

Pharmacodynamic interaction of green tea extract with hydrochlorothiazide against ischemia-reperfusion injury-induced myocardial infarction

Manodeep Chakraborty,
Jagadish Vasudev Kamath¹

Departments of Pharmacology,
Bhagwant University, Ajmer, Rajasthan,
¹Shree Devi College of Pharmacy,
Mangalore, Karnataka, India

J. Adv. Pharm. Technol. Res.

ABSTRACT

Globally, the rate of development of myocardial diseases and hypertension is very common, which is responsible for incremental morbidity and mortality statistics. Treatment of ischemic hypertensive patients with diuretics such as hydrochlorothiazide (HCTZ) can precipitate myocardial infarction due to hypokalemia. This study was undertaken to evaluate the pharmacodynamic interaction of green tea extract (GTE) with HCTZ against ischemia-reperfusion induced myocardial toxicity. Wistar albino rats of either sex were taken and pretreated with high (500 mg/kg, p.o.) and low (100 mg/kg, p.o.) dose of GTE for 30 days. Standard, high and low dose of interactive groups received HCTZ (10 mg/kg, p.o.) for last 7 days. Ischemia-reperfusion injury was induced by modified Lagendorff apparatus, and the effect of different treatments was evaluated by percentage recovery in terms of heart rate and developed tension, serum biomarkers, and heart tissue antioxidant levels. Prophylactic treatment groups, such as high and low dose of GTE and their interactive groups with HCTZ, exhibited significant percentage recovery in terms of heart rate and developed tension. Apart from that, significant increase in superoxide dismutase and catalase, decrease in thiobarbituric acid reactive species in heart tissue, as well as significant decrease in serum lactate dehydrogenase, creatinine phosphokinase-MB and N-acetylcysteine levels have also been documented. The present findings clearly suggest that GTE dose-dependently reduces myocardial toxicity due to ischemia, and combination with HCTZ can reduce the associated side-effects and exhibits myocardial protection.

Key words: Antioxidant, biomarker, green tea extract, hydrochlorothiazide, ischemic-reperfusion injury

INTRODUCTION

From the ancient times, in many societies, herbs and herb-based therapy played an important role to treat different

diseased condition and to improve the quality-of-life.^[1] When synthetic drug is combined with herb, it may influence the pharmacokinetic and pharmacodynamic profile of each other and can mimic, magnify or oppose the action of each other.^[2,3]

The treatment of ischemic hypertensive patients with diuretic like hydrochlorothiazide (HCTZ) require utmost care, because mild to moderate level of hypokalemia may lead to the development of cardiac arrhythmias.^[4] The combination of HCTZ with angiotensin-converting enzyme inhibitor-I, aldosterone antagonist or angiotensin Type-I receptor blocker can be beneficial.^[5]

Green tea is one of the most consumed beverages in the world. It is obtained from the nonfermented leaves of *Camellia sinensis* belonging to family *Theaceae*, which contains more catechins than black tea or oolong tea. Due to the presence of high levels of catechin, certain minerals

Address for correspondence:

Mr. Manodeep Chakraborty,
Department of Pharmacology, Shree Devi College of Pharmacy,
Airport Road, Kenjar, Mangalore - 574 142, Karnataka, India.
E-mail: manodeep.chakraborty@gmail.com

Access this article online

Quick Response Code:



Website:

www.japtr.org

DOI:

10.4103/2231-4040.137428

and vitamins increase the antioxidant potential of this type of tea. Since ancient times, green tea has been considered as a healthful beverage by the traditional Chinese medicine. Recent studies have been reported that green tea may contribute to the reduction in risk of cardiovascular diseases and some forms of cancer, as well as to the promotion of oral health. It plays an important role in the control of body weight and has shown anti-hypertensive, antibacterial, antiviral, neuroprotective and anti-fibrotic properties. It is reported that green tea gives protection from solar ultraviolet rays and increase bone mineral density.^[6,7]

Green tea has been reported to prevent left ventricular hypertrophy, hypertension, cardiovascular damage and endothelial dysfunction. One of the clinical study showed that the consumption of green tea is responsible for reducing the occurrence of cardiovascular disease by increasing plasma antioxidant capacity.^[8-10]

Until now, no study has been carried out indicating the combined effect of green tea and HCTZ on myocardial cell. Hence, this study has been designed to evaluate the combined effect of green tea and HCTZ against ischemia-reperfusion injury (IRI) induced myocardial stress.

