

Review Article

Role of Inflammatory Mediators in the Pathogenesis of Epilepsy

Tadayuki Shimada,¹ Takako Takemiya,² Hiroko Sugiura,¹ and Kanato Yamagata¹

¹Neural Plasticity Project, Tokyo Metropolitan Institute of Medical Science, 2-1-6 Kamikitazawa, Setagaya-ku, Tokyo 156-8506, Japan

²Medical Research Institute, Tokyo Women's Medical University, Shinjuku, Tokyo 162-8666, Japan

Correspondence should be addressed to Kanato Yamagata; yamagata-kn@igakuken.or.jp

Received 11 June 2014; Revised 4 August 2014; Accepted 5 August 2014; Published 13 August 2014

Academic Editor: Kiyoshi Matsumura

Copyright © 2014 Tadayuki Shimada et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Epilepsy is one of the most common chronic brain disorders worldwide, affecting 1% of people across different ages and backgrounds. Epilepsy is defined as the sporadic occurrence of spontaneous recurrent seizures. Accumulating preclinical and clinical evidence suggest that there is a positive feedback cycle between epileptogenesis and brain inflammation. Epileptic seizures increase key inflammatory mediators, which in turn cause secondary damage to the brain and increase the likelihood of recurrent seizures. Cytokines and prostaglandins are well-known inflammatory mediators in the brain, and their biosynthesis is enhanced following seizures. Such inflammatory mediators could be therapeutic targets for the development of new antiepileptic drugs. In this review, we discuss the roles of inflammatory mediators in epileptogenesis.

1. Introduction

Epilepsy is a chronic neurological disorder characterized by recurrent seizures and is often accompanied by cognitive deficits and mood disorders [1–3]. Approximately 50 million people worldwide have epilepsy. Because seizures are the result of uncontrollable neural excitation in the brain, epilepsy has been considered to primarily be a neuronal disease. The targeting of neuronal ion channels and both gamma-aminobutyric acid (GABA) and glutamate receptors has been the primary approach to eliminate seizures. However, studies that have focused exclusively on neurons fail to address the questions that arise from more complex models of epileptogenesis. To date, studies using animal models and human patients with epilepsy have shown that the pathogenesis of epilepsy may be associated with both neuronal and nonneuronal components such as glial cells [4], brain vasculature [5], and leucocytes from the periphery [6].

The aberrant regulation of glial functions can elicit seizures and promote epileptogenesis [4]. Glial abnormalities, including chronically activated astrocytes and microglia, glial scars, and various gliomas, are likely to form epileptic foci in the brain [4]. The mechanisms through which glial cells can promote epileptogenesis can include increased

neuronal excitability and inflammatory processes. The key roles of inflammatory processes in relation to epilepsy have been clarified over the last decade. Studies of the mechanisms of antiepileptic drugs (AEDs) have focused on ion channels, transporters, and excitatory/inhibitory neurotransmission [7]. However, the anti-inflammatory effects of AEDs have received recent attention due to their relevance to antiepileptic properties. For example, carbamazepine and levetiracetam can reduce the expression of inflammatory mediators in glial cell cultures [8, 9]. Levetiracetam can also normalize the resting membrane potential of astrocytes increased by inflammatory mediators [8]. One of the anticonvulsive effects of levetiracetam could be mediated through suppression of astroglial activation by inflammatory mediators [8].

In addition, dysfunction of the blood-brain barrier (BBB) may be responsible for abnormal neuronal firing. Disruption of the BBB causes the leakage of serum protein and leucocyte invasion into the brain. These exogenous inflammatory mediators have the potential to lower seizure thresholds [4, 5, 10, 11], which could alter channel sensitivity, neurotransmitter uptake or release, and glia-associated regulation of extracellular environments, such as potassium concentration [4, 5, 10, 11]. Accordingly, brain inflammation is one of the etiological factors that promote epileptogenesis and ictogenesis.

In this review, we discuss seizure-induced inflammatory mediators and the mechanisms through which these factors exacerbate epilepsy.

2. Inflammatory and Immune Responses in Epilepsy

Direct anti-inflammatory treatments have been reported to suppress some type of epileptic seizures that are resistant to conventional AEDs. For example, adrenocorticotrophic hormone (ACTH) has been a first-line treatment for infantile spasms [13]. Anti-inflammatory effects of increased steroid hormone by ACTH treatment could play a crucial role in the suppression of refractory epilepsy in West syndrome [14]. In addition, intravenous immunoglobulin (IVIg) can suppress seizures in some types of intractable epilepsy, an effect that may be partially mediated through a reduction in cytokines and a suppression of astrocyte activation [15, 16]. These drugs are also able to confer protection against seizures in mice with some types of epilepsy that are resistant to conventional AEDs [17]. Combined with antigliotic functions of conventional AEDs described above, anti-inflammatory medication could be a new promising treatment for refractory epilepsy.

Research with rodent models of epilepsy has uncovered roles for brain inflammation in epileptogenesis and ictogenesis. Pharmacological or electrical stimulation produces epileptic seizures accompanied by robust inflammatory responses in the brains of rodents [18–32]. The administration of proinflammatory or anti-inflammatory reagents has also been used to elucidate the effects of inflammatory mediators on seizure latency, frequency, duration, and severity [33, 34]. For instance, lipopolysaccharide, a provocative agent for inflammation, exacerbates seizure severity [33], whereas an inhibitory peptide against high-mobility group box-1 (HMGB1) inflammatory mediator decreased acute and chronic seizure recurrence [34]. Transgenic mouse models have also been used to evaluate the relationship between inflammatory mediators and seizure severity. Mice overexpressing cytokines, such as tumor necrosis factor- α (TNF- α) or interleukin-6 (IL-6), within astrocytes developed age-dependent neurological dysfunctions including a reduction in seizure threshold, spontaneous seizure frequency, and neuronal cell loss [35, 36]. These studies suggest the idea that brain inflammation could promote seizures and epileptogenesis.

