Early Developmental Disturbances of Cortical Inhibitory Neurons: Contribution to Cognitive Deficits in Schizophrenia

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Cognitive dysfunction is a disabling and core feature of schizophrenia. Cognitive impairments have been linked to disturbances in inhibitory (gamma-aminobutyric acid [GABA]) neurons in the prefrontal cortex. Cognitive deficits are present well before the onset of psychotic symptoms and have been detected in early childhood with developmental delays reported during the first year of life. These data suggest that the pathogenetic process that produces dysfunction of prefrontal GABA neurons in schizophrenia may be related to altered prenatal development. Interestingly, adult postmortem schizophrenia brain tissue studies have provided evidence consistent with a disease process that affects different stages of prenatal development of specific subpopulations of prefrontal GABA neurons. Prenatal ontogeny (ie, birth, proliferation, migration, and phenotypic specification) of distinct subpopulations of cortical GABA neurons is differentially regulated by a host of transcription factors, chemokine receptors, and other molecular markers. In this review article, we propose a strategy to investigate how alterations in the expression of these developmental regulators of subpopulations of cortical GABA neurons may contribute to the pathogenesis of cortical GABA neuron dysfunction and consequently cognitive impairments in schizophrenia.

Key words: parvalbumin/somatostatin/prefrontal cortex/ interneuron/GABA neuron/prenatal ontogeny/ postmortem/development

Introduction

Schizophrenia is frequently associated with a lifetime of impairment in social and occupational domains and premature mortality.^{1,2} While the diagnostic clinical features of schizophrenia include positive or psychotic symptoms and negative symptoms, poor long-term outcomes in individuals with schizophrenia have been principally linked to the severity of cognitive dysfunction in the illness.³ Cognitive impairments in schizophrenia include deficits in selective attention, declarative memory, working memory, and cognitive control.⁴ Cognitive dysfunction represents a core impairment in schizophrenia because cognitive deficits are common and relatively stable across the course of the illness, are present regardless of psychotic symptoms, and are found to a lesser degree in unaffected relatives.^{5,6} Furthermore, the lack of responsiveness of cognitive deficits to available antipsychotic medications^{7,8} indicates the need for greater insight into the pathogenetic processes that lead to the appearance of cognitive dysfunction in the illness.

Interestingly, cognitive disturbances are present prior to the onset of psychotic symptoms that typically occurs in late adolescence and early adulthood,⁹ which suggests that the disease process that underlies cognitive problems may disrupt the normal maturation of cortical circuits (previously reviewed).^{10,11} However, developmental delays have also been observed in early childhood,¹² even prior to the first birthday,^{13–15} in individuals who go on to develop schizophrenia. These and other findings have led to the hypothesis that the disease process that ultimately leads to the appearance of clinical symptoms of schizophrenia in late teens/early 20s could actually have its origins in utero.^{16,17} Consistent with this hypothesis, environmental insults that occur while in utero, such as maternal exposure to infection, have been linked to higher rates of schizophrenia.¹⁸ However, further investigation of the role of prenatal insults in the pathogenesis of schizophrenia requires a consideration of the components of cortical pathology that may subserve cognitive impairments and how disturbances in the prenatal ontogeny of this cortical circuitry may contribute to the nature of cortical circuitry dysfunction in the illness.

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Disturbances in Prefrontal GABA Neurons Contribute to Cognitive Dysfunction in Schizophrenia

Cognitive processes such as working memory are supported by the synchronized firing of groups of cortical pyramidal neurons at gamma frequencies (30-80 Hz).¹⁹ Individuals with schizophrenia show altered gamma oscillation activity in the prefrontal cortex while performing cognitive tasks.^{20,21} Gamma oscillations are subserved by a subpopulation of cortical inhibitory (gammaaminobutyric acid [GABA]) neurons that contain the calcium-binding protein parvalbumin and provide powerful perisomatic inhibitory control over pyramidal neuron output. Disturbing the function of parvalbumin neurons leads to lower gamma oscillatory power.^{22,23} Consequently, the presence of a disease-related disturbance in parvalbumin neurons would be predicted to interfere with the regulation of pyramidal neuron activity and gamma oscillations and lead to deleterious effects on cognition in schizophrenia.²⁴

