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## Identification of Trisubstituted-pyrazol Carboxamide Analogs as Novel and Potent Antagonists of Farnesoid X Receptor

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### Abstract

Farnesoid X receptor (FXR, NRIH4) plays a major role in the control of cholesterol metabolism. This suggests that antagonizing the transcriptional activity of FXR is a potential means to treat cholestasis and related metabolic disorders. Here we describe the synthesis, biological evaluation, and structure-activity relationship (SAR) studies of trisubstituted-pyrazol carboxamides as novel and potent FXR antagonists. One of these novel FXR antagonists, **4j** has an IC<sub>50</sub> of 7.5 nM in an FXR binding assay and 468.5 nM in a cell-based FXR antagonistic assay. Compound **4j** has no detectable FXR agonistic activity or cytotoxicity. Notably, **4j** is the most potent FXR antagonist identified to date; it has a promising *in vitro* profile and could serve as an excellent chemical tool to elucidate the biological function of FXR.

### 1. INTRODUCTION

The activation of Farnesoid X receptor (FXR, NRIH4),<sup>1</sup> in particular, gives bile acids the ability to modulate genomic signaling pathways. FXR is a ligand-dependent transcription factor that regulates gene networks involved in regulating lipid and cholesterol homeostasis.<sup>2</sup> As such, FXR is expressed primarily in tissues exposed to high concentrations of bile acids, such as the intestine, kidney, adrenal gland, and liver.<sup>3</sup> Bile acids are the purported endogenous agonists for FXR.<sup>3–4</sup> As the bile acid sensor, FXR regulates the expression of transporters and biosynthetic enzymes crucial for the physiological maintenance of bile acid homeostasis. Because of FXR's role in bile acid homeostasis, modulating FXR may be beneficial for treating all aspects of the metabolic syndrome, a complex disease cluster that includes risk factors such as dyslipidemia, insulin-resistance,

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increased blood pressure, visceral obesity and hypercoagulability.<sup>5</sup> Recent findings also suggest that FXR acts as a key metabolic regulator in the liver to maintain the homeostasis of liver metabolites.<sup>6</sup> FXR ligands have been investigated in preclinical studies for targeted therapy of metabolic diseases, but have shown limitations.<sup>7</sup> There is, therefore, a need for novel, potent and selective modulators as agonists or antagonists of FXR, both for potential clinical applications, as well as studies to better understand FXR's biological functions.

Several potent, selective FXR agonists have been reported recently, such as GW4064<sup>8</sup> and 6 $\alpha$ -ethylchenodeoxycholic acid (6-ECDCA).<sup>9</sup> However, preclinical development of FXR agonists is limited by the complex response, including possible undesirable effects, triggered by activation of FXR in the liver. For example, FXR activation inhibits bile acid synthesis and basolateral efflux of bile through indirect downregulation of cholesterol 7  $\alpha$ -hydroxylase, or cytochrome P450 7A1 (CYP7A1), the rate-limiting enzyme of the bile acid synthesis pathway.<sup>10</sup> In addition, although activation of FXR would reduce triglyceride levels in hypertriglyceridemic patients, it would also decrease the levels of high density lipoprotein and lead to accumulation of cholesterol as a consequence of the inhibited bile acid synthesis. Furthermore, FXR agonists interfere with the ability of constitutive androstane receptor to regulate basolateral transporters in hepatocytes.<sup>11</sup> These effects might worsen liver injury in a subset of patients who have obstructive cholestasis (a severe liver disease that impairs bile flow and causes irreversible liver damage), thereby limiting the preclinical development and possible clinical use of FXR agonists. In contrast, FXR antagonists might be useful for understanding the function of FXR and ultimately for treating liver disorders, if they targeted a specific cluster of genes and thus avoided the detrimental side effects mediated by FXR agonists. Treatment with FXR antagonists could potentially upregulate CYP7A1 expression and decrease overall cholesterol concentrations. This activity would occur through FXR's regulation of the expression of small heterodimer partner.<sup>2</sup> In addition, FXR can promote expression of an intestinal bile acid-binding protein (*I-BABP*) gene.<sup>12</sup> When expression of *I-BABP* is repressed by an antagonist of FXR, reabsorption of bile acids in the ileum is repressed, which may reduce the amount of bile acid that returns to the liver, and, thereby promote the expression of CYP7A1. Thus, an FXR antagonist could potentially induce repression of *I-BABP* expression in the ileum, leading to reduced levels of serum cholesterol, and suggesting potential as a therapeutic agent for hypercholesterolemia in humans. Therefore, an FXR-specific antagonist could be used to validate the clinical relevance of antagonizing FXR.

Despite the potential of FXR as a therapeutic target for metabolic diseases, few selective FXR antagonists (synthetic small molecules or natural products) are described in the literature. Recently, the natural product guggulsterone was identified as the first putative FXR antagonist.<sup>13-14</sup> However, the mechanism by which it antagonizes FXR is unclear. It appears that rather than being a true antagonist of FXR, guggulsterone is a unique FXR ligand that has antagonistic activity in coactivator association assays and can enhance the action of FXR agonists *in vivo*.<sup>15</sup> In addition to guggulsterone, several marine products, such as sulfated polyhydroxy sterols isolated from marine invertebrates, have been identified as putative FXR modulators.<sup>16</sup> This is exemplified by sulfated sterol. Theonellasterol has also been reported as an FXR antagonist. In rodent models, theonellasterol attenuated both liver

injury and the extent of liver necrosis caused by bile duct ligation.<sup>17</sup> In addition, several scalarane sesterterpenes, isolated from a marine sponge, inhibited FXR with IC<sub>50</sub> values between 2.4 and 24 μM.<sup>18</sup> Most recently, suvanine, a marine sponge sesterterpene, was reported by Di Leva et al. as an FXR antagonist.<sup>19</sup> Using suvanine as a template, Di Leva et al. developed suvanine derivative as a novel FXR antagonist (Figure 1).

The nonsteroidal antagonist AGN34 displayed an interesting profile of antagonist properties of the RXR/FXR heterodimer acting at RXR. AGN43 antagonizes FXR *in vitro* reporter assays but acts in a gene-selective manner *in vivo*.<sup>20</sup> Interestingly, the nonsteroidal compound 1,3,4-trisubstituted-pyrazolone derivative was recently identified as an FXR antagonist (Figure 1).<sup>21</sup> This compound strongly suppresses the regulating effects of CDCA on FXR target genes. These findings further suggest the feasibility of identifying FXR antagonists that regulate a specific subset of FXR responsive genes in a gene-specific fashion.<sup>22</sup>

We recently reported the discovery of an FXR antagonist through the use of a time-resolved fluorescence resonance energy transfer (TR-FRET) assay,<sup>23</sup> as well as several chemotypes identified as inhibitors of FXR. These chemotypes are represented by heterocyclic amide compounds, including substituted pyrazole compounds.<sup>24 a–b</sup> The pyrazole heterocycle has long been considered a privileged scaffold in drug discovery on the basis of its wide-ranging biological activities, including modulating G-protein coupled receptor modulators, inhibiting cyclooxygenase-2, and p38 MAP kinase.<sup>25</sup> A challenge in investigating pyrazole heterocycles as potential FXR antagonists is selecting a structural motif to serve as the scaffold of a focused library, given the diversity of structures that can be optimized to develop more potent and selective FXR antagonists. Therefore we started by assessing the antagonistic effect of a collection of known substituted pyrazole carboxamides (Enamine Small Molecule Compound Libraries) against FXR, and identified E16, a 1,3-disubstituted -pyrazole-4-carboxamide as a potent FXR antagonist. We then used E16 as the template for further structural optimization, which led to several novel compounds. These compounds were exemplified by 1-(3-Methoxybenzyl)-3-(3-methoxyphenyl)-N-(4-methyl-3-(morpholinylsulfonyl) phenyl)-1H-pyrazole-4-carboxamide **4j**, which had significantly improved FXR antagonistic activities as compared to E16. To our knowledge, **4j** is the most potent FXR antagonist to date. Our efforts in designing structurally novel, nonsteroidal FXR antagonists by modifying pyrazole carboxamides may represent an important direction in developing novel FXR antagonists.

## 2. RESULTS and DISCUSSION

### 2.1. Identification of a Substituted Pyrazole-4-carboxamide E16 as a Template for Designing Novel FXR Antagonists

We began by evaluating the FXR antagonistic activities of known or commercially available substituted pyrazole carboxamides. We first evaluated 16 commercially available substituted pyrazole carboxamides (Enamine Small Molecule Compound Libraries) by FXR biochemical TR-FRET binding, FXR cell-based agonistic and antagonistic, and cytotoxicity assays (Tables 1 and 2). Among the carboxamide analogs, E16 (Table 1), a 1,3,4-trisubstituted-pyrazole carboxamide analog, had the highest FXR binding affinity (IC<sub>50</sub> = 47

nM) and was a potent FXR antagonist ( $IC_{50} = 2.62 \mu\text{M}$ ) (Table 2 and Figure 2). None of the substituted pyrazole carboxamides tested displayed FXR agonistic activity (data not shown) or significant cytotoxicity ( $< 5\%$  of control) (Table 2).

## 2.2. Design

With the novel substituted pyrazole-4-carboxamide framework in hand, we started to design a series of new trisubstituted-pyrazole carboxamide analogs to evaluate their receptor binding affinity and antagonistic potency in modulating the transcriptional activity of FXR. Recently, several crystal structures of the FXR-ligand binding domain (LBD) in complex with agonists were reported.<sup>26</sup> However, no structural information is available on the binding mode of these FXR antagonists nor are co-crystal structures of a substituted pyrazole carboxamide framework in complex with the LBD of FXR available. Therefore, we could not directly identify the structural feature of the substituted pyrazole carboxamide framework that interacts with the ligand-dependent activation function 2 motif of the FXR LBD<sup>27</sup>. However, the flexible nature of the FXR side chains suggests the FXR LBD may have considerable flexibility to accommodate ligands of various shapes by changing its conformation in response to ligand binding.<sup>28</sup> Huang *et al.* predicted the binding poses of the 1,3,4-substituted-pyrazolone analogs in the FXR ligand binding pocket by means of molecular docking to address the flexibility of the FXR LBD.<sup>21</sup> It is recognized that the chemical structure of 1,3,4-substituted-pyrazolone is the most similar to the substituted 1,3,4-substituted-pyrazole, even though their functions are diverse.

To understand the structural basis for the SAR, we analyzed the published binding poses of the 1,3,4-substituted-pyrazolone derivative (Figure 1). The binding model showed that the 1,3-substituted alkyl rings of the pyrazolone core are crucial for the hydrophobic interactions between the two phenyl rings and the two aromatic residues (Phe329 and His294), even though the antagonistic potency of the pyrazolone derivative for FXR is still relatively low ( $IC_{50} = 8.96 \pm 3.62 \mu\text{M}$ ).<sup>21</sup> The SAR analysis of the 1,3,4-substituted-pyrazolone derivative demonstrated that a subtle interplay between steric/hydrophobic effects of the 1,3-substituent of the pyrazolone core seemed to be critical for high FXR antagonistic potency. These observations suggest that varying the 1,3-substituents of the pyrazole core could be critical for high antagonistic potency.

On the basis of the detailed SAR studies for the 16 substituted pyrazole carboxamides (Tables 1 and 2), the substituted morpholinosulfonyl moiety of aniline on the 4-position appeared to be necessary to improve the antagonistic activity against FXR (E16 in Table 1). To increase the diversity of substituted moieties ( $R_1$ ,  $R_2$ , and  $R_3$  in Figure 3) and optimize the 1,3,4-trisubstituted pyrazole-based molecules to improve their antagonistic activity against FXR, we investigated the alternative deconstructive hypothesis for fragment binding.<sup>29</sup> Retrospectively, 1,3,4-trisubstituted-pyrazole carboxamides can be considered as being formed by two fragments: the substituted- $N^1$ -C3- pyrazole core (F1) and the substituted morpholinosulfonyl aniline moiety chain (F2) (Figure 3).

On the basis of our preliminary results (Tables 1 and 2) suggesting the substituted  $N^1$ -C3-pyrazole moiety is sensitive to activity, various hydrophobic residues, such as alkyl or methoxy residues, substituted in the  $R_1$  or  $R_2$  positions of the phenyl-ring might

significantly improve the antagonistic activity (Tables 1 and 2). We also determined whether the substituted  $N^1$ -C3- pyrazole core (F1) and morpholinosulfonyl aniline (F2) could bind separately to FXR. We selected three commercially available products, 2-or 4-methyl -3-(morpholinosulfonyl)aniline and 3-(morpholinosulfonyl)aniline as F2 functionalities for this evaluation. A series of substituted F1 fragments was prepared (described in Experimental Section; compounds 3a–3e). These fragments failed to show an antagonistic effect against FXR. However, by connecting F1 moieties to F2 moieties through a linker of a single covalent bond by an amide bridge, the resulting carboxamide compounds had dramatically improved antagonistic activity against FXR, exemplified by compounds in series 4 (Figure 4). This suggested that 1,3,4-trisubstituted-pyrazole carboxamides can be created as potent antagonists of FXR by linking the F1 and F2 fragments.

