

Competition between recently potentiated synaptic inputs reveals a winner-take-all phase of synaptic tagging and capture

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Canonical models suggest that mechanisms of long-term memory consist of a synapse-specific, protein synthesis-independent induction phase (changes in synaptic weights/temporary tagging of such synapses) and, within adjacent dendritic compartments, a protein synthesis-dependent distribution phase that may accompany or immediately precede induction and whose protein products enable consolidation through synaptic capture. We now report that this distribution phase is competitive in a “winner-take-all” fashion when synapses potentiated at induction compete with each other for plasticity-related proteins. This finding highlights the importance of synaptic competition in creating stable long-lasting memory in neural networks without disruption.

Activity-dependent increases and decreases in synaptic efficacy, such as the physiological phenomena of long-term potentiation (LTP) and long-term depression, are considered to be prominent cellular mechanisms mediating learning and memory. Long-lasting persistence of synaptic strength requires protein synthesis that is achieved through transcription and translation (late LTP; L-LTP), whereas short-lasting changes do not involve protein synthesis (early LTP; E-LTP) (1). Based on this dependency on protein synthesis, a sequential model of memory has long been proposed involving the transition from E-LTP to L-LTP, and it remains an important framework for memory consolidation (2, 3). However, the newer concept of synaptic tagging and capture (STC) has changed our understanding of the necessity for a sequential framework. Instead, although memory encoding and tagging occur in real time in association with the event or stimulus to be remembered, the persistence of this trace depends also upon the capture of plasticity-related proteins (PRPs) whose synthesis can be triggered before, during, or immediately after memory encoding. The synthesis of PRPs, now thought to occur in relatively clustered dendritic domains (4, 5), creates the possibility for both synergistic (6) and competitive interactions between potential memories during the subsequent distribution phase. Synergistic effects are well-studied. However, although synaptic competition has been considered previously (7), the temporal dynamics of such competition are not well-understood. We now show that when the temporal persistence of synaptic potentiation is enabled on one pathway by virtue of the availability of PRPs from another earlier or later event, potentiation of a third pathway around the same time may trigger sufficient competition to prevent persistent potentiation on all pathways. Varying the timing of the potentiation of this third pathway enabled one or more pathways to persist whereas others do not. Thus, when the number of competing potential memories increases and the availability of PRPs is limited, a “winner-take-all” scenario appears to prevail whereby some traces persist in a stable manner whereas others do not. The use of multiple pathways models the likely situation in real life when the encoding of multiple memories can create synergistic interactions or competition for resources.

Results and Discussion

Initial interface chamber electrophysiological experiments, described in *SI Methods* (Fig. S1A and B), indicated that we could induce stable LTP lasting 12 h after strong tetanization (STET) and that a weakly tetanized pathway (WTET), induced within 30 min, would then also induce L-LTP (Fig. 1A). One of the prerequisites for consistently measuring L-LTP with multiple inputs that lasts more than 10 h is the slightly high extracellular K^+ concentration in the artificial cerebrospinal fluid that we used, similar to that of the original STC experiments reported by Frey and Morris (6). The stage was thus set for the key experiment, namely to induce LTP with STET, which should ordinarily result in stable L-LTP and weakly tetanize two pathways, rather than a single additional one, 30 and 45 min after STET later (Fig. 1B, blue and green filled circles). This arrangement should increase synaptic competition. The surprising result was that potentiation on all pathways declined, including the STET pathway, with potentiation decaying to baseline at 480 min. The same result was obtained when intervals of 30 and 60 min after STET were used (Fig. 1C). However, a different outcome prevailed with 30 and 75 min (Fig. 1D), with only the last tetanized pathway decaying to baseline. Synaptic competition appears, under the condition of our experiments, to occur over a period of about 60 min, but stabilization then occurs in a winner-take-all manner at the cost of memory persistence for a pathway tetanized later on (Fig. 1D, green filled circles).