MATERIALS AND METHODS

Chemicals

All chemicals used were of analytical grade and purchased from standard companies. Pure sample of HCTZ was gifted by Bangalore Test House (Bangalore, India). Biochemical kits were procured from Crest Biosystems (Goa, India).

Experimental animals

Healthy adult Wistar albino rats of either sex weighing 175-250 g, were housed in polypropylene cages, maintained under standardized condition (12 h L: D cycles, 25 ± 5°C) with paddy husk bedding at the Central Animal House, Shree Devi College of Pharmacy, Mangalore; were provided with standard pellet food and had free access to purified drinking water. The guidelines of Committee for the Purpose of Control and Supervision of Experiments on Animals, Ministry of Social Justice and Empowerment, Government of India were followed and prior permission was sought from the Institutional Animal Ethics Committee for conducting the study (SDCP/IAEC-19/2012-13).

Plant materials

Green tea (*C. sinensis*) leaves were purchased in the month of June, 2013 from the local market of Mangalore bearing the brand name green tea, manufactured by New hilltop traders (Vandiperiyar, Kerala). The authentication was done by Dr. Neoline J. Pinto, H.O.D., Department of Botany, St. Agnes College, Mangalore (SAC/MNG/SMP/Drug/2013-06/52). The Aqueous extract was prepared by mixing green tea leaves gently in distilled water maintained at 70°C to 80°C for

30 min. Thereafter, it was filtered and evaporated in the same temperature to get a thick gummy mass. The yield was found to be 24.76% (W/W). Extract was freshly dissolved in distilled water before giving each dose to animals.

Phytochemical estimations of the extract

Aqueous extract of green tea (GTE) was subjected to qualitative analysis to investigate the presence of various phytochemical constituents such as alkaloids, glycosides, steroids, flavonoids, gallic tannins, catecholic tannin, terpenoid, and saponins.^[11,12]

Acute toxicity study

Acute toxicity study was carried out according to Office of Prevention, Pesticide and Toxic Substance guidelines following the limit test procedure.^[13,14]

Mice were fasted overnight prior to the studies, and then divided into two groups of three each. Test dose of 2 g/kg body weight and 5 g/kg body weight were given orally to either group of mice, and then observed for 72 h for mortality. 1/10th and 1/50th of the maximum safe dose corresponding to 500 and 100 mg/kg orally were selected as high and low doses, respectively.

Experimental protocol

The animals were divided into six different treatment groups of eight animals each. Group I was served as IRI control and the Group II received HCTZ orally at a dose of 10 mg/kg for 7 days.^[5] Groups III and IV were prophylactically treated with GTE at a dose of 100 and 500 mg/kg for 30 days through oral route. The animals of Groups V and VI were treated with GTE at a dose of 100 and 500 mg/kg for 30 days, along with that HCTZ (10 mg/kg) was incorporated for last 7 days.

