroenterol* 2014 August 14; 20(30): 10238-10248 ISSN 1007-9327 (print) ISSN 2219-2840 (online) © 2014 Baishideng Publishing Group Inc. All rights reserved.

*TOPIC HIGHLIGHT*

## **WJG 20<sup>th</sup> Anniversary Special Issues (1): Hepatocellular carcinoma**

# **Hepatitis B virus, HBx mutants and their role in hepatocellular carcinoma**

Ashraf Ali, Hany Abdel-Hafiz, Mohd Suhail, Amany Al-Mars, Mohammad Khalid Zakaria, Kaneez Fatima, Sultan Ahmad, Esam Azhar, Adeel Chaudhary, Ishtiaq Qadri

Ashraf Ali, Mohd Suhail, Amany Al-Mars, Sultan Ahmad, Adeel Chaudhary, Ishtiaq Qadri, Department of Medical Biotechnology, King Fahad Medical Research Center, King Abdul Aziz University, PO Box 80216, Jeddah 21589, Saudi Arabia Hany Abdel-Hafiz, Division of Endocrinology, Department of Medicine and Metabolism, Anschutz Medical Campus, University of Colorado Denver, Aurora, CO 80045, United States Mohammad Khalid Zakaria, Department of Biochemistry, All India Institute of Medical Sciences, New Delhi 110029, India Kaneez Fatima, IQ Institute of Infection and Immunity, Lahore 54000, Pakistan Esam Azhar, Special Infectious Agents Unit-Biosafety Level 3, King Fahd Medical Research Center, King Abdul Aziz University, Jeddah 21589, Saudi Arabia Esam Azhar, Medical Laboratory Technology Department, Faculty of Applied Medical Sciences, King Abdul Aziz University, Jeddah 21589, Saudi Arabia Author contributions: Ali A and Qadri I conceived the idea, carried out the literature search, manuscript writing and final editing; Abdel-Hafiz H edited the manuscript while Suhail M, Mars A, Sultan A and Fatima K helped in referencing, editing and figure preparation and rectifying some grammatical errors; Financial and technical assistance to Ali A and Suhail M was provided by Azhar E and Chaudhary A.

Correspondence to: Ishtiaq Qadri, PhD, Professor, Head Medical Biotechnology, Head Liver Biology, Head Translational Research, Department of Medical Biotechnology, King Fahd Medical Research Center, King Abdul Aziz University, PO Box 80216, Jeddah 21589, Saudi Arabia. ishtiaq80262@yahoo.com

Telephone: +966-640-1000Fax: +966-6952-5321 Received: February 23, 2014 Revised: April 30, 2014 Accepted: May 25, 2014

Published online: August 14, 2014

### **Abstract**

Hepatocellular carcinoma (HCC) is one of the leading causes of death induced by cancer in the modern world and majority of the cases are related to chronic hepatitis B virus (HBV) infection. HBV-encoded X protein (HBx)

is known to play a pivotal role in the pathogenesis of viral induced HCC. HBx is a multifunctional protein of 17 kDa which modulates several cellular processes by direct or indirect interaction with a repertoire of host factors resulting in HCC. HBX might interfere with several cellular processes such as oxidative stress, DNA repair, signal transduction, transcription, protein degradation, cell cycle progression and apoptosis. A number of reports have indicated that HBx is one of the most common viral ORFs that is often integrated into the host genome and its sequence variants play a crucial role in HCC. By mutational or deletion analysis it was shown that carboxy terminal of HBx has a likely role in protein-protein interactions, transcriptional transactivation, DNA repair, cell, signaling and pathogenesis of HCC. The accumulated evidence thus far suggests that it is difficult to understand the mechanistic nature of HBx associated HCC, and HBx mediated transcriptional transactivation and signaling pathways may be a major determinant. This article addresses the role of HBx in the development of HCC with particular emphasis on HBx mutants and their putative targets.

© 2014 Baishideng Publishing Group Inc. All rights reserved.

**Key words:** Hepatitis B virus; Hepatocellular carcinoma; Transcription factors; Apoptosis; Epigenetics; Mutants; Tumor necrosis factor; Activating protein; Transforming growth factor; Mitogen activated protein kinase

**Core tip:** The available evidence supports a well-defined for the hepatitis B virus-encoded X protein (HBx) protein in viral-induced hepatocellular carcinoma. The progression cell cycle, transactivation potential, compromised DNA repair, inhibition the tumor suppressor gene and senescence-related factors are few of the key pathways by which HBx is known to promote pathogenesis. The activation and inhibition of cellular calcium and tyrosine kinase signaling pathways are some of the



other pathways modulated by HBx expression. Individually charged amino acids (K-130, V-131, D-120, 121) and or steches of amino acids (132-140) present in the carboxy-terminus of HBx protein seems to enhance the protein's ability to deregulates cellular processes.

Ali A, Abdel-Hafiz H, Suhail M, Al-Mars A, Zakaria MK, Fatima K, Ahmad S, Azhar E, Chaudhary A, Qadri I. Hepatitis B virus, HBx mutants and their role in hepatocellular carcinoma. *World J Gastroenterol* 2014; 20(30): 10238-10248 Available from: URL: http://www.wjgnet.com/1007-9327/full/v20/i30/10238.htm DOI: http://dx.doi.org/10.3748/wjg.v20.i30.10238

#### **INTRODUCTION**

Throughout the world, hepatocellular carcinoma (HCC) is the sixth most commonly occurring cancer and is the third leading cause of death due to its complex nature $l<sup>H</sup>$ . Majority of the HCC cases occur as a result of Hepatitis B virus (HBV) infection $^{[2]}$ . HCC is more common in Asia-Pacific region and Africa. Due to high frequency of reported HBV cases, persistent HBV exposure triggers progression of HCC. More than 50% of the sufferers of HCC have history of HBV infection<sup>[2]</sup>. The estimated annual worldwide incidence is around 500000 and 1000000. In the last two decades, annual incidence in the United States has also increased by 80**%**[3]. HBV is the smallest ds DNA virus known to infect humans. It is the archetype of the hepadnavirus family and similar viruses infecting different hosts such as ducks (DHBV), ground squirrels (GSHV), and woodchucks (WHV). HBV genome encodes four proteins which include the envelope protein (S/Pre-S), the core protein (C/pre-C), the polymerase (P), and the X protein (HBx). On the basis of variation within the nucleotide sequence it is classified into eight genotypes named  $A-H^{[4]}$ . Among them, B and C are mostly located in East Asia including China<sup>[5]</sup>. The HBx transcript is translated into a multifunctional protein which can regulate the activities of host cellular genes<sup>[6]</sup>. HBx triggers viral and cellular promoter indirectly by interaction with nuclear transcription factors $[7-9]$ . Additionally, HBx is found to be involved in the activation of numerous signal transduction cascades that are linked to cell proliferation and survival<sup>[10]</sup>. HBx mutants, especially those with mutations in the COOH-terminal have been implicated in  $HCC^{[11]}$ . Therefore, strategies involving targeting of HBx could help to achieve a significant level of therapeutic effect by inhibition of its functions which interferes with the cellular machinery and disrupts homeostasis.

#### **HBX**

HBx is a 154-amino acid regulatory protein with a molecular mass of approximately 17 kDa. As amino acid sequence of this gene is not homologous to any known

protein, it was assigned the name " $H B x''$ [12]. It is one of the most conserved proteins among different HBV subtypes and is found in almost all viruses of *Hepadnaviridae*. It is mostly localized within the cytoplasm and upto some extent in the nucleus of hepatic cells. HBx protein also play an important regulatory role in viral replication and subsequent infection<sup>[13]</sup>. HBx protein consists of two functional domains $^{[14]}$ . The amino-terminal domain is mapped to the first 50 amino acids including the dimerization region which is required for dimerization activity. The C-terminal transactivation domain is located between amino acid 53 and 142 which interacts with Xenopus anti-photoreceptor-1 (XAP-1)/UV-damaged DNA binding protein (UVDDB) and p53<sup>[15,16]</sup>. HBx is multi-functional protein that can transactivate viral and cellular promoters and enhancers through protein-protein interactions with several well defined targets (Figure 1). Instead of binding directly with DNA, HBX activates transcription by getting involved with several nuclear transcription factors like RNA polymerase binding protein (RBP5), transcriptional factor IIB (TFIIB), transcriptional factor IIH (TFIIH), a subunit of RNA polymerase, cAMP response elementbinding protein (CREB), CREB1-binding protein (CBP)/ p300, activating transcription factor 2 (ATF-2), activating protein (AP)-2, AP-1, and nuclear factor kappa B (NF $k$ B)<sup>[17,18]</sup>. Moreover, HBx can modulate cytoplasmic signal transduction pathways including Ras-Raf-mitogen activated protein kinase (Ras-Raf-MAPK), Janus kinase/STAT (JNK/STAT), focal adhesion kinase (FAK), proline-rich tyrosine kinase 2 (Pyk2), protein kinase C (PKC) and Srcdependent phosphatiylinositol-3 kinase (PI3K/Akt)<sup>[17,18]</sup>. Activation through transactivation of cellular signaling molecules can lead to hepatic cells proliferation. In addition, it has the tendency to directly inactivate or indirectly down-regulate various tumor suppressors, such as p53, or senescence-related factors[17,18].

