

Novel Integron-Mediated Fosfomycin Resistance Gene *fosK*

Hiromitsu Kitanaka,^a Jun-ichi Wachino,^a Wanchun Jin,^a Satoru Yokoyama,^a Masa-aki Sasano,^b Mitsuhiro Hori,^b Keiko Yamada,^a Kouji Kimura,^a Yoshichika Arakawa^a

Department of Bacteriology, Nagoya University Graduate School of Medicine, Nagoya, Japan^a; Clinical Microbiology Laboratory, Okazaki City Hospital, Okazaki, Japan^b

Fosfomycin (FOM) is an antibiotic produced by *Streptomyces fradiae* (1) and was approved for clinical use in Japan in 1980. FOM blocks MurA, which mediates bacterial peptidoglycan biosynthesis in its early step, showing a broad-spectrum antimicrobial activity against both Gram-positive and Gram-negative bacteria. FOM penetrates into bacterial cells via sugar transporters, such as GlpT and UhpT, located at the cytoplasmic membrane, and spontaneous FOM-resistant mutants appear due to a reduction or lack of these transporters. Moreover, several enzymes, such as FosA, FosB, FosC, FosD, FomX, FomA, and FomB, have been reported, and FosA was first characterized as a glutathione S-transferase of FOM (2) (Fig. 1). After our first report about FosA3 and FosC2 in 2010 (3), FosA3-producing *Escherichia coli* isolates were recovered from humans, livestock, and/or pets (4–7), and the *fosA3* gene has already transferred to *Klebsiella pneu-*

moniae (6) by a probable IS26 composite transposon carrying *fosA3*.

Acinetobacter soli HK001 was isolated from a blood culture of an infected human, and it showed very high resistance to FOM (MIC, >8,000 µg/ml) according to the agar dilution method recommended by the CLSI (8) in the presence of glucose-6-phosphate (G6P) (25 µg/ml), which induces UhpT. Four amplicons of class 1 integrons were found by PCR using 2 primers, 5'CS-

Published ahead of print 19 May 2014

Address correspondence to Yoshichika Arakawa, yarakawa@med.nagoya-u.ac.jp.

Copyright © 2014, American Society for Microbiology. All Rights Reserved.