Experimental procedure

Two hours after the last treatment, heart was isolated from the animals of different groups under ketamine (70 mg/kg, i.p.) and xylazine (10 mg/kg, i.p.) anesthesia. Isolated heart was subjected to modified Legendroff apparatus as mentioned in earlier reported article. The isolated heart was perfused with Krebs-Henseleit (K-H) solution, gassed with carbogen (95% O₂ and 5% CO₂) at 37°C, at a constant flow rate of 5 mL/min. The composition of K-H solution was (mM) NaCl 118, KCl 4.7, NaHCO₃ 25, NaHPO₄ 1.0, MgSO₄ 7H₂O 0.57, CaCl₂ 2.5, and glucose 11. The pH of K-H solution was adjusted to 7.4 to avoid K-H buffer acidosis that may occur after prolonged gassing with carbogen. The heart was allowed to equilibrate for 10 min and then regular recordings were taken for a perfusion period of 15 min. The heart was kept for equilibrium for a period of 10 min and then regular reading was taken for 15 min. The measurement of contractile force was determined with the help of force displacement transducer and recording was taken in digital physiograph (model no - DI-2, INCO, Ambala city, India). After the initial preischemic period, the isolated heart was subjected to no

flow global ischemia by stopping the flow of carbogenated K-H solution for 15 min. Then, the heart was reperfused for a period of 15 min. The heart rate and force of contraction was determined during the pre- and post-ischemic period, and the percentage recovery of heart rate and force of contraction was calculated. The levels of creatine kinase-MB (CK-MB), creatine kinase-N-acetylcysteine (CK-NAC), lactate dehydrogenase (LDH) activity were determined in perfusate collected during postischemic condition.^[15] Then half of the isolated hearts from all the groups were subjected for the preparation of heart tissue homogenate (HTH) using sucrose. Superoxide dismutase (SOD), catalase and thiobarbituric acid reactive substances (TBARS) were measured in HTH. For remaining heart samples, histological study was carried out.

Histological analysis

Heart sections were prepared from the remaining half of the heart samples in each group, stained with hematoxylin and eosin and change in histology were observed. The myocardial damage was determined by scoring method depending on the severity as follows, no change - 0 score, mild - 1 score (focal myocytes damage or small multifocal degeneration with a slight degree of inflammation), moderate - 2 score (extensive myofibrillar degeneration) and marked - 3 score (necrosis with diffuse inflammation).^[16]

Statistical analysis

Results are expressed as mean \pm standard error of the mean. Statistical significance was assessed using one-way analysis of variance followed by Tukey–Kramer multiple comparison tests. $P < 0.05$ was considered to be significant.

RESULTS

Preliminary phytochemical investigation

The preliminary phytochemical investigation of the GTE extract showed the presence of alkaloids, flavonoids, steroids, gallic tannins, and catecholic tannins. The percentage yield of GTE was found to be 24.76%.

Effect on lactate dehydrogenase, creatine kinase-MB and creatine kinase-N-acetylcysteine activities

The experimental group prophylactically treated with GTE-100, GTE-500 and GTE-500 + HCTZ showed a

significant decrease in LDH, CK-MB and CK-NAC levels in perfusate compared with IRI control alone.

High (GTE-500) and low dose (GTE-100) of GTE when incorporated with HCTZ, showed a significant decrease in LDH, CK-MB and CK-NAC activities in perfusate compared to HCTZ alone treated group [Table 1].

Effect on histological score

The groups pretreated with GTE-100, GTE-500 and GTE-500 combined with HCTZ, witnessed significant decrease in histological score compared to IRI control group. Groups treated with GTE-100 + HCTZ, GTE-500 + HCTZ showed a significant decrease in histological score compared to HCTZ alone treated group [Table 1].

Effect on superoxide dismutase, catalase and thiobarbituric acid reactive substances activities

Experimental groups such as GTE-100, GTE-500 and GTE-500 combined with HCTZ reported a significant increase in both SOD and catalase activities, and decrease in TBARS compared to IRI control group.

Groups treated with GTE-100 + HCTZ, GTE-500 + HCTZ showed a significant increase in both SOD and catalase, and decrease in TBARS activities compared to HCTZ alone treated group [Table 2].

Developed tension and heart rate

The treatment groups such as GTE-100, GTE-500 and GTE-500 + HCTZ showed significant recovery in ischemic heart in terms of heart rate and developed tension compared to IRI control. GTE-500 + HCTZ and GTE-100 + HCTZ showed significant recovery compared to HCTZ alone treated group [Table 2].

