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Effects of Growth Hormone Treatment in Adults with Prader-Willi Syndrome

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Abstract

Objective—Since limited data exist on adults with Prader-Willi syndrome (PWS) and growth hormone (GH) treatment, we report our experience on the effects of treatment for one year on body composition, physical activity, strength and energy expenditure, diet, general chemistry and endocrine data with quality of life measures.

Design—We studied 11 adults with PWS (6F:5M; average age = 32 yrs) over a 2 year period with GH treatment during the first year only. Electrolytes, IGF-I, glucose, thyroid, insulin, lipids, body composition, physical activity and strength, diet, energy expenditure and quality of life data were collected and analyzed statistically using linear modeling at baseline, at 12 months following GH therapy and at 24 months after treatment cessation for 12 months.

Results—Total lean muscle mass was significantly increased (p < 0.05) during GH treatment along with moderate-vigorous physical activity and plasma IGF-I and HDL levels, but returned to near baseline after treatment. Percent body fat decreased during the 12 months of GH treatment but increased after treatment.

Conclusions—Previously reported beneficial effects of GH treatment in children with PWS were found in our adults regarding body composition, physical activity and plasma HDL and IGF-I levels. Several beneficial effects diminished to near baseline after cessation of GH treatment for 12 months supporting the continuation of treatment in PWS into adulthood and possibly adults not previously treated during childhood.

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Keywords

Growth hormone treatment; Prader-Will syndrome adults; body composition; energy expenditure; physical activity

Introduction

Prader-Willi syndrome (PWS) is a complex genomic imprinting disorder affecting multiple body systems including hypotonia, poor suck and feeding problems, hypogonadism/ hypogenitalism, growth hormone (GH) deficiency with short stature and small hands/feet, hyperphagia and obesity, learning/behavior problems and a characteristic face.^{1–5} PWS is due to absence of paternally expressed genes in the 15q11-q13 chromosome region from a deletion, maternal disomy 15 or an imprinting defect.^{2, 3, 6}

Children with PWS grow at different rates than other children but most are born with a normal weight and length. Failure to thrive occurs but they gradually gain strength and muscle mass before onset of hyperphagia and obesity in early childhood. The fat mass increases more rapidly than muscle mass or stature in children with PWS without growth hormone treatment with more than twice the amount of body fat gained compared with others and often measuring about 50% of their total body weight.⁷ Weight control with exercise and dietary restrictions are key management issues.^{2,8,9}

Due to growth hormone deficiency reported in PWS, the Food and Drug Administration approved treatment in children genetically confirmed with PWS in 2000 without prior laboratory determination of growth hormone deficiency. GH treatment is now commonly prescribed in infancy and childhood to increase stature in PWS but also increases muscle and decreases fat thereby lowering risk factors for diabetes and cardiopulmonary problems. Increased alertness, physical activity and self-esteem in older children are potential effects of GH treatment^{10–15} which should improve the quality of life applicable in adulthood. In addition, Lindgren and Lindberg¹⁶ documented that GH treatment in children with PWS appear to normalize to an adult height with improvement in body composition, an important outcome for individuals with this disorder. PWS children appear to be highly sensitive to GH suggesting that growth factors (i.e., insulin-like growth factor I or IGF-I) should be monitored closely (e.g., every three months) and maintained within the normal range for age. There is a paucity of data on the use of GH in adults with PWS not previously treated and related to energy expenditure, physical activity, body composition, endocrine and quality of life measures.

The main objective of our study was to examine how GH treatment for one year would impact on body composition, quality of life, physical activity and energy balance in adults with PWS using novel approaches, and if loss of a positive effect, if present, during therapy disappears after cessation of treatment for 12 months. Anticipated outcomes:

1. Increase IGF-I level thereby lessoning risk factors for co-morbidity; improve quality of life measures; decrease fat, increase lean body mass and bone density determined by DEXA.

2. Increase energy expenditure as determined by whole-room calorimeter measures, diet records, physical activity monitors, strength and endurance measures.