Interestingly, some of the most consistently reported postmortem brain tissue findings in schizophrenia involve disturbances in parvalbumin neurons in the prefrontal cortex. For example, lower parvalbumin mRNA levels have been reported in prefrontal cortex gray matter homogenates in multiple different cohorts of schizophrenia subjects^{25–28} and do not appear to be attributable to antipsychotic medications.^{25,28} Furthermore, approximately half of parvalbumin neurons in the prefrontal cortex in schizophrenia fail to express detectable mRNA levels for the GABA synthesizing enzyme GAD67,²⁵ and protein levels of parvalbumin and GAD67 have been reported to be reduced in parvalbumin axon terminals in the illness.^{29,30} In addition, voltage-gated potassium channels are involved in the fast repolarization of neurons that allow high frequency firing such as gamma oscillations.³¹ Interestingly, transcript levels of several voltage-gated potassium channels that are selectively found in parvalbumin neurons, including Kv3.1 and KCNS3, which encode the Kv9.3 modulatory α subunit,^{31,32} are lower in the prefrontal cortex in schizophrenia.^{33,34} Taken together, these disturbances in biochemical markers important for the gamma oscillation-related function of parvalbumin neurons may provide a substrate for some of the cognitive disturbances observed in schizophrenia.

Alteration in Subpopulations of Cortical GABA Neurons Are Consistent With a Prenatal Origin in Schizophrenia

Recent studies have provided insight into the prenatal ontogeny (ie, neuronal birth, proliferation, migration, and phenotypic specification) of different subpopulations of cortical GABA neurons. In humans, (future) parvalbumin neurons, and another subpopulation of cortical GABA neurons that express the neuropeptide somatostatin, begin to be born and proliferate by the 8th week of gestation in the medial ganglionic eminence of primordial basal ganglia, then begin to migrate tangentially to the cerebral cortex.^{35–41} In contrast to parvalbumin and somatostatin neurons, the subpopulation of cortical GABA neurons that express the calciumbinding protein calretinin appears to largely originate from the caudal ganglionic eminence and possibly also from the subventricular zone of the dorsal pallium.³⁸⁻⁴⁴ Phenotypic specification involves the ongoing process of developing cell-type-specific expression patterns of biochemical markers, electrophysiological properties, and anatomical features that differentiate neurons into subclasses such as parvalbumin, somatostatin, and calretinin neurons. Furthermore, as discussed in the next section, many transcription factors and other molecular markers are required for various stages of prenatal ontogeny of cortical GABA neurons.

Interestingly, the pattern of evidence from postmortem human brain tissue studies suggests that disturbances in parvalbumin neurons may originate during prenatal ontogeny. First, lower mRNA levels for somatostatin, but not calretinin,^{25,28} have also been consistently observed in the prefrontal cortex across multiple cohorts of schizophrenia subjects.^{26-28,45} Furthermore, parvalbumin and somatostatin mRNAs have been reported to be primarily lower in the same schizophrenia subjects identified as having a "low-GABA-marker" molecular phenotype (approximately half of all schizophrenia subjects studied), while the other schizophrenia subjects show no abnormalities in either parvalbumin or somatostatin mRNA in the prefrontal cortex.²⁸ Thus, a selective disturbance in cortical parvalbumin and somatostatin neurons, but not calretinin neurons, in the same schizophrenia subjects may potentially be explained by a pathogenetic process early in development that selectively affects neurons originating from the medial ganglionic eminence.

Second, the characteristics of disturbances in parvalbumin and somatostatin neurons suggest that different stages of prenatal ontogeny may be disrupted in schizophrenia. For example, a full complement of parvalbumin neurons appears to present in the prefrontal cortex in schizophrenia.25,46 However, approximately half of parvalbumin neurons lack detectable mRNA levels of GAD67,²⁵ which suggests that parvalbumin neurons may migrate normally but fail to fully complete phenotypic specification.⁴⁷ Similarly, in situ hybridization studies of the prefrontal cortex in schizophrenia have also found that detectable somatostatin neurons express lower somatostatin mRNA levels.45,48 However, in contrast to the findings with parvalbumin neurons in schizophrenia, fewer somatostatin neurons were detectable in the gray matter, while more somatostatin neurons were detectable in the white matter of the prefrontal cortex in the illness.^{45,48} Thus, available data from postmortem human brain tissue studies are at least consistent with a disease

