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### **Systematic Review: The Association and Impact of Financial Conflicts of Interest in Basic Science Research**

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#### **Abstract**

**Background—** No prior study has evaluated financial relationships of investigators with pharmaceutical manufacturers for basic science. An example of the importance and impact of such relationships is in the evaluation of erythropoietin receptors'(EpoRs) effects on cancer cell lines, since studies have reported increased mortality when cancer patients receive erythropoiesis stimulating agents (ESAs).

**Purpose—**To assess the disclosed association that exist between pharmaceutical industry support and EpoRs effects on solid cancer cell lines.

**Data Sources—**MEDLINE and EMBASE (1988- July 2008) and two EpoR conferences sponsored by the National Institutes of Health.

**Study Selection—**All publications investigating EpoRs that met inclusion criteria were identified and included.

**Data Extraction—**Data were extracted on detection of EpoRs, presence of erythropoietininduced signaling events, presence of erythropoietin-induced changes in cellular function, nature of qualitative conclusions, and sources of funding for all 74 studies.

**Data Synthesis—In** comparison to studies of academic investigators with no disclosed funding support from ESA manufacturers (n=64), the studies from academic investigators with funding support from ESA manufacturers ( $n= 7$ ) and the laboratories directed by investigators employed by ESA manufacturers (n=3) were both less likely to identify: EpoR presence on solid tumor cells; erythropoietin-induced signaling events; erythropoietin-induced changes in cellular function; and less likely to conclude that their research had identified potentially harmful effects of erythropoietin on cancer cells. Additionally, presentations from industry-based investigator teams at NIH conferences were less likely to report EpoRs on cancer cell lines, downstream effects of erythropoietin, and cell proliferation and migration effects following EpoR administration.

**Conclusion—**Financial conflicts of interest impact the outcomes and presentation of basic science research data as well as publications.

#### **INTRODUCTION**

United States Senator Charles Grassley recently initiated conflict of interest probes of clinical investigators at Harvard Medical School, Columbia University, and Emory University.(1) These investigations focus on payments from pharmaceutical manufacturers to key opinion leaders. Grassley and Senator Charles Schumer introduced the Physician Payments Sunshine Act that requires transparency in relationships with pharmaceutical manufacturers. These actions stem from long-standing observations that clinical investigators who have financial support from pharmaceutical manufacturers are less likely than other clinical researchers to criticize the safety, effectiveness, or cost-effectiveness of drugs and devices distributed by sponsoring manufacturers (2–4) and more likely to endorse novel and less proven treatments.(5) A related and previously unanswered question concerns financial conflicts of interest for basic scientists.

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Prior studies provide some insight on academic-industrial relationships in the life sciences. In one review of relationships between academic institutions and industry, Blumenthal reported that these relationships facilitated technology transfer.(6) However, there is less scrutiny of academic conflicts of interest involving non-clinical research than clinical research. Moreover, partly due to less federal level regulatory requirements for disclosure of conflicts and relatively minimal monitoring or oversight of research conduct, academic institutions have embraced industrial sponsorship of basic science research and placed relatively less emphasis on disclosure, management, and monitoring of these relationships. In a survey of life science researchers, Blumenthal et al. reported that faculty members with industrial research support were at least as productive academically as those without such support and were more productive commercially, although they were more likely to restrict their communication with colleagues.(7) Bekelman et al. reported that financial relationships among industry, scientific investigators, and academic institutions were widespread, and that industry sponsorship was associated with pro-industry conclusions.(8) While Bekelman et al. primarily focused on clinical investigators with conflicts of interest, the study also noted that both clinical and basic science investigators with conflicts reported an unwillingness to publish certain findings, restrictions on publication efforts, or shifting research emphasis based on commercial considerations.(8) The use of ESAs in cancer patients represents a prime example of a hotly pursued basic science question that may ultimately have a large impact on the pharmaceutical companies as well as on clinical practice.