Because the F2 fragments have a hydrophobic region centered on the double hydrogen bond acceptor (sulfone) and had very weak antagonistic activity against FXR, it may provide considerable conformational rigidity to the receptor recognition after joining with F1 fragments.<sup>30</sup> Detailed SAR studies indicated that the position of the morpholinosulfonyl moiety of the F2 phenyl ring is important to modifications (Tables 1 and 2). Further analysis of the essential structural requirements for effective FXR antagonists showed that the meta-position-substituted morpholinosulfonyl moiety on the aniline amide moiety would influence the antagonistic activity. Therefore, we hypothesized that an appropriate 3-(morpholinosulfonyl)aniline moiety of the amide moiety needed to be fixed to act as a specific anchor during FXR binding recognition.

Variations of  $R_3$  on the F2 fragments could also affect the antagonistic potency. For example, introduction of a 4-methyl group on the F2 fragment resulted in greater antagonistic activity (Tables 1 and 2). Additionally, replacing the 4-methyl substituent on the aniline ring with a 2-methyl substituent conjugated with the same F1 resulted in lower antagonistic activities. Further, inclusion of an unmethylated 3-morpholinosulfonyl aniline substantially decreased the antagonistic potency (Tables 1 and 2). Taken together, this preliminary SAR analysis of a set of analogs provided important insight into the essential structural requirements that indicated 4-methyl-3-(morpholinosulfonyl)aniline in the F2 functionality would be an appropriate factor for effective FXR antagonists.

Guided by the SAR insights for the F2 fragments described above, we envisioned that F2 fragments might be a key pharmacophore region where an appropriate substituent, such as 4-methyl substituted aniline, would be crucial for hydrophobic interactions with FXR. We chose the 4-methyl -3-(morpholinosulfonyl)aniline as a fixed essential fragment and modified two regions of F1 fragments. We hypothesized that by subtly optimizing the steric/hydrophobic effects of the  $R_1$  substituents and fine-tuning the hydrogen-bonding capability of the  $R_2$  substituents, we could establish a focused library to develop novel FXR antagonists that could display high FXR binding affinity and potent cellular antagonistic effects against FXR.

### 2.3. Chemistry

**Synthesis of 1,3,4-Trisubstituted Pyrazole Nonsteroidal FXR Antagonists**—As indicated in Figure 4, the  $N^1$ - and C3-pyrazole moieties on the pyrazole core could be

explored with various electronic and hydrophobic substituted alkyls and phenyls. We hypothesized that potent analogs of 1,3-disubstituted-pyrazole-4-carboxamides may have structural features in common. From a parallel library synthesis perspective, compounds of this type are attractive because they contain only two vectors (denoted  $R_1$  and  $R_2$ ) that allow the introduction of considerable structural diversity. To explore novel carboxamide analogs of 1,3-disubstituted-pyrazole-4-carboxamides that could have high binding affinity and antagonistic effects against FXR, an efficient synthetic strategy was developed to optimize the  $N_1$ -C3-pyrazole core structure through substitutions at  $R_1$  and  $R_2$  as a focused two-component small array of pyrazole. Combining structural considerations and chemical feasibility, our initial strategy comprised traditional medicinal chemistry involving variations in both the electronic and steric properties of the appendages of the central pyrazole nucleus, which could act as a template to put substituents into the correct positions to fit the receptor. A scaffold expansion based on two diverse points given by the substituents and their positions on the pyrazole ring, as well as the substituents ( $R_1$  and  $R_2$ ) on the  $N^1$ ,C3- pyrazole rings, was therefore designed (Figures 5 and 6).

On the basis of the above design, two protocols for preparing the 1,3,4-trisubstituted-pyrazole carboxamides were developed to optimize the synthesis of compounds **4**. These protocols included preparing various structural isomers to determine whether the positions of the  $R_1$  and  $R_2$  could affect compound activity. The key intermediates **3** and **7** (Figures 5 and 6) could be prepared by two possible routes to allow access to final functional analogs. In the first route, lead optimization was initiated by exploring the phenyl functionality located on the  $N^1$ -position. A series of para- and meta-substitutions on the  $N^1$ - pyrazole ring was investigated (Figure 5). In general procedures for preparing phenyl hydrazones, synthesis often begins with commercially available substituted acetophenone **12** and substituted phenylhydrazines **11**. Condensation between acetophenone and substituted phenylhydrazines provides hydrazones **1**. A Vilsmeier-Haack reaction, which is the key reaction for the cyclization of hydrazones to pyrazoles, assembles the pyrazole nucleus scaffolds.<sup>31</sup> However, these conditions either do not form key aldehydes or have poor yields. We therefore modified the Vilsmeier-Haack reaction by adding the corresponding phenylhydrazones to an excess equivalence of  $\text{POCl}_3$  and DMF at 0 °C, then heated the reaction mixture to 60–70 °C with stirring over 18 h under  $\text{N}_2$  to afford the desired key cyclized aldehydes **2** at a 70–90% yield after hydrolysis. Oxidation of the substituted pyrazole-4-carboxaldehydes **2** with a Jones' reagent<sup>32</sup> yielded the corresponding substituted pyrazole-4-carboxylic acids **3**. Finally, the corresponding acids **3** were coupled with commercially available 4-methyl-3-(morpholinosulfonyl)aniline **13** through a Mitsunobu reaction<sup>33</sup> in the presence of cross-linking reagents (i.e., HOBt and EDCI) to produce the desired **4**. The synthetic pathways used to obtain the 5 analogs of 1,3,4-trisubstituted-1*H*-pyrazoles **4** are illustrated in Figure 5.

An alternative strategy to explore the SAR by introducing selected  $R_2$  functional groups to investigate the structure requirements for FXR antagonistic activity is shown in Figure 6. The introduction of  $R_2$  groups was carried out by treating commercially available  $\beta$ -keto esters **14** with triethyl orthoformate in the presence of  $\text{Ac}_2\text{O}$  as a catalyst to provide the corresponding ethoxymethylene intermediate **5**. Cyclization of **5** with hydrazine in dry

ethanol afforded the pyrazole core **6**. The *N*<sup>l</sup>-alkylation reaction of **6** with excess appropriate bromoalkyl and bromobenzyl was performed in the presence of cesium carbonate as the base and acetonitrile as solvent. The ethyl esters **6** were hydrolyzed under basic conditions to afford the corresponding carboxylic acid **7**. This was followed by a Mitsunobu coupling with 4-methyl-3-(morpholinomethyl)aniline in the presence of EDCI/DMAP as a fixed coupling reagent to produce the desired 14 products **4f–4s**. Notably, in this alternative pathway, the combination of an activating coupling agent (i.e., HOBt and EDCI) did not work well, so the combination of EDCI/DMAP as an alternative coupling reagent was used.

To explore the linker between the amide bridge and the aromatic core, we focused our efforts on modifying the chemical space in the amide moiety to examine if the amide bridge was sensitive to activity. Three amides of series **4** were altered to generate compounds **8**. We introduced a methyl moiety to assess the SAR. Compounds **4f**, **4m**, and **4o** were chosen to react with iodomethane in the presence of sodium hydride to give *N*-methylation compounds **8** (Figure 7). Because the amide position was hypothesized to be a site of potential metabolism,<sup>34</sup> further optimization involved reduction of amides. The amide groups in **4f**, **4m**, and **4o** were reduced to corresponding amines with 1N borane-THF complex to yield compounds **9** (Figure 7).

## 2.2. Biological Evaluation

**2.2.1. FXR TR-FRET Binding Assay**—The FXR binding affinities of E16, the 23 newly synthesized novel 1,3-trisubstituted-pyrazol-4-carboxamide analogs shown in Figures 5–7, and control compounds GW4064 and lithocholic acid (LCA) were tested through an *in vitro* FXR TR-FRET binding assay using DY246 as the fluorescent probe.<sup>23</sup> We chose TR-FRET assays because of their advantages of low background fluorescence interference and increased sensitivity as compared to other fluorescence-based assays, such as FI, FP and FRET assays.<sup>34</sup> We chose DY246 because it is a derivative of GW4064, which is a potent FXR agonist. In addition, DY246 acts as a potent FXR agonist (EC<sub>50</sub> of 550 nM) and has successfully been used as a fluorescent probe in a high throughput screening campaign to identify FXR antagonists.<sup>23</sup> In a representative assay, DMSO (vehicle and negative control), 10 μM GW4064 (positive control), and titrations of GW4064, LCA or other chemicals were incubated for 20 min with an appropriate mixture of GST-FXR-LBD, terbium (Tb)-anti-GST and DY246, after which the TR-FRET signals were collected and activity for each chemical was normalized to DMSO (negative control; 0% inhibition) and 10 μM GW4064 (positive control; 100% inhibition). The activities of compounds that displayed dose-dependent inhibitory effects were fit into a one-site competitive binding equation to derive IC<sub>50</sub> values. The TR-FRET binding activities are summarized in Tables 3, 4, and 5. Many of these compounds had significantly improved FXR binding affinity when compared to E16. Noticeably, **4j** had an IC<sub>50</sub> of 7.5 nM in the FXR TR-FRET binding assay, which was 5.26-fold more potent than E16 (IC<sub>50</sub> of 47 nM) (Tables 3, 4, and 5; Figure 8A).

**2.2.2. Cell-based FXR Transactivation Assay**—To investigate the cellular activities of the novel 1,3-substituted-pyrazol-4-carboxamide analogs, we used a GeneBLazer FXR cell-based assay, which detects the levels of a β-lactamase reporter controlled by a promoter containing the binding site of the DNA binding domain (DBD) of the GAL4 transcription

factor; the Gal4 DBD was fused to the LBD of FXR in this assay.<sup>36</sup> In typical test for agonists, DMSO (vehicle and negative control), 10  $\mu$ M GW4064 (positive control), and titrations of GW4064, LCA or the 23 newly synthesized novel 1,3-trisubstituted-pyrazol-4-carboxamide analogs were mixed with GeneBLAzer FXR-UAS-bla HEK 293T cells. Reporter assays were performed after a 16 h incubation. No agonistic activities were observed for any of the newly synthesized compounds (data not shown).

For the experiments to evaluate the antagonistic activity, titrations of compounds were tested in the presence of 400 nM GW4064. Assay controls included: DMSO alone (100% inhibition), 400 nM GW4064 alone (0% inhibition), and titrations of LCA in the presence of 400 nM GW4064 (as an assay reference). Activities of compounds that displayed activating or inhibitory activities in a dose-dependent manner were fit into a sigmoidal dose-response equation to derive individual EC<sub>50</sub> or IC<sub>50</sub> values. The tested chemicals' FXR activities are summarized in Tables 3, 4, and 5. Many of these compounds had significantly improved antagonistic activity when compared to E16. Noticeably, in the FXR antagonistic assay, **4j** had an IC<sub>50</sub> of 468.5 nM, which was 4.59-fold more potent than that of E16 (IC<sub>50</sub> of 2.62  $\mu$ M) (Tables 3, 4, and 5; Figure 8B).

**2.2.3. Cytotoxicity Assay**—For the FXR cell-based antagonistic assays, if a tested compound caused a signal reduction, it was usually considered an antagonist. However, signal reduction could also be caused non-specifically if a chemical was cytotoxic, although GeneBLAzer assays are relatively insensitive to cytotoxicity because of their use of ratiometric reading. To examine if certain chemical's antagonistic effect was caused by cytotoxicity, we used the same GeneBLAzer FXR cells in cytotoxicity assays. These assays were set up the same as for an FXR agonistic assay, with the exception of signal reading. Instead of detecting reporter activities, the CellTiter-Glo® assay was used to detect cell viability. Conditions including DMSO with or without cells were used as negative (0% inhibition) or positive (100% inhibition) controls, respectively. Cytotoxic activities are summarized in Tables 3, 4, and 5. All chemicals tested showed marginal toxicity (% inhibition < 5 %), therefore the antagonistic effect of the compounds was not caused by non-specific cytotoxicity.

### 2.3. SAR

We investigated the effect of introducing electron withdrawing groups, such as fluorine, chlorine, dichlorines, and trifluoromethyl atoms, at *ortho*, *meta*, or *para* positions of the *N*<sup>1</sup>-phenyl.<sup>37</sup> All of these derivatives (**4a**, **4b**, **4c**, **4d**, and **4e**) improved FXR antagonistic activity by about 3- to 4-fold as compared to E16 (Table 3), indicating that the electron withdrawing groups were well tolerated for antagonistic activity. This observation can be attributed to the higher interaction strength of fluorine or chlorine atoms compared to hydrogen, which enables a tighter fit.