process that interferes with the migration of some somatostatin neurons in cortical white matter early in development and leads to an incomplete phenotypic specification of the cortical parvalbumin and somatostatin neurons that complete the migration process. Consistent with this hypothesis, other studies have also observed a higher density of interstitial white matter neurons in the prefrontal cortex in schizophrenia.49-52 This evidence from postmortem human brain tissue studies, in combination with clinical findings of developmental delays in the first year of life,¹³⁻¹⁵ raises the hypothesis that selective disturbance in subpopulations of cortical GABA neurons may begin during the period of prenatal ontogeny in schizophrenia. Understanding potential pathogenetic mechanisms that could selectively disrupt the development of parvalbumin and somatostatin neurons next requires a consideration of the factors that regulate prenatal GABA neuron ontogeny and how disturbances in the expression levels of these ontogenetic factors in schizophrenia may contribute to cortical GABA neuron dysfunction in the illness.

Transcription Factors, Chemokine Receptors, and Other Molecular Markers Regulate the Prenatal Development of Cortical Parvalbumin and Somatostatin Neurons

Numerous transcription factors, which are proteins that selectively bind to specific regions of DNA and regulate the transcription of mRNAs, are required for the normal migration, phenotypic specification, maturation, and survival of specific subpopulations of cortical GABA neurons. For example, multiple cell-type-specific transcription factors such as Nkx2.1, Sox6, MafB, Zeb2 (also known as Sip1 and Zfhx1b), Dlx5, and Dlx6 are expressed early in gestation in the medial ganglionic eminence and regulate the migration, specification, and maturation of cortical parvalbumin and somatostatin, but not calretinin, neurons.^{37,53-60} Some molecular markers appear to be particularly important for the development of cortical parvalbumin neurons. For example, certain developmental regulators including the chemokine receptors CXCR4 and CXCR7 and the scaffold protein Disruptedin-Schizophrenia 1 (DISC1) are all strongly expressed in the medial ganglionic eminence and are necessary for the migration of cortical parvalbumin neurons to the cerebral cortex.^{61–65} In addition, the receptor tyrosine kinase for the trophic factor neuregulin 1, ErbB4, regulates the migration of cortical parvalbumin neurons and the development of excitatory synapses onto parvalbumin neurons.^{66–68} In contrast, the transcriptional coactivator peroxisome proliferator-activated receptor γ coactivator 1α begins to be expressed postnatally by parvalbumin neurons and is essential for complete development of the parvalbumin neuron phenotype.⁶⁹

While there is considerable overlap with the molecular markers that regulate parvalbumin neuron development,

some developmental regulators appear to be more specific for somatostatin neuron development. For example, ontogenetic transcription factor Dlx⁷⁰ does not appear to be required for the tangential migration of cortical GABA neurons, but is required for postnatal expression of GAD67.⁷¹ Postnatally, Dlx1 continues to be expressed by somatostatin neurons, but not parvalbumin neurons, and a complete loss of Dlx1 leads to a failure of cortical somatostatin, but not parvalbumin neurons, to survive into adulthood.⁷¹ In addition, after neuronal migration is complete, most cortical parvalbumin and somatostatin, but not calretinin, neurons begin to express the nuclear matrix and genome organizer Special AT-rich DNA Binding Protein 1 (SATB1).⁷² A complete loss of SATB1 does not appear to affect parvalbumin neurons but instead leads to substantially lower somatostatin mRNA and protein levels without affecting cell number, which indicates a role in the terminal differentiation and maturation of somatostatin neurons.72

Altered Expression of Transcription Factors and Other Developmental Regulators Might Contribute to Cortical Parvalbumin and Somatostatin Neuron Dysfunction in Schizophrenia