Inflammatory cytokines, including interleukin-1 β (IL-1 β) and HMGB1, are released from astrocytes and microglia after seizures [22–24, 37]. IL-1 β and HMGB1 activate IL-1R type I [38] and Toll-like receptor 4 (TLR4), respectively [39]. IL-1R/TLR signaling can regulate neuronal excitability, including the alteration of synaptic transmission, the reduction in GABA production, and the inhibition of outward current of Ca²⁺ channels [10, 40–42]. IL1R/TLR signaling may activate the Src kinase-mediated phosphorylation of N-methyl-D-aspartate (NMDA) receptor subunit 2B (NR2B) [43]. This phosphorylation can increase Ca²⁺ permeability of NMDA receptor [43]. Consequently, this NMDA-mediated Ca²⁺ influx could be enhanced in neurons, leading

to increased neuronal excitability and excitotoxicity [34, 44]. HMGB1/TLR4 signaling also targets NR2B [36]. Thus, the activation of IL-1 β /IL-1R or HMGB1/TLR4 signaling resulting from epileptic insults might result in a rapid change in neuronal excitability and a decreased seizure threshold (Figure 1(a)).

TNF- α is another inflammatory factor, and its expression is also upregulated following seizures [18, 36]. TNF- α is mainly released by microglia in the brain [45], and it can stimulate astrocytes to release glutamate [46]. An extracellular increase in glutamate concentration may stimulate glutamatergic neurons, thereby depolarizing their membrane potential. Taken together, postseizure production of inflammatory mediators can trigger neuronal hyperexcitability through modulations of ion channels and glutamate release in neurons and glia, respectively (Figure 1(a)).

In addition, the endothelial cells that make up the BBB could be another center for inflammatory processes in the brain. The activation of the IL-1 β system following seizures induces neuronal cell loss and a breakdown in the BBB [47]. This breakdown leads to BBB leakage and the entry of albumin into the brain. Albumin entry can induce a further upregulation in inflammatory mediators and reduce the potassium and glutamate uptake of astrocytes, which leads to increased neural excitability [48–50]. BBB leakage also initiates the invasion of leucocytes and an enhancement in inflammatory reactions [51]. These findings indicate that a breakdown in the BBB can increase neuronal excitability by enhancing inflammatory responses in the brain (Figure 1(a)).

Taken together, epileptic seizures can provoke inflammatory responses, which enhance calcium influx into neurons, activate glial cells to increase extracellular potassium and glutamate, and induce further inflammatory response via BBB break down. These inflammatory responses may promote neural hyperexcitability and decrease the seizure threshold. Consequently, epileptic seizures and inflammatory mediators can form a positive feedback loop, reinforcing each other (Figure 1(a)).

3. Prostaglandin Production in Epilepsy

In addition to inflammatory cytokines, prostaglandins (PGs) are major factors that stimulate inflammation processes. PGs are known to markedly increase following seizures and may contribute to epileptogenesis and reduction in seizure threshold [10, 52]. Phospholipase A₂ liberates arachidonic acid from membrane phospholipids. The enzyme cyclooxygenase (COX) converts arachidonic acid to PGH₂, and the specific PG synthase converts PGH₂ to various PGs such as thromboxane A₂, PGF_{2a}, PGE₂, PGI₂, or PGD₂. Consistent with the robust production of PGs in the brain following seizures, an inducible type of COX (COX-2), but not the constitutively expressed COX-1, is rapidly induced in the brain following seizures [16, 53].

Because PGs play an important role in inflammatory responses, the functions of PGs in epileptogenesis have been studied for a considerable amount of time [52, 54]; however, data on the roles of COX-2 in epilepsy appear