Recently, basic science studies have raised concern over the safety of administering erythropoiesis stimulating agents (ESAs) to cancer patients. Preclinical studies demonstrated erythropoietin receptors (EpoRs) on cell lines derived from solid tumors.(9, 10) Concern that EpoR signaling on tumor cells might contribute to tumor progression remained theoretical until 2003 when clinical trials identified increased tumor progression and mortality among ESA-treated head and neck cancer and breast cancer patients.(11, 12) Furthermore, a follow-up to the head and neck trial demonstrated that increased mortality with ESA administration was noted only in patients whose tumors expressed EpoRs, while patients lacking the receptor experienced decreased mortality with ESA treatment.(13) Safety concerns grew when six additional trials demonstrated increased mortality risks and four studies found increased tumor progression among ESA-treated patients.(14) These findings support the recent FDA labeling change that states ESAs are not indicated for cancer patients undergoing chemotherapy regimens with curative intent.(15, 16)

Basic science findings for EpoRs provide an opportunity to evaluate whether financial conflicts of interest might affect basic science research. Interpreting findings from investigations of EpoRs conducted by basic science investigators who do or do not have financial support from pharmaceutical manufacturers is central to evaluating this concern. Herein, we reviewed findings reported by academic investigators with or without pharmaceutical support and investigators employed by ESA manufacturers on the effects of ESAs on tumor cells, xenografts, and human cancers.

#### **METHODS**

We searched the MEDLINE and EMBASE (1988- July 2008) databases to identify research articles investigating EpoRs in solid tumors (Keywords: (Erythropoietin (Epo), erythropoietin receptor (EpoR)) AND (cancer, tumor, malignancy, carcinoma)). All publications investigating EpoRs were identified. Editorials, reviews, investigations concerning hematologic malignancies, articles that did not investigate presence of EpoRs, and studies that used artificially constructed EpoR complexes were excluded. Moreover, studies concerning tumor types for which ESA-stimulated signaling events were not investigated elsewhere in the literature were also excluded. Seventy-four articles were included in this study (Figure 1).

Data were abstracted on detection of EpoRs, presence of erythropoietin-induced signaling events, presence of erythropoietin-induced changes in cellular function, nature of qualitative conclusions, and sources of funding. Information on EpoR detection included: detection method, types of antibodies, negative and positive controls, band size, and location in the cell. For signaling events, information was obtained on positive and negative controls, and assessments of erythropoietin-induced changes in signaling proteins Janus-activatedkinase-2 (JAK2), signal transducer and activator of transcription-5 (STAT5), the Bcl-2 (Bcell lymphoma 2), family of proteins (including pro-apoptotic Bax, anti-apoptotic Bcl-2 and Bcl-xL), phosphoinositide 3-kinase (PI3K), Akt, Nuclear factor-kappa B (NF-κB), mitogenactivated protein kinase (MAPK), extracellular signal*-*regulated kinase-1 (ERK1), jun Nterminal kinase (JNK), Jagged-1, Notch-1, vascular endothelial growth factorm (VEGF), platelet derived growth factor-b (PDGF). For cellular function studies, information was obtained on erythropoietin-induced changes in cell survival, invasiveness, migration, apoptosis, hypoxia-inducible factor, EpoR expression, angiogenesis, tumor oxygenation, chemosensitization, proliferation, and growth in tumor samples, cancer cell lines or xenografted cancers. Conflicts of interest were abstracted through conflict statements and funding disclosures included in the publications. For publications that did not include funding acknowledgments as a matter of course, journal editors and first and last authors were queried regarding funding sources.

#### **Analyses**

All seventy-four articles were divided according to funding source. The first group, investigator teams with no conflicts of interest, included investigators with no disclosed funding support from ESA manufacturers. The second group, industry based investigator teams with conflicts of interest, included studies where greater than 75% of co-authors were employees of ESA manufacturers. The third group, academic based investigator teams with conflicts of interest, included studies from academic investigators who received funding support from ESA manufacturers (operationally defined as research grant support or academic investigators who received honoraria from ESA manufacturers, were consultants to ESA manufacturers, or who collaborated in the research study with active engagement in the research by investigators employed by ESA manufacturers).

Qualitative conclusions were scored as: unfavorable, neutral, favorable, or not present. Studies that explicitly reported potential adverse effects of ESAs in cancer patients were

scored as "unfavorable." Studies that explicitly emphasized potential therapeutic value of ESAs in cancer patients were scored as "favorable." Studies that presented both favorable and unfavorable information and did not validate one over the other were scored as "neutral." Studies which made no qualitative conclusions were scored as "not present"

The Chi-square statistical test was used to compare differences in study results according to funding source.