Materials and Methods

Subjects

We screened 12 genetically confirmed Caucasian adults with PWS and enrolled 11 adults (6F:5M; average age \pm SD = 32.3y \pm 11.1y; age range = 23y to 50y) for a non-controlled two year study on the effects of GH treatment for 12 months followed by an observation period of 12 months without treatment. The inclusion criteria for selection were as follows: Sixteen to 60 years of age; male or female with diagnosis of PWS confirmed by genetic testing; low plasma IGF-I level (e.g.,<25%ile) at baseline; and ability to provide informed consent or availability of a suitable legally authorized representative. The exclusion criteria were as follows: Pregnancy; previous GH treatment; untreated or uncontrolled endocrine disease (e.g., diabetes or thyroid disorder); history of severe scoliosis; uncontrolled high blood pressure, heart disease or history of stroke; weight greater than 95%ile based on gender and established age specific growth charts for PWS¹⁷) and severe breathing problems by history or sleep apnea found by sleep center polysomnography as a diagnostic tool used in sleep medicine. The adults with PWS enrolled in the study showed evidence of hypogonadism (i.e., males with small, undescended testicles and females with absent or sparse history of menstruation) with low plasma estrogen or testosterone levels.

The subjects were recruited from a multi-disciplinary clinic specializing in PWS directed by one of the authors (MGB) following informed consent with the study protocol approved by the local institutional review board. Height and weight were determined at each visit. Height (measured to the nearest 0.1 cm using a Forgue electric stadiometer) and weight (measured to the nearest 0.1 kg using a calibrated electronic scale, height) were obtained by one of the authors (CS). Heights that were discordant from previous measurements were confirmed by repeat measurement. Variations in height were attributed to lack of cooperation by the subjects. Age, weight, height, body mass index (BMI kg/m²), percent body fat and energy expenditure data are shown in Table 1. Nine PWS adults had the typical 15q11-q13 deletion; one with maternal disomy 15 and one with an imprinting defect determined by genetics testing.^{4, 5} During this study, daily subcutaneous injections of growth hormone (e.g., Genentech Nutropin AQ; 0.0125 mg/kg/day) were administered which were comparable to dosages reported previously in PWS adults and based on the treatment of adult GH deficiencies.¹⁸ Each subject and care provider underwent training for GH administration, recording of dosage and monitoring for potential side effects (edema, headaches and signs of allergic reactions). Compliance for growth hormone use was determined by monitoring the amount of GH unused in the kit when returned at each scheduled clinic visit (e.g., monthly). At the time of the physical examination and measurements, blood was collected for laboratory studies before a new package of GH and administration supplies were given to the subject for continuation of treatment. Growth hormone dosages were adjusted in the first 3 months of treatment to achieve plasma IGF-I levels within the therapeutic reference range for at least 9 months during the first year. Each assessment was undertaken over a two year

period (at baseline, at 12 months of GH therapy and at 24 months or one year following treatment).

Methods

Fasting Blood Samples and Body Composition Measures—Fasting blood samples were obtained for general chemistry (electrolytes), plasma IGF-I, glucose, thyroid, insulin and lipid levels at screening, baseline, 1 month, 2 months, 3 months, 6 months, 9 months, 12 months and 24 months using established protocols in the Children's Mercy Hospital clinically approved certified laboratories for patient care. Due to hyperphagia and food seeking behavior as major components of PWS, the fasting status of each subject could not be confirmed for all measurements. Dual energy x-ray absorptiometry (DEXA – General Electric Prodigy, Atlanta, GA) was used to determine body composition (lean muscle and fat) and bone mineral density following established protocols.¹⁹ Provocative growth hormone stimulation testing was not performed prior to enrolling in the study. All subjects were healthy and not undergoing medical care for chronic illnesses related to their disorder.