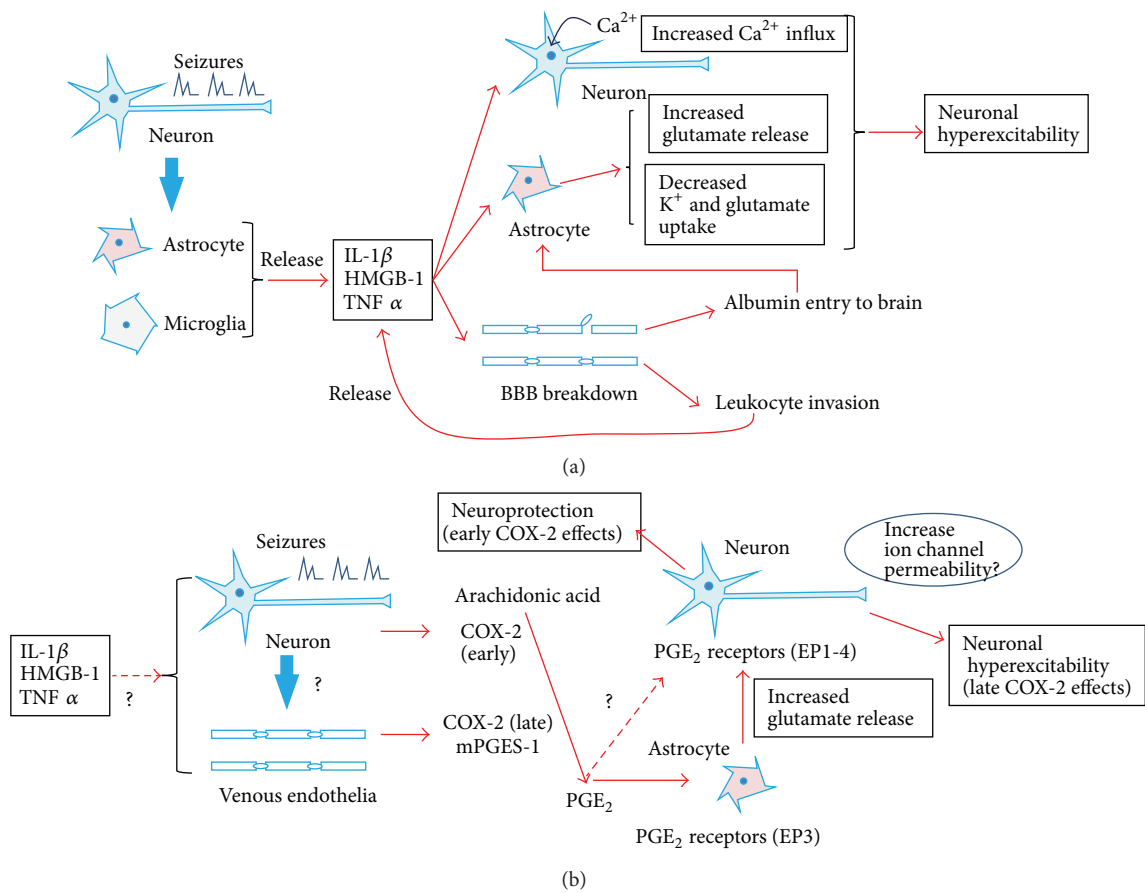


FIGURE 1: Proposed inflammatory mechanisms in epileptogenesis. (a) Epileptic seizures induce the release of cytokines from glial cells, thereby (1) increasing the influx of neuronal calcium; (2) enhancing extraneuronal glutamate concentration; (3) decreasing K⁺ and glutamate uptake by glia; and (4) impairing the BBB. BBB breakdown leads to albumin entry and leukocyte invasion into the brain, resulting in a further release of inflammatory cytokines. Such inflammatory responses cause an induction of neuronal hyperexcitability, reoccurrence of seizures, and finally the development of refractory epilepsy. (b) Seizures induce COX-2 in neurons (early phase) and vascular endothelial cells (late phase) and mPGES-1 in endothelial cells. These inducible PG synthases cooperate to produce PGE₂, most likely in endothelial cells. Endothelial PGE₂ might cause neuronal hyperexcitability by enhancing glutamate release from astrocytes via the glial EP3 receptor, whereas neuronal PGs may protect neurons against seizures.

to be bidirectional. For example, following an injection of kainic acid (KA), postseizure treatment with the selective COX-2 inhibitor NS-398 prevented neuronal cell loss in the hippocampus [55]. However, the administration of another COX-2 inhibitor, nimesulide, prior to an injection of KA strongly aggravated KA-induced seizures and increased the mortality rate [56]. Moreover, pretreatment with NS-398 increased KA-induced TUNEL-positive neuronal death [57]. These conflicting results might be partly explained by possible dual roles for COX-2, which has been shown to play early neuroprotective and late neurotoxic roles following seizures [58]. Thus, the timing of COX-2 inhibitor administration may be crucial when treating epilepsy (Figure 1(b)).

The bifunctional aspects of COX-2 in epileptogenesis can also be explained by the diversity of PGs. For instance, PGD₂ and PGF_{2 α} can exhibit anticonvulsive functions. PGD₂ synthase H-PGDS-knockout (KO) mice or PGD₂ receptor DPIR-KO mice showed more severe seizures after pentylentetrazol (PTZ) treatment than wild type (WT) mice, whereas

deficiencies of the other synthase L-PGDS or receptor DP2R did not alter seizure severity [59]. Additionally, the intracisternal administration of PGF_{2 α} after KA treatment reduced the seizure score and mortality [60]. By contrast, PGE₂ mainly functions as a promoter of epileptogenesis and ictogenesis. The administration of PGE₂ receptor (EP) antagonists reduced seizure severity and neuronal injury following pilocarpine- or PTZ-induced seizures [61–63]. Three PGE₂ synthase (PGES) genes have been identified, including membrane-bound PGES (mPGES)-1, mPGES-2, and cytosolic PGES (cPGES). mPGES-1 is induced in the venous endothelia of the brain following an injection of KA [64], and its expression is upregulated in conditions of inflammation, pain, and fever [16, 65, 66]. mPGES-2 and cPGES show constitutive levels of expression, and their physiological functions remain unknown. The finding that the activity of mPGES-1 is tightly coupled to that of COX-2 [67] could support the importance of PGE₂ function under epileptic conditions.