Based on this initial SAR, we next introduced longer benzyl chains or small alkyl groups to replace the *N*<sup>1</sup>-phenyl of the pyrazole moiety to see if it was a suitable position to find more potent antagonists for FXR. The alternative synthetic methodology for constructing the corresponding pyrazole-4-carboxylic acids worked well. Replacing the phenyl ring with



small hydrophobic substituents such as isopropyl (**4f**) or n-butyl (**4g**) allowed slight retention of the antagonistic activity toward FXR. Substituting with a methylcyclopropane (**4i**) decreased the activity by 2-fold for FXR as compared to E16. Interestingly, incorporating a bulkier group, such as benzo[*d*]thiazole (**4e**), led to 3-fold higher potency for FXR. Other substitutions, including longer and bulkier benzyl aromatic substituents such as 2-naphthyl (**4h**) and benzyl with an electron donating group *m*-methoxy (**4j**), enhanced antagonistic activity. Among the *N*<sup>1</sup>-alkylated analogs, **4j** exhibited the most potent FXR-antagonistic activity, having IC<sub>50</sub> values of 7.5 nM and 468.5 nM in a FXR TR-FRET binding assay and a FXR cell-based antagonistic assay, respectively. In addition, **4j** had no FXR agonistic activity and no cytotoxicity. These results suggest that an expanded hydrophobic moiety is necessary at the *N*<sup>1</sup>-position and the positive influence of the *m*-methoxybenzyl group may be generally preferred at the *N*<sup>1</sup>-position of the pyrazole ring.

Replacing the original C3-*p*-methyl phenyl of **4j** with the stronger electron-donating *m*-methoxy phenyl group resulted in analog **4o**, which had slightly decreased antagonistic activity (Table 3). This indicated that the binding pocket of FXR that hosted the *m*-methoxy phenyl group on the C3 position of pyrazole is not sufficiently wide to accept larger substituents for hydrogen bond interactions with FXR. Notably, replacing the original of C3-*p*-methyl phenyl (R<sub>2</sub>) of **4j** with a smaller isopropyl group (analog **4m**) significantly decreased the potency and affinity, demonstrating that the C3 moiety is very sensitive in terms of steric bulkiness and/or hydrophobicity. For instance, introducing a *p*-trifluoromethyl phenyl with electron withdrawing properties at the C3 position of the pyrazole ring gave products **4p** and **4q**, which had nanomolar IC<sub>50</sub> values for *in vitro* potency. **4p** and **4q** appeared to have similar antagonistic activities as their parent compounds **4f** and **4o**. In contrast, replacing the *p*-trifluoromethyl phenyl at the C3 position of the pyrazole ring with a linear 2-methoxy-ethyl group to obtain compounds **4r** and **4s**, resulted in severe loss of potency relative to that of **4f** and **4o**, indicating that an alkyl methoxy in the C3 position of pyrazole ring is not beneficial. Therefore, we speculate that an aromatic phenyl ring at the C3-position of the pyrazole ring is required not only to optimally bind to a hydrophobic area of FXR, but is also critical for  $\pi$ - $\pi$  interaction with the receptor to increase FXR antagonistic activity. These examples demonstrate the favorable interaction that can be gained from fine-tuning the hydrophobic shape complementarily.

We also modified compounds **4f**, **4m**, and **4o** with amide moieties. The carboxamide nitrogens of **4f**, **4m**, and **4o** were methylated to form *N*-methyl analogs **8a**, **8b**, and **8c** (Figure 7). Unfortunately, *N*-methylation of the CONH function of amide compounds **8** decreased the antagonistic activity (Table 4), as compared to the activity of the original amides, **8a**, **8b**, and **8c** were essentially inactive at 1  $\mu$ M against FXR (data not shown). Similarly, compounds **9** had weak activity at 1  $\mu$ M, at least on FXR (data not shown). Compound **9** had more than 300-fold less affinity and potency as compared with **4j**. The discrepancies in activity of these analogs between *in vitro* and *in vivo* might be due to their stability, solubility and /or permeability.

### 3. CONCLUSIONS

FXR is a nuclear receptor that functions as a physiological receptor for bile acids. A highly potent and selective antagonist of FXR would represent a novel option that could prove useful for treating dyslipidemia and cholestasis. Thus, there is considerable need to identify lead structures and pharmacophore in this relatively unexplored area. We investigated the preliminary SAR-based optimization of 1,3,4-trisubstituted-pyrazole carboxamides as FXR antagonists. In an effort to develop specific FXR antagonists, a traditional medicinal chemistry parallel strategy was used. A series of 1,3,4-trisubstituted-pyrazole carboxamide analogues were developed and analyzed for physicochemical properties and SAR based on *in vitro* data. All of these modified compounds were evaluated for FXR binding affinities in biochemical assays and antagonistic activity in cell-based transactivation assays. All amide compounds reported here behaved as potent FXR antagonists, and did not have FXR agonistic activity (data not shown). Of interest, compound **4j** had IC<sub>50</sub> values of 7.5 nM and 468.5 nM in FXR TR-FRET binding assays and FXR cell-based antagonistic assays, respectively, suggesting it is a promising candidate FXR antagonist. In addition, compound **4j** does not have any FXR agonistic activity and cytotoxicity, suggesting that expansion of R<sub>1</sub> and R<sub>2</sub> on the pyrazole scaffold increases hydrogen bond acceptor interactions with FXR. This work has resulted in the most potent antagonist to date for FXR, **4j**, which may emerge as novel chemical tool to explore the effect of FXR inhibition in pharmacological settings. Studies aimed at further profiling **4j** *in vitro* and expanding the SAR of these analogs with the goal of developing FXR antagonists that can be delivered orally and are active *in vivo* are under way.

### 4. EXPERIMENTAL SECTION

#### 4.1. Chemistry

**General procedures**—Organic reagents were purchased from commercial suppliers unless otherwise noted and were used without further purification. All solvents were analytical or reagent grade. All reactions were carried out in flame-dried glassware under argon or nitrogen. Melting points were determined and reported automatically by an optoelectronic sensor in open capillary tubes and were uncorrected. <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra were measured at 500 MHz and 125 MHz, respectively, using CDCl<sub>3</sub> or CD<sub>3</sub>OD as solvents and tetramethylsilane (Me<sub>4</sub>Si) as the internal standard. Flash column chromatography was performed using Sigma-Aldrich silica gel 60 (200–400 mesh), carried out under moderate pressure with columns of an appropriate size packed and eluted with appropriate eluents. All reactions were monitored by thin layer chromatography (TLC) on precoated plates (silica gel HLF). TLC spots were visualized either by exposure to iodine vapor or by irradiation with UV light. Organic solvents were removed under vacuum by a rotary evaporator. Elemental analyses were performed by Columbia Analytical Services Inc, Tucson, Arizona.

**4.1.1. (E)-1-(4-fluorophenyl)-2-(1-p-tolylolethylidene)hydrazine (1a):** To a solution of 4'-methylacetophenone (0.5 g, 3.73 mmol) in 8 mL of acetic acid and 5 mL of MeOH, 4-fluorophenylhydrazine hydrochloride (0.6 g, 3.73 mmol) was added. The reaction mixture

was stirred at room temperature for 18 h. The reaction mixture was diluted with an additional 30 mL of water and subsequently filtered. The solid was dried *in vacuo* to afford 0.63 g of beige solid **1a** (70% yield): mp 101.8 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 7.68 (d, 2H), 7.19 (d, 3H), 7.07 (t, 2H), 6.98 (d, 2H), 2.37 (s, 3H), 2.23 (s, 3H).

**4.1.2. (E)-1-(3-Chlorophenyl)-2-(1-p-tolylolethylidene)hydrazine (1b):** To a solution of 4'-methylacetophenone (0.5 g, 3.73 mmol) in 8 mL of acetic acid and 5 mL of water/MeOH, 3-chlorophenylhydrazine hydrochloride (0.67 g, 3.73 mmol) was added. The reaction mixture was stirred at room temperature for 18 h. The reaction mixture was diluted with an additional 30 mL of water and subsequently filtered. The solid was dried *in vacuo* to afford 0.58 g of beige solid **1b** (60% yield): mp 100.5 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 7.68 (t, 2H), 7.20 (t, 5H), 6.98 (s, 1H), 6.83 (d, 1H), 2.38 (s, 3H), 2.28 (s, 3H).

**4.1.3. (E)-1-(2,4-dichlorophenyl)-2-(1-p-tolylolethylidene)hydrazine (1c):** To a solution of 4'-methylacetophenone (0.5 g, 3.73 mmol) in 8 mL of acetic acid and 5 mL of water/MeOH, 2,4-dichlorophenylhydrazine hydrochloride (0.8 g, 3.73 mmol) was added. The reaction mixture was stirred at room temperature for 18 h. The reaction mixture was diluted with an additional 30 mL of water and subsequently filtered. The solid was dried *in vacuo* to afford 0.98 g of beige solid **1c** (90% yield): mp 122.7 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 7.71 (t, 2H), 7.61 (d, 2H), 7.29 (s, 1H), 7.021(d, 3H), 2.38 (s, 3H), 2.28 (s, 3H).

**4.1.4. (E)-1-(3-Trifluoromethylphenyl)-2-(1-p-tolylolethylidene)hydrazine (1d):** To a solution of 4'-methylacetophenone (0.5 g, 3.73 mmol) in 8 mL of acetic acid and 5 mL of water/MeOH, 3-trifluoromethylphenylhydrazine hydrochloride (0.66 g, 3.73 mmol) was added. The reaction mixture was stirred at room temperature for 18 h. The reaction mixture was diluted with an additional 30 mL of water and subsequently filtered. The solid was dried *in vacuo* to afford 1.01 g of beige solid **1d** (93% yield): mp 103.7 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 7.72 (d, 2H), 7.44 (m, 4H), 7.28 (d, 2H), 7.14 (s, 1H), 2.41 (s, 3H), 2.27 (s, 3H).

**4.1.5. (E)-2-(2-(1-(p-tolyl)ethylidene)hydrazinyl)benzo[d]thiazole (1e):** To a solution of 4'-methylacetophenone (0.5 g, 3.73 mmol) in 8 mL of acetic acid and 1 mL of water, 2-hydrazinobenthiazole (0.62 g, 3.73 mmol) was added. The reaction mixture was stirred at room temperature for 18 h. The reaction mixture was diluted with an additional 20 mL of water and subsequently filtered. The solid was dried *in vacuo* to afford 1 g of beige solid **1e** (99% yield): mp 112.6 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 7.74 (d, 2H), 7.68 (d, 1H), 7.53 (d, 1H), 7.36 (d, 1H), 7.25 (d, 2H), 7.16 (d, 1H), 2.41 (s, 3H), 2.17 (s, 3H).

**4.1.6. 1-(4-fluorophenyl)-3-p-tolyl-1H-pyrazole-4-carbaldehyde (2a):** A solution of DMF (3 mL) and POCl<sub>3</sub> (1.5 mL, 16.4 mmol) was made in an ice bath under argon and stirred for 10 min. **1a** (0.21 g, 0.87 mmol) dissolved in dichloroethane (5 mL) was added to this solution; the mixture was warmed to 70 °C and stirred for an additional 18 h. The reaction mixture was then cooled to room temperature, slowly poured into saturated aqueous NaHCO<sub>3</sub> (200 mL) under ice cold conditions, and then warmed to room temperature and stirred for 3 h. The aqueous layer was extracted from the reaction mixture with EtOAc. The organic layers were washed with water and brine, dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated *in*

*vacuo* to yield 0.21 g (88%) of the title compound: mp 143.8 °C, which was used without further purification. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 10.05 (s, 1H), 8.47 (s, 1H), 7.78 (m, 1H), 7.71 (d, 2H), 7.33 (d, 2H), 7.19 (m, 3H), 2.44 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 187.0, 156.5, 132.7, 130.9, 130.2, 129.8, 123.9, 123.1, 118.1, 117.8, 23.3.

**4.1.7. 1-(3-Chlorophenyl)-3-*p*-tolyl-1*H*-pyrazole-4-carbaldehyde (2b):** A solution of DMF (3 mL) and POCl<sub>3</sub> (1.5 mL, 16.4 mmol) was made in an ice bath under argon and stirred for 10 min. **1b** (0.58 g, 2.24 mmol) dissolved in dichloroethane (5 mL) was added to this solution; the mixture was heated to 70 °C and stirred for an additional 18 h. The reaction mixture was then cooled to room temperature and slowly poured into saturated aqueous NaHCO<sub>3</sub> (200 mL) under ice cold conditions, then warmed to room temperature and stirred for 3 h. The aqueous layer was extracted with EtOAc. The organic layers were washed with water, brine, dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated *in vacuo* to yield 0.64 g (88%) of the title compound: mp 118.7 °C, which was used without further purification. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 10.05 (s, 1H), 8.53 (s, 1H), 7.88 (s, 1H), 7.67 (d, 3H), 7.46 (t, 1H), 7.31 (d, 3H), 2.44 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 186.5, 159.3, 141.1, 132.1, 130.9, 130.2, 129.3, 121.6, 118.8, 23.0.