Dietary Intake—All participants were asked to complete a 3-day food record with the assistance of their care provider in order to determine the average caloric intake. Because of the range of cognitive abilities in individuals with PWS and their food seeking behavior, obtaining accurate measurements of dietary intake would be difficult. Thus, to increase the accuracy of caloric intake, the 3-day food records were sent to the participant as well as the primary caregiver for completion. The primary caregiver was asked to assist each participant in recording all foods consumed during 3 days (2 week days and 1 weekend day). Primary caregivers were provided detailed instructions on completion of the 3-day food records and were asked to track all food consumption during the three day periods. To increase accuracy of the records, caregivers were asked to provide labels and/or packaging from all packaged foods. Upon completion, registered dietitians followed up with the participants to ensure the accuracy of the records and reviewed the records for completeness and nutrient calculations performed on the recorded data using the Nutrition Data System for Research Software 2010 (University of Minnesota, Minneapolis, MN).

Strength Index—Voluntary physical activity was measured by CSA accelerometers and strength by the one repetition of maximum method (1RM) with two exercises (bench and leg press) on all subjects following established protocols.²⁰ Assessments of muscular strength (assessed by 1RM) were obtained at baseline, at 12 and 24 months using bench press (Paramount Fitness Corporation, Los Angeles, CA) and leg press (Hammer Strength, Shiller Park, IL) according to the protocol described by Lemmer et al.²⁰ The participants initially completed a light 5 repetition warm-up period with minimal weight. After a brief rest for 60 seconds, a weight estimated to be less than the participant's 1RM was chosen and the participant was instructed to lift the weight through a full range of motion. If the lift was successful, the weight was increased then followed by a 60 second rest. They were then asked to lift the increased weight again and 1RMs obtained in less than 5 lifts for exercise. This procedure was repeated until a weight was reached that the participant could no longer lift. The maximum weight lifted through the full range of motion was recorded as their 1RM.

Indirect Calorimeter and Metabolic Measurements—A whole-room indirect calorimeter as described previously^{19,21} was used to continuously measure energy expenditure over an 8 hour period at baseline, and at the end of year one following GH treatment and at year two. Twenty-four hour energy expenditure was then extrapolated from the 8 hour measurement.²¹

The completed 3-day food records were used to develop standardized menus to mimic the participant's habitual diet (kcal and macronutrient composition) while in the whole-room calorimeter. Participants were asked to follow the menus for the 3 day period prior to their baseline metabolic measurement which were used in the whole-room calorimeter. These same menus were then provided to each participant prior to each subsequent visit and they were asked to follow these menus during the 3 days prior to testing.

Participants entered the whole-room calorimeter at 8:00am and remained in the room until 4:00pm. Upon entering the calorimeter, participants were asked to lie quietly, but not sleep, for the first 60 minutes of the whole-room stay in order to measure their resting metabolic rate (RMR). The participants were then allowed to watch TV, read or play games while in the room. They were not allowed to sleep while in the calorimeter. Research staff checked on the participants at least every 30 minutes to ensure that the participants were awake. Standardized meals scheduled to be consumed during the testing period were provided to the participant at the appropriate times.

Urine was collected during the calorimeter stay for the determination of total nitrogen concentration²² to assess protein oxidation.^{23,24} Using measured oxygen consumption and carbon dioxide production, respiratory quotient (RQ), energy expenditure and substrate oxidation were calculated using the equations of Jequier et al.²⁵

Physical Activity Measurements—Daily physical activity was measured with accelerometers [Actigraph GTM1 (Actigraph, Pensacola, FL)] at baseline and at year one and year two and mailed to the participants with specific instructions on wearing the device approximately 10 days prior to visit. Participants wore the accelerometer on a belt over their non-dominant hip for seven consecutive days prior to their visit except when sleeping, swimming or bathing. Upon arrival to the laboratory, the accelerometers were collected and data downloaded. Data were collected in one-minute epochs and a minimum of 12 hours required to be considered a valid monitored day. Intensity cut-points were based upon the National Health and Nutrition Examination Survey reported by Troiano et al.²⁶ with sedentary (99 counts/minute), light (100 to 2019 counts/minute), moderate (2020 to 5999 counts/minute) and vigorous (6000 counts/min) recorded on each subject.

