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## When Deletions Gain Functions: Commandeering Epigenetic Mechanisms

Justin Chen<sup>1</sup> and William A. Weiss<sup>1,\*</sup>

<sup>1</sup>Departments of Neurology, Pediatrics, Neurosurgery, and the Helen Diller Family Comprehensive Cancer Center, University of California, San Francisco, CA 94158 USA

### Abstract

Recurrent chromosomal deletions in cancer are typically thought to harbor tumor suppressors. In a recent publication in *Nature*, Northcott and colleagues identify a novel region of structural variation in medulloblastoma that leads to oncogenic activation of *GFI1B* and *GFI1* by repositioning these genes next to super-enhancers.

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Genomic instability is one of the enabling hallmarks of cancer and can lead to extensive chromosomal abnormalities (Hanahan and Weinberg, 2011). Sites of recurrent genomic aberrations have long been thought to harbor genes important for tumor development; indeed, oncogenes such as *MYC* and *ERBB2* are found in amplifications, the *BCR-ABL* fusion gene results from a chromosomal translocation, and tumor suppressors such as *RB1*, *PTEN*, and *TP53* are frequently lost in deleted regions.

Medulloblastoma is the most common malignant pediatric brain tumor, with large-scale genomic and transcriptomic analyses identifying four distinct molecular subgroups (Taylor et al., 2012). Northcott, Lee, Zichner, and coworkers (Northcott et al., 2014) recently analyzed whole-genome sequencing of primary group 3 and 4 medulloblastoma samples for somatic structural variants (SVs). Rather than limiting their search to recurrent amplifications or deletions, they analyzed all chromosomal breakpoints and identified a novel region of interest spanning over 400 kb on chromosome 9q34.13. A single gene at this locus, growth factor independence 1B (*GFI1B*), was found to be overexpressed concomitant with proximal SV. Despite the variety of observed SVs ranging from deletions (Figure 1A), tandem duplication, and/or inversions, these SVs resulted in repositioning *GFI1B* next to super-enhancers (Figure 1). Additionally, after observing mutually exclusive expression patterns within group 3 tumors between *GFI1B* and its paralog, growth factor independence 1 (*GFI1*), the authors identified that *GFI1* was also subject to SV, similarly varied between interchromosomal translocations and tandem duplications (Figure 1B). These also led

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\*Correspondence: waweiss@gmail.com.

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to similar displacement of GFI1 to regions adjacent to other enhancers or super-enhancers (Figure 1C).

Enhancers are short stretches of genomic DNA that serve to bind activators and function in cis to drive transcription of nearby genes (Ong and Corces, 2011). Recently, super-enhancers have been identified as exceptionally large euchromatic regions that serve as a concentrated site of activator and transcription factor binding and stimulate higher transcriptional activity than typical enhancers. While enhancers are highly prevalent throughout the genome, super-enhancers are scattered sparsely at a few hundred sites, residing at key cell-identity genes where they define cell types by driving specific expression patterns (Hnisz et al., 2013; Whyte et al., 2013). Super-enhancers are also thought to be dynamic, forming at essential oncogenes during tumorigenesis where they remain exquisitely sensitive to bromodomain inhibition (Lovén et al., 2013).

This idea of dynamic super-enhancers works in concert with data showing epigenetic heterogeneity and plasticity play integral roles in the acquisition of drug resistance in cancer (Sharma et al., 2010). With a low somatic mutation rate (0.52 per Mb) and frequent alteration of chromatin modifiers across all subgroups (Jones et al., 2012), medulloblastoma may be the perfect candidate to observe this phenomenon. Northcott et al., however, describe “enhancer hijacking” as a mechanism in which genomic instability leads to the utilization of existing epigenetic structure to drive oncogene expression. Thus, as epigenetic plasticity represents a complementary approach to the acquisition of somatic mutations in the pathogenesis of cancer, this study leads to intriguing questions about the state of the epigenome in medulloblastoma. Is the utilization of enhancer hijacking (rather than dynamic generation of a new super-enhancer) simply due to the enrichment of somatic copy number aberrations in these subgroups (Northcott et al., 2012)? Are there fundamental differences between medulloblastoma and other cancers that lead to a relatively static epigenome in medulloblastoma, or are super-enhancers not so readily plastic or dynamic?