## **ROLE OF HBX PROTEIN-INDUCED HEPATOCARCINOGENESIS**

#### *HBx interferes with nucleotide excision repair*

One of the key factors playing a role in the commencement and development of HCC is DNA damage and accumulation of errors<sup>[19]</sup>. Previously, we and others have shown that HBx protein inhibits DNA repair pathway. Acidic amino acids residues 120 and 121 of HBx were found to be critical for interaction with TFIIH and modulation of DNA repair process. It has been shown by *in vitro* and *in vivo* studies that HBx expression did not affect cell growth and tumor formation unless coupled with other factors such as exposure to hepatocarcinogen (diethynitrosamine) or UV irradiation<sup>[20]</sup>. HBx is not directly implicated in cancer progression but plays a major role as a promoting factor in HCC development. HBx protein is also known to interfere with nucleotide excision repair (NER) pathway through both p53 dependent and independent mechanisms. Studies have demonstrated that HBx protein inhibits cell cycle control



**Figure 1 Schematic depicting the potential contributions of hepatitis B virus -encoded protein in different cellular processes.** It influences the apoptosis, transcription, DNA repair and epigenetic changes as well as affecting transactivation mechanism. MAPK: Mitogen activated protein kinase; TGF: Transforming growth factor; IL: Interleukin; PKC: Protein kinase C; ATF: Activating transcription factor; NF-κB: Nuclear factor kappa B; ASPP: Apoptosis-stimulating protein of p53; ERK: Extracellular signal-regulated kinases; JNK: Janus kinase; HIF: Hypoxia-inducible factor.

checkpoints and facilitate accumulation of host mutations by interfering with  $NER^{[21-23]}$ . HBx exerts these effects by binding to several proteins involved in DNA repair pathways and inhibiting their repair capacity. HBx de-regulated factors include human homologue of UV-DDB, XPB and XPD components of TFIIH. These are essential components for NER in both p53-proficient andp53-deficient hepatocytes<sup>[24,25]</sup>. HBx has been shown to repress XPB and XPD indirectly and inhibit the DNA binding properties of the transcription factor Sp1 $[22,26,27]$ . It has been earlier noted that HBx inactivates p53 and also impair DNA repair. The COOH-terminus of p53 serves as a scaffold for HBx binding which ultimately results in the localization of p53 from nucleolus to cytoplasm. This leads to uncontrolled cell cycle progression and DNA repair. HBx also represses transcriptional activity of p53 by disrupting cross talk among several cellular factors[28]. Furthermore, it also disrupts interaction of p53 with TFIIH resulting in compromised TFIIH induced helicase activity during assembly of the DNA repair complex<sup>[29,30]</sup>. HBx binding to the carboxy (COOH) terminal domain of p53 is known block its association with XPB and XPD.

#### *Effects of HBx proteins on anti- and pro-apoptotic pathways*

HBx contributes to the progression of HCC by affecting apoptosis in different ways. HBx has been shown to either inhibit or have no effect on apoptosis<sup>[31-33]</sup>. Such

contradictory manifestations are likely due to the differential pattern of HBx expression and depend upon the cell types involved. It has been suggested that increased quantity of HBx promote apoptosis, whereas low levels inhibits this phenomenon<sup>[34]</sup>. One of the examples of its anti-apoptotic feature involves complex formation and sequestration of p53 in the cytoplasm and blocking its entrance into the nucleus<sup>[35-38]</sup>. As a result, p53 fails to up-regulate its various downstream effector molecules like Bax*,* p21*,* or Fas*,* which are required in the apoptotic pathway thus leading to increased cell survival<sup>[35]</sup>. HBx also prevents p53-induced apoptosis through HBx-P3K-Akt-Bad pathway, by inhibiting caspase 3 function, associated with H-ras oncogene by induction of phosphatidyl inositol-3 kinase and  $\overrightarrow{AKt}$  pathway<sup>[39]</sup>. Increase in the concentration of anti-apoptotic protein survivin is another example in this category which acts by p38/MAPK pathway<sup>[40]</sup>. HBx upregulation of survivin expression was found to be HURP-dependent. Another molecule which is highly expressed in hepatocellular carcinoma and mediates actions of HBx opposite to that of p53 is cyclooxygenase-2  $(COX-2)^{[41]}$ . Activation of  $COX-2/$ PGE2 pathway leads to elimination of p53-induced apoptosis imposed by HBX expression. In contrast, it is demonstrated by cell culture studied in HeLa, Hep3B, and HepG2 cells that HBx induces apoptosis directly by inducing cytoplasmic cytochrome c, caspase-3-like activity and nuclear fragmentation. HBx induced apoptosis is evident in HBx transgenic livers, primary hepatocytes

and in mice having p53 null mutation. Down-regulation of *C-myc* expression by HBx increased the response of PLC/PRF/5 cells to TNF- $\alpha$  induced apoptosis<sup>[42,43]</sup>. TRAIL-induced apoptosis mediated by HBx has also been shown in the hepatocytes<sup>[44]</sup>. A recent report shows that only the complete HBx protein, and not the one with the truncated C-terminus, induces apoptosis, which further indicate that the COOH-terminal region is necessary for its pro-apoptotic functions<sup>[45]</sup>.

#### *Epigenetic modification*

The importance of epigenetic modification in hepatocellular carcinoma was revealed when researchers found out that the methylation of at least one tumor suppressor gene was present in majority of tumors studied. However, its prevalence in HBx induced HCCs is not  $clear^{[46]}$ . Additional studies have shown that HBx can induce epigenetic changes and inactivate tumor suppressor genes and/or activate oncogenes thereby promoting the development of  $HCC^{[47]}$ . Moreover, these hyper methylated genes show direct role in cell growth, cell cycle regulation, dedifferentiation, apoptosis and xenobiotic metabolism<sup>[48]</sup>, altogether which contributes to HBVrelated HCC progression. HBx may modify transcription of DNA methyltransferase (DNMT)1 and DNMT3 and may act as potentially critical epigenetic deregulating agent. These epigenetic changes induced by HBx in several tumor suppressor genes have been highlighted. For example, HBx represses the E*-*cadherin (a tumor suppressor gene) by inducing DNMT1 transcription<sup>[49,50]</sup> and also influences Wnt signaling pathway. Such repressive effect is brought about by Lysine-30 located in the transactivating domain of HBx. HBx induces hypermethylation of p16 which show positive correlation with HCC<sup>[51,52]</sup>. Activation of DNMT1 transcription by HBx results in hypermethylation of p16 and ultimately its repression<sup>[53]</sup>. This activation is mediated by  $pRb-$ E2f pathway. Genomic hypomethylation and regional hypermethylation of the tumor promoter gene IGFBP-3 are known contributors of HCC. Promoter region of IGFBP is hypermethylated by DNMT1, DNMT3A1 and DNMT3A2[53]. Activation of *Ras* signaling pathway is responsible for Induction of  $DNMT1^{[54]}$ . Tumor suppressor p53 also display a key role in regulation of the DNMTs expression. It regulates apoptosis by interacting with apoptosis-stimulating protein of p53 (ASPP)1 and ASPP2 family of proteins<sup>[55,56]</sup>. ASPP2 has a much important role than ASPP1 in HBx protein induced HCC. Promoter hypermethylation of glutathione S-transferase P1, which helps in mounting a protective state against injury from oxidant carcinogens, is shown to be mediated by HBx. In summary the data is consistent with the hypothesis that epigenetic events induced by HBx play an important role in the pathogenesis of HCC, and also that of several other tumors. It is assumed that epigenetic down-regulation of retinoic acid receptorbeta 2 (RAR-β2) by HBx protein is an important step in HCC progression<sup>[57,58]</sup>. HBx decreases the expression of

retinoic acid receptor-beta two (RAR-β2) in human HCC cells by hypermethylating its promoter through DNA methyltransferase 1 and 3a. HBx abolishes the crucial potent effect of retinoic acid to down-regulate G1 checkpoint regulators such as p16, p21, and p27. This ultimately leads to activation of E2F1 and the generation of HCC tumor<sup>[59]</sup>.

#### *Telomerase activity*

Induction of Telomerase plays an important role in transformation of cancer cells as studies have shown that majority of hepatocellular carcinomas display a high level of telomerase activity[60]. Several studies were carried out to establish the role of HBx in telomerase but such effects has been controversial. HBx can increase the telomerase activity in cells transfected by virus, protein-positive HCC tissues or cultured HCC cells by increasing expression of TERT<sup>[61]</sup>. These studies demonstrate that HBx induced *C-myc* have a role in the process of telomerase activation and prolonging the life-span of normal cells. On the other hand Su  $et$   $a^{[62]}$  and his coworkers have shown that HBx inhibit telomerase activity and suggested that the controversial results could be attributed to various isoforms of HBx. They found that the transcriptional suppression of human telomerase achieved by HBx isoform is by increased binding of MAZ to its promoter $[63, 64]$ . However, more data is required to authenticate the function of telomerase in HBx induced HCC.