Quality of Life Assessments—Two quality of life instruments were selected for administration to the PWS adults. The Quality of Life—Assessment of Growth Hormone Deficiency in Adults (QoL-AGHDA) questionnaire was administered initially to the PWS adults by one of the authors (BKS) by reading questions along with the possible choices to each participant. No additional explanations of the study questions were required for each subject to participate successfully in the assessments. The QoL-AGHDA is a 25-item disease-specific instrument used to assess patients' perceptions about their physical, social,

and mental health for adults with growth hormone deficiency.^{27,28} A total score was produced by summing across all items; a higher QoL-AGHDA score indicates a poorer quality of life. This instrument has been shown to be both reliable and valid with a high level of internal consistency. The short form (SF) – 36 was also used which is a validated self-administered questionnaire that assesses eight health dimensions, including physical functioning, role limitations related to physical functioning, general health perception, vitality, pain, social functioning, emotional well-being and role limitations due to emotional health.²⁹ Dimension scores were standardized between 0 and 100, with higher scores representing a better quality of life and previously used on individuals with cognitive impairment without comprising reliability and validity³⁰ and also utilized with GH-treated PWS adults.³¹ The SF-36 scale was also administered at baseline, at 12 months following growth hormone therapy and at 24 months after one-year cessation from growth hormone

Data Analysis

treatment.

Our primary endpoint was the longitudinal comparisons of body composition, blood chemistry and physical activity measures between different stages of growth hormone treatment over a 2-year period (baseline, at 12 months while on GH treatment and at 24 months after treatment). We used mixed modeling³² to handle non-independence of the data where the outcome measures were collected at multiple time points. The model effects and the error covariance components for the repeated measures were estimated using the restricted maximum likelihood (REML) procedure. In the mixed models, total lean gram, total percent body fat, average counts of moderate-vigorous bouts per day, average length of moderate-vigorous bouts per day, IGF-I, HDL and quality of life scores were contrasted between three measurement periods (baseline, 12 months, 24 months) adjusting for the participants' gender. The compound symmetry error structure provided a smaller AIC [(Akaikes information criteria) or model fit index used to select a model among a finite set of models] and BIC score [(Bayesian information criteria) or another model fit index used for model selection, which introduces a penalty term for the number of parameters in the model] than did the completely general (unstructured), first-order autoregressive and variance component error structures and thus chosen for the model system. All analyses were conducted using SAS 9.231 (SAS Institute, Cary, NC).

Results

The data were analyzed using mixed modeling to handle interdependency among observations and descriptive characteristics grouped by gender (Table 1). All PWS adults (45% were male) were Caucasian with an average age (\pm SD) of $32.3y\pm11.1y$. Figure 1 presents the adjusted means of each outcome measure at baseline, 12 months and 24 months. In the mixed model system, total lean gram, number of bouts of moderate-vigorous physical activity per day, the average plasma IGF-I levels, which were low or outside the reference range at baseline in our subjects and HDL levels were increased during the growth hormone treatment phase of the study (baseline to 12 months), but after treatment (12 to 24 months) the measures decreased back to baseline levels. The adverse events that might be related to growth hormone therapy included one episode of mild edema of the lower leg in a single

patient (50 year old male) during the first month of GH treatment which stabilized during the rest of the treatment period and impaired glucose tolerance in four subjects defined as one plasma glucose level over 100mg/dl. The average pretreatment plasma glucose level in these subjects was 82 ± 6 mg/dl before and 99 ± 22 mg/dl during GH treatment. In all instances, the elevated glucose level was sporadic. One individual had a plasma glucose level over 100mg/dl (i.e., 102mg/dl) before GH treatment and impaired glucose tolerance noted during treatment. A second individual had a plasma glucose level over 100mg/dl (i.e., 102mg/dl) before GH treatment and normal plasma glucose during treatment. All subjects with impaired glucose tolerance during the treatment phase had normal plasma glucose on follow-up measurements. One individual had well controlled type 2 diabetes and the overall glycemic control did not deteriorate during treatment. The remainder of the subjects had glucose levels less than 100mg/dl before, during, and after treatment. The elevated plasma glucose levels did not correlate with the plasma IGF1 levels. The participants lowered their total percent body fat during the first 12 months of GH treatment, but they regained a fat mass more than what they reduced during the 12 months following GH treatment. These changes were significant at 0.05 alpha level, except for the moderate-vigorous physical activity changes from 12 to 24 months and pain and HDL changes from baseline to 12 months. There were no significant differences between time points for diet records, resting metabolic rate, 8 hour energy expenditure or respiratory quotient (Table 2).