Taken together, the critical and varied roles of these (and many more) ontogenetic transcriptional regulators and chemokine receptors in the different stages of development of cortical parvalbumin and somatostatin neurons suggest that a disturbance in their expression or function could plausibly produce the pattern of deficits in parvalbumin and somatostatin neurons observed in the prefrontal cortex in schizophrenia. Interestingly, recent studies have reported abnormalities in these developmental regulators in schizophrenia. For example, lower Dlx1 mRNA levels have been reported in the orbital frontal cortex in subjects with schizophrenia.73 Given the important role that Dlx1 has been reported to play in the postnatal development and survival of cortical somatostatin neurons,⁷¹ deficits in Dlx1 may potentially contribute to some of the disturbances reported in somatostatin neurons, including lower somatostatin mRNA levels and fewer detectable somatostatin neurons, in schizophrenia.45 In contrast, higher mRNA levels for a splice variant of ErbB4, termed JMa, have been reported in the prefrontal cortex in multiple cohorts of schizophrenia subjects.74-76 Because the ErbB4-JMa splice variant is extracellular and is susceptible to proteolytic cleavage,⁷⁷ Weickert and colleagues have hypothesized that higher ErbB4-JMa levels may interfere with neuregulin signaling in schizophrenia,⁷⁶ which may in turn disrupt the migration of, and development of excitatory inputs to, cortical parvalbumin neurons in the disorder.^{66–68} Furthermore, DISC1, which is important for the tangential migration of future cortical parvalbumin neurons,^{64,65} was previously identified as a susceptibility gene for schizophrenia.⁷⁸ In addition.

recent genome-wide association studies have implicated Zeb2, another ontogenetic transcription factor involved in cortical parvalbumin and somatostatin neuron development,^{59,60} as a candidate gene that provides increased risk for schizophrenia.⁷⁹

However, the prenatal ontogeny of cortical parvalbumin and somatostatin neurons cannot be directly studied in schizophrenia and instead requires a multifaceted approach to investigate the role of prenatal disturbances in the development of cortical GABA neuron dysfunction in schizophrenia. First, the use of postmortem human brain tissue studies allows the identification of developmental factors that are consistently altered in expression in schizophrenia and the determination of whether the pattern of deficits in developmental factors is consistent with the observed alterations in cortical parvalbumin and somatostatin neurons in schizophrenia. Second, the use of postmortem brain tissue studies from a wide range of postnatal ages of monkeys, which have a more similar composition of cortical GABA neuron subpopulations and extended period of postnatal development to humans, allows a determination of whether altered expression of transcription factors in schizophrenia is more similar to an altered postnatal developmental trajectory, as has been reported for other parvalbumin neuron markers.^{10,11,80} Finally, animal models that mirror the magnitude and cell-type specificity of the deficits in ontogenetic factors observed in schizophrenia are needed to provide construct validity for models of disrupted prenatal ontogenv of cortical parvalbumin and somatostatin neurons in schizophrenia.

In this issue, we utilize this cross-species approach to investigate another important transcription factor, Lhx6, that is selectively expressed by, and involved in the prenatal developmental of, cortical parvalbumin and somatostatin neurons.^{32,81–83} Investigating the expression level of Lhx6 in the prefrontal cortex in schizophrenia subjects, the pattern of postnatal development of Lhx6 in the prefrontal cortex of monkeys, and the effects of a partial, cell-type specific loss of Lhx6 that mimics Lhx6 deficits in schizophrenia helps provide a framework for the process of investigating potential disruptions in prenatal development in schizophrenia. This approach may serve as a useful strategy for defining the pathogenesis of cortical circuit dysfunction in schizophrenia and for identifying the molecular mechanisms that are vulnerable to environmental events at different stages of development. This knowledge may inform both the type and timing of preemptive interventions for cognitive dysfunction in schizophrenia.