#### *Trans-activating mechanisms of HBx*

It is widely acknowledged that HBx protein acts as a potent transactivator and can affect various viral and cellular promoters and enhancers<sup>[65]</sup>. It regulates transcriptional activity basically *via* direct protein-protein interaction. The transactivation functions of HBx is exercised in cytoplasm by signaling pathways, and in the nucleus by DNA-binding proteins<sup>[66]</sup>. A wide variety of cellular and viral genes, class Ⅱ and Ⅲ promoters/enhancers, and proto-oncogenes such as *c-jun*, *c-fos*, and *c-myc* are upregulated by HBx. Besides, it activates number of transcriptional factors and genes like *NF*κ*B*, *AP-1*, *ATF*/*cAMP* and *CREB* in the nucleus<sup>[67,68]</sup>. The repertoire of genes and proteins which are linked with proliferation of cells also include interleukin-8 (IL-8), tumor necrosis factor (TNF), transforming growth factor (TGF)-β1, and early growth response factor (EGRF)<sup>[69]</sup>. In cytoplasm HBx activates signal transduction cascades involving RAS/RAF/ MAPK, JAK/STAT, and Src kinases $^{[27,70]}$ . Induction of RAS/RAF/MAPK by HBx leads to activation of AP-1 and NFκB transcription factors which causes the de-regulation of various checkpoints which control the cell cy $cle^{[71,72]}$ . HBx may also affect angiogenic pathways which play an important role in HCC development. HBx can induce up-regulation of vascular endothelial growth factor (VEGF) besides upregulating metalloproteinases (MMPs, MMP2, MMP9 and MMP14) which can be instrumental in promoting invasion and metastasis $[73]$ . Apart from this, there are several other ways by which HBx induces HCC

like stimulation of a number of signaling pathways associated with enhancing motility and overall survival chance of tumor cells, up-regulation of inflammatory mediators and an increased possibility for men to develop liver cancer by influencing androgen receptor pathway. There are many reports which advocate that COOH-terminal of HBx is critical for its transactivational activity and that it regulates cell proliferation and viability<sup>[74]</sup>. Deletion of its COOH-terminal has been documented in patients with chronic HBV infection<sup>[36]</sup>. The carcinogenic potential of C-terminus truncated HBx has also been implicated in Wnt signaling pathways which may have influence upon the development of HCC<sup>[68]</sup>.

## **EFFECT OF HBX ON MICRO RNA EXPRESSION ASSOCIATED WITH HCC**

MicroRNAs are noncoding RNAs which are involved in the regulation of gene expression. Additionally, they play crucial roles in numerous pathobiological processes including tumor formation. Therefore, their possible role in the causation of hepatocellular carcinoma is not unexpected. However, the question as to how HBx actually regulates miRNA expression during HCC has no easy answer. However, Wu *et al*<sup>[75]</sup> in 2014 have shown that HBx down regulates microRNA-15b *via* fucosyl transferase 2 induced Globo H (a cancer-associated carbohydrate antigen) expression ultimately influencing HCC proliferation. In a recently published article Zhang *et al*<sup> $76$ </sup> 2013 have found that miR-205 is down regulated in 33 samples of HCC tissues in contrast to adjacent healthy areas of the liver. They suggested that miR-205 could be a potential tumor-suppressive gene in case of HCC. According to them, HBx inhibit tumor suppressor miR-205 and increases hepato carcinogenesis by hypermethylation of miR-205 promoter. Wei *et al*<sup>[77]</sup> have demonstrated HBx induces epigenetic repression of miR-132 by methylation of DNA and suggested that it could be a promising biochemical marker for HCC. They have also shown that miR-101 is down regulated by HBx which was in turn induced by HBX targeting the methyl transferase 3A (*DNMT3A*) gene. In a mouse model of liver cancer, it was observed that miR-148a is repressed by HBx and this leads to cancer growth and eventual metastasis. Expression of miR-148a in hepatoma cells reduces hematopoietic pre-B cell leukemia transcription factor-interacting protein. This caused the suppression of AKT and extracellular signal-regulated kinases (ERK) induced mammalian target of rapamycin inhibition involving AKT/ERK/Fork head box protein O4/ATF5 pathways<sup>[78]</sup>. Recently Qiu *et al*<sup>[79]</sup> have highlighted that HBx downregulates PDCD4 by upregulation of miR-21. HBx suppresses EGFR by miR-7 which confirms the role of HBX-miR-7-EGFR as a critical signaling pathway in controlling cell growth in HCC<sup>[80]</sup>. HBx perturbs the *in vitro* expression of miRNA in cancerous hepatocytes of the host liver, especially by down regulating the mi-16 family. Its suppression was c-Myc mediated and is a necessary requirement for the HBx-induced transformation of HepG2 cells *in vitro*<sup>[81]</sup>. Another microRNA, miR-152 is frequently down-regulated during HBx expression and it is also known to regulate DNMT1 in HBV-related HCC. Additionally, tumor-suppressive role of miR-152 in the epigenetic irregularity of HBV-related HCC has been observed $^{\{82\}}$ . Wild-type HBx and the high proliferationinducing mutant HBx can influence the expression profile of miR-338-3p and miR-551b by its down regulation in L02 cells. Here, the cell growth inhibition occurs by direct modulation of cyclinD1, cyclinG1, and  $E2F^{[83,84]}$ . In another study showing linkage of mi RNA with HBx, it was found that HBx could inhibit apoptosis of HepG2 cells through down-regulation of miR-192 which induces apoptosis of  $HepG2$  cells<sup>[85]</sup>.

### **MUTATIONS OF HBX**

In the HCC tissue samples, negative with hepatitis B surface antigen, there seem to be a preferential accumulation of X transcripts suggesting the possible role of HBX in neoplastic transformation<sup>[86]</sup>. Several mutants of HBx have been isolated from hepatic tissues and sera taken from sufferers of  $HCC^{[87,88]}$ . The incidence of HBx mutations in such circumstances have been found to be reasonably high which underscores the point that the HBx mutants after all may be highly significant in the pathogenesis of hepatocellular carcinoma<sup>[89,90]</sup> (Table 1). Deletions of 3' is one of the most frequently reported mutations of HBx, and such mutations are more frequently observed in hepatocarcinoma cells rather than untransformed liver cells which highlights the potential role of these mutations in tumor development. Poussin *et al*<sup>911</sup> have analyzed the HBx expression correlation in tumorous *vs* non-tumorous tissues of human origin. They observed that HBV X gene was truncated at its 3' end in five out of nine tumors from the liver and only one out of eight liver tissues with no tumors. A high rate of mutation was revealed in X genes from three tumors upon Sequence analysis. This was in contrast to the lower rate of mutation observed in the X genes of adjoining non tumorous tissues. In an investigation carried out by Tu *et al*<sup>45</sup>*,* sera and tissues were taken from tumor as well as healthy liver regions of patients with HBV related HCC. They conducted a detailed analysis of the structure and functioning of HBx sequences. With the exception of one, six out of seven tumor samples showed that the HBx sequences in these tissues and especially those harvested from HBV integrant have a deletion at COOHterminal part. Many of these shortened sequences were divested of their transcriptional capacity. Additionally, their function of acting as a brake to cell division and transformation seemed to be compromised. In fact COOH shortened HBX improved the transforming character of ras and myc oncogenes<sup>[45]</sup>. Centromere protein A (CENP-A) is known to be frequently over-expressed in HCC. A positive correlation between HBx COOH terminus mutation and expression of CENP-A has been docu-



#### **Table 1 Hepatitis B virus -encoded X protein mutants and their role in liver injury and hepatocellular carcinoma**

aa: Amino acid; A31: Alanine 31; M-130: Methionine 130; I: Isoleucine; V: Valine; Ser: Serine(S); Met: Methionine; R: Arginine; LOX-5: Arachidonate 5-lipoxygenase; SREBP: Sterol Regulatory Element-Binding Protein; HBV: Hepatitis B virus; HBx: HBV-encoded X protein; HCC: Hepatocellular carcinoma.