Discussion

Growth hormone replacement in children with PWS has well-defined benefits and risks, although data are limited for adults with PWS. However, adults with PWS share similarities with GH deficient patients (GHD) regarding reduced lean body mass (LBM) and increased fat mass (FM). GH treatment in non-PWS GHD adults improves body composition and has beneficial effects on muscle strength and physical performance. The reports on GH treatment in adults with PWS, to date, indicate improved body composition with increased LBM, a reduction in total body fat, subcutaneous adiposity and visceral fat but with small increases in fasting glucose after 12 months of treatment.³³ Positive effects of GH treatment on cognitive functioning were noted with improved verbal reasoning and visuospatial skills and motor performance in children.^{34–36}

We report a novel approach in PWS by measuring energy balance and comparing data obtained from a monitored whole room calorimeter, daily physical activity measures and quality of life assessments. Quality of life assessments (i.e., SF-36) were recorded in relationship to repeated body composition, blood chemistry and hormone levels, dietary records and strength indices in the selected PWS adults at baseline, after 12 months of GH treatment and at 24 months while off GH treatment for 12 months.

Other GH treatment trials reported in PWS men and women included randomized treatment with GH or a placebo for one year followed for two years with open-labeled GH treatment.³⁷ At one year, IGF-I levels were increased by 1.5 fold and body composition improved. We also observed an increase (about 4 fold) over baseline in plasma IGF-I levels after 12 months of GH treatment. Positive effects on body composition (i.e., decreased visceral and thigh fat and increased muscle mass) were reported by Sode-Carlsen et al.^{37–39}

in their Scandinavian cohort of PWS adults after two years of GH treatment. The beneficial effects occurred without serious adverse events while lipid and glucose metabolism data were unchanged. We also observed the positive impact on body composition with decreased total percent body fat after GH treatment but returned to near baseline after treatment cessation for 12 months. One of our PWS males did experience mild leg edema during treatment. High density lipoprotein levels did increase in our PWS adults during GH treatment but returned to a lower (less beneficial) level after treatment cessation.

Mogul et al.¹⁸ in 2008 reported the effectiveness and safety of GH in 38 GH-deficient PWS adults with a mean age of 30.5 years over a 12-month period using both six-month doseoptimization and six-month stable treatment periods. Human recombinant GH was initially administered at 0.2 mg/day with monthly 0.2 mg increments to a maximum of 1.0 mg/day, as tolerated, comparable to the dosage used in our study. They reported that lean body mass increased by about 3 kg and percent body fat decreased by 3% in their 30 PWS adults who completed the 6-12 month trial of GH treatment. Mildly progressive ankle edema was the most serious treatment adverse event reported in 5 of their 38 PWS adults. The total lean body mass in our study increased significantly by 2.3 kg during GH treatment but returned to a near baseline level after cessation of treatment for 12 months. The mean fasting glucose and insulin levels in their study were normal at baseline and remained in the normal reference range during GH treatment which agreed with the results found in our study. Although glucose levels were higher at 12 months of GH treatment in our PWS adults compared to baseline (e.g., 116 vs 101 mg/dl, respectively), the levels were not significantly different (see Table 2). No PWS adults in our study were diagnosed with diabetes during GH treatment.