#### **RESULTS**

Our literature review of EpoR in solid tumors identified 64 investigations from academic investigators who indicated no financial support from ESA manufacturers, three from industry-based investigators employed by ESA manufacturers, and seven from academicbased investigators who received support from ESA manufacturers (including three academically-associated studies that included co-investigators employed by ESA manufacturers). Prior to 2003 (the year when the first two trials identifying increased rates of tumor progression and death with ESA administration to cancer patients were published), one-third of studies on EpoRs in the solid tumor setting from academic-based investigators who indicated no financial conflicts of interest had been published, two-thirds of such studies from academic-based investigators with financial conflicts of interest had been published, and none of the studies were from industry-based investigators had been reported.

#### **Published basic science findings**

**Group I: Reports from academic investigative teams who did not receive financial support from ESA manufacturers—**Sixty-four studies from investigators without financial support from ESA manufacturers predominantly reported results in support of potentially harmful effects of erythropoietin on solid tumors (Table 1). Fifty-seven studies investigated that EpoR was present on cancer cells. All of these studies positively identified EpoRs (Table 1). To detect EpoR protein, investigators employed immunohistochemistry, immunocytochemistry, and radiolabeled ligands. Reverse transcriptase PCR and real-time PCR were used to detect EpoR DNA. Most EpoR bands were approximately 66 kiloDaltons in weight (range from 59 to 110 kiloDaltons). Twenty-two studies evaluated EpoR in malignant and paired benign tissues; 20 identified EpoRs in malignant cells and reported either no EpoRs on paired benign tissue, or EpoR levels lower than in the malignant counterpart.(9, 17–35) Two studies identified EpoR in both samples: one did not report on comparative levels(36) and the second reported similar levels in brain tissue.(37) Of 34 studies evaluating signaling events in response to the administration of erythropoietin, 31 identified these events (Table 1). Signaling proteins investigated included JAK2 pathways (three of three studies identified activated JAK2(29, 30, 38) and six of seven demonstrated inhibition of erythropoietin-induced downstream effects with concurrent administration of JAK2 inhibitor, (30, 38–43) PI3-kinase (five studies, all demonstrated inhibition of erythropoietin-induced downstream effects with concurrent administration of PI3K inhibitor)(10, 37, 43–45), and MAP-kinase (one of two studies identified activated MAPK (46)' (24) and one study demonstrated inhibition of erythropoietin-induced downstream effects with concurrent administration of MAPK inhibitor(39)). Forty studies evaluated cell

function and regulation changes following erythropoietin administration, including mitogenic effects (13 studies, 10 identified effects) (9, 26, 37, 43, 47–52) (32, 46, 53), invasiveness and migration (five studies, all identified effects)(29, 30, 38, 39, 42), angiogenesis (five studies, two identified effects)(51, 52, 54, 55), growth (18 studies, 9 identified effects) (37, 38, 43, 47, 49–52, 56) (10, 32, 46, 48, 53, 57–60) and cytoprotective effects (19 studies, 12 identified effects). (10, 18, 24, 26, 27, 29, 41, 44, 45, 61, 62) (40, 46, 50, 54, 55, 57, 63) Other investigations evaluated the effects of erythropoiesis stimulating agents on chemotherapy effectiveness, including tumor protection (14 studies- 8 reported protection (9, 18, 24, 26, 27, 29, 41, 62), six reported no effects (10, 24, 46, 50, 57, 62), and two reported increased chemotherapy efficacy (40, 54), some studies reported different results within their manuscript and thus were counted twice) and effects of erythropoietin on apoptosis (four studies, all reported reduced apoptosis).(44, 61, 62, 64) These studies utilized primary tumor samples and established tumor cell lines including cancers of the breast (17 studies), cervix (9 studies), neuroblastoma (7 studies), kidney (6 studies), ovary (7 studies), and prostate (7 studies) (Table 1).