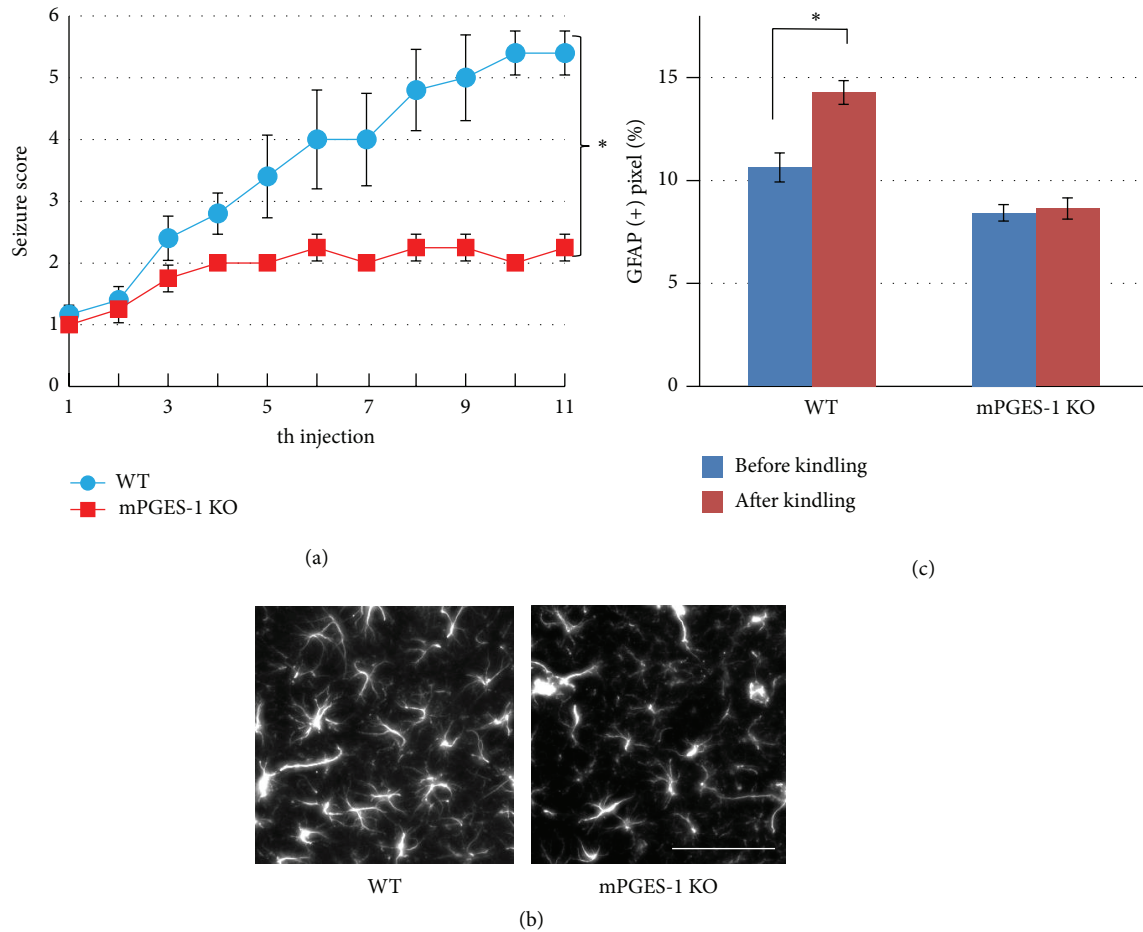


FIGURE 2: mPGES-1-KO mice show a less severe aggravation of epileptic seizures. (a) Development of epileptic convulsive seizures following consecutive treatments with PTZ. Two-month-old female C57/BL6 background mice of WT or mPGES-1-KO mice were injected intraperitoneally with a subconvulsive dose (35 mg/kg) of PTZ every other day. After each PTZ injection, the convulsive behaviors were observed for 30 min and the resultant seizures were classified and scored as follows: 0: normal behavior; 1: immobilization; 2: facial, forelimb, or hindlimb myoclonus; 3: continuous whole body myoclonus; 4: rearing, tonic seizure; 5: tonic-chronic seizure; and 6: death [12]. mPGES-1-KO mice showed significant reduction in seizure score compared with that in WT mice (*: $P < 0.05$, repeated-measured ANOVA). (b) Representative images of the CA3 region of hippocampal sections from WT and mPGES-1-KO mice after PTZ-induced chemical kindling. Mice were fixed with 4% paraformaldehyde. Serial sections (30 μm) were generated, and immunostaining was performed with a rabbit anti-GFAP antibody (DAKO). Gliosis was observed only in the sections from wild type mice. Scale bar = 20 μm . (c) Quantification of gliosis development: 200 $\mu\text{m} \times 200 \mu\text{m}$ squares were randomly overlaid on the CA3 region of hippocampal slices and the percentage of pixels that had a higher intensity of GFAP signal than the threshold was compared. The GFAP-positive area was significantly increased after kindling in WT mice, whereas no increase was observed in mPGES-1-KO mice (* $P < 0.01$, two-way ANOVA followed by Tukey's test). Six squares were taken from one brain slice. Four slices were examined under each condition.

Several lines of evidence suggest that PGE₂ could have an important role in epileptic neuronal cell death and the reduction in seizure threshold. mPGES-1-KO mice show little postictal production of PGE₂, and the lack of increased PGE₂ production resulted in the promotion of neuronal survival following KA injection [64, 68]. Furthermore, the intraventricular injection of PGE₂ decreased the latency to methylmalonate- (MMA-) induced seizures and increased the amplitude of spikes measured by an electroencephalogram (EEG) in rats [69]. Thus, COX-2-coupled PGE₂ production can promote seizures through mechanisms that drive epileptogenesis.

To determine whether mPGES-1-derived PGE₂ is directly involved in epileptogenesis, we used mPGES-1-KO mice. To mimic the gradual exacerbation of epileptic seizures, a subconvulsive dose of PTZ was administered to WT or mPGES-1-KO mice every other day (PTZ chemical kindling). WT mice showed rapid increases in seizure severity, whereas mPGES-1-KO mice showed a significantly slower elevation in seizure score (Figure 2(a)). Immunohistochemical analyses revealed that glial fibrillary acidic protein- (GFAP-) positive astrocytes were increased in the hippocampal CA3 region of the wild type mice after the kindling, indicating that PGE₂ production promotes reactive astrogliosis

(Figure 2(b)). Interestingly, mPGES-1-KO mice did not show any increase in GFAP-positive astrocytes after the kindling (Figure 2(c)). Reactive astrogliosis is one of the pathological hallmarks of temporal lobe epilepsy, which include segmental neuronal loss and increases in GFAP-positive cells in the hippocampus and associated temporal lobe structures [4]. Our results indicate that mPGES-1-KO mice are resistant to the development of PTZ kindling and also to kindling-induced astrogliosis.

How can PGE₂ regulate hyperexcitability and epileptogenesis? PGE₂ acts on four G protein-coupled receptors named EP1, EP2, EP3, and EP4. EP receptors have been shown to be localized in the brain cells [70, 71]; however, it still remains unclear on which cells PGE₂ acts to produce its epileptogenic effects. By contrast, pharmacological approaches have preceded the clarification of brain PGE₂ function. The administration of a selective EP2 antagonist prevented neuronal cell loss after pilocarpine-induced seizures [62]. Furthermore, the pharmacological inhibition of other receptors, EP1, EP3, or EP4, resulted in increases in PTZ-induced seizure latency [63]. These results suggest that PGE₂ may cause neuronal cell death and neuronal hyperexcitability via hippocampal EP receptors (Figure 1(b)).

The effects of PGE₂ on neurons might be partly similar to those of other inflammatory mediators in terms of their enhancement of neuronal excitability. Several studies have shown that PG synthesis is upregulated by IL-1 β treatment. In a lung fibroblast cell line, IL-1 β induced mPGES-1 mRNA expression [72]. Additionally, IL-1 β treatment increased COX-2 mRNA levels in cultured murine primary astrocytes [73]. Thus, inflammatory cytokines may promote PGE₂ synthesis in the brain following epileptic seizures.

PGE₂ can act on neuronal EP receptors directly. The application of PGE₂ led to significant increases in firing frequency and excitatory postsynaptic potential (EPSP) amplitude in rat hippocampal slice cultures [74]. PGE₂-induced increases in neuronal excitability may be partly attributable to an inhibition of potassium currents, resulting in boosted Ca²⁺ influx [74]. However, given that the mPGES-1 enzyme is preferentially expressed in the brain microvasculature [65], it may be more likely that PGE₂ acts on glial cells but not on neurons. PGE₂ appears to act on glial processes surrounding brain microvessels. We previously reported that the EP3 receptor was localized to astroglial end feet around microvessels [64]. Additionally, mPGES-1-KO mice showed decreased astrogliosis compared to WT mice following KA treatment [64]. These findings suggest that endothelial PGE₂ may act on glial EP3 receptors and alter glial function. In fact, glutamate release from astrocytes was drastically reduced in mPGES-1-KO mice compared to WT mice [66, 75]. Thus, PGE₂ is both an inflammatory mediator and a promoter of extracellular gliotransmission via astrocytic EP receptors [46] (Figure 1(b)).