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References

- 1. Insel TR, Scolnick EM. Cure therapeutics and strategic prevention: raising the bar for mental health research. *Mol Psychiatry*. 2006;11:11–17.
- Saha S, Chant D, McGrath J. A systematic review of mortality in schizophrenia: is the differential mortality gap worsening over time? *Arch Gen Psychiatry*. 2007;64:1123–1131.
- Green MF. Cognitive impairment and functional outcome in schizophrenia and bipolar disorder. J Clin Psychiatry. 2006;67(suppl 9):3–8; discussion 36.
- 4. Lesh TA, Niendam TA, Minzenberg MJ, Carter CS. Cognitive control deficits in schizophrenia: mechanisms and meaning. *Neuropsychopharmacology*. 2011;36:316–338.
- 5. Egan MF, Goldberg TE, Gscheidle T, et al. Relative risk for cognitive impairments in siblings of patients with schizophrenia. *Biol Psychiatry*. 2001;50:98–107.
- Keefe RS, Fenton WS. How should DSM-V criteria for schizophrenia include cognitive impairment? *Schizophr Bull*. 2007;33:912–920.
- Keefe RS, Bilder RM, Davis SM, et al.; CATIE Investigators; Neurocognitive Working Group. Neurocognitive effects of antipsychotic medications in patients with chronic schizophrenia in the CATIE Trial. Arch Gen Psychiatry. 2007;64:633–647.
- Keefe RS, Sweeney JA, Gu H, et al. Effects of olanzapine, quetiapine, and risperidone on neurocognitive function in early psychosis: a randomized, double-blind 52-week comparison. *Am J Psychiatry*. 2007;164:1061–1071.
- Woodberry KA, Giuliano AJ, Seidman LJ. Premorbid IQ in schizophrenia: a meta-analytic review. *Am J Psychiatry*. 2008;165:579–587.
- 10. Beneyto M, Lewis DA. Insights into the neurodevelopmental origin of schizophrenia from postmortem studies of prefrontal cortical circuitry. *Int J Dev Neurosci*. 2011;29:295–304.
- Hoftman GD, Lewis DA. Postnatal developmental trajectories of neural circuits in the primate prefrontal cortex: identifying sensitive periods for vulnerability to schizophrenia. *Schizophr Bull*. 2011;37:493–503.
- 12. Jones P, Rodgers B, Murray R, Marmot M. Child development risk factors for adult schizophrenia in the British 1946 birth cohort. *Lancet*. 1994;344:1398–1402.
- Ridler K, Veijola JM, Tanskanen P, et al. Fronto-cerebellar systems are associated with infant motor and adult executive functions in healthy adults but not in schizophrenia. *Proc Natl Acad Sci U S A*. 2006;103:15651–15656.
- 14. Sørensen HJ, Mortensen EL, Schiffman J, Reinisch JM, Maeda J, Mednick SA. Early developmental milestones and risk of schizophrenia: a 45-year follow-up of the Copenhagen Perinatal Cohort. *Schizophr Res.* 2010;118:41–47.
- 15. Clarke MC, Tanskanen A, Huttunen M, et al. Increased risk of schizophrenia from additive interaction between infant motor developmental delay and obstetric complications: evidence

from a population-based longitudinal study. *Am J Psychiatry*. 2011;168:1295–1302.

- Weinberger DR. Implications of normal brain development for the pathogenesis of schizophrenia. *Arch Gen Psychiatry*. 1987;44:660–669.
- Murray RM, O'Callaghan E, Castle DJ, Lewis SW. A neurodevelopmental approach to the classification of schizophrenia. *Schizophr Bull*. 1992;18:319–332.
- Brown AS, Derkits EJ. Prenatal infection and schizophrenia: a review of epidemiologic and translational studies. *Am J Psychiatry*. 2010;167:261–280.
- 19. Howard MW, Rizzuto DS, Caplan JB, et al. Gamma oscillations correlate with working memory load in humans. *Cereb Cortex*. 2003;13:1369–1374.
- Cho RY, Konecky RO, Carter CS. Impairments in frontal cortical gamma synchrony and cognitive control in schizophrenia. *Proc Natl Acad Sci U S A*. 2006;103:19878–19883.
- Minzenberg MJ, Firl AJ, Yoon JH, Gomes GC, Reinking C, Carter CS. Gamma oscillatory power is impaired during cognitive control independent of medication status in first-episode schizophrenia. *Neuropsychopharmacology*. 2010;35:2590–2599.
- Whittington MA, Traub RD, Faulkner HJ, Jefferys JG, Chettiar K. Morphine disrupts long-range synchrony of gamma oscillations in hippocampal slices. *Proc Natl Acad Sci* USA. 1998;95:5807–5811.
- 23. Gulyás AI, Szabó GG, Ulbert I, et al. Parvalbumin-containing fast-spiking basket cells generate the field potential oscillations induced by cholinergic receptor activation in the hippocampus. *J Neurosci.* 2010;30:15134–15145.
- 24. Lewis DA, Curley AA, Glausier JR, Volk DW. Cortical parvalbumin interneurons and cognitive dysfunction in schizophrenia. *Trends Neurosci.* 2012;35:57–67.
- Hashimoto T, Volk DW, Eggan SM, et al. Gene expression deficits in a subclass of GABA neurons in the prefrontal cortex of subjects with schizophrenia. *J Neurosci*. 2003;23:6315–6326.
- Mellios N, Huang HS, Baker SP, Galdzicka M, Ginns E, Akbarian S. Molecular determinants of dysregulated GABAergic gene expression in the prefrontal cortex of subjects with schizophrenia. *Biol Psychiatry*. 2009;65:1006–1014.
- 27. Fung SJ, Webster MJ, Sivagnanasundaram S, Duncan C, Elashoff M, Weickert CS. Expression of interneuron markers in the dorsolateral prefrontal cortex of the developing human and in schizophrenia. *Am J Psychiatry*. 2010;167:1479–1488.
- Volk DW, Matsubara T, Li S, et al. Deficits in transcriptional regulators of cortical parvalbumin neurons in schizophrenia. *Am J Psychiatry*. 2012;169:1082–1091.
- 29. Curley AA, Lewis DA. Cortical basket cell dysfunction in schizophrenia. *J Physiol*. 2012;590:715–724.
- Glausier JR, Fish KN, Lewis DA. Altered parvalbumin basket cell inputs in the dorsolateral prefrontal cortex of schizophrenia subjects. *Mol Psychiatry*. 2014;19:30–36.
- Rudy B, McBain CJ. Kv3 channels: voltage-gated K+ channels designed for high-frequency repetitive firing. *Trends Neurosci*. 2001;24:517–526.
- 32. Georgiev D, Gonzalez-Burgos G, Kikuchi M et al. Selective expression of KCNS3 potassium channel alpha-subunit in parvalbumin-ontaining GABA neurons in the human prefrontal cortex. *PLoS One* 2012;7:e43904
- 33. Georgiev D, Arion D, Enwright JF, et al. Lower gene expression for KCNS3 potassium channel subunit in parvalbumincontaining neurons in the prefrontal cortex in schizophrenia. *Am J Psychiatry*. 2014;171:62–71.