**Group II: Reports from industry-based investigative teams—**Three studies from investigators employed by ESA manufacturers reported findings concerning presence of EpoR in tumor cells. One investigated erythropoietin-induced growth in tumor cells.(65–67) Elliot et al. investigated the specificity of commercially available antibodies used in EpoR studies. These antibodies identified proteins with weights larger than the expected molecular weight of EpoR. To validate this discrepancy, the authors performed protein sequence analysis on the proteins detected by the antibodies and found an abundance of non-EpoR proteins, including heat shock protein 70 (a protein commonly found in more aggressive tumors).(65) The M20 antibody was an exception to this non-specificity, and accurately detected EpoR in immunoblotting, but not in immunnohistochemical, studies. The authors conclude that caution should be used when interpreting results of studies using common antibodies. In a second study, Sinclair et al.(66) reported gene amplification of EpoR loci in solid tumor samples occurred at a frequency similar to that of non-oncogenes; EpoR transcript levels in tumors and tumor cell lines were low in comparison to bone marrow and equivalent to or lower than those reported for normal tissues of tumor origin and when EpoR mRNA was detected, it was not on the cell surface.(66) Investigators employed by another ESA manufacturer evaluated effects of erythropoietin administration alone or in combination with anticancer therapy on breast cancer cell lines.(67) Immunoblotting, flow cytometry, and immunohistochemistry evaluations identified cytosolic EpoR expression. Tumor growth assessments in breast cancer xenograft models found no evidence of migration, proliferation, or activation of mitogen-activated protein kinase and AKT following erythropoietin treatment and that treatment with erythropoietin alone or with paclitaxel resulted in equivalent tumor burdens compared with vehicle-based controls.

**Group III: Reports from investigative teams comprised of academics where one or more co-authors disclosed funding support from ESA manufacturers—** Seven studies were reported by academic investigators who had funding support from ESA manufacturers.(68–73) One study identified mRNA EpoR expression in neuroblastoma cell lines; erythropoietin administration did not induce tumor proliferation of the cell lines.(70)

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Using tissue microarrays, Brown et al. reported that preabsorption of C20 antibodies with synthetic heat shock protein peptides resulted in suppression of cytoplasmic staining in formalin fixed non-small cell lung cancer tissues.(69) Also, Western blots identified three components of which one was lost after C20 pre-absorption and one (the putative EpoR component) was retained. Laugsch et al. investigated by real-time RT-PCR, immunofluorescence microscopy, Western blotting, and cell growth analysis whether several human cancer cell lines possessed functional EpoRs.(71) These researchers detected EpoR mRNA in all cell lines, although neither hypoxia nor erythropoietin treatment altered EpoR mRNA expression. Four commercial antibodies cross-reacted with several proteins. Depending on the antibody used, EpoR was localized to the plasma membrane, the cytoplasm, or the nucleus. Experiments with small interfering RNA showed that EpoR protein was not expressed by tumor cells except for UT7/erythropoietin leukemia cells, which served as an EpoR positive control line, and by cells transfected with the human EpoR gene. The authors reported that erythropoietin increased signaling and proliferation in the UT7 control line and did not result in activation of signaling proteins or increased cell proliferation in tumor cells.(71) Westphal et al. reported EpoR and protein expression in various tumor cell lines using RT-PCR, Western blots, and immocytochemistry, although erythropoietin treatment did not result in increased proliferation of EpoR-positive tumor cell lines.(72) In aggregate, these findings were similar to findings from ESA manufacturers, raising concern over the significance of studies from investigators without pharmaceutical support who reported on EpoR expression in malignancies.

Three studies identified off-target effects that could improve tumor responsiveness to chemotherapy and radiation therapy- increased tumor oxygenation, cis-platinum sensitization, or angiogenesis inhibition following erythropoietin exposure.(68, 73, 74) Silver et al. demonstrated cis-platin sensitizing effects of erythropoietin on human ovarian cancer xenografts in mice.(73) Blackwell et al. found that hypoxic measurements were lower in non-anemic mice that received erythropoietin after tumor implantation versus mice with tumors that received erythropoietin before tumor implantation or that received placebo. (68) These findings suggested that erythropoietin administration might improve tumor oxygenation independent of hemoglobin effects. Hale et al. found that in SK-OV-3 ovarian cancer cell lines, erythropoietin-treatment decreased hypoxia-induced HIF-1-alpha protein levels and VEGF transcription, with no effect on cell growth and in MCF-7 breast cancer cell lines, erythropoietin inhibited HIF-1-alpha signaling.(74) These findings suggested that erythropoietin may have anti-angiogenic properties.