The next step was to characterize some of the determinants of synaptic competition. Competition is likely to be determined by the availability of PRPs and the number of tags that can capture

Significance

Our findings will refine the existing concept of a simple, linear phase model of memory in terms of temporal constraints on associative memory formation. We provide evidence for an in-between situation in which a “winner-take-all” process shapes the process of stable long-term memory storage. We show that when the temporal persistence of plasticity is enabled on one pathway by virtue of the availability of proteins from another earlier or later event, potentiation of a further third pathway around the same time may trigger sufficient competition to prevent persistent potentiation on all pathways. This mimics the daily situation of hippocampal neurons, which have to cope with the learning of new events under conditions of competition that can threaten stabilization.

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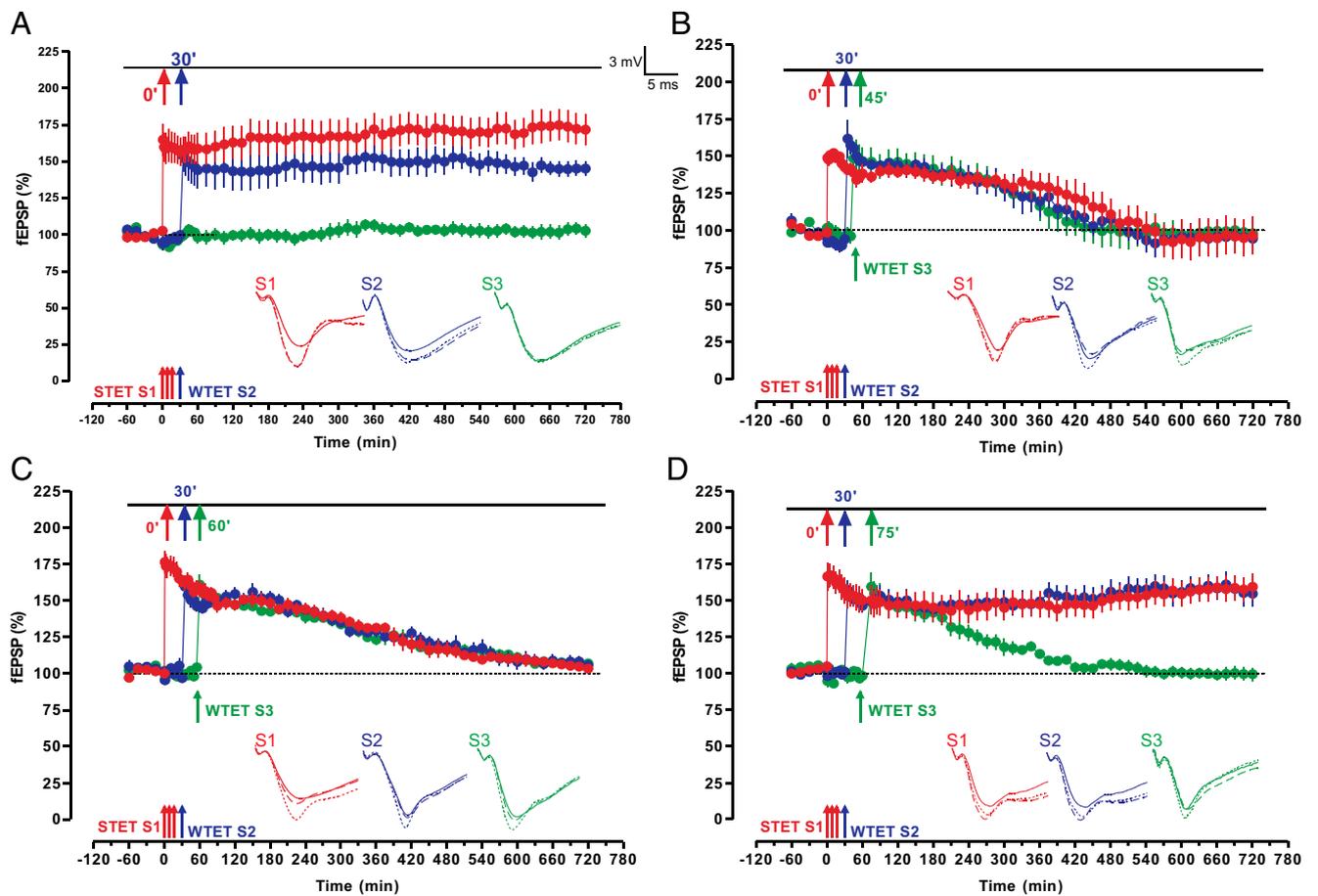