mented in HCC tissues<sup>[92]</sup>. Carboxy-terminal truncated HBx were frequently detected in HCC tissues by Ma *et*   $a^{[93]}$ . They found that the truncated HBx is able to effectively transform immortalized liver cell line. Differential expression of key genes implicated in the control of cell cycle and apoptosis have been observed by them. Based on these studies, it can be concluded that the length of COOH terminus truncation is important for HBx's transcription activity. The truncated HBx exhibiting deletion within 14 amino acids had no effect on its transactivation property<sup>[94]</sup>. Thus it could be concluded that the last 14 amino acids at the carboxy terminus of HBx does not contribute towards transactivation activity[94]. HBx is divided into six regions of which A, C, and E regions show a more conserved state. To decipher the regions of HBx which have a significant role in transactivation, Kumar *et al*[94] effected a range of 10 deletion mutants and four single point mutants which related to corresponding conserved cysteine residues. The mutant gene expression was also examined by the use of HBx specific monoclonal antibody. It was found by them that deletion of region A which happens to be extremely conserved did not in anyways affect transactivation. Also the non-substitution of the N terminal cysteine produced no difference to transactivation. Removal of the regions C and D however led to an important loss of function. Kumar *et al*<sup>[94]</sup> inferred and stated that residues 132-140 of region E and C137 seemed to be crucial for transactivation. They also showed that a fusion cassette containing residues 58-140 proved to be an efficient transactivator, suggesting an im-

portant role of these residues in transcriptional activity<sup>[94]</sup>. C-terminal deletional mutant of HBx (deleted at nucleotides 382-400) was shown to down-regulate a miRNA (miR338-3p) in HBx infected cells which prolonged cell proliferation and increased the risk of HCC development<sup>[83,84]</sup>. Wnt-5a is correlated with the HCC development. HBx mutants deprived of thirty amino acids from their C-terminus induced the expression of Wnt-5a gene by more than 10 folds in Huh7 cells. However, the expression of Wnt-5a was suppressed when the same cells were transfected with HBx mutant deleted with forty amino acids from the same C-terminus. Such changes in Wnt-5a expression levels were also observed in patients with HCC when compared with appropriate non-tumorous counterparts. This suggests that HBx mutants might play a role in the development and progression of HCC through the Wnt-5a pathway as HBx showed negative correlation with Wnt-5a<sup>[95]</sup>. Sirma *et al*<sup>[74]</sup> studied the influence of HBx and its natural mutants on cellular growth, proliferation and survival. They explained that the loss of antiproliferative and apoptotic role of HBx by the natural mutants may propel the hepatocytes to go into uncontrollable cell division and probably help in malignant transformation which could ultimately lead to HCC.

Some point mutations have also been implicated in affecting the functions of HBx and HCC development. HBx with point mutations K130M/V131I has been shown to induce hypoxia-inducible factor  $1\alpha$  which has a role in the development of solid tumors under hypoxic environment conditions in HCC. A novel HBx associated

mutation at amino acids L30F/S144A was recorded in 13 out of 44 HCC tissue samples<sup>[96]</sup>. Chen *et al*<sup>[88]</sup> highlighted more than 50 varying type of mutations involving the HBx gene. They found that HBx mutation in tissue and serum (12 and 9 sites respectively) showed significant association with HCC. This point to a possible role in HCC development. Insertion mutation at position 204: Insert 204AGGCCC was found to be coexisting with point mutation at 260 (G-->A) and 264 (G/C/T-->A). This opened a significant window in the understanding of HBX mutation in that, it also pointed that the nuclear localization of HBx protein in tumorous hepatocytes was closely associated with the afore mentioned characteristic alignment of HBx. The above mutation seems to have connection with the nuclear localization of HBx protein, suggesting a link with HCC development. With the help of innovative laser capture micro dissection technology, Iavarone *et al*<sup>89</sup> consistently identified mutations at aa 127, 130 and 131, though they are not able to find any clear point mutation profile between tumor and healthy tissue samples. On the other hand, deletion in *HBx* gene which was detected in several sufferers was more commonly associated with tumor derived sequences (6/18). This is in contrast to healthy tissue derived sequences in which the no is less (1 out of 20). The researchers concluded that there is a consistent presence of characteristic HBx encoding sequences in clonally proliferating cells. This strengthens the hypothesis that HBx deletions may be critically involved in the development of liver cancer. Another study which identified mutations linked with the progression of HCC is mutation of nucleotides T1653, T1689, and/or T1762/A1764. Double mutant T1762/ A1764 along with the adjacent T1766/A1768 mutation was found to significantly elevate the risk of HCC in HBV-infected patients<sup>[90]</sup>. Recently, a natural mutant of HBx (HBxDelta127) was discovered with shortening of 27 amino acids at the COOH-terminal (deletion of 382 to 401 bp). This can potentially induce growth and proliferation of hepatoma cells. HBx Delta127 significantly enhanced the transcription of FAS in human hepatoma HepG2 and H7402 cells by activating SREBP-1c which in turn activates  $5$ -LOX<sup>[97]</sup>. In an important discovery it was revealed that cancer associated HBx variants clearly blunted transactivation and proapoptotic functions. Although their ability to block p53-mediated apoptosis remained unaltered, suggesting that mutations in HBx may contribute to the development of HCC. This result is consistent with the hypothesis that certain mutations in HBx and p53 (at codon 249) may cooperate in contributing to liver carcinogenesis<sup>[98]</sup>. Diet incorporated fungal aflatoxin B1 has been instrumental in the causation of HCC in Africa and China region, yet how HBx mutants interplay with toxins is still not known. Exposure to aflatoxin B1 can result in induction of mutation in the p53 gene, a G to T transversion in codon 249 (p53ser249 mutation) inactivates the tumor suppressor gene. This mutation is found in majority of HCC patients in regions having high aflatoxin  $β1$  exposure<sup>[99]</sup>. It was shown by Belloni  $et$   $d^{[100]}$  that HBV mutants that lack the ability to express HBx are replication deficient. Such replication incompetence is restored by exogenously expressing HBx transcomplements. Several evidences suggest that mutations in HBx contribute heavily towards the development of HCC. However, in what profound and exact manner these structural and point mutation affects functions and behavior of HBX need to be further explored.

#### **CONCLUSION**

HBx modulates a variety of cellular activities such as transcription, cell cycle progression, DNA damage repair, cell proliferation, and apoptosis. The localization of HBx is consistent with its dual specificity role. HBx predominantly localizes in the cytoplasm and to some extent within the nucleus and mitochondria. This dynamic cellular distribution and compartmentalization could attribute to HBx multiple functions at different stages of viral infection and HCC progression. Although extensive study has been done to elucidate the implications of HBx in hepatocellular carcinoma, its precise role in carcinogenic manifestations is yet to be deciphered completely and is appropriate to be the main focus of the future studies. The difficulty in addressing its role arises due to the lack of convenient and clinically relevant models to study viral replication. However, shedding some light on its indirect effects, it can be concluded that HBx helps in the progression of HCC since it can modulate cellular microenvironment by acting on transcription factors, cell cycle regulators, apoptosis and DNA repair mechanisms. This conclusion can be further supported by the fact that the duck HBV, that lacks an X open reading frame, does not induce cancer in ducks. Thus, HBx represents one of the central players of oncogenesis in liver and could be an attractive therapeutic target for HCC suppression. HBx mutants especially those with mutation in the COOHterminal end have been implicated in HCC, therefore therapeutic strategies targeting HBx could be effective at multiple stages of HCC development.

#### **ACKNOWLEDGMENTS**

We thank King Fahd Medical Research Center (KFMRC) and Center of Genomic Medicine (CEGMR) for financial support. We thank Mr. Mohammad Sanaullah M.Y. Gazdar (Head KFMRC Library), Ms. Shylu Mathew of CEGMR, Dr. Zubair Alam of KFMRC and Dr. Zainab Younis and Ms. Nadia Rashid of IQ Institute of Infection and Immunity, Lahore Pakistan for helpful discussion and editing.

#### **REFERENCES**

- 1 **Forner A**, Llovet JM, Bruix J. Hepatocellular carcinoma. *Lancet* 2012; **379**: 1245-1255 [PMID: 22353262]
- 2 **Lupberger J**, Hildt E. Hepatitis B virus-induced oncogenesis. *World J Gastroenterol* 2007; **13**: 74-81 [PMID: 17206756]
- 3 **Altekruse SF**, McGlynn KA, Reichman ME. Hepatocellular carcinoma incidence, mortality, and survival trends in the United States from 1975 to 2005. *J Clin Oncol* 2009; **27**:

1485-1491 [PMID: 19224838 DOI: 10.1200/JCO.2008.20.7753]