**4.1.8. 1-(2,4-Dichlorophenyl)-3-*p*-tolyl-1*H*-pyrazole-4-carbaldehyde (2c):** A solution of DMF (3 mL) and POCl<sub>3</sub> (1.5 mL, 16.4 mmol) was made in an ice bath under argon and stirred for 10 min. **1c** (0.61 g, 2.08 mmol) dissolved in dichloroethane (5 mL) was added to this solution; the mixture was heated to 70 °C and stirred for an additional 18 h. The reaction mixture was then cooled to room temperature and slowly poured into saturated aqueous NaHCO<sub>3</sub> (200 mL) under ice cold conditions and then warmed to room temperature and stirred for 3 h. The aqueous layer was extracted with EtOAc. The organic layers were washed with water, brine, dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated *in vacuo* to yield 0.53 g (78%) of the title compound: mp 85.8 °C, which was used without further purification. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 10.05 (s, 1H), 8.48 (s, 1H), 7.66 (m, 3H), 7.54 (s, 1H), 7.42 (d, 1H), 7.37 (d, 2H), 2.38 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 186.6, 156.59 140.9, 137.4, 136.8, 134.1, 132.0, 131.7, 130.8, 129.4, 127.2, 105.7, 22.7.

**4.1.9. 3-*p*-Tolyl-1-(3-(trifluoromethyl)phenyl)-1*H*-pyrazole-4-carbaldehyde (2d):** A solution of DMF (3 mL) and POCl<sub>3</sub> (1.5 mL, 16.4 mmol) was made in an ice bath under argon and stirred for 10 min. **1d** (0.75 g, 2.56 mmol) dissolved in dichloroethane (7 mL) was added to this solution; the mixture was heated to 70 °C and stirred for an additional 18 h. The reaction mixture was then cooled to room temperature and slowly poured into saturated aqueous NaHCO<sub>3</sub> (200 mL) under ice cold conditions and then warmed to room temperature and stirred for 3 h. The aqueous layer was extracted with EtOAc. The organic layers were washed with water and brine, dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated *in vacuo* to yield 0.79 g (93%) of the title compound (mp 155.1 °C), which was used without further purification. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 10.08 (s, 1H), 8.60 (s, 1H), 8.13 (s, 1H), 7.99 (q, 1H), 7.74 (d, 2H), 7.67 (d, 2H), 7.35 (d, 2H), 2.45 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 186.6, 157.3, 141.7, 140.8, 132.3, 131.8, 130.9, 130.3, 126.6, 126.2, 123.8, 22.8.

**4.1.10. 1-(2,3-Dihydro-1H-inden-1-yl)-3-(p-tolyl)-1H-pyrazole-4-carbaldehyde (2f):**

A solution of DMF (3 mL) and POCl<sub>3</sub> (1.5 mL, 16.4 mmol) was made in an ice bath under argon and stirred for 10 min. **1f** (1.0 g, 3.55 mmol) dissolved in dichloroethane (5 mL) was added to the solution; the mixture was heated to 60 °C and stirred for an additional 18 h. The reaction mixture was then cooled to room temperature, slowly poured into saturated aqueous NaHCO<sub>3</sub> (200 mL) under ice cold conditions and then warmed to room temperature and stirred for an additional 5 h. The aqueous layer was extracted with EtOAc. The organic layers were washed with water, brine, dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated *in vacuo* to yield 0.74 g (70%) of the title compound: mp 145.3 °C, which was used without further purification. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 10.11 (s, 1H), 9.08 (s, 1H), 7.98 (d, 1H), 7.89 (d, 1H), 7.79 (d, 2H), 7.55 (t, 1H), 7.44 (t, 1H), 7.35 (d, 2H), 2.41 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 186.0, 157.1, 152.0, 141.5, 134.0, 130.9, 130.3, 128.3, 124.4, 123.2, 23.4.

**4.1.11. 1-(4-fluorophenyl)-3-p-tolyl-1H-pyrazole-4-carboxylic acid (3a):**

To a solution of **2a** (0.21 g, 0.75 mmol) dissolved in 16 mL of acetone, 5 mL of Jones' reagent was added. The reaction mixture was stirred at room temperature and monitored by TLC. The reaction mixture was poured into water and extracted with EtOAc. The combined organic layers were washed with water and brine, dried over Na<sub>2</sub>SO<sub>4</sub>, and concentrated *in vacuo* to afford 0.16 g (72%) of the title acid: mp 202.0 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.51 (s, 1H), 7.75 (m, 3H), 7.32 (d, 2H), 7.18 (t, 4H), 2.41 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 170.7, 164.5, 155.9, 140.6, 135.1, 132.6, 130.9, 130.6, 130.2, 130.1, 122.8, 118.2, 22.7. Anal. Calcd for C<sub>17</sub>H<sub>13</sub>FN<sub>2</sub>O<sub>2</sub>: C, 68.91; H, 4.41; N, 9.45. Found: C, 68.13; H, 4.43; N, 8.94.

**4.1.12. 1-(3-Chlorophenyl)-3-p-tolyl-1H-pyrazole-4-carboxylic acid (3b):**

To a solution of **2b** (0.51 g, 1.72 mmol) dissolved in 16 mL of acetone, 5 mL of Jones' reagent was added. The reaction mixture was stirred at room temperature and monitored by TLC. The reaction mixture was poured into water and extracted with EtOAc. The combined organic layers were washed with water and brine, dried over Na<sub>2</sub>SO<sub>4</sub>, and concentrated *in vacuo* to afford 0.51 g (95%) of the title acid: mp 194.4 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.57 (s, 1H), 7.86 (d, 1H), 7.77 (t, 2H), 7.66 (d, 1H), 7.44 (t, 1H), 7.34 (d, 1H), 7.28 (m, 2H), 2.41 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 168.8, 156.9, 147.7, 141.6, 140.3, 137.2, 134.7, 132.1, 130.6, 129.1, 121.3, 118.7, 22.8. Anal. Calcd for C<sub>17</sub>H<sub>13</sub>FCIN<sub>2</sub>O<sub>2</sub>: C, 65.29; H, 4.19; N, 8.96. Found: C, 64.65; H, 3.37; N, 8.80.

**4.1.13. 1-(2,4-Dichlorophenyl)-3-p-tolyl-1H-pyrazole-4-carboxylic acid (3c):**

To a solution of **2c** (0.53 g, 1.6 mmol) dissolved in 16 mL of acetone, 5 mL of Jones' reagent was added. The reaction mixture was stirred at room temperature and monitored by TLC. The reaction mixture was poured into water and extracted with EtOAc. The combined organic layers were washed with water and brine, dried over Na<sub>2</sub>SO<sub>4</sub>, and concentrated *in vacuo* to afford 0.51 g (91%) of the title acid: mp 211.7 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.52 (s, 1H), 7.75 (d, 2H), 7.66 (d, 1H), 7.58 (s, 1H), 7.41 (d, 1H), 7.24 (d, 2H), 2.43 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 171.6, 155.7, 139.3, 131.9, 130.6, 130.2, 134.7, 132.1, 130.6, 129.8, 22.7. Anal. Calcd for C<sub>17</sub>H<sub>12</sub>Cl<sub>2</sub>N<sub>2</sub>O<sub>2</sub>: C, 58.81; H, 3.48; N, 8.07. Found: C, 58.46; H, 3.40; N, 7.68.

**4.1.14. 3-(*p*-Tolyl)-1-(3-(trifluoromethyl)phenyl)-1*H*-pyrazole-4-carboxylic acid (3d):**

To a solution of **2d** (0.79 g, 2.39 mmol) dissolved in 16 mL of acetone, 5 mL of Jones' reagent was added. The reaction mixture was stirred at room temperature and monitored by TLC. The reaction mixture was poured into water and extracted with EtOAc. The combined organic layers were washed with water and brine, dried over Na<sub>2</sub>SO<sub>4</sub>, and concentrated *in vacuo* to afford 0.81 g (98%) of the title acid: mp 181.8 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.64 (s, 1H), 8.09 (s, 1H), 7.98 (d, 1H), 7.79 (d, 2H), 7.63 (d, 2H), 7.31 (t, 2H), 2.45 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 168.2, 155.7, 140.8, 140.5, 134.8, 131.7, 130.7, 130.2, 129.8, 125.5, 123.7, 117.9, 114.5, 22.8. Anal. Calcd for C<sub>18</sub>H<sub>13</sub>F<sub>3</sub>N<sub>2</sub>O<sub>2</sub>: C, 62.43; H, 3.78; N, 8.09. Found: C, 61.95; H, 3.74; N, 7.62.

**4.1.15. 1-(Benzo[d]thiazol-2-yl)-3-(*p*-tolyl)-1*H*-pyrazole-4-carboxylic acid (3e):**

To a solution of **2e** (0.71 g, 2.31 mmol) dissolved in 16 mL of acetone, 5 mL of Jones' reagent was added. The reaction mixture was stirred at room temperature and monitored by TLC. The reaction mixture was poured into water and extracted with EtOAc. The combined organic layers were washed with water and brine, dried over Na<sub>2</sub>SO<sub>4</sub>, and concentrated *in vacuo* to afford 0.41 g (57%) of the title acid: mp 208.0 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 9.09 (s, 1H), 7.98 (d, 1H), 7.90 (d, 1H), 7.81 (d, 2H), 7.54 (d, 1H), 7.43 (d, 1H), 7.35 (d, 2H), 2.45 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 171.2, 160.1, 151.6, 141.3, 134.0, 131.3, 130.3, 129.0, 126.9, 124.2, 123.5, 23.0.

**General Procedure of Mitsunobu Reaction**—EDCI and HOBT were added to a solution of **3** in CH<sub>2</sub>Cl<sub>2</sub>. The reaction mixture was stirred for 1–2 h. 4-methyl-3-(morpholinosulfonyl)aniline was added to the reaction and the reaction mixture was stirred at room temperature overnight and quenched with water. Layers were separated and the aqueous layer was extracted with EtOAc. The combined organic solution was dried over Na<sub>2</sub>SO<sub>4</sub> and then evaporated under a vacuum. The resulting yellow solids were purified by flash column chromatography to afford **4a–4e**.

**4.1.16. 1-(4-fluorophenyl)-*N*-(4-methyl-3-(morpholinosulfonyl)phenyl)-3-*p*-tolyl-1*H*-pyrazole-4-carboxamide (4a):**

To a solution of **3a** (0.2 g, 0.67 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (10 mL) was added EDCI (0.22 g, 1.16 mmol) and HOBT (0.16 g, 1.16 mmol). The reaction mixture was stirred for 1 h. To the reaction was added 4-methyl-3-(morpholinosulfonyl)aniline (0.17 g, 0.68 mmol). The reaction mixture was stirred at room temperature for 12 h and quenched with water. Layers were separated, and the aqueous layer was extracted with EtOAc. The combined organic solution was dried over Na<sub>2</sub>SO<sub>4</sub> and then evaporated under a vacuum. The resulting yellow solid was purified by flash column chromatography (Hexanes/EtOAc 1:1) to afford **4a** as a white solid (0.09 g, 26% yield), mp 95.9 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.52 (s, 1H), 7.73 (s, 1H), 7.72 (d, 2H), 7.64 (t, 2H), 7.54 (d, 2H), 7.36 (d, 2H), 7.27 (d, 3H), 3.72 (d, 4H), 3.17 (d, 4H), 2.57 (s, 3H), 2.47 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 162.1, 152.6, 141.4, 137.5, 137.4, 134.8, 134.6, 132.7, 131.2, 130.7, 130.2, 125.2, 122.7, 122.3, 119.5, 117.9, 117.7, 67.7, 46.8, 23.9, 22.6. Anal. Calcd for C<sub>28</sub>H<sub>27</sub>FN<sub>4</sub>O<sub>4</sub>S: C, 62.91; H, 5.09; N, 10.48; F, 3.55. Found: C, 62.75; H, 4.57; N, 10.46; F, 3.4.

**4.1.17. 1-(3-Chlorophenyl)-N-(4-methyl-3-(morpholinosulfonyl)phenyl)-3-(p-tolyl)-1H-pyrazole-4-carboxamide (4b):** To a solution of **3b** (0.2 g, 0.67 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (10 mL) was added EDCI (0.22 g, 1.16 mmol) and HOBT (0.15 g, 1.16 mmol). The reaction mixture was stirred for 1 h. To the reaction was added 4-methyl-3-(morpholinosulfonyl)aniline (0.17 g, 0.68 mmol). The reaction mixture was stirred at room temperature for 12 h and quenched with water. The organic layer was collected and the aqueous layer was extracted with EtOAc. The combined organic solution was dried over Na<sub>2</sub>SO<sub>4</sub>, evaporated under a vacuum. The resulting yellow solid was purified by flash column chromatography (H/EtOAc 1:1) to afford **4b** as a white solid (0.07 g, 19% yield), mp 204.9 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.60 (s, 1H), 7.88 (s, 1H), 7.72 (d, 1H), 7.62(m, 3H), 7.45 (d, 2H), 7.41 (m, 3H), 7.24 (d, 2H), 3.75 (d, 4H), 3.21 (d, 4H), 2.59 (s, 3H), 2.50 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 162.1, 152.7, 147.6, 141.7, 137.9, 134.8, 132.8, 132.0, 131.3, 130.7, 128.9, 125.1, 122.3, 121.3, 118.5, 67.7, 64.9, 23.0, 21.5. Anal. Calcd for C<sub>28</sub>H<sub>27</sub>ClN<sub>4</sub>O<sub>4</sub>S: C, 61.03; H, 4.94; N, 10.17; Cl, 6.43. Found: C, 60.78; H, 4.61; N, 9.94; Cl, 6.7.