- 34. Yanagi M, Joho RH, Southcott SA, Shukla AA, Ghose S, Tamminga CA. Kv3.1-containing K(+) channels are reduced in untreated schizophrenia and normalized with antipsychotic drugs. *Mol Psychiatry*. 2014;19:573–579.
- Xu Q, Cobos I, De La Cruz E, Rubenstein JL, Anderson SA. Origins of cortical interneuron subtypes. J Neurosci. 2004;24:2612–2622.
- Butt SJ, Fuccillo M, Nery S, et al. The temporal and spatial origins of cortical interneurons predict their physiological subtype. *Neuron*. 2005;48:591–604.
- Cobos I, Long JE, Thwin MT, Rubenstein JL. Cellular patterns of transcription factor expression in developing cortical interneurons. *Cereb Cortex*. 2006;16(suppl 1):i82–i88.
- Fertuzinhos S, Krsnik Z, Kawasawa YI, et al. Selective depletion of molecularly defined cortical interneurons in human holoprosencephaly with severe striatal hypoplasia. *Cereb Cortex*. 2009;19:2196–2207.
- 39. Zecevic N, Hu F, Jakovcevski I. Interneurons in the developing human neocortex. *Dev Neurobiol*. 2011;71:18–33.
- 40. Hansen DV, Lui JH, Flandin P, et al. Non-epithelial stem cells and cortical interneuron production in the human ganglionic eminences. *Nat Neurosci.* 2013;16:1576–1587.
- Ma T, Wang C, Wang L, et al. Subcortical origins of human and monkey neocortical interneurons. *Nat Neurosci.* 2013;16:1588–1597.
- 42. Letinic K, Zoncu R, Rakic P. Origin of GABAergic neurons in the human neocortex. *Nature*. 2002;417:645–649.
- 43. Zecevic N, Chen Y, Filipovic R. Contributions of cortical subventricular zone to the development of the human cerebral cortex. *J Comp Neurol*. 2005;491:109–122.
- 44. Jakovcevski I, Mayer N, Zecevic N. Multiple origins of human neocortical interneurons are supported by distinct expression of transcription factors. *Cereb Cortex*. 2011;21:1771–1782.
- 45. Morris HM, Hashimoto T, Lewis DA. Alterations in somatostatin mRNA expression in the dorsolateral prefrontal cortex of subjects with schizophrenia or schizoaffective disorder. *Cereb Cortex*. 2008;18:1575–1587.
- Woo TU, Miller JL, Lewis DA. Schizophrenia and the parvalbumin-containing class of cortical local circuit neurons. *Am J Psychiatry*. 1997;154:1013–1015.
- Volk DW, Lewis DA. Prenatal ontogeny as a susceptibility period for cortical GABA neuron disturbances in schizophrenia. *Neuroscience*. 2013;248C:154–164.
- Yang Y, Fung SJ, Rothwell A, Tianmei S, Weickert CS. Increased interstitial white matter neuron density in the dorsolateral prefrontal cortex of people with schizophrenia. *Biol Psychiatry*. 2011;69:63–70.
- Akbarian S, Bunney WE Jr, Potkin SG, et al. Altered distribution of nicotinamide-adenine dinucleotide phosphatediaphorase cells in frontal lobe of schizophrenics implies disturbances of cortical development. *Arch Gen Psychiatry*. 1993;50:169–177.
- Anderson SA, Volk DW, Lewis DA. Increased density of microtubule associated protein 2-immunoreactive neurons in the prefrontal white matter of schizophrenic subjects. *Schizophr Res.* 1996;19:111–119.
- Kirkpatrick B, Messias NC, Conley RR, Roberts RC. Interstitial cells of the white matter in the dorsolateral prefrontal cortex in deficit and nondeficit schizophrenia. *J Nerv Ment Dis.* 2003;191:563–567.
- 52. Eastwood SL, Harrison PJ. Interstitial white matter neuron density in the dorsolateral prefrontal cortex and parahippocampal gyrus in schizophrenia. *Schizophr Res.* 2005;79:181–188.