Fig. 1. Synaptic competition and its duration. (A) Time duration of late LTP (12 h) in S1 (red filled circles), followed by early LTP in S2 by WTET (blue filled circles); here, S2 expressed STC. In addition, stable control baseline potentials up to 12 h were recorded from S3 ($n = 7$) (for more details, see *SI Methods*). (B) Induction of L-LTP at 0 min in S1 (red filled circles) followed by E-LTP in S2 at 30 min (blue filled circles) and another E-LTP in S3 at 45 min (green filled circles). Here, if two weak plasticity forms (E-LTP in S2 and E-LTP in S3) compete for PRPs from a strong plasticity pathway (L-LTP), all plasticity forms decay to baseline after 4 h. This result provides strong evidence for synaptic competition in a physiological situation ($n = 7$). (C) The experimental design was the same as in B but E-LTP in S3 (green filled circles) was induced at 60 min ($n = 7$). (D) The experimental design was the same as in B and C but E-LTP in S3 was induced at 75 min (green filled circles). Interestingly, no synaptic competition was observed between S1 and S2, but S3 was not transformed to L-LTP ($n = 7$). (Insets) Averages of analog traces recorded from synaptic inputs S1 and S2, 30 min before (continuous line), 90 min after (dotted line), and 12 h (hatched line) after the induction of the corresponding plasticity. Error bars indicate \pm SEM; fEPSP, field excitatory post synaptic potentials; n , number of experiments. (Calibration bar for all analog sweeps, 3 mV/5 ms.)

them. Given that depotentiation (DP) 5 min after the induction of E-LTP can effectively reset synaptic tags (8, 9), we applied STET on distal synaptic input S1 and then, 30 min later, WTET on S2, followed 5 min later by DP stimuli (1 Hz, 250 pulses; Fig. 2A, blue filled circles). This depotentiated E-LTP should have erased a competing synaptic tag (10). Accordingly, when WTET was then applied to S3 45 min after STET, persistent potentiation was observed on both S1 and S3 (Fig. 2A)—a strikingly different result from that shown in Fig. 1B. However, if the depotentiated pathway S2 was repotentiated at 60 min after STET on S1, reinstating competition, the outcome was potentiation declining across all pathways (Fig. 2B), analogous to that shown in Fig. 1B. Moreover, reactivation of S2 at 75 min did not affect the stable potentiation seen in S1 and S3 (Fig. 2C, red and green), whereas pathway S2 itself, being repotentiated at a time point outside the synergy/competition window of the PRP distribution phase, fails to stabilize. It was intriguing to us to check whether the PRPs provided after setting the competing partners could also be involved in competition. To do this, we provided STET in S3 at 60 or 75 min after the initial WTET in S1 (Fig. 3A and B; weak-before-strong protocol). Interestingly, the outcome was similar to that in Fig. 1B and D. These intriguing

observations provide evidence that competitive tag setting by coincidental weak activation of synapses within the temporal vicinity of strongly activated synapses triggers a graded decay of long-lasting L-LTP into a shorter form of LTP despite strong tetanic activation of that pathway. This graded and slow decline of L-LTP is a feature of the competitive maintenance mechanism. Thus, under the conditions of our experiments, the molecular boundary seems to be rather sharp and lies between 60 and 75 min poststimulus (for a statistical analyses see *Tables S1–S5*).

These observations reveal a winner-take-all component of the “synergy/competition phase” of protein synthesis-dependent LTP in which the potentiated synaptic population is destined either for persistent potentiation or for a time-dependent decay. Synaptic competition in the late stage of LTP has been reported in the presence of protein synthesis inhibitors to reduce the availability of PRPs, and the suggestion has been made that such competition could provide a means for selective information storage when multiple inputs converge (7). Our results go beyond this by identifying that competition occurs even in the early phase of synaptic memory consolidation, with the outcome that either a pathway bifurcates into a stable potentiated state or it decays to baseline. This finding is in agreement

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