doi:10.1128/AAC.03131-14

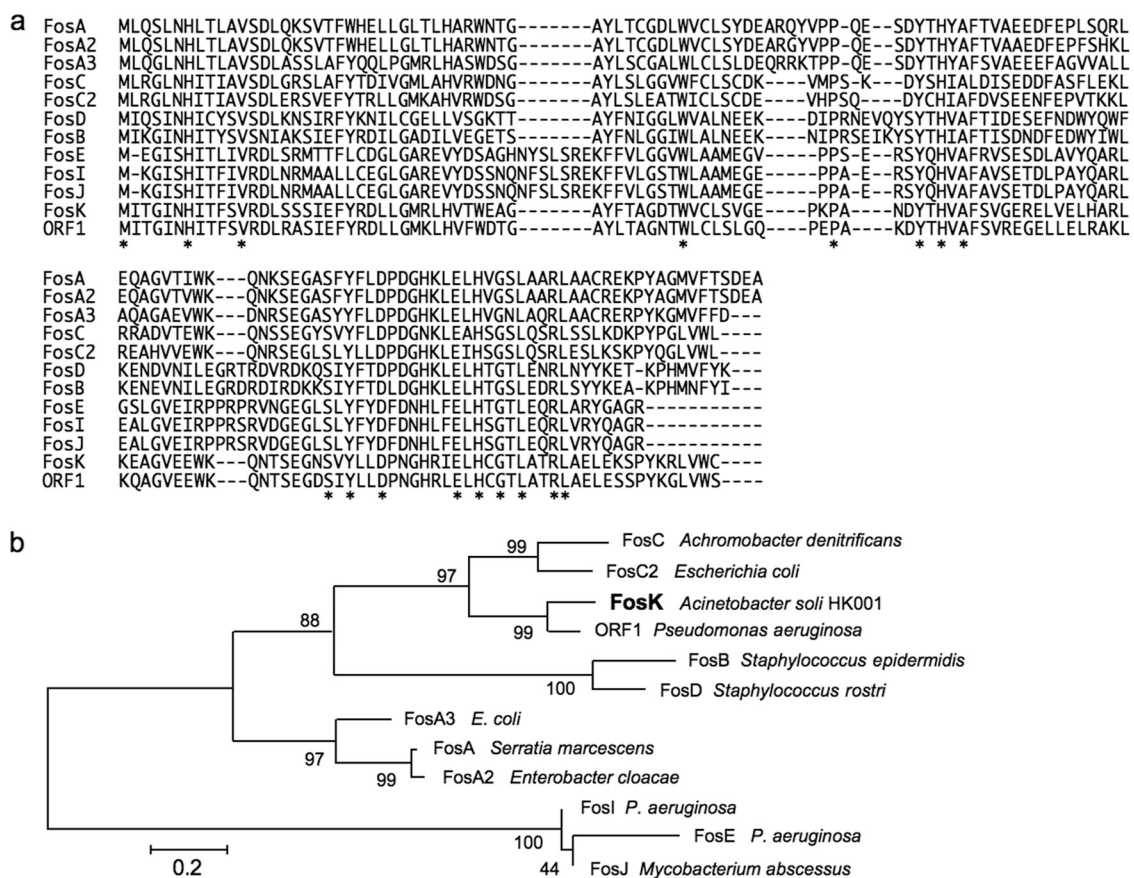


FIG 1 (a) Predicted amino acid sequences of FosK and other fosfomycin-modifying enzymes. *, amino acid residue conserved among the 12 fosfomycin resistance determinants. (b) Phylogenetic relationships among the 12 glutathione S-transferases, including probable ones calculated by MEGA 5 (<http://www.megasoftware.net/>). GenBank or Protein Data Bank accession numbers are indicated for the following proteins: FosA (AAA98399), FosA2 (ACC85616), FosA3 (AB522970), FosB (CAA38136), FosC (AAZ14834), FosC2 (AB522969), FosD (AHB87392), FosE (BAO48025), FosI (BAO47999), FosJ (YP_006316014), FosK (AB917040), and ORF1 (AAP50248).

TABLE 1 MICs of fosfomycin for *A. soli* HK001 and *E. coli* DH10B transformed with the *fosK* gene

| Strain | FOM MIC ($\mu\text{g/ml}$) ^a |
|---|---|
| <i>Acinetobacter soli</i> HK001 | >8,192 |
| <i>E. coli</i> DH10B | 1 |
| <i>E. coli</i> DH10B(pBCSK+) | 2 |
| <i>E. coli</i> DH10B(pBCSK+:: <i>fosK</i>) | >2,048 |
| <i>E. coli</i> ATCC 25922 | 2 |

^a FOM, fosfomycin. MICs were measured by the agar dilution method recommended by the CLSI.

Class1-integron (5'-GGCATCCAAGCAGCAAG-3') and 3'-CS-Class1-integron (5'-AAGCAGACTTGACCTGA-3'). An amplicon of 1.2 kb was excised and purified. Its nucleotide sequence was directly determined and revealed an *aacA4* gene and a new gene cassette located between *intI1* and the 3'-CS (conserved sequence). The new cassette encoded a protein with significant similarity to other Fos proteins (Fig. 1) and was named FosK. The deduced amino acid sequence of FosK showed 81% identity in its amino acid sequence to open reading frame 1 (ORF1) of *Pseudomonas aeruginosa* (9). Moreover, 52%, 52%, 51%, 50%, 48%, and 47% amino acid identities were observed between FosK and FosC2, FosD, FosA3, FosA, FosA2, and FosC, respectively, suggesting their close phylogenetic relationship (Fig. 1). The *fosK* gene was again amplified by PCR using total bacterial DNA and a high-fidelity DNA polymerase, PrimeSTAR HS (TaKaRa Bio Inc., Ohtsu, Japan), together with primers F2-BamHI (5'-CGGGATCCCCGACATGGTTCAAACACGCCAGGC-3') and R2-HindIII (5'-TACCCAAGCTTGGGTTTTGGGGCGGACTTGTA GC-3'). The amplicon was ligated with pBCSK+ and cleaved by BamHI and HindIII, and *E. coli* DH10B was transformed with the recombinant plasmids. Then FOM-resistant transformants were selected. After nucleotide sequencing of the insert on both strands, a clone carrying no mutation in the *fosK* gene was finally chosen. The FOM MIC for the transformant harboring intact *fosK* was augmented to >2,048 $\mu\text{g/ml}$ from 1 $\mu\text{g/ml}$ for the recipient with G6P (25 $\mu\text{g/ml}$) (Table 1).