In a separate study, Angulo et al.⁴⁰ in 2007 evaluated the final adult height in children with PWS and with and without GH. They found that administration of growth hormone in infancy and/or early childhood restored linear growth and final adult height without significant adverse events. In contrast, children with PWS who had not received growth hormone treatment had a significant decrease in growth velocity, with a mean final adult height under 2 standard deviations below the relevant population mean and with a higher risk for type 2 diabetes mellitus.

Lastly, endocrine and metabolic aspects of adults with PWS were studied by Hoybye⁴¹ in 2004 with special emphasis on the effect of growth hormone on a cohort of 19 young adults with clinical PWS having a mean BMI of 35 kg/m². At baseline the activity of the GH-IGF-I system was impaired and bone mineral density was low, as similarly seen in our PWS adults. Seventeen adults with PWS subsequently completed 12 months of GH treatment without significant adverse effects. Their study showed that adults with PWS had a partial GH deficiency, and GH treatment showed beneficial effects on body composition in adults with PWS without significant side-effects.

Bertella et al.³¹ reported their experience with quality of life measures in a study of 13 adults with PWS treated with GH using the SF-36 quality of life instrument to assess eight health dimensions. They found significant improvements with GH treatment in physical

functioning, body pain and general mental health. We also recorded higher scores for 4 of the 8 health dimension variables tested during GH treatment in our PWS study.

In conclusion, our study on GH treatment in adults with PWS of both sexes showed a positive impact on several variables including total lean gram weight, moderate-vigorous bouts of physical activity and plasma IGF-I and HDL levels during the growth hormone treatment interval but fasting glucose levels increased during treatment but not significantly different compared with baseline. The average fasting glucose levels were also high (outside of the normative reference range at 12 months) following GH treatment. The average baseline insulin levels were also high in comparison to reference ranges for our PWS adults. However, after cessation of GH treatment for 12 months, several variables returned close to baseline levels. In contrast, our PWS adults showed lower total percent body fat after 12 months of growth hormone treatment, but they regained more body fat than what they lost during the 12 months off treatment. All changes were significant at 0.05 alpha levels, except for the number of moderate-vigorous bouts of spontaneous physical activity recorded changes from 12 to 24 months and HDL levels from baseline to 12 months after GH treatment. Considering the extremely low physical activity levels at baseline in the PWS adults, the lack of change in the number of moderate-vigorous bouts of spontaneous physical activity from 12 to 24 months can be viewed as a positive outcome. An increase in moderate-vigorous spontaneous activity, regardless of size, has the potential to have positive long term benefits on weight management and overall health. The quality of life instruments were used to assess PWS adults receiving growth hormone and showed improvement with higher scores recorded for 4 of the 8 variables tested between baseline and at 12 or 24 months implying improvement during GH treatment for emotional and physical health, energy, social and particularly pain sensation. Interestingly, individuals with PWS are known to have decreased pain sensation and the change in pain perception during GH treatment will require further investigation.

There were no significant differences identified between time point measures for resting metabolic rate, 8 hour energy expenditure, or respiratory quotient using the whole room calorimeter despite an increase in lean body mass from the effects of GH treatment in our study. In the literature, the energy requirement in non-PWS individuals for lean body mass is estimated at about 15 to 25 kcal per kg per day^{42,43} and the lean body mass increased by 2.3 kg on average in our PWS adults following GH treatment. Therefore, the potential increase in resting metabolic rate would be estimated at 34–57 kcal which would account for little change in the resting metabolic rate detectable in our study. A second potential factor to consider for the lack of difference in energy expenditure measures may be the testing conditions. Although the whole room indirect calorimetry does allow for accurate measurements of energy expenditure, the space restrictions may limit spontaneous physical activity due to space restrictions in the whole room chamber could result in lower 24 hour metabolic rate calculations when compared to free-living arrangements outside of the chamber.