Effect on histopathological study

Under ischemia, the isolated hearts showed different pathological changes such as severe myocardial edema, separation of fibers, and loss of striation. HCTZ alone treated group also associated with these severe pathological changes. GTE-100, GTE-500 and GTE-500 + HCTZ showed a significant decrease in histopathological score. GTE-100 + HCTZ and GTE-500 + HCTZ both documented

Table 1: Effect on CK-NAC, CK-MB, LDH in perfusate using IRI in isolated rat heart

Treatment	CK-NAC (U/L)	CK-MB (U/L)	LDH (U/L)	Histological score
IRI	1450.00 \pm 28.86	797.10 \pm 60.26	849.03 \pm 32.18	2.91 \pm 0.17
HCTZ	1373.33 \pm 14.53	699.00 \pm 15.26	782.10 \pm 25.28	2.59 \pm 0.15
GTE-100	608.90 \pm 81.14 ^{###}	399.00 \pm 15.86 ^{###}	549.63 \pm 76.01 ^{###}	1.41 \pm 0.09 ^{###}
GTE-500	332.23 \pm 7.86 ^{###}	262.26 \pm 9.53 ^{###}	310.00 \pm 20.81 ^{###}	1.27 \pm 0.12 ^{###}
GTE-100+HCTZ	1030.00 \pm 25.16 [*]	616.80 \pm 34.24 [*]	649.93 \pm 18.99 [*]	2.03 \pm 0.21 ^{**}
GTE-500+HCTZ	913.33 \pm 3.33 ^{#####}	568.33 \pm 13.08 ^{#####}	589.33 \pm 23.67 ^{#####}	1.95 \pm 0.16 ^{###}

All values are mean \pm SEM, $n=6$, ^{###} $P < 0.001$, ^{**} $P < 0.01$, ^{*} $P < 0.05$ compared to IRI control and ^{###} $P < 0.001$, ^{**} $P < 0.01$, ^{*} $P < 0.05$ compared to HCTZ. GTE-100: Green tea extract-100 mg/kg, GTE-500: Green tea extract-500 mg/kg, HCTZ: Hydrochlorothiazide-10 mg/kg, SEM: Standard error of mean, CK-MB: Creatine kinase-MB, CK-NAC: Creatine kinase-N-acetylcysteine, LDH: Lactate dehydrogenase, IRI: Ischemia-reperfusion injury

significant reduction in histopathological score compared to HCTZ alone treated group [Figure 1].

DISCUSSION

The aim of this study was to elucidate the pharmacodynamic interaction of GTE with HCTZ using ischemia-reperfusion induced myocardial injury. Observed results suggested that GTE (100 and 500 mg/kg, p.o.) showed beneficial results dose-dependently. Apart from that, GTE when combined with HCTZ indicated better results compared to HCTZ alone treated group against ischemia-reperfusion induced myocardial injury.

In this experimental model, myocardial toxicity was induced by ischemia-reperfusion. Ischemia, which is an acute or chronic form of cardiac disability, occurred due to imbalance between the supply and demand of oxygen in the blood. In IRI model, global ischemia causes immediate biological alteration, such as increase in intracellular Na⁺,

which results an increase in intracellular Ca²⁺ via Na⁺/Ca²⁺ exchange. Increased intracellular Ca²⁺ is responsible for the formation of irreversible damage in the cardiac cell at the end of 15 min global ischemia.^[17-19]

Hydrochlorothiazide alters the renal tubular mechanisms of electrolyte reabsorption. The direct action of HCTZ is responsible for an increase in the excretion of sodium and chloride. The indirect action causes reduction in plasma volume, consequent increase in urinary potassium loss, plasma renin activity, aldosterone secretion, and decrease in serum potassium loss. It has been observed that in patients with cardiac ischemia, heart failure or left ventricular hypertrophy, the likelihood of cardiac arrhythmia is increased because of mild-to-moderate hypokalemia. It has been reported that HCTZ induced hypokalemia is responsible for the increase in serum biomarker levels, such as LDH, CK-MB, and decrease in heart tissue antioxidant levels like SOD and catalase in rat. Therefore, it is proved that their undesirable metabolic consequences have been