**4.1.18. 1-(2,4-Dichlorophenyl)-N-(4-methyl-3-(morpholinosulfonyl)phenyl)-3-(p-tolyl)-1H-pyrazole-4-carboxamide (4c):** To a solution of **3c** (0.2 g, 0.58 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (5 mL) was added EDCI (0.22 g, 1.16 mmol) and HOBT (0.16 g, 1.16 mmol). The reaction mixture was stirred for 1 h. To the reaction was added 4-methyl-3-(morpholinosulfonyl)aniline (0.15 g, 0.58 mmol). The reaction mixture was stirred at room temperature for 12 h and quenched with water. The organic layer was collected and the aqueous layer was extracted with EtOAc. The combined organic solution was dried over Na<sub>2</sub>SO<sub>4</sub>, evaporated under a vacuum. The resulting yellow solid was purified by flash column chromatography (H/EtOAc 1:1) to afford **4c** as a white solid (0.06 g, 20% yield), mp 99.8 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.52 (s, 1H), 7.73 (s, 1H), 7.62 (m, 3H), 7.45 (d, 2H), 7.38 (m, 3H), 7.24 (d, 2H), 3.73 (d, 4H), 3.19 (d, 4H), 2.57 (s, 3H), 2.46 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 162.0, 152.6, 141.9, 139.9, 137.7, 134.7, 134.3, 132.0, 131.3, 130.8, 129.5, 127.4, 125.2, 122.0, 118.3, 116.6, 67.7, 46.8, 22.9, 21.8. Anal. Calcd for C<sub>28</sub>H<sub>26</sub>Cl<sub>2</sub>N<sub>4</sub>O<sub>4</sub>S: C, 57.44; H, 4.48; N, 9.57; Cl, 12.22. Found: C, 57.11; H, 4.27; N, 8.91; Cl, 12.2.

**4.1.19. N-(4-Methyl-3-(morpholinosulfonyl)phenyl)-3-(p-tolyl)-1-(3-(trifluoromethyl)phenyl)-1H-pyrazole-4-carboxamide (4d):** To a solution of **3d** (0.2 g, 0.58 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (10 mL) was added EDCI (0.34 g, 1.76 mmol) and HOBT (0.24 g, 1.76 mmol). The reaction mixture was stirred for 1 h. To the reaction was added 4-methyl-3-(morpholinosulfonyl)aniline (0.2 g, 0.78 mmol). The reaction mixture was stirred at room temperature for 16 h and quenched with water. The organic layer was collected and the aqueous layer was extracted with EtOAc. The combined organic solution was dried over Na<sub>2</sub>SO<sub>4</sub>, evaporated under a vacuum. The resulting yellow solid was purified by flash column chromatography (H/EtOAc 1:1) to afford **4d** as a white solid (0.1 g, 31% yield), mp 192.7 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.67 (s, 1H), 8.12 (s, 1H), 7.92 (d, 1H), 7.72 (s, 1H), 7.64 (d, 4H), 7.51 (t, 3H), 7.41 (d, 2H), 3.74 (d, 4H), 3.19 (d, 4H), 2.58 (s, 3H), 2.50 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 165.0, 151.6, 141.9, 139.9, 137.7, 134.7, 134.3, 132.0, 131.3, 130.8, 129.5, 127.4, 125.2, 124.1, 118.3, 116.6, 67.8, 46.9, 23.9, 23.6. Anal. Calcd for C<sub>29</sub>H<sub>27</sub>F<sub>3</sub>N<sub>4</sub>O<sub>4</sub>S: C, 59.58; H, 4.66; N, 9.58; F, 9.75. Found: C, 60.07; H, 4.38; N, 9.53; F, 10.

**4.1.20. 1-(Benzo[d]thiazol-2-yl)-N-(4-methyl-3-(morpholinofonyl)phenyl)-3-(p-tolyl)-1H-pyrazole-4-carboxamide (4e):** To a solution of **3e** (0.2 g, 0.59 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (5 mL) was added EDCI (0.14 g, 0.71 mmol) and HOBT (0.1 g, 0.7 mmol). The reaction mixture was stirred for 1 h. To the reaction was added 4-methyl-3-(morpholinofonyl)aniline (0.15 g, 0.59 mmol). The reaction mixture was stirred at room temperature for 12 h and quenched with water. The organic layer was collected and the aqueous layer was extracted with EtOAc. The combined organic solution was dried over Na<sub>2</sub>SO<sub>4</sub>, evaporated under a vacuum. The resulting yellow oil was purified by flash column chromatography (H/EtOAc 1:1) to afford **4e** as a pale-yellow solid (0.07 g, 21% yield), mp 153.1 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.51 (s, 1H), 8.02 (s, 1H), 7.97 (d, 2H), 7.79 (d, 1H), 7.71 (t, 2H), 7.55 (t, 1H), 7.40 (dd, 4H), 3.74 (d, 4H), 3.19 (d, 4H), 2.63 (s, 3H), 2.44 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 165.0, 159.6, 144.5, 135.4, 137.7, 131.3, 131.0, 130.7, 130.2, 128.5, 126.1, 123.1, 123.0, 67.7, 47.7, 23.0, 21.7. Anal. Calcd for C<sub>29</sub>H<sub>27</sub>N<sub>5</sub>O<sub>4</sub>S<sub>2</sub>: C, 60.71; H, 4.74; N, 12.21. Found: C, 60.57; H, 4.2; N, 12.6.

**General Procedure of Mitsunobu Reaction for producing 4f–4s**—EDCI and DMAP were added to a solution of **7** in CH<sub>2</sub>Cl<sub>2</sub>. To the reaction was added 4-methyl-3-(morpholinofonyl)aniline. The reaction mixture was stirred at room temperature for 6 h and quenched with water and then treated with 10% HCl. The organic layer was collected, and the aqueous layer was extracted with EtOAc. The combined organic solution was dried over Na<sub>2</sub>SO<sub>4</sub>, evaporated under a vacuum. The resulting yellow oil was purified by flash column chromatography to afford **4f–4s**.

#### Selected examples

**4.1.21. 1-Isopropyl-N-(4-methyl-3-(morpholinofonyl)phenyl)-3-(p-tolyl)-1H-pyrazole-4-carboxamide (4f):** To a solution of **7a** (0.11 g, 0.43 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (5 mL) was added EDCI (0.14 g, 0.73 mmol) and DMAP (0.09 g, 0.73 mmol). To the reaction was added 4-methyl-3-(morpholinofonyl)aniline (0.11 g, 0.43 mmol). The reaction mixture was stirred at room temperature for 6 h, quenched with water and then treated with 10% HCl. The organic layer was collected, and the aqueous layer was extracted with EtOAc. The combined organic solution was dried over Na<sub>2</sub>SO<sub>4</sub>, evaporated under a vacuum. The resulting yellow oil was purified by flash column chromatography (H/EtOAc 1:1) to afford **4f** as a white solid (0.11 g, 50% yield), mp 198.2 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.13 (s, 1H), 7.69 (s, 1H), 7.54 (d, 2H), 7.44 (d, 2H), 7.35 (d, 2H), 7.26 (d, 1H), 4.55 (m, 1H), 3.74 (d, 4H), 3.17 (d, 4H), 2.56 (s, 3H), 2.46 (s, 3H), 1.59 (d, 6H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 163.2, 145.3, 140.8, 135.2, 134.7, 130.3, 130.1, 126.1, 121.8, 120.9, 117.9, 109.9, 67.7, 56.5, 46.7, 24.1, 22.9, 21.1. Anal. Calcd for C<sub>25</sub>H<sub>30</sub>N<sub>4</sub>O<sub>4</sub>S · ½ H<sub>2</sub>O: C, 61.07; H, 6.15; N, 11.39. Found: C, 61.26; H, 6.31; N, 11.29.

**4.1.22. 1-Butyl-N-(4-methyl-3-(morpholinofonyl)phenyl)-3-(p-tolyl)-1H-pyrazole-4-carboxamide (4g):** To a solution of **7b** (0.2 g, 0.77 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (10 mL) was added EDCI (0.18 g, 0.92 mmol) and HOBT (0.12 g, 0.92 mmol). The reaction mixture was stirred for 1 h. To the reaction was added 4-methyl-3-(morpholinofonyl)aniline (0.2 g, 0.77 mmol). The reaction mixture was stirred at room temperature for 12 h and quenched with water. The organic layer was collected, and the aqueous layer was extracted with EtOAc.



The combined organic solution was dried over Na<sub>2</sub>SO<sub>4</sub>, evaporated under a vacuum. The resulting yellow oil was purified by flash column chromatography (H/EtOAc 1:1) to afford **4g** as a white solid (0.2 g, 53% yield), mp 177.4 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.07 (s, 1H), 7.69 (s, 1H), 7.54 (d, 2H), 7.44 (d, 2H), 7.35 (d, 2H), 7.23 (s, 1H), 4.18 (m, 2H), 3.74 (d, 4H), 3.17 (d, 4H), 2.56 (s, 3H), 2.46 (s, 3H), 1.92 (t, 2H), 1.41 (t, 2H), 0.98 (t, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 162.7, 151.3, 140.9, 137.6, 134.8, 131.7, 131.5, 130.7, 125.2, 122.1, 120.9, 116.9, 67.7, 53.9, 46.8, 33.5, 22.8, 21.4, 21.1, 14.9. Anal. Calcd for C<sub>26</sub>H<sub>32</sub>N<sub>4</sub>O<sub>4</sub>S: C, 62.88; H, 6.49; N, 11.28. Found: C, 62.65; H, 6.27; N, 11.08.

**4.1.23. *N*-(4-Methyl-3-(morpholinosulfonyl)phenyl)-1-(naphthalen-2-ylmethyl)-3-(*p*-tolyl)-1*H*-pyrazole-4-carboxamide (4h):** To a solution of **7c** (0.2 g, 0.78 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (10 mL) was added EDCI (0.18 g, 0.94 mmol) and DMAP (0.11 g, 0.94 mmol). To the reaction was added 4-methyl-3-(morpholinosulfonyl)aniline (0.2 g, 0.78 mmol). The reaction mixture was stirred at room temperature for 6 h and then treated with 10% HCl. The organic layer was collected, and the aqueous layer was extracted with EtOAc. The combined organic solution was dried over Na<sub>2</sub>SO<sub>4</sub>, evaporated under a vacuum. The resulting yellow oil was purified by flash column chromatography (H/EtOAc 1:1) to afford **4h** as a white solid (0.2 g, 50% yield), mp 144.9 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.13 (s, 1H), 7.85 (m, 4H), 7.68 (s, 1H), 7.53 (m, 4H), 7.45 (m, 2H), 7.35 (d, 2H), 7.20 (d, 2H), 5.51 (s, 2H), 3.72 (t, 4H), 3.15 (t, 4H), 2.51 (s, 3H), 2.46 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 162.2, 151.2, 141.2, 137.4, 136.9, 135.6, 134.8, 133.7, 131.3, 130.8, 130.4, 129.4, 129.2, 129.1, 128.1, 127.1, 125.1, 122.1, 67.8, 58.4, 46.9, 22.9, 21.6. Anal. Calcd for C<sub>33</sub>H<sub>32</sub>N<sub>4</sub>O<sub>4</sub>S: C, 68.25; H, 5.55; N, 9.65. Found: C, 68.00; H, 6.04; N, 9.48.

**4.1.24. 1-(Cyclopropylmethyl)-*N*-(4-methyl-3-(morpholinosulfonyl)phenyl)-3-(*p*-tolyl)-1*H*-pyrazole-4-carboxamide (4i):** To a solution of **7d** (0.2 g, 0.78 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (10 mL) was added EDCI (0.18 g, 0.94 mmol) and DMAP (0.11 g, 0.94 mmol). To the reaction was added 4-methyl-3-(morpholinosulfonyl)aniline (0.2 g, 0.78 mmol). The reaction mixture was stirred at room temperature for 6 h and then treated with 10% HCl. The organic layer was collected, and the aqueous layer was extracted with EtOAc. The combined organic solution was dried over Na<sub>2</sub>SO<sub>4</sub>, evaporated under a vacuum. The resulting yellow solid was purified by crystallization from EtOAc gave **4i** as a white solid (0.3 g, 78% yield), mp 210.0 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.23 (s, 1H), 7.71 (d, 1H), 7.54 (d, 2H), 7.46 (d, 2H), 7.35 (d, 2H), 7.21 (d, 1H), 4.05 (d, 2H), 3.74 (t, 4H), 3.17 (t, 4H), 2.51 (s, 3H), 1.36 (brs, 1H), 0.74 (q, 2H), 0.45 (t, 2H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 162.5, 150.8, 141.0, 137.6, 137.1, 131.3, 130.8, 125.0, 122.1, 67.8, 58.9, 46.9, 23.2, 21.9, 12.0, 5.6. Anal. Calcd for C<sub>26</sub>H<sub>30</sub>N<sub>4</sub>O<sub>4</sub>S · ¼ H<sub>2</sub>O: C, 61.98; H, 6.00; N, 11.02. Found: C, 61.63; H, 5.82; N, 10.84.