- Batista-Brito R, Rossignol E, Hjerling-Leffler J, et al. The cellintrinsic requirement of Sox6 for cortical interneuron development. *Neuron.* 2009;63:466–481.
- Azim E, Jabaudon D, Fame RM, Macklis JD. SOX6 controls dorsal progenitor identity and interneuron diversity during neocortical development. *Nat Neurosci.* 2009;12:1238–1247.
- 55. Sussel L, Marin O, Kimura S, Rubenstein JL. Loss of Nkx2.1 homeobox gene function results in a ventral to dorsal molecular respecification within the basal telencephalon: evidence for a transformation of the pallidum into the striatum. *Development*. 1999;126:3359–3370.
- Xu Q, Tam M, Anderson SA. Fate mapping Nkx2.1lineage cells in the mouse telencephalon. J Comp Neurol. 2008;506:16–29.
- Du T, Xu Q, Ocbina PJ, Anderson SA. NKX2.1 specifies cortical interneuron fate by activating Lhx6. *Development*. 2008;135:1559–1567.
- Wang Y, Dye CA, Sohal V, et al. Dlx5 and Dlx6 regulate the development of parvalbumin-expressing cortical interneurons. *J Neurosci.* 2010;30:5334–5345.
- 59. van den Berghe V, Stappers E, Vandesande B, et al. Directed migration of cortical interneurons depends on the cell-autonomous action of Sip1. *Neuron*. 2013;77:70–82.
- McKinsey GL, Lindtner S, Trzcinski B, et al. Dlx1&2dependent expression of Zfhx1b (Sip1, Zeb2) regulates the fate switch between cortical and striatal interneurons. *Neuron*. 2013;77:83–98.
- Wang Y, Li G, Stanco A, et al. CXCR4 and CXCR7 have distinct functions in regulating interneuron migration. *Neuron*. 2011;69:61–76.
- Sánchez-Alcañiz JA, Haege S, Mueller W, et al. Cxcr7 controls neuronal migration by regulating chemokine responsiveness. *Neuron*. 2011;69:77–90.
- Meechan DW, Tucker ES, Maynard TM, LaMantia AS. Cxcr4 regulation of interneuron migration is disrupted in 22q11.2 deletion syndrome. *Proc Natl Acad Sci U S A*. 2012;109:18601–18606.
- Steinecke A, Gampe C, Valkova C, Kaether C, Bolz J. Disrupted-in-Schizophrenia 1 (DISC1) is necessary for the correct migration of cortical interneurons. *J Neurosci.* 2012;32:738–745.
- 65. Lee FH, Zai CC, Cordes SP, Roder JC, Wong AH. Abnormal interneuron development in disrupted-in-schizophrenia-1 L100P mutant mice. *Mol Brain*. 2013;6:20.
- Flames N, Long JE, Garratt AN, et al. Short- and long-range attraction of cortical GABAergic interneurons by neuregulin-1. *Neuron*. 2004;44:251–261.
- Fazzari P, Paternain AV, Valiente M, et al. Control of cortical GABA circuitry development by Nrg1 and ErbB4 signalling. *Nature*. 2010;464:1376–1380.
- Ting AK, Chen Y, Wen L, et al. Neuregulin 1 promotes excitatory synapse development and function in GABAergic interneurons. *J Neurosci.* 2011;31:15–25.