Table 2: Effect on SOD, catalase, TBARS in heart tissue homogenate and percentage recovery of developed tension and heart rate using IRI in isolated rat heart

Treatment	SOD (U/mg)	Catalase (U/mg)	TBARS (U/mg)	Percentage recovery	
				Heart rate	Developed tension
IRI	1.93±0.23	1.54±0.27	19.66±2.90	51.29±2.85	26.79±0.92
HCTZ	2.13±0.14	1.93±0.32	19.43±0.44	51.10±1.77	25.16±2.27
GTE-100	4.50±0.28 ^{###}	4.53±0.20 ^{###}	10.00±0.57 [#]	68.13±1.78 ^{###}	59.57±3.47 ^{###}
GTE-500	5.73±0.14 ^{###}	7.36±0.24 ^{###}	7.00±0.57 ^{###}	73.60±0.47 ^{###}	77.39±3.60 ^{###}
GTE-100+HCTZ	2.73±0.14 ^{**}	2.80±0.15 ^{**}	17.83±0.44 ^{**}	56.87±3.45 ^{**}	32.87±3.45 ^{**}
GTE-500+HCTZ	3.40±0.45 ^{***}	3.76±0.12 ^{***}	13.33±0.44 ^{***}	59.76±1.76 ^{***}	42.76±1.76 ^{***}

All values are mean±SEM, n=6, ^{###}P<0.001, ^{**}P<0.001, [#]P<0.05 compared to IRI control and ^{***}P<0.001, ^{**}P<0.01 compared to HCTZ. GTE-100: Green tea extract-100 mg/kg, GTE-500: Green tea extract-500 mg/kg, HCTZ: Hydrochlorothiazide-10 mg/kg, SEM: Standard error of mean, SOD: Super oxide dismutase, TBARS: Thiobarbituric acid reactive substances, IRI: Ischemia-reperfusion injury