In conclusion, epileptic seizures rapidly induce COX-2 mRNA in neurons and afterwards upregulate both COX-2 and mPGES-1 mRNAs in venous endothelia [16, 64–66]. Treatment with NS-398 and nimesulide prior to seizure induction aggravates neuronal damage in the hippocampus,

whereas postseizure treatments with NS-398 prevent the damages [55, 56]. Neuronal COX-2-derived PGs appear to protect neurons against seizures, whereas endothelial PG (PGE₂) might participate in seizure-induced neuronal cell death by stimulating astrocytes to release glutamate [58]. Thus, PG-mediated inflammation may also form a positive feedback loop to exacerbate epileptic seizures in a late phase after seizures (Figure 1(b)). Epileptic seizures rapidly induce COX-2 in excitatory neurons and increase brain PGE₂ levels [64]. However, this neuronal upregulation of COX-2 and PGE₂ is transient and simultaneous robust increases in brain PGD₂ levels could surpass the effects of PGE₂, resulting in suppression of seizures. By contrast, late induction of endothelial COX-2 and mPGES-1 lasts longer than early phase and this sustained production of PGE₂ may promote neurodegeneration via astrocytes. Thus, time course and cell type-specific expression of COX-2 could cause the dual effects of PG production on epileptogenesis.

4. Conclusion and Perspective

Most current AEDs act on ion channels that directly control neuronal excitability. These medications have at least two major problems. First, even with optimal AED therapy, 20~30% of patients have poor seizure control and become intractable. Second, as these medications act as general central nervous system (CNS) depressants and must be taken chronically for seizure suppression, they also have marked inhibitory effects on cognitive development. Therefore, the development of new AEDs that can modulate seizures through another mechanism is required for refractory epilepsy treatment.

Animal models and clinical evidence have emphasized the involvement of inflammatory mediators in seizure susceptibility and epileptogenesis [76–78]. Robust and general inflammatory responses in the brain lower the seizure threshold, enhance neuronal excitability, increase BBB permeability, and promote epileptogenesis. COX-2 and mPGES-1 are known to play key roles in the inflammatory responses to insults and consequently increase postseizure inflammation and the resulting hyperexcitability of brain neurons. COX-2 inhibitors such as celecoxib have been developed and prescribed for chronic inflammatory pain and rheumatoid arthritis. Hence, they can be used in conjunction with AEDs to treat brain inflammation and reduce neuronal hyperexcitability. Although celecoxib suppresses epileptogenesis, for example, the development of PTZ-kindling, this inhibitor may be more effective for postseizure inflammation than for the prevention of seizure incidence. However, given that chronic celecoxib treatment can be accompanied by serious cardiovascular side effects [79], persistent COX-2 inhibition would not be beneficial as a therapeutic strategy for intractable epilepsy.

By contrast, the regulation of specific PG signaling should offer more selective actions and fewer complicating adverse effects. Although PGE₂ synthase inhibitors may be good candidates for epilepsy treatment, specific inhibitors are still under development. Recently, attention has been paid to EP

receptors. An EP2 inhibitor appears to suppress neurodegenerative pathology following epileptic seizures, and the inhibition of the other EP receptors, EP1, EP3, and EP4, might delay seizure induction [61–63]. The discovery of selective EP receptor antagonist(s) could contribute to avoid the serious side effects of chronic COX-2 inhibition and provide more selective therapies for refractory epilepsy.

Modulators of PG receptor function could also improve our understanding of the roles of PGs in the brain immune system and inflammation. Consequently, more precise molecular roles of PGs in epileptogenesis will need to be clarified. This knowledge will help to develop newer and better treatments and medications for epilepsy.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Acknowledgments

The authors thank Dr. Shin Yasuda for helping with the software used for statistical analysis. This work was partly supported by JSPS KAKENHI Grant nos. 24700349, 24659093, and 25293239 and MEXT KAKENHI Grant nos. 25110737 and 23110525.