- 4 **Sun TT**, Chu YY. Carcinogenesis and prevention strategy of liver cancer in areas of prevalence. *J Cell Physiol Suppl* 1984; **3**: 39-44 [PMID: 6430924 DOI: 10.1002/jcp.1041210407]
- 5 **Chen X**, Dai B, Liu Z, Gao J, Ji Z, Guo J, Chen G, Deng Z, Shao Z. A novel B/C inter-genotype recombinant of hepatitis B virus identified in north-west China. *J Gen Virol* 2014; **95**: 153-155 [PMID: 24129025 DOI: 10.1099/vir.0.054023-0]
- **Bouchard MJ**, Schneider RJ. The enigmatic X gene of hepatitis B virus. *J Virol* 2004; **78**: 12725-12734 [PMID: 15542625 DOI: 10.1128/JVI.78.23.12725-12734.2004]
- 7 **Edamoto Y**, Hara A, Biernat W, Terracciano L, Cathomas G, Riehle HM, Matsuda M, Fujii H, Scoazec JY, Ohgaki H. Alterations of RB1, p53 and Wnt pathways in hepatocellular carcinomas associated with hepatitis C, hepatitis B and alcoholic liver cirrhosis. *Int J Cancer* 2003; **106**: 334-341 [PMID: 12845670 DOI: 10.1002/ijc.11254]
- 8 **Tommasi S**, Pinto R, Pilato B, Paradiso A. Molecular pathways and related target therapies in liver carcinoma. *Curr Pharm Des* 2007; **13**: 3279-3287 [PMID: 18045179 DOI: 10.2174 /138161207782360663]
- 9 **Kew MC**. Hepatitis B virus x protein in the pathogenesis of hepatitis B virus-induced hepatocellular carcinoma. *J Gastroenterol Hepatol* 2011; **26** Suppl 1: 144-152 [PMID: 21199526 DOI: 10.1111/j.1440-1746.2010.06546.x]
- 10 **McCubrey JA**, Steelman LS, Abrams SL, Lee JT, Chang F, Bertrand FE, Navolanic PM, Terrian DM, Franklin RA, D' Assoro AB, Salisbury JL, Mazzarino MC, Stivala F, Libra M. Roles of the RAF/MEK/ERK and PI3K/PTEN/AKT pathways in malignant transformation and drug resistance. *Adv Enzyme Regul* 2006; **46**: 249-279 [PMID: 16854453 DOI: 10.1016/j.advenzreg.2006.01.004]
- 11 **Ng SA**, Lee C. Hepatitis B virus X gene and hepatocarcinogenesis. *J Gastroenterol* 2011; **46**: 974-990 [PMID: 21647825 DOI: 10.1007/s00535-011-0415-9]
- 12 **Miller RH**, Robinson WS. Common evolutionary origin of hepatitis B virus and retroviruses. *Proc Natl Acad Sci USA* 1986; **83**: 2531-2535 [PMID: 3458214 DOI: 10.1073/ pnas.83.8.2531]
- 13 **Murakami S**. Hepatitis B virus X protein: a multifunctional viral regulator. *J Gastroenterol* 2001; **36**: 651-660 [PMID: 11686474 DOI: 10.1007/s005350170027]
- 14 **Gong DY**, Chen EQ, Huang FJ, Leng XH, Cheng X, Tang H. Role and functional domain of hepatitis B virus X protein in regulating HBV transcription and replication in vitro and in vivo. *Viruses* 2013; **5**: 1261-1271 [PMID: 23698398 DOI: 10.3390/v5051261]
- 15 **Lin Y**, Nomura T, Yamashita T, Dorjsuren D, Tang H, Murakami S. The transactivation and p53-interacting functions of hepatitis B virus X protein are mutually interfering but distinct. *Cancer Res* 1997; **57**: 5137-5142 [PMID: 9371515]
- 16 **Becker SA**, Lee TH, Butel JS, Slagle BL. Hepatitis B virus X protein interferes with cellular DNA repair. *J Virol* 1998; **72**: 266-272 [PMID: 9420223]
- 17 **Feitelson MA**, Lee J. Hepatitis B virus integration, fragile sites, and hepatocarcinogenesis. *Cancer Lett* 2007; **252**: 157-170 [PMID: 17188425 DOI: 10.1016/j.canlet.2006.11.010]
- 18 **Diao J**, Garces R, Richardson CD. X protein of hepatitis B virus modulates cytokine and growth factor related signal transduction pathways during the course of viral infections and hepatocarcinogenesis. *Cytokine Growth Factor Rev* 2001; **12**: 189-205 [PMID: 11325602 DOI: 10.1016/ S1359-6101(00)00034-4]
- 19 **Qadri I**, Fatima K, AbdeL-Hafiz H. Hepatitis B virus X protein impedes the DNA repair via its association with transcription factor, TFIIH. *BMC Microbiol* 2011; **11**: 48 [PMID: 21375739 DOI: 10.1186/1471-2180-11-48]
- 20 **Slagle BL**, Lee TH, Medina D, Finegold MJ, Butel JS. Increased sensitivity to the hepatocarcinogen diethylnitrosamine in transgenic mice carrying the hepatitis B virus X

gene. *Mol Carcinog* 1996; **15**: 261-269 [PMID: 8634084]

- 21 **Bellamy CO**, Clarke AR, Wyllie AH, Harrison DJ. p53 Deficiency in liver reduces local control of survival and proliferation, but does not affect apoptosis after DNA damage. *FASEB J* 1997; **11**: 591-599 [PMID: 9212083]
- 22 **Lee TH**, Elledge SJ, Butel JS. Hepatitis B virus X protein interacts with a probable cellular DNA repair protein. *J Virol* 1995; **69**: 1107-1114 [PMID: 7815490]
- 23 **Capovilla A**, Carmona S, Arbuthnot P. Hepatitis B virus X-protein binds damaged DNA and sensitizes liver cells to ultraviolet irradiation. *Biochem Biophys Res Commun* 1997; **232**: 255-260 [PMID: 9125143 DOI: 10.1006/bbrc.1997.6269]
- 24 **Qadri I**, Conaway JW, Conaway RC, Schaack J, Siddiqui A. Hepatitis B virus transactivator protein, HBx, associates with the components of TFIIH and stimulates the DNA helicase activity of TFIIH. *Proc Natl Acad Sci USA* 1996; **93**: 10578-10583 [PMID: 8855220 DOI: 10.1073/pnas.93.20.10578]
- 25 **Groisman IJ**, Koshy R, Henkler F, Groopman JD, Alaoui-Jamali MA. Downregulation of DNA excision repair by the hepatitis B virus-x protein occurs in p53-proficient and p53-deficient cells. *Carcinogenesis* 1999; **20**: 479-483 [PMID: 10190565 DOI: 10.1093/carcin/20.3.479]
- 26 **Benn J**, Schneider RJ. Hepatitis B virus HBx protein deregulates cell cycle checkpoint controls. *Proc Natl Acad Sci USA* 1995; **92**: 11215-11219 [PMID: 7479968 DOI: 10.1073/ pnas.92.24.11215]
- 27 **Lee YI**, Lee S, Lee Y, Bong YS, Hyun SW, Yoo YD, Kim SJ, Kim YW, Poo HR. The human hepatitis B virus transactivator X gene product regulates Sp1 mediated transcription of an insulin-like growth factor II promoter 4. *Oncogene* 1998; **16**: 2367-2380 [PMID: 9620554 DOI: 10.1038/sj.onc.1201760]
- 28 **Dewantoro O**, Gani RA, Akbar N. Hepatocarcinogenesis in viral Hepatitis B infection: the role of HBx and p53. *Acta Med Indones* 2006; **38**: 154-159 [PMID: 16953033]
- 29 **Madden CR**, Finegold MJ, Slagle BL. Altered DNA mutation spectrum in aflatoxin b1-treated transgenic mice that express the hepatitis B virus x protein. *J Virol* 2002; **76**: 11770-11774 [PMID: 12388740 DOI: 10.1128/JVI.76.22.11770-11774.2002]