- Lucas EK, Markwardt SJ, Gupta S, et al. Parvalbumin deficiency and GABAergic dysfunction in mice lacking PGClalpha. J Neurosci. 2010;30:7227–7235.
- Anderson SA, Eisenstat DD, Shi L, Rubenstein JL. Interneuron migration from basal forebrain to neocortex: dependence on Dlx genes. *Science*. 1997;278:474–476.
- Cobos I, Calcagnotto ME, Vilaythong AJ, et al. Mice lacking Dlx1 show subtype-specific loss of interneurons, reduced inhibition and epilepsy. *Nat Neurosci.* 2005;8:1059–1068.
- Denaxa M, Kalaitzidou M, Garefalaki A, et al. Maturationpromoting activity of SATB1 in MGE-derived cortical interneurons. *Cell Rep.* 2012;2:1351–1362.
- Joshi D, Fung SJ, Rothwell A, Weickert CS. Higher gammaaminobutyric acid neuron density in the white matter of orbital frontal cortex in schizophrenia. *Biol Psychiatry*. 2012;72:725–733.
- 74. Silberberg G, Darvasi A, Pinkas-Kramarski R, Navon R. The involvement of ErbB4 with schizophrenia: association and expression studies. *Am J Med Genet B Neuropsychiatr Genet*. 2006;141B:142–148.
- Law AJ, Kleinman JE, Weinberger DR, Weickert CS. Diseaseassociated intronic variants in the ErbB4 gene are related to altered ErbB4 splice-variant expression in the brain in schizophrenia. *Hum Mol Genet*. 2007;16:129–141.
- Joshi D, Fullerton JM, Weickert CS. Elevated ErbB4 mRNA is related to interneuron deficit in prefrontal cortex in schizophrenia. J Psychiatr Res. 2014;53:125–132.
- Junttila TT, Sundvall M, Määttä JA, Elenius K. Erbb4 and its isoforms: selective regulation of growth factor responses by naturally occurring receptor variants. *Trends Cardiovasc Med.* 2000;10:304–310.
- Millar JK, Wilson-Annan JC, Anderson S, et al. Disruption of two novel genes by a translocation co-segregating with schizophrenia. *Hum Mol Genet*. 2000;9:1415–1423.
- Ripke S, O'Dushlaine C, Chambert K, et al.; Multicenter Genetic Studies of Schizophrenia Consortium; Psychosis Endophenotypes International Consortium; Wellcome Trust Case Control Consortium 2. Genome-wide association analysis identifies 13 new risk loci for schizophrenia. *Nat Genet*. 2013;45:1150–1159.
- Hoftman GD, Volk DW, Bazmi HH et al. Altered cortical expression of GABA-related genes in schizophrenia: Illness progression vs developmental disturbance [published online ahead of print December 22, 2013]. Schizophr Bull doi:10.1093/schbul/sbt178.
- Liodis P, Denaxa M, Grigoriou M, Akufo-Addo C, Yanagawa Y, Pachnis V. Lhx6 activity is required for the normal migration and specification of cortical interneuron subtypes. J Neurosci. 2007;27:3078–3089.
- 82. Zhao Y, Flandin P, Long JE, Cuesta MD, Westphal H, Rubenstein JL. Distinct molecular pathways for development of telencephalic interneuron subtypes revealed through analysis of Lhx6 mutants. *J Comp Neurol*. 2008;510:79–99.
- Neves G, Shah MM, Liodis P, et al. The LIM homeodomain protein Lhx6 regulates maturation of interneurons and network excitability in the mammalian cortex. *Cereb Cortex*. 2013;23:1811–1823.