References

- [1] O. Devinsky, "Effects of seizures on autonomic and cardiovascular function," *Epilepsy Currents*, vol. 4, no. 2, pp. 43–46, 2004.
- [2] J. M. Pellock, "Understanding co-morbidities affecting children with epilepsy," *Neurology*, vol. 62, no. 5, supplement 2, pp. S17–S23, 2004.
- [3] J. E. Jones, J. K. Austin, R. Caplan, D. Dunn, S. Plioplys, and J. A. Salpekar, "Psychiatric disorders in children and adolescents who have epilepsy," *Pediatrics in Review*, vol. 29, no. 2, pp. e9–e14, 2008.
- [4] J. Wetherington, G. Serrano, and R. Dingledine, "Astrocytes in the epileptic brain," *Neuron*, vol. 58, no. 2, pp. 168–178, 2008.
- [5] A. Friedman, D. Kaufer, and U. Heinemann, "Blood-brain barrier breakdown-inducing astrocytic transformation: novel targets for the prevention of epilepsy," *Epilepsy Research*, vol. 85, no. 2-3, pp. 142–149, 2009.
- [6] J. Greenwood, S. Etienne-Manneville, P. Adamson, and P. Couraud, "Lymphocyte migration into the central nervous system: implication of ICAM-1 signalling at the blood-brain barrier," *Vascular Pharmacology*, vol. 38, no. 6, pp. 315–322, 2002.
- [7] R. L. MacDonald and M. A. Rogawski, "Cellular effects of antiepileptic drugs," in *Epilepsy: A Comprehensive Textbook*, pp. 1433–1445, Wolters Kluwer/Lippincott Williams & Wilkins, 2nd edition, 2008.
- [8] A. Haghikia, K. Ladage, D. Hinkerohe et al., "Implications of antiinflammatory properties of the anticonvulsant drug levetiracetam in astrocytes," *Journal of Neuroscience Research*, vol. 86, no. 8, pp. 1781–1788, 2008.
- [9] I. Matoth, F. Pinto, C. Sicsic, and T. Brenner, "Inhibitory effect of carbamazepine on inflammatory mediators produced by stimulated glial cells," *Neuroscience Research*, vol. 38, no. 2, pp. 209–212, 2000.
- [10] B. Viviani, F. Gardoni, and M. Marinovich, "Cytokines and Neuronal Ion Channels in Health and Disease," *International Review of Neurobiology*, vol. 82, pp. 247–263, 2007.
- [11] A. Vezzani, J. French, T. Bartfai, and T. Z. Baram, "The role of inflammation in epilepsy," *Nature Reviews Neurology*, vol. 7, no. 1, pp. 31–40, 2011.
- [12] K. Itoh, M. Watanabe, K. Yoshikawa, Y. Kanaho, L. J. Berliner, and H. Fujii, "Magnetic resonance and biochemical studies during pentylenetetrazole-kindling development: the relationship between nitric oxide, neuronal nitric oxide synthase and seizures," *Neuroscience*, vol. 129, no. 3, pp. 757–766, 2004.
- [13] F. Vigeveno and M. R. Cilio, "Vigabatrin versus ACTH as first-line treatment for infantile spasms: a randomized, prospective study," *Epilepsia*, vol. 38, no. 12, pp. 1270–1274, 1997.
- [14] N. A. Shumiloff, W. M. Lam, and K. B. Manasco, "Adrenocorticotropic hormone for the treatment of West syndrome in children," *Annals of Pharmacotherapy*, vol. 47, no. 5, pp. 744–754, 2013.
- [15] A. R. Crow, S. Song, J. W. Semple, J. Freedman, and A. H. Lazarus, "A role for IL-1 receptor antagonist or other cytokines in the acute therapeutic effects of IVIg?" *Blood*, vol. 109, no. 1, pp. 155–158, 2007.
- [16] K. Yamagata, K. I. Andreasson, W. E. Kaufmann, C. A. Barnes, and P. F. Worley, "Expression of a mitogen-inducible cyclooxygenase in brain neurons: regulation by synaptic activity and glucocorticoids," *Neuron*, vol. 11, no. 2, pp. 371–386, 1993.
- [17] M. Maroso, S. Balosso, T. Ravizza et al., "Interleukin-1 β biosynthesis inhibition reduces acute seizures and drug resistant chronic epileptic activity in mice," *Neurotherapeutics*, vol. 8, no. 2, pp. 304–315, 2011.
- [18] M. Minami, Y. Kuraishi, and M. Satoh, "Effects of kainic acid on messenger RNA levels of IL-1 β , IL-6, TNF α and LIF in the rat brain," *Biochemical and Biophysical Research Communications*, vol. 176, no. 2, pp. 593–598, 1991.
- [19] C. Eriksson, R. Tehranian, K. Iverfeldt, B. Winblad, and M. Schultzberg, "Increased expression of mRNA encoding interleukin-1 β and caspase-1, and the secreted isoform of interleukin-1 receptor antagonist in the rat brain following systemic kainic acid administration," *Journal of Neuroscience Research*, vol. 60, no. 2, pp. 266–279, 2000.
- [20] M. G. de Simoni, C. Perego, T. Ravizza et al., "Inflammatory cytokines and related genes are induced in the rat hippocampus by limbic status epilepticus," *European Journal of Neuroscience*, vol. 12, no. 7, pp. 2623–2633, 2000.
- [21] A. Vezzani, D. Moneta, M. Conti et al., "Powerful anticonvulsant action of IL-1 receptor antagonist on intracerebral injection and astrocytic overexpression in mice," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 97, no. 21, pp. 11534–11539, 2000.
- [22] N. P. Turrin and S. Rivest, "Innate immune reaction in response to seizures: implications for the neuropathology associated with epilepsy," *Neurobiology of Disease*, vol. 16, no. 2, pp. 321–334, 2004.
- [23] B. Voutsinos-Porche, E. Koning, H. Kaplan et al., "Temporal patterns of the cerebral inflammatory response in the rat lithium-pilocarpine model of temporal lobe epilepsy," *Neurobiology of Disease*, vol. 17, no. 3, pp. 385–402, 2004.
- [24] J. A. Gorter, E. A. van Vliet, E. Aronica et al., "Potential new antiepileptogenic targets indicated by microarray analysis in