- 30 **Smith ML**, Chen IT, Zhan Q, O'Connor PM, Fornace AJ. Involvement of the p53 tumor suppressor in repair of u.v.-type DNA damage. *Oncogene* 1995; **10**: 1053-1059 [PMID: 7700629]
- 31 **Terradillos O**, Pollicino T, Lecoeur H, Tripodi M, Gougeon ML, Tiollais P, Buendia MA. p53-independent apoptotic effects of the hepatitis B virus HBx protein in vivo and in vitro. *Oncogene* 1998; **17**: 2115-2123 [PMID: 9798683 DOI: 10.1038/ sj.onc.1202432]
- 32 **Kim H**, Lee H, Yun Y. X-gene product of hepatitis B virus induces apoptosis in liver cells. *J Biol Chem* 1998; **273**: 381-385 [PMID: 9417092 DOI: 10.1074/jbc.273.1.381]
- 33 **Miao J**, Chen GG, Chun SY, Lai PP. Hepatitis B virus X protein induces apoptosis in hepatoma cells through inhibiting Bcl-xL expression. *Cancer Lett* 2006; **236**: 115-124 [PMID: 15990224 DOI: 10.1016/j.canlet.2005.05.014]
- 34 **Ye L**, Dong N, Wang Q, Xu Z, Cai N, Wang H, Zhang X. Progressive changes in hepatoma cells stably transfected with hepatitis B virus X gene. *Intervirology* 2008; **51**: 50-58 [PMID: 18334850 DOI: 10.1159/000120289]
- 35 **Wang XW**, Gibson MK, Vermeulen W, Yeh H, Forrester K, Stürzbecher HW, Hoeijmakers JH, Harris CC. Abrogation of p53-induced apoptosis by the hepatitis B virus X gene. *Cancer Res* 1995; **55**: 6012-6016 [PMID: 8521383]
- 36 **Elmore LW**, Hancock AR, Chang SF, Wang XW, Chang S, Callahan CP, Geller DA, Will H, Harris CC. Hepatitis B virus X protein and p53 tumor suppressor interactions in the modulation of apoptosis. *Proc Natl Acad Sci USA* 1997; **94**: 14707-14712 [PMID: 9405677 DOI: 10.1073/pnas.94.26.14707]
- 37 **Ueda H**, Ullrich SJ, Gangemi JD, Kappel CA, Ngo L, Feitelson MA, Jay G. Functional inactivation but not structural mutation of p53 causes liver cancer. *Nat Genet* 1995; **9**: 41-47 [PMID: 7704023 DOI: 10.1038/ng0195-41]
- 38 **Wang XW**, Forrester K, Yeh H, Feitelson MA, Gu JR, Harris CC. Hepatitis B virus X protein inhibits p53 sequence-specific DNA binding, transcriptional activity, and association with transcription factor ERCC3. *Proc Natl Acad Sci USA* 1994; **91**: 2230-2234 [PMID: 8134379 DOI: 10.1073/pnas.91.6.2230]
- 39 **Lee YI**, Kang-Park S, Do SI, Lee YI. The hepatitis B virus-X protein activates a phosphatidylinositol 3-kinase-dependent survival signaling cascade. *J Biol Chem* 2001; **276**: 16969-16977 [PMID: 11278872 DOI: 10.1074/jbc.M011263200]
- 40 **Kuo TC**, Chao CC. Hepatitis B virus X protein prevents apoptosis of hepatocellular carcinoma cells by upregulating SATB1 and HURP expression. *Biochem Pharmacol* 2010; **80**: 1093-1102 [PMID: 20541537 DOI: 10.1016/j.bcp.2010.06.003]
- 41 **Cheng AS**, Yu J, Lai PB, Chan HL, Sung JJ. COX-2 mediates hepatitis B virus X protein abrogation of p53-induced apoptosis. *Biochem Biophys Res Commun* 2008; **374**: 175-180 [PMID: 18601905 DOI: 10.1016/j.bbrc.2008.06.098]
- 42 **Su F**, Schneider RJ. Hepatitis B virus HBx protein sensitizes cells to apoptotic killing by tumor necrosis factor alpha. *Proc Natl Acad Sci USA* 1997; **94**: 8744-8749 [PMID: 9238048 DOI: 10.1073/pnas.94.16.8744]
- 43 **Chan DW**, Ng IO. Knock-down of hepatitis B virus X protein reduces the tumorigenicity of hepatocellular carcinoma cells. *J Pathol* 2006; **208**: 372-380 [PMID: 16353167 DOI: 10.1002/ path.1901]
- 44 **Liu YG**, Liu SX, Liang XH, Zhang Q, Gao LF, Han LH, Cao YL, Hou N, Du J, Sun WS. Blockade of TRAIL pathway ameliorates HBV-induced hepatocyte apoptosis in an acute hepatitis model. *Biochem Biophys Res Commun* 2007; **352**: 329-334 [PMID: 17126290 DOI: 10.1016/j.bbrc.2006.11.024]
- 45 **Tu H**, Bonura C, Giannini C, Mouly H, Soussan P, Kew M, Paterlini-Bréchot P, Bréchot C, Kremsdorf D. Biological impact of natural COOH-terminal deletions of hepatitis B virus X protein in hepatocellular carcinoma tissues. *Cancer Res* 2001; **61**: 7803-7810 [PMID: 11691796]
- 46 **Yang B**, Guo M, Herman JG, Clark DP. Aberrant promoter methylation profiles of tumor suppressor genes in hepatocellular carcinoma. *Am J Pathol* 2003; **163**: 1101-1107 [PMID: 12937151 DOI: 10.1016/S0002-9440(10)63469-4]
- 47 **Tian Y**, Yang W, Song J, Wu Y, Ni B. Hepatitis B virus X protein-induced aberrant epigenetic modifications contributing to human hepatocellular carcinoma pathogenesis. *Mol Cell Biol* 2013; **33**: 2810-2816 [PMID: 23716588 DOI: 10.1128/ MCB.00205-13]
- 48 **Pogribny IP**, Rusyn I. Role of epigenetic aberrations in the development and progression of human hepatocellular carcinoma. *Cancer Lett* 2014; **342**: 223-230 [PMID: 22306342 DOI: 10.1016/j.canlet.2012.01.038]
- 49 **Lee JO**, Kwun HJ, Jung JK, Choi KH, Min DS, Jang KL. Hepatitis B virus X protein represses E-cadherin expression via activation of DNA methyltransferase 1. *Oncogene* 2005; **24**: 6617-6625 [PMID: 16007161 DOI: 10.1038/sj.onc.1208827]
- 50 **Liu J**, Lian Z, Han S, Waye MM, Wang H, Wu MC, Wu K, Ding J, Arbuthnot P, Kew M, Fan D, Feitelson MA. Downregulation of E-cadherin by hepatitis B virus X antigen in hepatocellullar carcinoma. *Oncogene* 2006; **25**: 1008-1017 [PMID: 16247464 DOI: 10.1038/sj.onc.1209138]
- 51 **Jung JK**, Arora P, Pagano JS, Jang KL. Expression of DNA methyltransferase 1 is activated by hepatitis B virus X protein via a regulatory circuit involving the p16INK4a-cyclin D1- CDK 4/6-pRb-E2F1 pathway. *Cancer Res* 2007; **67**: 5771-5778 [PMID: 17575144 DOI: 10.1158/0008-5472.CAN-07-0529]
- 52 **Zhu R**, Li BZ, Li H, Ling YQ, Hu XQ, Zhai WR, Zhu HG. Association of p16INK4A hypermethylation with hepatitis B virus X protein expression in the early stage of HBV-associated hepatocarcinogenesis. *Pathol Int* 2007; **57**: 328-336 [PMID: 17539963 DOI: 10.1111/j.1440-1827.2007.02104.x]
- 53 **Zhu YZ**, Zhu R, Fan J, Pan Q, Li H, Chen Q, Zhu HG. Hepatitis B virus X protein induces hypermethylation of p16(INK4A) promoter via DNA methyltransferases in the