**Groups I, II, and III: A Comparison—**The findings differed according to whether the studies were reported by academic investigators who did not receive financial support from ESA manufacturers, academic investigators who received financial support from ESA manufacturers, and investigators employed by ESA manufacturers with respect to several fundamental issues: EpoR presence on solid tumor cells (100%, 60%, and 67%, respectively,  $(p<0.04)$ ; erythropoietin-induced signaling events (94%, 0%, and 0%, respectively), p=0.001); and erythropoietin-induced changes in cellular function (57%, 0%, and 0%, respectively; p=0.007) (Figure 2). Qualitative statements about clinical implications were included in 42 reports from investigators who did not have funding from ESA

manufacturers, five reports from investigators who had received funding support from ESA manufacturers, and two reports from investigators who directed laboratories supported by ESA manufacturers. Among these studies, statements concluding that the investigations had identified potentially harmful effects of erythropoietin on cancer cells were included in 57% of reports from academic investigators who did not have funding from ESA manufacturers, 0% of the reports from academic investigators who had received financial support from ESA manufacturers, and 0% of the reports from investigators who directed laboratories supported by ESA manufacturers (p=0.008). In contrast, statements indicating that the findings identified potentially beneficial anti-tumor effects of ESAs were included in 14% of reports from academic investigators who did not have funding from ESA manufacturers, 0% of reports from investigators employed by ESA manufacturers, and 60% of reports from academic investigators who had received financial support from ESA manufacturers  $(p<0.04)$ .

**NIH Conferences on EpoRs—**Workshops on EpoRs have been convened by the National Cancer Institute in 2007 and the National Institutes of Diabetes and Diseases of the Kidney (NIDDK) in 2008.(75, 76) Participants included 12 investigators from academic institutions who reported EpoR findings and four investigators employed by ESA manufacturers.(Figure 3). Academic investigators were co-authors for 27 of the 64 published manuscripts on EpoRs from academic investigators who did not have financial support from ESA manufacturers and one of the seven published manuscripts on EpoRs reported by academic investigators who had received financial support from ESA manufacturers. Investigators employed by ESA manufacturers were authors for all three published manuscripts on EpoRs conducted at the basic science laboratories of ESA manufacturers. Presentations reported on cell lines established from lung, head and neck, melanoma, ovarian, brain, cervical, and breast cancers, xenograft animal models, or in one case, findings from clinical specimens. These investigations identified EpoR mRNA and protein on various tumor cells and demonstrated that erythropoietin/EpoR signaling axis activation in cancer cells involved PI3K-Akt, JAK-STAT and NF-kB. Tumor cells could utilize the erythropoietin/EpoR-signaling axis in autocrine or paracrine fashion and recruitment of exogenous erythropoietin increased proliferation, anti-apoptosis, invasion, chemotherapy resistance, and angiogenesis. In contrast, investigators employed by ESA manufacturers reported that their investigations of tumor cell lines had identified a protein band corresponding to heat shock protein-70, and not EpoRs and that their investigations of animal models did not identify erythropoietin-induced angiogenesis or tumor promotion. Overall, in comparison to presentations from investigators employed by ESA manufacturers, presentations from academic investigators were more likely to report EpoRs on cancer cell lines (100% versus 25%; p<0.01), downstream effects of erythropoietin (88% versus 0%; p<0.04), and cell proliferation and migration effects following EpoR administration (91% versus 0%; p<0.01) (Table 2).

#### **Discussion**

The difference in findings for EpoR investigations reported by investigators with and without financial support from ESA manufacturers provides empirical evidence that

conflicts of interest exist in the basic science setting that impact outcomes. This observation is unexpected as it runs counter to the popular belief that the scientific process is reproducible and protects against variable outcomes of laboratory studies. In interpreting our findings, several factors should be considered.

Three areas of disagreement for studies of EpoR have been debated. At the level of protein detection, investigators employed by one ESA manufacturer noted that the C20 antibody, commonly used in academic investigators' studies, identified bands representing proteins between 64 and 68 kiloDaltons in size. Academic investigators reported that degradation of these EpoRs could result in identification of a 59 kiloDalton EpoR band (the size of the band noted in reports from Elliot et al.). Academic investigators noted that one report published by the ESA manufacturers of western blot analyses with a less commonly evaluated antibody (M-20) confirmed the presence of EpoRs in breast, cervical and brain tumor cells. Also, several studies reported by academic investigators reported functional EpoRs on cancer cells with methods that did not involve antibodies. A second difference centers around the level of changes in cellular function of erythropoietin exposed cancer cells that should be considered biologically significant. Investigators employed by ESA manufacturers operationally defined a 2-fold difference in cellular function between tumor cells and control cells to be biologically significant. Conversely, academic investigators did not establish a threshold level, positing that reproducible, statistically significant differences could have clinical implications. A third issue concerned the choice of positive and negative controls. Investigators employed by ESA manufacturers considered the carrier protein included in proprietary formulations of ESAs to be the appropriate negative control. Academic investigators noted that while they did not use specific proprietary compounds as controls, adequate negative controls, such as the addition of soluble EpoR to the medium, had been employed in their studies.