- a rat model for temporal lobe epilepsy,” *Journal of Neuroscience*, vol. 26, no. 43, pp. 11083–11110, 2006.
- [25] K. H. Jung, K. Chu, S. T. Lee et al., “Cyclooxygenase-2 inhibitor, celecoxib, inhibits the altered hippocampal neurogenesis with attenuation of spontaneous recurrent seizures following pilocarpine-induced status epilepticus,” *Neurobiology of Disease*, vol. 23, no. 2, pp. 237–246, 2006.
- [26] K. Yoshikawa, Y. Kita, K. Kishimoto, and T. Shimizu, “Profiling of eicosanoid production in the rat hippocampus during kainic acid-induced seizure: dual phase regulation and differential involvement of COX-1 and COX-2,” *The Journal of Biological Chemistry*, vol. 281, no. 21, pp. 14663–14669, 2006.
- [27] E. Aronica and J. A. Gorter, “Gene expression profile in temporal lobe epilepsy,” *Neuroscientist*, vol. 13, no. 2, pp. 100–108, 2007.
- [28] F. Dhote, A. Peinnequin, P. Carpentier et al., “Prolonged inflammatory gene response following soman-induced seizures in mice,” *Toxicology*, vol. 238, no. 2–3, pp. 166–176, 2007.
- [29] B. Lee, H. Dziema, K. H. Lee, Y. Choi, and K. Obrietan, “CRE-mediated transcription and COX-2 expression in the pilocarpine model of status epilepticus,” *Neurobiology of Disease*, vol. 25, no. 1, pp. 80–91, 2007.
- [30] S. K. Kulkarni and A. Dhir, “Cyclooxygenase in epilepsy: from perception to application,” *Drugs of Today*, vol. 45, no. 2, pp. 135–154, 2009.
- [31] L. Holtman, E. A. van Vliet, P. M. Edelbroek, E. Aronica, and J. A. Gorter, “Cox-2 inhibition can lead to adverse effects in a rat model for temporal lobe epilepsy,” *Epilepsy Research*, vol. 91, no. 1, pp. 49–56, 2010.
- [32] N. Polascheck, M. Bankstahl, and W. Löscher, “The COX-2 inhibitor parecoxib is neuroprotective but not antiepileptogenic in the pilocarpine model of temporal lobe epilepsy,” *Experimental Neurology*, vol. 224, no. 1, pp. 219–233, 2010.
- [33] S. Auvin, D. Shin, A. Mazarati, and R. Sankar, “Inflammation induced by LPS enhances epileptogenesis in immature rat and may be partially reversed by IL1RA,” *Epilepsia*, vol. 51, supplement 3, pp. 34–38, 2010.
- [34] M. Maroso, S. Balosso, T. Ravizza et al., “Toll-like receptor 4 and high-mobility group box-1 are involved in ictogenesis and can be targeted to reduce seizures,” *Nature Medicine*, vol. 16, no. 4, pp. 413–419, 2010.
- [35] I. L. Campbell, C. R. Abraham, E. Masliah et al., “Neurologic disease induced in transgenic mice by cerebral overexpression of interleukin 6,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 90, no. 21, pp. 10061–10065, 1993.
- [36] L. Probert, K. Akassoglou, G. Kassiotis, M. Pasparakis, L. Alexopoulou, and G. Kollias, “TNF- α transgenic and knockout models of CNS inflammation and degeneration,” *Journal of Neuroimmunology*, vol. 72, no. 2, pp. 137–141, 1997.
- [37] A. Vezzani and T. Granata, “Brain inflammation in epilepsy: experimental and clinical evidence,” *Epilepsia*, vol. 46, no. 11, pp. 1724–1743, 2005.
- [38] K. Kuno and K. Matsushima, “The IL-1 receptor signaling pathway,” *Journal of Leukocyte Biology*, vol. 56, no. 5, pp. 542–547, 1994.
- [39] J. S. Park, F. Gamboni-Robertson, Q. He et al., “High mobility group box 1 protein interacts with multiple Toll-like receptors,” *The American Journal of Physiology—Cell Physiology*, vol. 290, no. 3, pp. C917–C924, 2006.
- [40] K. M. Rodgers, M. R. Hutchinson, A. Northcutt, S. F. Maier, L. R. Watkins, and D. S. Barth, “The cortical innate immune response increases local neuronal excitability leading to seizures,” *Brain*, vol. 132, no. 9, pp. 2478–2486, 2009.
- [41] S. Wang, Q. Cheng, S. Malik, and J. Yang, “Interleukin-1 β inhibits γ -aminobutyric acid type A (GABA(A)) receptor current in cultured hippocampal neurons,” *The Journal of Pharmacology and Experimental Therapeutics*, vol. 292, no. 2, pp. 497–504, 2000.
- [42] M. Schäfers and L. Sorkin, “Effect of cytokines on neuronal excitability,” *Neuroscience Letters*, vol. 437, no. 3, pp. 188–193, 2008.
- [43] B. Viviani, S. Bartesaghi, F. Gardoni et al., “Interleukin-1 β enhances NMDA receptor-mediated intracellular calcium increase through activation of the Src family of kinases,” *Journal of Neuroscience*, vol. 23, no. 25, pp. 8692–8700, 2003.
- [44] A. Vezzani, M. Maroso, S. Balosso, M. Sanchez, and T. Bartfai, “IL-1 receptor/Toll-like receptor signaling in infection, inflammation, stress and neurodegeneration couples hyperexcitability and seizures,” *Brain, Behavior, and Immunity*, vol. 25, no. 7, pp. 1281–1289, 2011.
- [45] T. Renno, M. Krakowski, C. Piccirillo, J.-. Lin, and T. Owens, “TNF- α expression by resident microglia and infiltrating leukocytes in the central nervous system of mice with experimental allergic encephalomyelitis: Regulation by Th1 cytokines,” *Journal of Immunology*, vol. 154, no. 2, pp. 944–953, 1995.
- [46] P. Bezzi, M. Domercq, L. Brambilla et al., “CXCR4-activated astrocyte glutamate release via TNFa: amplification by microglia triggers neurotoxicity,” *Nature Neuroscience*, vol. 4, no. 7, pp. 702–710, 2001.
- [47] T. Ravizza, B. Gagliardi, F. Noé, K. Boer, E. Aronica, and A. Vezzani, “Innate and adaptive immunity during epileptogenesis and spontaneous seizures: evidence from experimental models and human temporal lobe epilepsy,” *Neurobiology of Disease*, vol. 29, no. 1, pp. 142–160, 2008.
- [48] M. Morin-Brureau, A. Lebrun, M. Rousset et al., “Epileptiform activity induces vascular remodeling and zonula occludens 1 downregulation in organotypic hippocampal cultures: role of VEGF signaling pathways,” *The Journal of Neuroscience*, vol. 31, no. 29, pp. 10677–10688, 2011.
- [49] L. Librizzi, F. Noè, A. Vezzani, M. de Curtis, and T. Ravizza, “Seizure-induced brain-borne inflammation sustains seizure recurrence and blood-brain barrier damage,” *Annals of Neurology*, vol. 72, no. 1, pp. 82–90, 2012.
- [50] U. Heinemann, D. Kaufer, and A. Friedman, “Blood-brain barrier dysfunction, TGF β signaling, and astrocyte dysfunction in epilepsy,” *GLIA*, vol. 60, no. 8, pp. 1251–1257, 2012.
- [51] R. M. Ransohoff, P. Kivisäkk, and G. Kidd, “Three or more routes for leukocyte migration into the central nervous system,” *Nature Reviews Immunology*, vol. 3, no. 7, pp. 569–581, 2003.
- [52] H. Rantala, R. Tarkka, and M. Uhari, “Systematic review of the role of prostaglandins and their synthetase inhibitors with respect to febrile seizures,” *Epilepsy Research*, vol. 46, no. 3, pp. 251–257, 2001.
- [53] V. L. Marcheselli and N. G. Bazan, “Sustained induction of prostaglandin endoperoxide synthase-2 by seizures in hippocampus: inhibition by a platelet-activating factor antagonist,” *The Journal of Biological Chemistry*, vol. 271, no. 40, pp. 24794–24799, 1996.
- [54] T. Takemiya, K. Matsumura, and K. Yamagata, “Roles of prostaglandin synthesis in excitotoxic brain diseases,” *Neurochemistry International*, vol. 51, no. 2–4, pp. 112–120, 2007.