early stage of HBV-associated hepatocarcinogenesis. *J Viral Hepat* 2010; **17**: 98-107 [PMID: 19732323 DOI: 10.1111/ j.1365-2893.2009.01156.x]

- 54 **Park IY**, Sohn BH, Yu E, Suh DJ, Chung YH, Lee JH, Surzycki SJ, Lee YI. Aberrant epigenetic modifications in hepatocarcinogenesis induced by hepatitis B virus X protein. *Gastroenterology* 2007; **132**: 1476-1494 [PMID: 17408664 DOI: 10.1053/j.gastro.2007.01.034]
- 55 **Samuels-Lev Y**, O'Connor DJ, Bergamaschi D, Trigiante G, Hsieh JK, Zhong S, Campargue I, Naumovski L, Crook T, Lu X. ASPP proteins specifically stimulate the apoptotic function of p53. *Mol Cell* 2001; **8**: 781-794 [PMID: 11684014 DOI: 10.1016/S1097-2765(01)00367-7]
- 56 **Zhao J**, Wu G, Bu F, Lu B, Liang A, Cao L, Tong X, Lu X, Wu M, Guo Y. Epigenetic silence of ankyrin-repeat-containing, SH3-domain-containing, and proline-rich-region- containing protein 1 (ASPP1) and ASPP2 genes promotes tumor growth in hepatitis B virus-positive hepatocellular carcinoma. *Hepatology* 2010; **51**: 142-153 [PMID: 20034025 DOI: 10.1002/ hep.23247]
- Jung JK, Park SH, Jang KL. Hepatitis B virus X protein overcomes the growth-inhibitory potential of retinoic acid by downregulating retinoic acid receptor-beta2 expression via DNA methylation. *J Gen Virol* 2010; **91**: 493-500 [PMID: 19828754 DOI: 10.1099/vir.0.015149-0]
- Niu D, Zhang J, Ren Y, Feng H, Chen WN. HBx genotype D represses GSTP1 expression and increases the oxidative level and apoptosis in HepG2 cells. *Mol Oncol* 2009; **3**: 67-76 [PMID: 19383368 DOI: 10.1016/j.molonc.2008.10.002]
- 59 **Yokota T**, Suda T, Igarashi M, Kuroiwa T, Waguri N, Kawai H, Mita Y, Aoyagi Y. Telomere length variation and maintenance in hepatocarcinogenesis. *Cancer* 2003; **98**: 110-118 [PMID: 12833463 DOI: 10.1002/cncr.11428]
- Zhang X, Dong N, Zhang H, You J, Wang H, Ye L. Effects of hepatitis B virus X protein on human telomerase reverse transcriptase expression and activity in hepatoma cells. *J Lab Clin Med* 2005; **145**: 98-104 [PMID: 15746653 DOI: 10.1016/ j.lab.2004.11.018]
- Liu YC, Chen CJ, Wu HS, Chan DC, Yu JC, Yang AH, Cheng YL, Lee SC, Harn HJ. Telomerase and c-myc expression in hepatocellular carcinomas. *Eur J Surg Oncol* 2004; **30**: 384-390 [PMID: 15063891 DOI: 10.1016/j.ejso.2004.01.003]
- 62 **Su JM**, Lai XM, Lan KH, Li CP, Chao Y, Yen SH, Chang FY, Lee SD, Lee WP. X protein of hepatitis B virus functions as a transcriptional corepressor on the human telomerase promoter. *Hepatology* 2007; **46**: 402-413 [PMID: 17559154 DOI: 10.1002/hep.21675]
- Her S, Bell RA, Bloom AK, Siddall BJ, Wong DL. Phenylethanolamine N-methyltransferase gene expression. Sp1 and MAZ potential for tissue-specific expression. *J Biol Chem* 1999; **274**: 8698-8707 [PMID: 10085109 DOI: 10.1074/ jbc.274.13.8698]
- 64 **Qu ZL**, Zou SQ, Cui NQ, Wu XZ, Qin MF, Kong D, Zhou ZL. Upregulation of human telomerase reverse transcriptase mRNA expression by in vitro transfection of hepatitis B virus X gene into human hepatocarcinoma and cholangiocarcinoma cells. *World J Gastroenterol* 2005; **11**: 5627-5632 [PMID: 16237755]
- 65 **Kremsdorf D**, Soussan P, Paterlini-Brechot P, Brechot C. Hepatitis B virus-related hepatocellular carcinoma: paradigms for viral-related human carcinogenesis. *Oncogene* 2006; **25**: 3823-3833 [PMID: 16799624 DOI: 10.1038/sj.onc.1209559]
- 66 **Nakatake H**, Chisaka O, Yamamoto S, Matsubara K, Koshy R. Effect of X protein on transactivation of hepatitis B virus promoters and on viral replication. *Virology* 1993; **195**: 305-314 [PMID: 8337816 DOI: 10.1006/viro.1993.1381]
- 67 **Chirillo P**, Falco M, Puri PL, Artini M, Balsano C, Levrero M, Natoli G. Hepatitis B virus pX activates NF-kappa B-dependent transcription through a Raf-independent pathway. *J Virol* 1996; **70**: 641-646 [PMID: 8523586]
- 68 **Kim HR**, Lee SH, Jung G. The hepatitis B viral X protein activates NF-kappaB signaling pathway through the up-regulation of TBK1. *FEBS Lett* 2010; **584**: 525-530 [PMID: 19958770 DOI: 10.1016/j.febslet.2009.11.091]
- 69 **Andrisani OM**, Barnabas S. The transcriptional function of the hepatitis B virus X protein and its role in hepatocarcinogenesis (Review). *Int J Oncol* 1999; **15**: 373-379 [PMID: 10402250]
- 70 **Benn J**, Schneider RJ. Hepatitis B virus HBx protein activates Ras-GTP complex formation and establishes a Ras, Raf, MAP kinase signaling cascade. *Proc Natl Acad Sci USA* 1994; **91**: 10350-10354 [PMID: 7937954 DOI: 10.1073/pnas.91.22.10350]
- 71 **Benn J**, Su F, Doria M, Schneider RJ. Hepatitis B virus HBx protein induces transcription factor AP-1 by activation of extracellular signal-regulated and c-Jun N-terminal mitogenactivated protein kinases. *J Virol* 1996; **70**: 4978-4985 [PMID: 8764004]
- 72 **Um HR**, Lim WC, Chae SY, Park S, Park JH, Cho H. Raf-1 and protein kinase B regulate cell survival through the activation of NF-kappaB in hepatitis B virus X-expressing cells. *Virus Res* 2007; **125**: 1-8 [PMID: 17188775 DOI: 10.1016/ j.virusres.2006.11.007]
- 73 **Liu LP**, Liang HF, Chen XP, Zhang WG, Yang SL, Xu T, Ren L. The role of NF-kappaB in Hepatitis b virus X proteinmediated upregulation of VEGF and MMPs. *Cancer Invest* 2010; **28**: 443-451 [PMID: 20073580 DOI: 10.3109/0735790090 3405959]
- 74 **Sirma H**, Giannini C, Poussin K, Paterlini P, Kremsdorf D, Bréchot C. Hepatitis B virus X mutants, present in hepatocellular carcinoma tissue abrogate both the antiproliferative and transactivation effects of HBx. *Oncogene* 1999; **18**: 4848-4859 [PMID: 10490818 DOI: 10.1038/sj.onc.1202867]
- 75 **Wu CS**, Yen CJ, Chou RH, Chen JN, Huang WC, Wu CY, Yu YL. Downregulation of microRNA-15b by hepatitis B virus X enhances hepatocellular carcinoma proliferation via fucosyltransferase 2-induced Globo H expression. *Int J Cancer* 2014; **134**: 1638-1647 [PMID: 24122375 DOI: 10.1002/ijc.28501]
- 76 **Zhang T**, Zhang J, Cui M, Liu F, You X, Du Y, Gao Y, Zhang S, Lu Z, Ye L, Zhang X. Hepatitis B virus X protein inhibits tumor suppressor miR-205 through inducing hypermethylation of miR-205 promoter to enhance carcinogenesis. *Neoplasia* 2013; **15**: 1282-1291 [PMID: 24339740]
- 77 **Wei X**, Tan C, Tang C, Ren G, Xiang T, Qiu Z, Liu R, Wu Z. Epigenetic repression of miR-132 expression by the hepatitis B virus x protein in hepatitis B virus-related hepatocellular carcinoma. *Cell Signal* 2013; **25**: 1037-1043 [PMID: 23376496 DOI: 10.1016/j.cellsig.2013.01.019]
- 78 **Xu X**, Fan Z, Kang L, Han J, Jiang C, Zheng X, Zhu Z, Jiao H, Lin J, Jiang K, Ding L, Zhang H, Cheng L, Fu H, Song Y, Jiang Y, Liu J, Wang R, Du N, Ye Q. Hepatitis B virus X protein represses miRNA-148a to enhance tumorigenesis. *J Clin Invest* 2013; **123**: 630-645 [PMID: 23321675 DOI: 10.1172/ JCI64265]
- 79 **Qiu X**, Dong S, Qiao F, Lu S, Song Y, Lao Y, Li Y, Zeng T, Hu J, Zhang L, Zhang L, Fan H. HBx-mediated miR-21 upregulation represses tumor-suppressor function of PDCD4 in hepatocellular carcinoma. *Oncogene* 2013; **32**: 3296-3305 [PMID: 23604124 DOI: 10.1038/onc.2013.150]
- 80 **Chen YJ**, Chien PH, Chen WS, Chien YF, Hsu YY, Wang LY, Chen JY, Lin CW, Huang TC, Yu YL, Huang WC. Hepatitis B Virus-Encoded X Protein Downregulates EGFR Expression via Inducing MicroRNA-7 in Hepatocellular Carcinoma Cells. *Evid Based Complement Alternat Med* 2013; **2013**: 682380 [PMID: 23840262 DOI: 10.1155/2013/682380]
- 81 **Wu G**, Yu F, Xiao Z, Xu K, Xu J, Tang W, Wang J, Song E. Hepatitis B virus X protein downregulates expression of the miR-16 family in malignant hepatocytes in vitro. *Br J Cancer* 2011; **105**: 146-153 [PMID: 21629246 DOI: 10.1038/ bjc.2011.190]
- 82 **Huang J**, Wang Y, Guo Y, Sun S. Down-regulated microR-

NA-152 induces aberrant DNA methylation in hepatitis B virus-related hepatocellular carcinoma by targeting DNA methyltransferase 1. *Hepatology* 2010; **52**: 60-70 [PMID: 20578129 DOI: 10.1002/hep.23660]