**4.1.25. 1-(3-Methoxybenzyl)-*N*-(4-methyl-3-(morpholinosulfonyl)phenyl)-3-(*p*-tolyl)-1*H*-pyrazole-4-carboxamide (4j, DY268):** To a solution of **7e** (0.1 g, 0.31 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (8 mL) was added EDCI (0.071 g, 0.37 mmol) and DMAP (0.045 g, 0.37 mmol). To the reaction was added 4-methyl-3-(morpholinosulfonyl)aniline (0.08 g, 0.31 mmol). The reaction mixture was stirred at room temperature for 6 h and then treated with 10% HCl. The organic layer was collected, and the aqueous layer was extracted with EtOAc. The combined organic solution was dried over Na<sub>2</sub>SO<sub>4</sub>, evaporated under a vacuum. The resulting yellow

oil was purified by flash column chromatography (H/EtOAc 1:1) to afford **4j** as a white solid (0.08 g, 47% yield), mp 183.3 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.01 (s, 1H), 7.69 (s, 1H), 7.55 (d, 2H), 7.44 (d, 2H), 7.34 (m, 3H), 7.22 (d, 2H), 6.93 (d, 1H), 6.87 (s, 1H), 5.32 (s, 2H), 3.85 (s, 3H), 3.74 (t, 4H), 3.17 (t, 4H), 2.56 (s, 3H), 2.46 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 162.6, 160.0, 141.4, 137.1, 135.5, 134.8, 131.5, 131.3, 130.8, 125.1, 122.1, 115.5, 115.4, 67.7, 58.0, 56.7, 46.8, 22.8, 21.6. Anal. Calcd for C<sub>30</sub>H<sub>32</sub>N<sub>4</sub>O<sub>5</sub>S: C, 64.27; H, 5.75; N, 9.99. Found: C, 64.01; H, 5.77; N, 9.63.

**4.1.26. *N*-(4-methyl-3-(morpholinofonyl)phenyl)-3-(*p*-tolyl)-1-(3-**

**(trifluoromethyl)benzyl)-1*H*-pyrazole-4-carboxamide (4k):** To a solution of **7f** (0.11 g, 0.34 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (5 mL) was added EDCI (0.078 g, 0.41 mmol) and DMAP (0.05 g, 0.41 mmol). To the reaction was added 4-methyl-3-(morpholinofonyl)aniline (0.087 g, 0.34 mmol). The reaction mixture was stirred at room temperature for 6 h and then treated with 10% HCl. The organic layer was collected and the aqueous layer was extracted with EtOAc. The combined organic solution was dried over Na<sub>2</sub>SO<sub>4</sub>, evaporated under a vacuum. The resulting yellow solid was purified by crystallization from EtOAc gave **4k** as a white solid (0.1 g, 56% yield), mp 179.3 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.07 (s, 1H), 7.68 (s, 1H), 7.59 (s, 1H), 7.51 (t, 4H), 7.45 (t, 2H), 7.43 (d, 2H), 7.21 (d, 2H), 5.41 (s, 2H), 3.72 (t, 4H), 3.15 (t, 4H), 2.51 (s, 3H), 2.46 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 162.4, 151.6, 141.4, 137.3, 135.7, 134.8, 134.6, 132.8, 131.3, 130.8, 126.2, 125.1, 122.2, 118.4, 67.7, 57.4, 46.8, 22.8, 21.6. Anal. Calcd for C<sub>30</sub>H<sub>29</sub>F<sub>3</sub>N<sub>4</sub>O<sub>4</sub>S: C, 60.19; H, 4.88; N, 9.36, F, 9.52. Found: C, 60.45; H, 4.70; N, 9.15, F, 9.20.

**4.1.27. 1-(4-Hydroxybutyl)-*N*-(4-methyl-3-(morpholinofonyl)phenyl)-3-(*p*-tolyl)-1*H*-**

**pyrazole-4-carboxamide (4l):** To a solution of **7g** (0.09 g, 0.34 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (5 mL) was added EDCI (0.078 g, 0.41 mmol) and DMAP (0.05 g, 0.41 mmol). To the reaction was added 4-methyl-3-(morpholinofonyl)aniline (0.087 g, 0.34 mmol). The reaction mixture was stirred at room temperature for 6 h and then treated with 10% HCl. The organic layer was collected, and the aqueous layer was extracted with EtOAc. The combined organic solution was dried over Na<sub>2</sub>SO<sub>4</sub>, evaporated under a vacuum. The resulting yellow oil was purified by flash column chromatography (CHCl<sub>3</sub>/MeOH 95:5) to afford **4l** as a white solid (0.05 g, 27% yield), mp 57.8 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.01 (s, 1H), 7.68 (s, 1H), 7.50 (d, 2H), 7.49 (t, 1H), 7.15 (t, 4H), 4.21 (d, 2H), 4.20 (d, 2H), 3.72 (t, 4H), 3.11 (t, 4H), 2.51 (s, 3H), 2.46 (s, 3H), 1.94 (t, 2H), 1.26 (t, 2H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 162.2, 154.9, 141.3, 139.8, 137.6, 135.2, 134.8, 131.8, 131.3, 130.6, 129.9, 125.2, 122.3, 67.7, 63.3, 53.8, 46.8, 30.7, 27.2, 21.6. Anal. Calcd for C<sub>26</sub>H<sub>32</sub>N<sub>4</sub>O<sub>4</sub>S: C, 60.71; H, 4.74; N, 12.21. Found: C, 60.57; H, 4.2; N, 12.26.

**4.1.28. (Z)-Ethyl 3-ethoxy-2-(4-methylbenzoyl)acrylate (5a):** Ethyl 3-oxo-3-(*p*-tolyl)propanoate (1.0 g, 4.85 mmol) and triethylorthoformate (1.15 g, 7.76 mmol) were heated to reflux and stirred for 30 min. Acetic anhydride (1.5 g, 14.55 mmol) was then added, and refluxing continued for 12 h. Then the mixture was cooled and diluted with EtOAc, and water was added to the solution, which was then stirred for 10 min to decompose excess triethylorthoformate. The aqueous layer was extracted with EtOAc. The organic layers were washed with water and brine, dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated *in vacuo* to yield 1.21 g

(94%) of the title compound, which was used without further purification.  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  7.82 (d, 2H), 7.69 (s, 1H), 7.27 (d, 2H), 4.17 (q, 2H), 4.11 (q, 2H), 2.40 (s, 3H), 1.28 (t, 3H), 1.17 (t, 3H).  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta$  192.9, 162.7, 145.5, 136.2, 130.8, 130.5, 130.3, 113.7, 73.1, 61.9, 23.1, 16.6, 15.6.

**4.1.29. (Z)-Ethyl 2-(ethoxymethylene)-4-methyl-3-oxopentanoate (5b):** Ethyl isobutyryl acetate (2.0 g, 12.64 mmol) and triethylorthoformate (3.0 g, 20.22 mmol) were heated to reflux for 30 min. Acetic anhydride (3.8 g, 37.92 mmol) was then added, and refluxing continued for about 12 h. The reaction mixture was cooled and diluted with EtOAc, and water was added to the solution, which was then stirred for 10 min to decompose excess triethylorthoformate. The aqueous layer was extracted with EtOAc. The organic layers were washed with water and brine, dried over  $\text{Na}_2\text{SO}_4$  and concentrated *in vacuo* to yield 2.49 g (92%) of the title compound, which was used without further purification.  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  7.52 (s, 1H), 4.29 (q, 2H), 4.19 (q, 2H), 3.14 (m, 1H), 1.37 (t, 3H), 1.32 (t, 3H), 1.16 (s, 3H), 1.11 (s, 3H).  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta$  203.4, 165.8, 163.1, 114.7, 73.4, 62.2, 42.1, 20.2, 19.3, 16.6, 15.6.

**4.1.30. (Z)-Ethyl 3-ethoxy-2-(3-methoxybenzoyl)acrylate (5c):** Ethyl (3-methoxybenzoyl) acetate (1.0 g, 4.5 mmol) and triethylorthoformate (1.07 g, 7.2 mmol) were heated to reflux for 30 min. Acetic anhydride (1.37 g, 13.5 mmol) was then added, and refluxing continued for about 12 h. The reaction mixture was cooled and diluted with EtOAc, and water was added to the solution, which was then stirred for 10 min to decompose excess triethylorthoformate. The aqueous layer was extracted with EtOAc. The organic layers were washed with water and brine, dried over  $\text{Na}_2\text{SO}_4$  and concentrated *in vacuo* to yield 1.1 g (88%) of the title compound, which was used without further purification.  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  7.68 (s, 1H), 7.47 (t, 2H), 7.35 (t, 1H), 7.11 (t, 1H), 4.17 (q, 2H), 4.11 (q, 2H), 1.16 (t, 3H), 1.06 (t, 3H).  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta$  193.3, 167.1, 162.9, 161.2, 140.1, 130.7, 130.5, 123.7, 121.4, 114.1, 73.2, 62.0, 56.8, 16.6, 15.6.

**4.1.31. (Z)-Ethyl 3-ethoxy-2-(3-(trifluoromethyl)benzoyl)acrylate (5d):** Ethyl (4-trifluoromethylbenzoyl) acetate (1.0 g, 3.84 mmol) and triethylorthoformate (0.91 g, 6.15 mmol) were heated to reflux for 30 min. Acetic anhydride (1.18 g, 11.53 mmol) was then added, and refluxing continued for about 12 h. The reaction mixture was cooled and diluted with EtOAc, and water was added to the solution, which was then stirred for 10 min to decompose excess triethylorthoformate. The aqueous layer was extracted with EtOAc. The organic layers were washed with water and brine, dried over  $\text{Na}_2\text{SO}_4$  and concentrated *in vacuo* to yield 1.08 g (89%) of the title compound, which was used without further purification.  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  8.06 (d, 1H), 7.97 (d, 1H), 7.87 (s, 1H), 7.74 (d, 2H), 4.16 (q, 2H), 4.11 (q, 2H), 1.27 (t, 6H).  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta$  193.2, 164.1, 130.7, 130.3, 127.8, 127.3, 126.8, 114.1, 73.6, 62.1, 16.6, 15.5.

**4.1.32. (Z)-Methyl 2-(ethoxymethylene)-5-methoxy-3-oxopentanoate (5e):** Methyl 5-methoxy-3-oxovalerate (2.0 g, 12.48 mmol) and triethylorthoformate (3.0 g, 19.96 mmol) were heated to reflux for 30 min. Acetic anhydride (3.82 g, 37.44 mmol) was then added, and refluxing continued for about 6 h. The reaction mixture was cooled and diluted with

EtOAc, and water was added to the solution, which was then stirred for 10 min to decompose excess triethylorthoformate. The aqueous layer was extracted with EtOAc. The organic layers were washed with water and brine, dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated *in vacuo* to yield 2.27 g (84%) of the title compound, which was used without further purification. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 7.90 (s, 1H), 4.25 (q, 2H), 3.66 (t, 2H), 3.35 (s, 3H), 3.23 (t, 2H), 1.31 (t, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 200.5, 185.9, 165.7, 108.1, 67.7, 67.6, 60.3, 51.4, 38.8, 15.2.

**4.1.33. Ethyl 3-(*p*-tolyl)-1*H*-pyrazole-4-carboxylate (6a):** A solution of hydrazine monohydrate (0.2 ml, 4.2 mmol) in dry ethanol (1 mL) was added dropwise at 0 °C to a solution of (*Z*)-Ethyl 3-ethoxy-2-(4-methylbenzoyl)acrylate (**5a**) (1.0 g, 3.81 mmol) in ethanol (5 mL). The reaction mixture was stirred at room temperature for 12 h and the resulting solid was filtered, washed with water and cold ethanol, and dried to afford 0.77 g (88%) of **6a** as a white solid, mp 83.5 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.04 (s, 1H), 7.59 (d, 2H), 7.27 (d, 2H), 4.27 (q, 2H), 2.40 (s, 3H), 1.31 (t, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 164.8, 149.3, 142.8, 140.8, 130.4, 130.3, 127.9, 113.1, 61.6, 22.8, 15.7.

**4.1.34. Ethyl 3-isopropyl-1*H*-pyrazole-4-carboxylate (6b):** A solution of hydrazine monohydrate (0.67 g, 13.40 mmol) in dry ethanol (1 mL) was added dropwise at 0 °C to a solution of (*Z*)-ethyl 2-(ethoxymethylene)-4-methyl-3-oxopentanoate (**5b**) (2.49 g, 11.62 mmol) in ethanol (10 mL). The reaction mixture was stirred at room temperature for 12 h and the solvent was removed, solidified by pet. ether to afford 1.28 g (61%) of **6-2** as a white solid, mp 71.5 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 11.46 (brs, 1H), 7.96 (s, 1H), 4.32 (q, 2H), 3.69 (m, 1H), 1.36 (s, 6H), 1.33 (t, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 165.1, 142.1, 111.6, 61.3, 27.1, 22.9, 15.8.