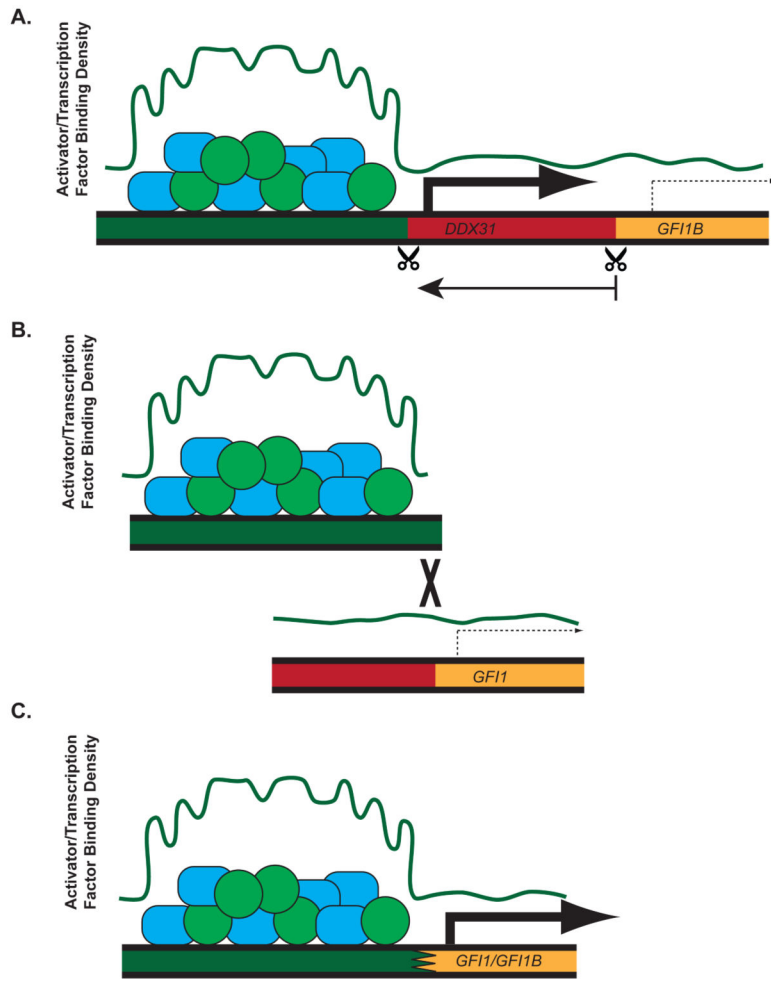
Another question that remains to be explored is whether there is an underlying function within the GFI1 gene family that promotes enhancer hijacking. GFI1 and GFI1B are highly homologous transcriptional repressors that are expressed in the hematopoietic compartment and are known proto-oncogenes in leukemia and lymphoma. Since both GFI1 and GFI1B are subjected to transcriptional autoregulation and are able to repress each other's expression, does the underlying SV represent the only mechanism of escape from a complex net of feedback loops? Northcott et al. show that GFI1/GFI1B cooperate with MYC to drive medulloblastoma in an orthotopic xenograft mouse model, despite the fact that neither alone was able to promote tumorigenesis in this model. Curiously, GFI1 activation, but not GFI1B, correlated with MYC expression in medulloblastoma, whereas both cooperated with MYC in this preclinical in-vivo model. Whether and how they cooperate to make a permissive environment for such SV remains to be determined.

Each of the four medulloblastoma subgroups differ in age distribution, gender, and outcome. Two of these subgroups (groups 1 and 2) are driven predominantly by a single prominent signaling pathway (WNT or SHH), while group 3 and group 4 tumors show more complex genetics and signaling. Group 3 tumors generally have the poorest prognosis, and by

identifying these super-enhancer activated oncogenes, the authors identify a possible therapeutic avenue using bromodomain inhibitors. Furthermore, this study was successful in identifying oncogenes at regions containing, in part, common deletions. Despite knowing for 15 years that translocations in Burkitt's lymphoma can lead to activation of MYC by juxtaposing it next to the IGH enhancer, the current paradigm still focuses on the assumption that the type of SV defines the function of the gene of interest: gains representing oncogenic loci and losses representing tumor suppressor loci. While large efforts have been spent to identify tumor suppressors in commonly deleted regions, Northcott, Lee, Zichner, and colleagues ask us to rethink these strategies for tumors where SVs have clearly delineated and recurrent breakpoints, especially if studies to date have not yielded strong candidates.

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**Figure 1. GFI1 and GFI1B hijack enhancers via structural variation**  
In medulloblastoma, GFI1B (as shown in A) and GFI1 (as shown in B) normally reside in heterochromatic regions with little to no expression. Structural variation including deletions (A) or interchromosomal translocations (B) can lead to juxtaposition of these genes next to enhancers or super-enhancers (as shown in C) which drive oncogenic expression due to their increased concentrations of bound activators and transcription factors (plotted in green and represented by circles and rectangles).