- 83 **Fu X**, Tan D, Hou Z, Hu Z, Liu G. miR-338-3p Is Down-Regulated by Hepatitis B Virus X and Inhibits Cell Proliferation by Targeting the 3'-UTR Region of CyclinD1. *Int J Mol Sci* 2012; **13**: 8514-8539 [PMID: 22942717 DOI: 10.3390/ijms13078514]
- 84 **Fu X**, Tan D, Hou Z, Hu Z, Liu G, Ouyang Y, Liu F. The effect of miR-338-3p on HBx deletion-mutant (HBx-d382) mediated liver-cell proliferation through CyclinD1 regulation. *PLoS One* 2012; **7**: e43204 [PMID: 22912826 DOI: 10.1371/ journal.pone.0043204]
- 85 **Xie QH**, He XX, Chang Y, Jiang X, Lin JS. [HBx gene downregulates miR-192 expression and inhibits apoptosis of human hepatoma cell line HepG2]. *Zhonghua Gan Zang Bing Za Zhi* 2011; **19**: 857-860 [PMID: 22433310 DOI: 10.3760/cma. j.issn.1007-3418.2011.11.015]
- Paterlini P, Poussin K, Kew M, Franco D, Brechot C. Selective accumulation of the X transcript of hepatitis B virus in patients negative for hepatitis B surface antigen with hepatocellular carcinoma. *Hepatology* 1995; **21**: 313-321 [PMID: 7843699 DOI: 10.1002/hep.1840210208]
- 87 **Wang WL**, London WT, Feitelson MA. Hepatitis B x antigen in hepatitis B virus carrier patients with liver cancer. *Cancer Res* 1991; **51**: 4971-4977 [PMID: 1654208]
- 88 **Chen GG**, Li MY, Ho RL, Chak EC, Lau WY, Lai PB. Identification of hepatitis B virus X gene mutation in Hong Kong patients with hepatocellular carcinoma. *J Clin Virol* 2005; **34**: 7-12 [PMID: 16087118 DOI: 10.1016/j.jcv.2005.01.006]
- 89 **Iavarone M**, Trabut JB, Delpuech O, Carnot F, Colombo M, Kremsdorf D, Bréchot C, Thiers V. Characterisation of hepatitis B virus X protein mutants in tumour and nontumour liver cells using laser capture microdissection. *J Hepatol* 2003; **39**: 253-261 [PMID: 12873823 DOI: 10.1016/ S0168-8278(03)00217-4]
- 90 **Guo X**, Jin Y, Qian G, Tu H. Sequential accumulation of the mutations in core promoter of hepatitis B virus is associated with the development of hepatocellular carcinoma in Qidong, China. *J Hepatol* 2008; **49**: 718-725 [PMID: 18801591 DOI: 10.1016/j.jhep.2008.06.026]
- Poussin K, Dienes H, Sirma H, Urban S, Beaugrand M, Franco D, Schirmacher P, Bréchot C, Paterlini Bréchot P. Expression of mutated hepatitis B virus X genes in human hepatocellular carcinomas. *Int J Cancer* 1999; **80**: 497-505 [PMID: 9935147 DOI: 10.1002/(SICI)1097-0215(19990209)80:4<497:: AID-IJC3>3.0.CO;2-8]
- 92 **Liu L**, Li Y, Zhang S, Yu D, Zhu M. Hepatitis B virus X protein mutant upregulates CENP-A expression in hepatoma cells. *Oncol Rep* 2012; **27**: 168-173 [PMID: 21956590 DOI: 10.3892/or.2011.1478]
- 93 **Ma NF**, Lau SH, Hu L, Xie D, Wu J, Yang J, Wang Y, Wu MC, Fung J, Bai X, Tzang CH, Fu L, Yang M, Su YA, Guan XY. COOH-terminal truncated HBV X protein plays key role in hepatocarcinogenesis. *Clin Cancer Res* 2008; **14**: 5061-5068 [PMID: 18698024 DOI: 10.1158/1078-0432.CCR-07-5082]
- 94 **Kumar V**, Jayasuryan N, Kumar R. A truncated mutant (residues 58-140) of the hepatitis B virus X protein retains transactivation function. *Proc Natl Acad Sci USA* 1996; **93**: 5647-5652 [PMID: 8643631 DOI: 10.1073/pnas.93.11.5647]
- 95 **Liu X**, Wang L, Zhang S, Lin J, Zhang S, Feitelson MA, Gao H, Zhu M. Mutations in the C-terminus of the X protein of hepatitis B virus regulate Wnt-5a expression in hepatoma Huh7 cells: cDNA microarray and proteomic analyses. *Carcinogenesis* 2008; **29**: 1207-1214 [PMID: 18477650 DOI: 10.1093/ carcin/bgn111]
- 96 **Wang Q**, Zhang T, Ye L, Wang W, Zhang X. Analysis of hepatitis B virus X gene (HBx) mutants in tissues of patients suffered from hepatocellular carcinoma in China. *Cancer Epidemiol* 2012; **36**: 369-374 [PMID: 22178505 DOI: 10.1016/

j.canep.2011.11.006]

- 97 **Wang Q**, Zhang WY, Ye LH, Zhang XD. A mutant of HBx (HBxDelta127) promotes hepatoma cell growth via sterol regulatory element binding protein 1c involving 5-lipoxygenase. *Acta Pharmacol Sin* 2010; **31**: 367-374 [PMID: 20173757 DOI: 10.1038/aps.2010.5]
- 98 **Jiang W**, Wang XW, Unger T, Forgues M, Kim JW, Hussain SP, Bowman E, Spillare EA, Lipsky MM, Meck JM, Cavalli LR, Haddad BR, Harris CC. Cooperation of tumor-derived HBx mutants and p53-249(ser) mutant in regulating cell proliferation, anchorage-independent growth and aneuploidy in a telomerase-immortalized normal human hepatocytederived cell line. *Int J Cancer* 2010; **127**: 1011-1020 [PMID: 20017137 DOI: 10.1002/ijc.25118]
- 99 **Hsu IC**, Metcalf RA, Sun T, Welsh JA, Wang NJ, Harris CC. Mutational hotspot in the p53 gene in human hepatocellular carcinomas. *Nature* 1991; **350**: 427-428 [PMID: 1849234 DOI: 10.1038/350427a0]
- 100 **Belloni L**, Pollicino T, De Nicola F, Guerrieri F, Raffa G, Fanciulli M, Raimondo G, Levrero M. Nuclear HBx binds the HBV minichromosome and modifies the epigenetic regulation of cccDNA function. *Proc Natl Acad Sci USA* 2009; **106**: 19975-19979 [PMID: 19906987 DOI: 10.1073/pnas.0908365106]
- 101 **Yeh CT**, Shen CH, Tai DI, Chu CM, Liaw YF. Identification and characterization of a prevalent hepatitis B virus X protein mutant in Taiwanese patients with hepatocellular carcinoma. *Oncogene* 2000; **19**: 5213-5220 [PMID: 11077437 DOI: 10.1038/sj.onc.1203903]
- 102 **Hsia CC**, Yuwen H, Tabor E. Hot-spot mutations in hepatitis B virus X gene in hepatocellular carcinoma. *Lancet* 1996; **348**: 625-626 [PMID: 8774611 DOI: 10.1016/S0140-6736(05)64851-9]
- 103 **Hsia CC**, Nakashima Y, Tabor E. Deletion mutants of the hepatitis B virus X gene in human hepatocellular carcinoma. *Biochem Biophys Res Commun* 1997; **241**: 726-729 [PMID: 9434776 DOI: 10.1006/bbrc.1997.7882]
- 104 **Wang Q**, Zhang W, Liu Q, Zhang X, Lv N, Ye L, Zhang X. A mutant of hepatitis B virus X protein (HBxDelta127) promotes cell growth through a positive feedback loop involving 5-lipoxygenase and fatty acid synthase. *Neoplasia* 2010; **12**: 103-115 [PMID: 20126469]
- 105 **Tang H**, Delgermaa L, Huang F, Oishi N, Liu L, He F, Zhao L,

Murakami S. The transcriptional transactivation function of HBx protein is important for its augmentation role in hepatitis B virus replication. *J Virol* 2005; **79**: 5548-5556 [PMID: 15827169 DOI: 10.1128/JVI.79.9.5548-5556.2005]

- 106 **Lin X**, Xu X, Huang QL, Liu YQ, Zheng DL, Chen WN, Lin JY. Biological impacts of "hot-spot" mutations of hepatitis B virus X proteins are genotype B and C differentiated. *World J Gastroenterol* 2005; **11**: 4703-4708 [PMID: 16094714]
- 107 **Tangkijvanich P**, Sa-Nguanmoo P, Mahachai V, Theamboonlers A, Poovorawan Y. A case-control study on sequence variations in the enhancer II/core promoter/precore and X genes of hepatitis B virus in patients with hepatocellular carcinoma. *Hepatol Int* 2010; **4**: 577-584 [PMID: 21063480 DOI: 10.1007/s12072-010-9197-z]
- 108 **Zhang H**, Shan CL, Li N, Zhang X, Zhang XZ, Xu FQ, Zhang S, Qiu LY, Ye LH, Zhang XD. Identification of a natural mutant of HBV X protein truncated 27 amino acids at the COOH terminal and its effect on liver cell proliferation. *Acta Pharmacol Sin* 2008; **29**: 473-480 [PMID: 18358094 DOI: 10.1111/j.1745-7254.2008.00764.x]
- 109 **Gouas DA**, Villar S, Ortiz-Cuaran S, Legros P, Ferro G, Kirk GD, Lesi OA, Mendy M, Bah E, Friesen MD, Groopman J, Chemin I, Hainaut P. TP53 R249S mutation, genetic variations in HBX and risk of hepatocellular carcinoma in The Gambia. *Carcinogenesis* 2012; **33**: 1219-1224 [PMID: 22759751 DOI: 10.1093/carcin/bgs135]
- 110 **Muroyama R**, Kato N, Yoshida H, Otsuka M, Moriyama M, Wang Y, Shao RX, Dharel N, Tanaka Y, Ohta M, Tateishi R, Shiina S, Tatsukawa M, Fukai K, Imazeki F, Yokosuka O, Shiratori Y, Omata M. Nucleotide change of codon 38 in the X gene of hepatitis B virus genotype C is associated with an increased risk of hepatocellular carcinoma. *J Hepatol* 2006; **45**: 805-812 [PMID: 17050029 DOI: 10.1016/j.jhep.2006.07.025]
- 111 **Kwun HJ**, Jang KL. Natural variants of hepatitis B virus X protein have differential effects on the expression of cyclindependent kinase inhibitor p21 gene. *Nucleic Acids Res* 2004; **32**: 2202-2213 [PMID: 15107488 DOI: 10.1093/nar/gkh553]
- 112 **Qadri I**, Maguire HF, Siddiqui A. Hepatitis B virus transactivator protein X interacts with the TATA-binding protein. *Proc Natl Acad Sci USA* 1995; **92**: 1003-1007 [PMID: 7862623 DOI: 10.1073/pnas.92.4.1003]

**P- Reviewer**: Gallego-Duran R, Luo GH, TanakaT **S- Editor**: Nan J **L- Editor**: A **E- Editor**: Wang CH







# Published by **Baishideng Publishing Group Inc**

8226 Regency Drive, Pleasanton, CA 94588, USA Telephone: +1-925-223-8242 Fax: +1-925-223-8243 E-mail: bpgoffice@wjgnet.com Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx http://www.wjgnet.com





 **© 2014 Baishideng Publishing Group Inc. All rights reserved.**