Several beneficial effects of growth hormone treatment noted previously in children with PWS were also observed in our PWS adults, specifically improved body composition, increased physical activity (number and duration of spontaneous bouts of activity or

movement) and higher morning fasting plasma HDL (12 months compared with 24 months) and IGF-I levels. Several of the measures appeared to regress to baseline after GH treatment cessation for 12 months. Due to better recognition, care and treatment including prevention of extreme obesity and co-morbidities, it is expected that GH treatment in PWS adults may exceed 50 years as affected patients are now living to an advanced age. Our study is not definitive due to sample size and length of treatment but supports the accumulating data on the use of GH in treating PWS adults with or without prior GH therapy to produce beneficial effects on body composition, physical activity, lipid profiles and possibly on quality of life with minimal adverse effects but glucose and insulin levels should be followed during GH treatment as recommended in treating non-PWS GH deficient patients. The authors do encourage an expanded study with a larger group of patients including those at an older age and obtain long term outcome data greater than two years duration on potential side effects (e.g., increased glucose levels, insulin resistance, edema) before a firm decision can be made regarding use of long term GH treatment and cost effectiveness in PWS adults. The implications and cost of long term GH treatment to each individual and to society and added health risks will warrant more investigation and discussion.

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Figure 1.

Multiple Parameters Showing Significant Differences in Individuals with Prader-Willi Syndrome Receiving Growth Hormone Treatment

*p < 0.05; **p < 0.01 for fixed effect within-subject (time) comparison using 12 months as a reference point.

Table 1

Baseline Descriptive Characteristics of Individuals with Prader-Willi Syndrome Grouped by Gender

Variable	Females (N=6)	Males (N=5)
Age (yrs)	33.8 ± 12.2	30.4 ± 10.9
Height (cm)**	144.7 ± 1.6	154.2 ± 2.5
Weight (kg)**	72.5 ± 5.4	98.9 ± 7.2
BMI	34.5 ± 2.2	41.6 ± 2.9
Percent Body Fat	51.0 ± 2.3	51.5 ± 2.9
24 hr EE (kcal)*	1 929 ± 124	2654 ± 274

BMI=Body mass index. EE=Energy expenditure. Values are means and standard errors. Independent samples t-test was used for comparison;

* p < 0.05,

** p < 0.01.

Table 2

Adjusted Means for Each Outcome Measure at Baseline, at 12 Months of Growth Hormone Treatment and at 24 Months When Off Treatment for 12 months for Individuals with Prader-Willi Syndrome

	Total					
	Baseline		12 months		24 months	
Variables	Μ	SE	M	SE	М	SE
Weight (kg)	84.6	4.9	85.2	5.0	88.0	5.2
Height (cm)	149.1	1.3	149.6	1.4	149.5	1.4
Body mass index (BMI)	37.7	2.0	37.6	2.0	38.8	2.1
Abdomen circumference (cm)	112.4	5.3	114.8	5.8	112.6	6.2
Hip circumference (cm)	114.8	4.1	109.8	4.7	112.8	5.1
Body composition						
Total bone mineral density (BMD)	1.14	0.05	1.12	0.05	1.15	0.05
Total lean (g)	39 955 ^{**}	2 727	42 301	2 749	40.386^{*}	2 785
Total tissue fat (g)	81 941	4 830	82 644	4 959	85 157	5 170
Total regional fat (g)	41 987	3 118	40 561	3 259	44 959	3 489
Percent body fat	51.2^{*}	2.1	48.5	2.2	52.1**	2.3
Whole-room chamber						
Energy intake (%)						
Fat	50.0	4.1	53.6	4.6	58.0	5.1
Carbohydrate	46.3	4.0	43.4	4.5	39.3	4.9
Protein	3.7	0.4	3.0	0.5	2.8	0.6
Energy expenditure (kcal)						
8 hr	738	49	752	53	806	58
24 hr (extrapolated)	2 215	146	2 257	160	2 417	174
Metabolic rate (kcal/min)	1.54	0.10	1.57	0.11	1.68	0.12