**4.1.35. Ethyl 3-(3-methoxyphenyl)-1*H*-pyrazole-4-carboxylate (6c):** A solution of hydrazine monohydrate (0.22 g, 4.35 mmol) in dry ethanol (1 mL) was added dropwise at 0 °C to a solution of (*Z*)-ethyl 3-ethoxy-2-(3-methoxybenzoyl)acrylate (**5c**) (1.1 g, 3.95 mmol) in ethanol (5 mL). The reaction mixture was stirred at room temperature for 12 h, and the solvent was removed. The crude was resolved in EtOAc, washed with water, and concentrated to afford 0.97 g (98%) of **6c** as a red oil, which was used without further purification. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.05 (s, 1H), 7.36 (t, 1H), 7.29 (t, 3H), 6.98 (d, 1H), 4.28 (q, 2H), 3.81 (s, 3H), 1.30 (t, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 164.6, 160.7, 141.6, 132.2, 130.7, 122.8, 116.5, 116.2, 112.8, 61.6, 56.8, 15.7.

**4.1.36. Ethyl 3-(4-(trifluoromethyl)phenyl)-1*H*-pyrazole-4-carboxylate (6d):** A solution of hydrazine monohydrate (0.22 g, 4.35 mmol) in dry ethanol (1 mL) was added dropwise at 0 °C to a solution of (*Z*)-ethyl 3-ethoxy-2-(3-methoxybenzoyl)acrylate (**5d**) (1.08 g, 3.41 mmol) in ethanol (5 mL). The reaction mixture was stirred at room temperature for 12 h and the solvent was removed. The crude was resolved in EtOAc, washed with water, and concentrated to afford a red oil which was solidified by EtOAc to give 0.79 g (82%) of **6d** as a pale red solid, mp 177.2 °C. <sup>1</sup>H NMR (MeOD) δ 8.12 (s, 1H), 7.84 (d, 2H), 7.67 (d, 2H), 4.28 (q, 2H), 1.29 (t, 3H). <sup>13</sup>C NMR (MeOD) δ 164.7, 136.1, 131.2, 131.1, 130.7, 127.4, 126.7, 126.2, 125.7, 112.6, 61.4, 14.4.

**4.1.37. Methyl 3-(2-methoxyethyl)-1H-pyrazole-4-carboxylate (6e):** A solution of hydrazine monohydrate (0.57 g, 11.54 mmol) in dry ethanol (1 mL) was added dropwise at 0 °C to a solution of (Z)-methyl 2-(ethoxymethylene)-5-methoxy-3-oxopentanoate (**5e**) (2.27 g, 10.49 mmol) in ethanol (7 mL). The reaction mixture was stirred at room temperature for 12 h, and the solvent was removed. The crude was resolved in EtOAc, washed with water, and concentrated to afford a red oil which was solidified by EtOAc to give 1.72 g (89%) of **6e** as a yellow oil. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 7.90 (s, 1H), 4.24 (q, 2H), 3.76 (s, 1H), 3.65 (t, 2H), 3.33 (s, 3H), 1.28 (t, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 165.2, 148.3, 141.5, 122.4, 72.1, 60.3, 52.5, 27.3.

**4.1.38. 1-Isopropyl-3-(p-tolyl)-1H-pyrazole-4-carboxylic acid (7a):** A solution of **6a** (0.5 g, 2.17 mmol), 1-Bromo-2-methyl-propane (0.55 g, 3.25 mmol) and cesium carbonate (2.12 g, 6.51 mmol) in acetonitrile (10 mL) was stirred at room temperature for 16 h, and the reaction was stopped. Water (10 mL) was added into the reaction mixture, and the aqueous layer was extracted with ethyl acetate (3 × 15 mL). The combined organic layers were washed with brine (3 × 10 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated to give crude. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.00 (s, 1H), 7.67 (d, 2H), 7.22 (d, 2H), 4.54 (m, 1H), 4.25 (q, 2H), 2.38 (s, 3H), 1.56 (d, 6H), 1.31 (t, 3H). The crude residue in ethanol (12 mL) was treated with 1N aqueous potassium hydroxide (7.5 mL) at 75 °C for 2.5 h. The reaction was cooled, acidified with 5N HCl and the ethanol removed *in vacuo*. The solids were filtered, washed with water, hexanes and dried *in vacuo* to afford 0.31 g (58%) of **7-1** as a beige powder, mp 129.0 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.06 (s, 1H), 7.66 (d, 2H), 7.21 (d, 2H), 4.53 (m, 1H), 2.38 (s, 3H), 1.56 (d, 6H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 169.8, 154.6, 139.7, 134.6, 131.0, 130.6, 130.1, 55.9, 24.1, 24.0, 22.9.

**4.1.39. 1-Butyl-3-(p-tolyl)-1H-pyrazole-4-carboxylic acid (7b):** A solution of **6a** (0.5 g, 2.17 mmol), 1-Bromobutane (0.29 g, 3.25 mmol) and cesium carbonate (2.12 g, 6.51 mmol) in acetonitrile (10 mL) was stirred at room temperature for 16 h, and the reaction was stopped. Water (10 mL) was added into the reaction mixture and the aqueous layer was extracted with ethyl acetate (3 × 15 mL). The combined organic layers were washed with brine (3 × 10 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated. The crude residue in ethanol (12 mL) was treated with 1N aqueous potassium hydroxide (7.5 mL) at 75 °C for 3.5 h. The reaction was cooled, acidified with 5N HCl and the ethanol removed *in vacuo*. Solvent was removed and the resulting yellow oil was purified by flash column chromatography (CHCl<sub>3</sub>/MeOH 9:1) to afford **7-2** as pale-yellow oil (0.5 g, 50% yield). <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.06 (s, 1H), 7.66 (d, 2H), 7.21 (d, 2H), 4.53 (m, 1H), 2.38 (s, 3H), 1.56 (d, 6H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 169.8, 154.6, 139.7, 134.6, 131.0, 130.6, 130.1, 55.9, 24.1, 24.0, 22.9.

**4.1.40. 1-(Naphthalen-2-ylmethyl)-3-(p-tolyl)-1H-pyrazole-4-carboxylic acid (7c):** A solution of **6a** (0.18 g, 0.78 mmol), 2-(bromomethyl)naphthalene (0.26 g, 1.17 mmol) and cesium carbonate (0.76 g, 2.34 mmol) in acetonitrile (5 mL) was stirred at room temperature for 16 h, and the reaction was stopped. Water (10 mL) was added into the reaction mixture and the aqueous layer was extracted with ethyl acetate (3 × 15 mL). The combined organic layers were washed with brine (3 × 10 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated.

The crude residue in ethanol (10 mL) was treated with 1N aqueous potassium hydroxide (7.5 mL) at 75°C for 3.5 h. The reaction was cooled, acidified with 5N HCl and the ethanol removed *in vacuo* to afford **7c** as a pale-yellow solid (0.2 g, 77% yield), mp 151.2 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.06 (s, 1H), 7.75 (m, 3H), 7.71 (d, 2H), 7.53 (d, 2H), 7.48 (m, 1H), 7.20 (m, 3H), 5.45 (s, 2H), 2.40 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 169.3, 155.2, 139.8, 134.7, 131.3, 130.6, 130.5, 130.4, 129.9, 129.7, 129.6, 129.3, 129.2, 129.1, 128.9, 128.0, 59.9, 22.8.

**4.1.41. 1-(Cyclopropylmethyl)-3-(*p*-tolyl)-1H-pyrazole-4-carboxylic acid (7d):** A

solution of **6a** (0.5 g, 2.17 mmol), bromomethylcyclopropane (0.43 g, 3.25 mmol) and cesium carbonate (2.12 g, 6.51 mmol) in acetonitrile (10 mL) was stirred at room temperature for 16 h. Water (10 mL) was added into the reaction mixture and the aqueous layer was extracted with ethyl acetate (3 × 15 mL). The combined organic layers were washed with brine (3 × 10 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated to give the crude. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.09 (s, 1H), 7.68 (d, 2H), 7.26 (d, 2H), 4.24 (q, 2H), 4.02 (t, 2H), 2.42 (s, 3H), 1.33 (brs, 1H), 1.22 (t, 3H), 0.72 (t, 2H), 0.48 (t, 2H). The crude residue in ethanol (10 mL) was treated with 1N aqueous potassium hydroxide (7.5 mL) at 75°C for 3.5 h. The reaction was cooled, acidified with 5N HCl and the ethanol removed *in vacuo*. The solids were filtered, washed with water hexanes and dried over *in vacuo* to afford **7d** as a pale-yellow solid (0.3 g, 55% yield), mp 142.6 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.18 (s, 1H), 7.69 (d, 2H), 7.23 (d, 2H), 4.04 (d, 2H), 2.44 (s, 3H), 1.36 (brs, 1H), 0.74 (t, 2H), 0.44 (t, 2H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 169.7, 154.9, 143.7, 139.7, 136.6, 130.6, 130.5, 130.1, 112.0, 58.7, 22.8, 12.0, 5.5, 5.3.

**4.1.42. 1-(3-Methoxybenzyl)-3-(*p*-tolyl)-1H-pyrazole-4-carboxylic acid (7e):** A solution of **6a** (0.1 g, 0.43 mmol), 3-methoxybenzyl bromide (0.13 g, 0.65 mmol) and cesium carbonate (0.42 g, 1.29 mmol) in acetonitrile (5 mL) was stirred at room temperature for 16 h. Water (10 mL) was added to the reaction mixture, and the aqueous layer was extracted with ethyl acetate (3 × 15 mL). The combined organic layers were washed with brine (3 × 10 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated to give the crude. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.05 (s, 1H), 7.90 (d, 2H), 7.24 (m, 3H), 6.89 (d, 2H), 6.84 (s, 1H), 5.29 (s, 2H), 4.21 (q, 2H), 3.75 (s, 3H), 2.41 (s, 3H), 1.21 (t, 3H). The crude residue in ethanol (5 mL) was treated with 1N aqueous potassium hydroxide (3.5 mL) at 75°C for 3.5 h. The reaction was cooled, acidified with 5N HCl and the solvent removed *in vacuo*. The solids were filtered, washed with hexanes and dried over *in vacuo* to afford **7e** as a white solid (0.1 g, 77% yield), mp 169.6 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 7.95 (s, 1H), 7.68 (d, 2H), 7.31 (t, 1H), 7.22 (t, 2H), 6.90 (d, 2H), 6.84 (s, 1H), 5.29 (b, 2H), 3.73 (s, 3H), 2.41 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 168.9, 155.2, 139.8, 137.3, 131.5, 131.2, 130.6, 130.5, 130.1, 121.8, 115.4, 115.3, 57.8, 56.7, 22.7.

**4.1.43. 3-(*p*-Tolyl)-1-(3-(trifluoromethyl)benzyl)-1H-pyrazole-4-carboxylic acid (7f):** A

solution of **6a** (0.1 g, 0.43 mmol), 3-(trifluoromethyl)benzyl bromide (0.15 g, 0.65 mmol) and cesium carbonate (0.42 g, 1.29 mmol) in acetonitrile (5 mL) was stirred at room temperature for 6 h, and the reaction was stopped. Water (10 mL) was added to the reaction mixture, and the aqueous layer was extracted with ethyl acetate (3 × 15 mL). The combined organic layers were washed with brine (3 × 10 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated to give the crude. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 7.97 (s, 1H), 7.68 (d, 2H), 7.58 (s, 1H),

7.51 (m, 3H), 7.23 (d, 2H), 5.39 (s, 2H), 4.26 (q, 2H), 2.41 (s, 3H), 1.26 (t, 3H). The crude residue in ethanol (5 mL) was treated with 1N aqueous potassium hydroxide (3.5 mL) at 75°C for 5.5 h. The reaction was cooled, acidified with 5N HCl and the ethanol removed *in vacuo*. The solids were filtered, washed with water hexanes and dried over *in vacuo* to afford **7f** as a white solid (0.11 g, 77% yield), mp 149.9 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.01 (s, 1H), 7.67 (d, 2H), 7.52 (s, 1H), 7.49 (d, 2H), 7.21 (m, 3H), 5.39 (s, 2H), 2.41 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 169.2, 156.4, 143.9, 140.1, 137.65, 132.7, 131.1, 130.7, 130.6, 130.1, 126.8, 126.2, 119.5, 118.5, 57.2, 22.7.

**4.1.44. 1-(4-Hydroxybutyl)-3-(p-tolyl)-1H-pyrazole-4-carboxylic acid (7g):** A solution of **6a** (0.1 g, 0.43 mmol), 4-bromo-1-butanol (0.10 g, 0.65 mmol) and cesium carbonate (0.42 g, 1.29 mmol) in acetonitrile (5 mL) was stirred at room temperature for 6 h, and the reaction was stopped. Water (10 mL) was added to the reaction mixture, and the aqueous layer was extracted with ethyl acetate (3 × 15 mL). The combined organic layers were washed with brine (3 × 10 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated to give the crude. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 7.98 (s, 1H), 7.65 (d, 2H), 7.20 (d, 2H), 4.24 (q, 2H), 4.09 (t, 2H), 3.45 (t, 2H), 2.41 (s, 3H), 1.67 (t, 2H), 1.57 (t, 2H), 1.42 (t, 3H). The crude residue in ethanol (5 mL) was treated with 1N aqueous potassium hydroxide (3.5 mL) at 75°C for 5.5 h. The reaction was cooled; the ethanol was removed *in vacuo*, acidified with 5N HCl, and dried *in vacuo* to afford **7g** as an oil (0.09 g, 82% yield). <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 7.99 (s, 1H), 7.65 (d, 2H), 7.19 (d, 2H), 4.14 (q, 2H), 3.62 (t, 2H), 2.40 (s, 3H), 1.96 (t, 2H), 1.55 (t, 2H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 168.8, 154.9, 139.7, 137.3, 131.1, 130.6, 130.1, 130.7, 112.0, 63.1, 53.7, 30.5, 27.9, 22.8.