FOM was recently considered to be a potent agent for treatment of infections caused by multidrug-resistant bacteria, such as extended-spectrum β -lactamase (ESBL)-producing *E. coli* and *K. pneumoniae* (10). FOM has also been approved for veterinary use in various countries (11). The *fosK* gene, together with *aacA4*, is mediated by a class 1 integron, and thus this genetic element will be further transmitted into various *Enterobacteriaceae*. Since *fosK* confers on bacteria a very high level of resistance to fosfomycin, we should diligently monitor the prevalence and trend of *fosK* as well as of *fosA3* in both human and animals going forward.

Nucleotide sequence accession number. The *fosK* gene has been assigned accession number AB917040.

ACKNOWLEDGMENT

This study was supported by grant no. H24-Shinko-Ippan-010.

REFERENCES

- Rogers TO, Birnbaum J. 1974. Biosynthesis of fosfomycin by *Streptomyces fradiae*. Antimicrob. Agents Chemother. 5:121–132. <http://dx.doi.org/10.1128/AAC.5.2.121>.
- Bernat BA, Laughlin LT, Armstrong RN. 1997. Fosfomycin resistance protein (FosA) is a manganese metallothione transferase related to glyoxalase I and the extradiol dioxygenases. Biochemistry 36:3050–3055. <http://dx.doi.org/10.1021/bi963172a>.
- Wachino J, Yamane K, Suzuki S, Kimura K, Arakawa Y. 2010. Prevalence of fosfomycin resistance among CTX-M-producing *Escherichia coli* clinical isolates in Japan and identification of novel plasmid-mediated fosfomycin-modifying enzymes. Antimicrob. Agents Chemother. 54:3061–3064. <http://dx.doi.org/10.1128/AAC.01834-09>.
- Hou J, Yang X, Zeng Z, Lv L, Yang T, Lin D, Liu JH. 2013. Detection of the plasmid-encoded fosfomycin resistance gene *fosA3* in *Escherichia coli* of food-animal origin. J. Antimicrob. Chemother. 68:766–770. <http://dx.doi.org/10.1093/jac/dks465>.
- Pan YS, Yuan L, Zong ZY, Liu JH, Wang LF, Hu GZ. 2014. A multidrug-resistance region containing *bla*_{CTX-M-65}, *fosA3* and *rmtB* on conjugative IncFII plasmids in *Escherichia coli* ST117 isolates from chicken. J. Med. Microbiol. 63:485–488. <http://dx.doi.org/10.1099/jmm.0.070664-0>.
- Lee SY, Park YJ, Yu JK, Jung S, Kim Y, Jeong SH, Arakawa Y. 2012. Prevalence of acquired fosfomycin resistance among extended-spectrum β -lactamase-producing *Escherichia coli* and *Klebsiella pneumoniae* clinical isolates in Korea and IS26-composite transposon surrounding *fosA3*. J. Antimicrob. Chemother. 67:2843–2847. <http://dx.doi.org/10.1093/jac/dks319>.
- Sato N, Kawamura K, Nakane K, Wachino J, Arakawa Y. 2013. First detection of fosfomycin resistance gene *fosA3* in CTX-M-producing *Escherichia coli* isolates from healthy individuals in Japan. Microb. Drug Resist. 19:477–482. <http://dx.doi.org/10.1089/mdr.2013.0061>.
- Clinical and Laboratory Standards Institute. 2012. Methods for dilution antimicrobial susceptibility testing of bacteria that grow aerobically. Approved standard, 8th ed. Document M7-A9. Clinical and Laboratory Standards Institute, Wayne, PA.
- Yatsuyanagi J, Saito S, Konno T, Harata S, Suzuki N, Amano K. 2005. The ORF1 gene located on the class-1-integron-associated gene cassette actually represents a novel fosfomycin resistance determinant. Antimicrob. Agents Chemother. 49:2573. <http://dx.doi.org/10.1128/AAC.49.6.2573.2005>.
- Neuner EA, Sekeres J, Hall GS, van Duin D. 2012. Experience with fosfomycin for treatment of urinary tract infections due to multidrug-resistant organisms. Antimicrob. Agents Chemother. 56:5744–5748. <http://dx.doi.org/10.1128/AAC.00402-12>.
- Soraci AL, Perez DS, Martinez G, Dieguez S, Tapia MO, Amanto F, Harkes R, Romano O. 2011. Disodium-fosfomycin pharmacokinetics and bioavailability in post weaning piglets. Res. Vet. Sci. 90:498–502. <http://dx.doi.org/10.1016/j.rvsc.2010.07.011>.