	Total					
	Baseline		12 months		24 months	
Variables	Μ	SE	М	SE	Μ	SE
Respiratory quotient (RQ)	0.85	0.01	0.83	0.01	0.82	0.01
Resting metabolic rate (RMR) (kcal)	1 845	161	1 919	203	1 914	278
RMR RQ (during first hour)	0.82	0.02	0.80	0.03	0.82	0.04
Physical activity						
Physical activity/Average counts/min	177	27	256	33	215	49
Activity min⁄day						
Sedentary	612	20	547	24	581	37
Light	214	12	218	14	225	21
Moderate	10.7	4.0	22.8	4.8	15.7	7.3
Vigorous	0.07	0.52	1.06	0.62	0.18	0.93
Bouts/day						
Moderate/Vigorous	0.18^{**}	0.12	0.73	0.13	0.26	0.19
Vigorous	0.00	0.02	0.04	0.02	0.00	0.04
Bout lengths/day (min)						
Moderate/Vigorous	4.6^*	2.4	12.5	2.8	6.1	4.0
Vigorous	0.00	0.44	06.0	0.53	0.00	0.80
Strength index	93	11	104	12	106	15
Laboratory						
Insulin-like growth factor I (IGF-I) (ng/ml) (reference range 98–553)	93**	37.6	385	48.6	140^{**}	53.1
Glucose (mg/dl) (range 65–110)	101	12	116	13	98	14
Insulin (mcU/ml) (range 2–18)	22.3	4.0	26.3	5.3	15.9	5.8
HOMA $\mathring{\tau}$ (insulin resistance): glucose (mg/dl) \times insulin (mcU/ml) /405	5.7	1.5	1.0	1.9	7.2	1.9
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	Total					
	Baseline		12 months		24 months	
Variables	Μ	SE	Μ	SE	Μ	SE
Free T4 (ng/dl) (range 0.8–1.9)	2.7	0.8	2.2	1.5	2.1	1.5
Thyroid stimulating hormone (TSH) (mcU/ml) (range 0.35–5.50)	1.3	0.2	3.1	0.7	0.7	0.7
Cholesterol (mg/dl) (range 140–200)	180	10.3	176	11.6	164	13.0
Triglyceride (mg/dl) (range 30–200)	123	10.0	116	11.8	113	12.9
High density lipoprotein (HDL) (mg/dl)	51	3.5	55	3.8	46*	4.1
Low density lipoprotein (LDL) (mg/dl)	108	8.1	95	9.0	93	10.3
Very low density lipoprotein (VLDL) (mg/dl)	24.2	2.0	23.1	2.3	22.6	2.6
Dietary						
Daily total energy intake (kcal)	1 419	235	1 442	244	1 291	267
Daily fat kcal (%)	29.2	2.0	30.6	2.4	35.1	3.2
Daily carbohydrate kcal (%)	52.9	3.0	54.8	3.5	48.2	4.9
Daily protein kcal (%)	20.0	1.3	17.7	1.6	19.6	2.5
Quality of life	9.4	1.9	8.7	2.0	10.5	2.2
SF-36						
Dimension 1 Physical functioning	72	9	80	7	70	6
Dimension 2 Role limitations/physical health	74	11	69	12	75	17
Dimension 3 Role limitations/emotional health	82	12	99	13	76	16
Dimension 4 Energy/fatigue	53	7	67	8	52	11
Dimension 5 Emotional well being	81	5	84	5	88	9
Dimension 6 Social functioning	81	S	85	S	92	9

	Total					
	Baseline	1	2 months		24 months	
Variables	М	SE	Μ	SE	Μ	SE
Dimension 7 Pain	72	~	82	6	60	10^*
Dimension 8 General health	73	9	67	9	75	8
* * / 0.5.						

p < 0.05;

p > 0.01 for fixed effect within-subject (time) comparison using 12 months as a reference point M = mean; SE = standard error *

⁺Homeostasis model assessment (HOMA), an indicator of insulin resistance is represented by the following equation: glucose (mg/dl) × insulin (mcU/ml) /405 (see Matthews et al., 1985, for description)⁴⁴