Recommendations for adjudication of the methodological differences vary. An investigator employed by an ESA manufacturer suggested that academic and ESA manufacturers should develop a consensus statement on the types of positive and negative controls and characteristics of reagents used in studies of erythropoietin and EpoRs.(77) Academic researchers countered that the peer-review system is the usual venue for quality control, and adopting a single set of study conditions might result in laboratory costs too expensive for academic laboratories. ESA manufacturers offered to provide \$5 million to the NIH Foundation to support a Request for Application for basic science studies of EpoRs in tumor cells. The Board of Scientific Advisors of the NIH raised ethical concerns over accepting these funds.

While studies have identified conflict of interest concerns with clinical studies evaluating efficacy, safety, and cost-effectiveness of pharmaceuticals, conflicts in basic science studies may be more worrisome. Basic science manuscripts from investigators who have received financial support from pharmaceutical manufacturers undergo corporate review to maximally protect disclosure of proprietary information. Inclusion of proprietary information might be dis-allowed by the industry sponsor. Also, unlike in clinical research, in basic science research, some manuscripts are more commonly authored solely by investigators employed by pharmaceutical manufacturers.(5) Moreover, the pharmaceutical

manufacturers, as sponsors, often decide which investigations may be submitted for peerreview and often influence the decision on the targeted journal for publication.

We conclude that financial conflict of interest considerations impact the interpretation of basic science studies in presentation and publications. As many universities have established basic science research partnerships with pharmaceutical manufacturers, transparency in these collaborations is paramount to allow for continued free exchange of scientific knowledge.

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#### **Figure 2.**

Percentage of published studies identifying presence of EpoR, erythropoietin-induced changes in signaling, erythropoietin-induced changes in cellular function and qualitative conclusions by conflict of interest type.

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#### **Figure 3.**

Percentage of research presentations at National Cancer Institute and the National Institute of Diabetes and Digestive and Kidney Diseases national meetings identifying presence of EpoR, erythropoietin-induced changes in signaling, and erythropoietin-induced changes in cellular function by conflict of interest type.

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## **Table 1**

List of studies included in comprehensive review. Tumor type investigated, findings regarding presence of erythropoietin receptors, ESA induced changes List of studies included in comprehensive review. Tumor type investigated, findings regarding presence of erythropoietin receptors, ESA induced changes in signaling, ESA-induced changes in cellular function, qualitative conclusions and descriptions of conflicts of interest are summarized. in signaling, ESA-induced changes in cellular function, qualitative conclusions and descriptions of conflicts of interest are summarized.





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Bold: At least ONE author presented at NIDDK or NIH Meeting Sept 2008 White: Authors classified as academics with no conflicts of interest (n= 64) Light Orange: Authors classified as academics with conflicts of interest (n= 7) Dark Orange: Authors classified as employed by ESA manufacturers (n= 3)

White: Authors classified as academics with no conflicts of interest (n= 64) Bold: At least ONE author presented at NIDDK or NIH Meeting Sept 2008

Light Orange: Authors classified as academics with conflicts of interest (n= 7) Dark Orange: Authors classified as employed by ESA manufacturers (n=3)

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# **Table 2**

signaling, erythropoietin-induced -induced changes in cellular function, qualitative conclusions and descriptions of conflicts of interest are summarized. signaling, erythropoietin-induced -induced changes in cellular function, qualitative conclusions and descriptions of conflicts of interest are summarized. Summary of presentations given at NCI and NIDDK<sup>\*</sup> meetings regarding presence of erythropoietin receptors, erythropoietin-induced changes in Summary of presentations given at NCI and NIDDK*\** meetings regarding presence of erythropoietin receptors, erythropoietin-induced changes in



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NIDDK refers to the Workshop on Erythropoietin Receptor (Epo-R) Expression and Function in Non-Hematopoietic Tissues hosted by the National Institute of Diabetes and Digestive and Kidney Diseases in Bethesda, MD (Sep 8–9, 2008). NCI refers to Erythropoietic Stimulating Agents and Tumor Growth Workshop hosted by National Cancer Institute in Rockville, MD (Dec 18–19, 2007).

<sup>\*\*</sup> Based on the 74 articles on EpoRs included in Table 1A. Based on the 74 articles on EpoRs included in Table 1A.