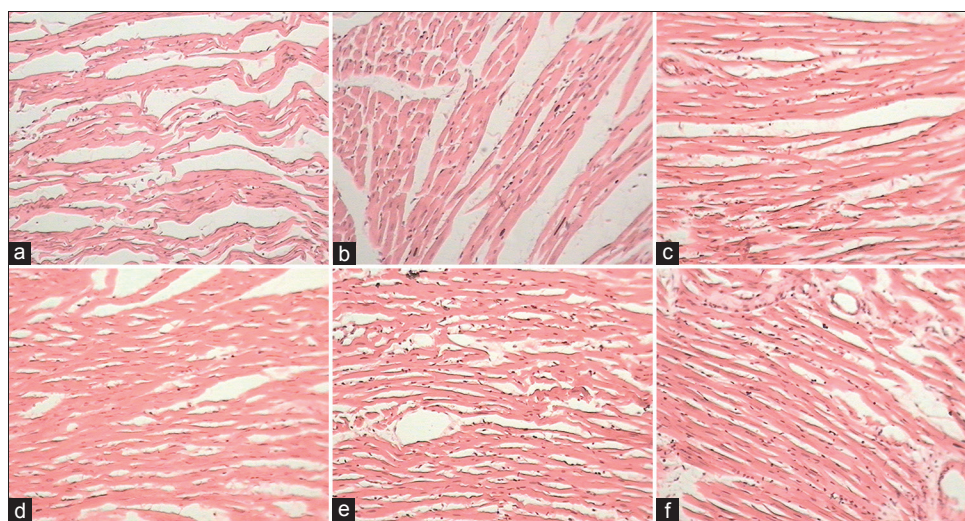


Figure 1: Microscopic section of (a) ischemia-reperfusion injury control – (1) Myocardial edema, (2) separation of fibers, (3) loss of striation, (4) diffuse inflammation (5) necrosis. (b) Hydrochlorothiazide (HCTZ) treated – (1) Myocardial edema, (2) separation of fibers, (3) loss of striation. (c) Green tea extract-100 (GTE)-100 – (1) Myocardial edema, (2) separation of fibers, (3) loss of striation. (d) GTE-500 – (1) Myocardial edema, (2) separation of fibers. (e) GTE-100 + HCTZ – (1) Myocardial edema, (2) separation of fibers, (3) loss of striation. (f) GTE-500 + HCTZ – (1) Myocardial edema, (2) separation of fibers, (3) loss of striation (H and E, ×400)

suspected to contribute an increase in cardiovascular morbidity and mortality. Hence, search for concurrently administered safe therapeutic medicament continues, which can ameliorate the hypokalemia in patients with ischemic heart diseases.^[5,20]

Camellia sinensis (green tea) is having a rich source of polyphenols, which is responsible for strong free radical-scavenging activity. Green tea has reported its potency against cardiovascular disease risk factors. It reduces body weight by interfering within the sympathoadrenal system, reduction in fatty acid synthesis and decrease in cholesterol absorption. It possesses antithrombotic activity by inhibiting platelet aggregation. Green tea inhibits low density lipid oxidation, reduce adhesion molecule expression and reduce systolic as well as diastolic blood pressures.^[21]

In this study, ischemia-reperfusion induced injury in the myocardial cell generates oxygen free radicals. Oxygen free radical is extensively associated with the loss of membrane integrity, which causes leaking of different biomarker enzymes from the cell. Apart from that, antioxidants, such as SOD and catalase also come out, which is responsible for inhibition of free radicals neutralization. As a result, further accumulation of free radicals takes place, leading to exaggerated tissue damage.^[22]

Prophylactic treatment with GTE in both high and low dose, reported to have a beneficial role against the ischemic condition in myocardial cell. GTE dose-dependently decreased serum biomarker levels and increased antioxidant levels in tissue. Apart from that, GTE is also responsible for the significant recovery in terms of heart rate and developed tension. The combination group of GTE (500 and 100 mg/kg) and HCTZ demonstrated better results compared to HCTZ alone. Comparatively, high dose of GTE (500 mg/kg) was found to be more beneficial to reduce HCTZ induced myocardial toxicity. Histopathological study showed that high and low doses of GTE and their combination with HCTZ demonstrated a significant reduction in the histological score by reducing inflammation and restoring myocardial cell integrity, which supports the other findings of this study.

CONCLUSION

It can be concluded that GTE exhibited dose-dependent protection against ischemia-reperfusion induced myocardial injury. Apart from that, combination of GTE reduced significant myocardial side-effects associated with HCTZ. This observation can be very important for ischemic hypertensive patients, where HCTZ cannot be given as monotherapy due to potential myocardial side-effects. Future studies can be carried out to establish the fact clinically.