- [55] T. Takemiya, M. Maehara, K. Matsumura, S. Yasuda, H. Sugiura, and K. Yamagata, "Prostaglandin E₂ produced by late induced COX-2 stimulates hippocampal neuron loss after seizure in the CA3 region," *Neuroscience Research*, vol. 56, no. 1, pp. 103–110, 2006.
- [56] T. Kunz and E. H. Oliw, "Nimesulide aggravates kainic acid-induced seizures in the rat," *Pharmacology and Toxicology*, vol. 88, no. 5, pp. 271–276, 2001.
- [57] E. J. Baik, E. J. Kim, S. H. Lee, and C. Moon, "Cyclooxygenase-2 selective inhibitors aggravate kainic acid induced seizure and neuronal cell death in the hippocampus," *Brain Research*, vol. 843, no. 1-2, pp. 118–129, 1999.
- [58] G. E. Serrano, N. Lelutiu, A. Rojas et al., "Ablation of cyclooxygenase-2 in forebrain neurons is neuroprotective and dampens brain inflammation after status epilepticus," *Journal of Neuroscience*, vol. 31, no. 42, pp. 14850–14860, 2011.
- [59] M. K. Kaushik, K. Aritake, S. Kamauchi et al., "Prostaglandin D₂ is crucial for seizure suppression and postictal sleep," *Experimental Neurology*, vol. 253, pp. 82–90, 2014.
- [60] J. I. Chung, A. Y. Kim, S. H. Lee, and E. J. Baik, "Seizure susceptibility in immature brain due to lack of COX-2-induced PGF_{2α}," *Experimental Neurology*, vol. 249, pp. 95–103, 2013.
- [61] J. Jiang, Y. Quan, T. Ganesh, W. A. Pouliot, F. E. Dudek, and R. Dingledine, "Inhibition of the prostaglandin receptor EP2 following status epilepticus reduces delayed mortality and brain inflammation," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 110, no. 9, pp. 3591–3596, 2013.
- [62] J. Jiang, T. Ganesh, Y. Du et al., "Small molecule antagonist reveals seizure-induced mediation of neuronal injury by prostaglandin E2 receptor subtype EP2," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 109, no. 8, pp. 3149–3154, 2012.
- [63] M. S. Oliveira, A. F. Furian, L. M. Rambo et al., "Modulation of pentylenetetrazol-induced seizures by prostaglandin E2 receptors," *Neuroscience*, vol. 152, no. 4, pp. 1110–1118, 2008.
- [64] T. Takemiya, K. Matsumura, H. Sugiura et al., "Endothelial microsomal prostaglandin E synthase-1 exacerbates neuronal loss induced by kainate," *Journal of Neuroscience Research*, vol. 88, no. 2, pp. 381–390, 2010.
- [65] K. Yamagata, K. Matsumura, W. Inoue et al., "Coexpression of microsomal-type prostaglandin E synthase with cyclooxygenase-2 in brain endothelial cells of rats during endotoxin-induced fever," *Journal of Neuroscience*, vol. 21, no. 8, pp. 2669–2677, 2001.
- [66] T. Takemiya, K. Matsumura, H. Sugiura et al., "Endothelial microsomal prostaglandin synthase-1 facilitates neurotoxicity by elevating astrocytic Ca²⁺ levels," *Neurochemistry International*, vol. 58, no. 4, pp. 489–496, 2011.
- [67] M. Murakami, H. Naraba, T. Tanioka et al., "Regulation of prostaglandin E2 biosynthesis by inducible membrane-associated prostaglandin E2 synthase that acts in concert with cyclooxygenase-2," *Journal of Biological Chemistry*, vol. 275, no. 42, pp. 32783–32792, 2000.
- [68] P. Ciceri, Y. Zhang, A. F. Shaffer et al., "Pharmacology of celecoxib in rat brain after kainate administration," *The Journal of Pharmacology and Experimental Therapeutics*, vol. 302, no. 3, pp. 846–852, 2002.
- [69] M. G. S. S. Salvadori, C. R. R. Banderó, A. C. Jesse et al., "Prostaglandin E2 potentiates methylmalonate-induced seizures," *Epilepsia*, vol. 53, no. 1, pp. 189–198, 2012.
- [70] P. J. Cimino, C. D. Keene, R. M. Breyer, K. S. Montine, and T. J. Montine, "Therapeutic targets in prostaglandin E2 signaling for neurologic disease," *Current Medicinal Chemistry*, vol. 15, no. 19, pp. 1863–1869, 2008.
- [71] N. G. Carlson, M. A. Rojas, J. D. Black et al., "Microglial inhibition of neuroprotection by antagonists of the EP1 prostaglandin E2 receptor," *Journal of Neuroinflammation*, vol. 6, article 5, 2009.
- [72] J. N. Walters, J. S. Bickford, K. J. Newsom et al., "Regulation of human microsomal prostaglandin synthase-1 by IL-1 β requires a distal enhancer element with a unique role for C/EBP β ," *Biochemical Journal*, vol. 443, no. 2, pp. 561–571, 2012.
- [73] M. K. O'Banion, J. C. Miller, J. W. Chang, M. D. Kaplan, and P. D. Coleman, "Interleukin-1 β induces prostaglandin G/H synthase-2 (cyclooxygenase-2) in primary murine astrocyte cultures," *Journal of Neurochemistry*, vol. 66, no. 6, pp. 2532–2540, 1996.
- [74] C. Chen and N. G. Bazan, "Endogenous PGE2 regulates membrane excitability and synaptic transmission in hippocampal CA1 pyramidal neurons," *Journal of Neurophysiology*, vol. 93, no. 2, pp. 929–941, 2005.
- [75] T. Takemiya and K. Yamagata, "Intercellular signaling pathway among endothelia, astrocytes and neurons in excitatory neuronal damage," *International Journal of Molecular Sciences*, vol. 14, no. 4, pp. 8345–8357, 2013.
- [76] A. Vezzani and A. Friedman, "Brain inflammation as a biomarker in epilepsy," *Biomarkers in Medicine*, vol. 5, no. 5, pp. 607–614, 2011.
- [77] A. Friedman and R. Dingledine, "Molecular cascades that mediate the influence of inflammation on epilepsy," *Epilepsia*, vol. 52, no. 3, pp. 33–39, 2011.
- [78] D. Xu, S. D. Miller, and S. Koh, "Immune mechanisms in epileptogenesis," *Frontiers in Cellular Neuroscience*, vol. 7, article 195, 2013.
- [79] M. R. Weir, R. S. Sperling, A. Reicin, and B. J. Gertz, "Selective COX-2 inhibition and cardiovascular effects: a review of the rofecoxib development program," *The American Heart Journal*, vol. 146, no. 4, pp. 591–604, 2003.