REFERENCES

1. Waltner-Law ME, Wang XL, Law BK, Hall RK, Nawano M, Granner DK. Epigallocatechin gallate, a constituent of green tea, represses hepatic glucose production. *J Biol Chem* 2002;277:34933-40.
2. Fugh-Berman A, Ernst E. Herb-drug interactions: Review and assessment of report reliability. *Br J Clin Pharmacol* 2001;52:587-95.
3. Hoes AW, Grobbee DE, Peet TM, Lubsen J. Do non-potassium-sparing diuretics increase the risk of sudden cardiac death in hypertensive patients? Recent evidence. *Drugs* 1994;47:711-33.
4. Fugh-Berman A. Herb-drug interactions. *Lancet* 2000;355:134-8.
5. Asdaq SM, Inamdar MN. The potential for interaction of hydrochlorothiazide with garlic in rats. *Chem Biol Interact* 2009;181:472-9.
6. Cabrera C, Artacho R, Giménez R. Beneficial effects of green tea: A review. *J Am Coll Nutr* 2006;25:79-99.
7. Brown MD. Green tea (*Camellia sinensis*) extract and its possible role in the prevention of cancer. *Altern Med Rev* 1999;4:360-70.
8. Papparella I, Ceolotto G, Montemurro D, Antonello M, Garbisa S, Rossi G, et al. Green tea attenuates angiotensin II-induced cardiac hypertrophy in rats by modulating reactive oxygen species production and the Src/epidermal growth factor receptor/Akt signaling pathway. *J Nutr* 2008;138:1596-601.
9. Antonello M, Montemurro D, Bolognesi M, Di Pascoli M, Piva A, Grego F, et al. Prevention of hypertension, cardiovascular damage and endothelial dysfunction with green tea extracts. *Am J Hypertens* 2007;20:1321-8.
10. Nakagawa K, Ninomiya M, Okubo T, Aoi N, Juneja LR, Kim M, et al. Tea catechin supplementation increases antioxidant capacity and prevents phospholipid hydroperoxidation in plasma of humans. *J Agric Food Chem* 1999;47:3967-73.
11. Yanagi S, Matsumura K, Marui A, Morishima M, Hyon SH, Ikeda T, et al. Oral pretreatment with a green tea polyphenol for cardioprotection against ischemia-reperfusion injury in an isolated rat heart model. *J Thorac Cardiovasc Surg* 2011;141:511-7.
12. Bajerska J, Wozniowicz M, Jeszka J, Drzymala-Czyz S, Walkowiak J. Green tea aqueous extract reduces visceral fat and decreases protein availability in rats fed with a high-fat diet. *Nutr Res* 2011;31:157-64.
13. Finar IL. *Organic Chemistry*. 4th ed. London: ELBS; 1993. p. 518.
14. Mukherjee PK. *Quality Control of Herbal Drugs: An Approach to Evaluation of Botanicals*. 1st ed. New Delhi: Business Horizons; 2002.
15. Inamdar MN, Venkataraman BV, Aleem MA. A simple and improved perfusion apparatus for isolated hearts. *Indian J Pharmacol* 1994;26:262-5.
16. Chakraborty M, Asdaq SM. Interaction of *Semecarpus anacardium* L. with propranolol against isoproterenol induced myocardial damage in rats. *Indian J Exp Biol* 2011;49:200-6.
17. Mouhieddine S, Tresallet N, Boucher F, de Leiris J. Ultrastructural basis of the free-radical scavenging effect of indapamide in experimental myocardial ischemia and reperfusion. *J Cardiovasc Pharmacol* 1993;22 Suppl 6:S47-52.
18. James TN, Keyes JW. *Studies of the dying myocardial cell. The Etiology of Myocardial Infarction*. Boston, Mass: Little, Brown and Co.; 1963. p. 189-205.
19. Jennings RB, Schaper J, Hill ML, Steenbergen C Jr, Reimer KA. Effect of reperfusion late in the phase of reversible ischemic injury. Changes in cell volume, electrolytes, metabolites, and ultrastructure. *Circ Res* 1985;56:262-78.
20. Asdaq SM, Inamdar MN. Pharmacodynamic interaction of garlic with hydrochlorothiazide in rats. *Indian J Physiol Pharmacol* 2009;53:127-36.

21. Hernández Figueroa TT, Rodríguez-Rodríguez E, Sánchez-Muniz FJ. The green tea, a good choice for cardiovascular disease prevention? Arch Latinoam Nutr 2004;54:380-94.
22. Asdaq SM, Inamdar MN, Asad M. Pharmacodynamic interaction of garlic with propranolol in ischemia-reperfusion induced myocardial damage. Pak J Pharm Sci 2010;23:42-7.

How to cite this article: Chakraborty M, Kamath JV. Pharmacodynamic interaction of green tea extract with hydrochlorothiazide against ischemia-reperfusion injury-induced myocardial infarction. J Adv Pharm Technol Res 2014;5:134-9.

Source of Support: Nil, **Conflict of Interest:** Nil.