**4.1.45. 3-Isopropyl-1-(3-methoxybenzyl)-1H-pyrazole-4-carboxylic acid (7h):** A solution of **6b** (0.5 g, 2.74 mmol), 3-methoxybenzyl bromide (0.83 g, 4.11 mmol) and cesium carbonate (2.68 g, 8.22 mmol) in acetonitrile (10 mL) was stirred at room temperature for 16 h and the reaction was stopped. Water (10 mL) was added into the reaction mixture and the aqueous layer was extracted with ethyl acetate (3 × 15 mL). The combined organic layers were washed with brine (3 × 10 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated to give crude. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 7.73 (s, 1H), 7.29 (d, 1H), 6.87 (t, 1H), 6.83 (d, 1H), 6.77 (s, 1H), 5.22 (s, 2H), 4.26 (q, 2H), 3.82 (s, 3H), 3.54 (m, 1H), 1.35 (d, 6H), 1.26 (t, 3H). The crude residue in ethanol (12 mL) was treated with 1N aqueous potassium hydroxide (7.5 mL) at 75°C for 4.5 h. The reaction was cooled; the ethanol was removed *in vacuo*, acidified with 5N HCl, and dried *in vacuo* to afford **7h** as a white solid (0.59 g, 78% yield), mp 111.9 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 7.79 (s, 1H), 7.29 (t, 1H), 6.88 (d, 1H), 6.84 (d, 1H), 6.78 (s, 1H), 5.23 (s, 2H), 3.79 (s, 3H), 3.55 (m, 1H), 1.32 (d, 6H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 169.6, 163.0, 161.5, 138.0, 136.4, 131.4, 121.7, 115.3, 115.1, 111.8, 57.5, 56.7, 28.0, 23.3.

**4.1.46. 3-Isopropyl-1-(3-methoxybenzyl)-1H-pyrazole-4-carboxylic acid (7i):** A solution of **6c** (0.3 g, 1.22 mmol), 2-iodopropane (0.31 g, 1.83 mmol) and cesium carbonate (1.19 g, 3.66 mmol) in acetonitrile (5 mL) was stirred at room temperature for 16 h, and the reaction was stopped. Water (10 mL) was added to the reaction mixture, and the aqueous layer was extracted with ethyl acetate (3 × 15 mL). The combined organic layers were washed with brine (3 × 10 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated to give crude. <sup>1</sup>H NMR

(CDCl<sub>3</sub>) δ 8.01 (s, 1H), 7.39 (d, 1H), 7.32 (t, 2H), 6.92 (d, 1H), 4.26 (q, 2H), 4.14 (m, 1H), 3.85 (s, 3H), 1.43 (d, 6H), 1.17 (t, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 164.6, 161.6, 142.5, 133.5, 130.8, 123.5, 117.0, 116.1, 61.4, 56.7, 55.8, 51.9, 24.2, 15.7. The crude residue in ethanol (12 mL) was treated with 1N aqueous potassium hydroxide (7.5 mL) at 75°C for 12 h. The reaction was cooled; the ethanol was removed *in vacuo*, acidified with 5N HCl, and dried *in vacuo* to afford **7i** as a yellow oil (0.25 g, 78% yield). <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.08 (s, 1H), 7.39 (d, 2H), 7.32 (t, 1H), 6.95 (d, 1H), 4.51 (m, 1H), 3.85 (s, 3H), 1.58 (d, 6H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 169.6, 160.6, 154.3, 143.5, 134.9, 134.7, 130.9, 123.3, 116.9, 115.9, 56.7, 55.9, 24.1, 24.0.

**4.1.47. 1-(3-Methoxybenzyl)-3-(3-methoxyphenyl)-1H-pyrazole-4-carboxylic acid (7j):**

A solution of **6c** (0.3 g, 1.22 mmol), benzyl-3-methoxybenzene (0.37 g, 1.83 mmol) and cesium carbonate (1.19 g, 3.66 mmol) in acetonitrile (5 mL) was stirred at room temperature for 6 h, and the reaction was stopped. Water (10 mL) was added to the reaction mixture and the aqueous layer was extracted with ethyl acetate (3 × 15 mL). The combined organic layers were washed with brine (3 × 10 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated to give crude. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 7.92 (s, 1H), 7.39 (s, 1H), 7.32 (t, 3H), 6.94 (d, 3H), 6.84 (s, 1H), 5.31 (s, 2H), 4.24 (q, 2H), 3.85 (s, 3H), 3.76 (s, 3H), 1.86 (t, 3H). The crude residue in ethanol (12 mL) was treated with 1N aqueous potassium hydroxide (7.5 mL) at 75°C for 12 h. The reaction was cooled; the ethanol was removed *in vacuo*, acidified with 5N HCl, and dried *in vacuo* to afford **7j** as a yellow oil (0.27 g, 66% yield). <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 7.96 (s, 1H), 7.38 (d, 2H), 7.32 (t, 3H), 6.92 (d, 2H), 6.84 (s, 1H), 5.29 (s, 2H), 3.83 (s, 3H), 3.73 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 168.7, 160.6, 161.1, 143.5, 137.7, 137.4, 134.6, 131.5, 130.4, 123.2, 121.8, 116.2, 115.9, 115.4, 115.3, 57.9, 56.7.

**4.1.48. 1-Isopropyl-3-(4-(trifluoromethyl)phenyl)-1H-pyrazole-4-carboxylic acid (7k):**

A solution of **6d** (0.3 g, 1.06 mmol), 2-iodopropane (0.36 g, 1.83 mmol) and cesium carbonate (1.19 g, 3.66 mmol) in acetonitrile (5 mL) was stirred at room temperature for 6 h, and the reaction was stopped. Water (10 mL) was added to the reaction mixture, and the aqueous layer was extracted with ethyl acetate (3 × 15 mL). The combined organic layers were washed with brine (3 × 10 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated to give crude. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.00 (s, 1H), 7.92 (s, 1H), 7.93 (d, 2H), 7.64 (d, 2H), 4.25 (q, 2H), 4.10 (m, 1H), 1.61 (d, 6H), 1.28 (t, 3H). The crude residue in ethanol (10 mL) was treated with 2N aqueous potassium hydroxide (5.5 mL) at 75°C for 12 h. The reaction was cooled; the ethanol was removed *in vacuo*. The residue was taken up with EtOAc and water and acidified with 5N HCl to pH 3. Organic phases were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated *in vacuo* to afford **7k** as a yellow solid (0.21 g, 67% yield), mp 161.1 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 10.17 (brs, 1H), 8.10 (s, 1H), 7.93 (d, 2H), 7.62 (d, 2H), 4.51 (brs, 1H), 1.52 (s, 6H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 168.1, 151.5, 143.9, 142.1, 135.9, 133.5, 132.7, 130.3, 129.6, 125.3, 54.7, 22.4, 22.3.

**4.1.49. 1-(3-Methoxybenzyl)-3-(4-(trifluoromethyl)phenyl)-1H-pyrazole-4-carboxylic acid (7l):** A solution of **6d** (0.2 g, 0.7 mmol), 3-methoxy benzyl bromide (0.21 g, 1.05 mmol) and cesium carbonate (0.68 g, 2.1 mmol) in acetonitrile (5 mL) was stirred at room temperature for 12 h. Water (10 mL) was added to the reaction mixture, and the aqueous



layer was extracted with ethyl acetate (3×15 mL). The combined organic layers were washed with brine (3 × 10 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated to give crude. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 7.96 (t, 2H), 7.66 (d, 2H), 7.29 (d, 2H), 6.88 (d, 2H), 6.83 (s, 1H), 5.28 (s, 2H), 4.23 (q, 2H), 3.79 (s, 3H), 1.26 (t, 3H). The crude residue in ethanol (5 mL) was treated with 2N aqueous potassium hydroxide (4 mL) at 75°C for 12 h. The reaction was cooled; the ethanol was removed *in vacuo*. The residue was taken up with EtOAc/water and acidified with 5N HCl to pH 3. Organic phases were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated *in vacuo* to afford **7a** as a yellow solid (0.21 g, 80% yield), mp 141.7 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 7.92 (s, 1H), 7.87 (d, 2H), 7.23 (t, 1H), 6.84 (d, 2H), 6.77 (s, 1H), 5.24 (s, 2H), 3.74 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 166.1, 160.8, 159.6, 142.4, 139.2, 135.9, 135.6, 130.0, 129.5, 129.1, 125.4, 119.8, 113.6, 113.0, 112.7, 64.8, 55.0.

**4.1.50. 3-Isopropyl-1-(3-methoxybenzyl)-N-(4-methyl-3-**

**(morpholinosulfonyl)phenyl)-1H-pyrazole-4-carboxamide (4m):** To a solution of **7h** (0.12 g, 0.44 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (5 mL) was added EDCI (0.1 g, 0.53 mmol) and DMAP (0.07 g, 0.53 mmol). To the reaction was added 4-methyl-3-(morpholinosulfonyl)aniline (0.11 g, 0.44 mmol). The reaction mixture was stirred at room temperature for 6 h and then treated with 10% HCl. The organic layer was collected, and the aqueous layer was extracted with EtOAc. The combined organic solution was dried over Na<sub>2</sub>SO<sub>4</sub>, evaporated under a vacuum. The resulting yellow oil was purified by flash column chromatography (H/EtOAc 1:1) to afford **4m** as a white solid (0.18 g, 83% yield), mp 179.7 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.10 (s, 1H), 7.79 (dd, 3H), 7.31 (m, 2H), 6.83 (t, 2H), 6.82 (s, 1H), 5.28 (s, 2H), 3.81 (s, 3H), 3.66 (t, 4H), 3.59 (m, 1H), 3.11 (t, 4H), 2.60 (s, 3H), 1.39 (d, 6H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 163.2, 161.2, 160.0, 138.8, 138.5, 136.3, 134.5, 135.1, 131.8, 131.4, 126.1, 122.6, 122.7, 121.7, 115.2, 115.1, 67.6, 57.4, 56.7, 46.8, 28.4, 23.6, 21.5. Anal. Calcd for C<sub>26</sub>H<sub>32</sub>N<sub>4</sub>O<sub>5</sub>S: C, 60.92; H, 6.29; N, 10.93. Found: C, 60.45; H, 5.97; N, 10.51.

**4.1.51. 1-Isopropyl-3-(3-methoxyphenyl)-N-(4-methyl-3-**

**(morpholinosulfonyl)phenyl)-1H-pyrazole-4-carboxamide (4n):** To a solution of **7i** (0.25 g, 0.96 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (5 mL) was added EDCI (0.22 g, 1.15 mmol) and DMAP (1.15 g, 1.15 mmol). To the reaction was added 4-methyl-3-(morpholinosulfonyl)aniline (0.25 g, 0.96 mmol). The reaction mixture was stirred at room temperature for 12 h and then treated with 10% HCl. The organic layer was collected, and the aqueous layer was extracted with EtOAc. The combined organic solution was dried over Na<sub>2</sub>SO<sub>4</sub>, evaporated under a vacuum. The resulting yellow oil was purified by flash column chromatography (H/EtOAc 1:1) to afford **4n** as a white solid (0.26 g, 55% yield), mp 158.0 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.15 (s, 1H), 7.72 (s, 1H), 7.53 (s, 1H), 7.47 (d, 2H), 7.24 (d, 2H), 7.19 (s, 1H), 7.05 (m, 1H), 4.57 (m, 1H), 3.86 (s, 3H), 3.74 (t, 4H), 3.17 (t, 4H), 2.57 (s, 3H), 1.61 (s, 6H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 163.2, 161.7, 150.6, 142.2, 137.7, 136.9, 134.8, 134.5, 132.9, 132.6, 131.7, 125.6, 123.6, 123.1, 122.0, 117.6, 67.7, 56.8, 56.0, 46.9, 24.2, 21.7. Anal. Calcd for C<sub>25</sub>H<sub>30</sub>N<sub>4</sub>O<sub>5</sub>S · ¼ H<sub>2</sub>O: C, 59.53; H, 5.92; N, 11.10. Found: C, 59.76; H, 5.67; N, 11.10.

**4.1.52. 1-(3-Methoxybenzyl)-3-(3-methoxyphenyl)-N-(4-methyl-3-**

**(morpholinosulfonyl)phenyl)-1H-pyrazole-4-carboxamide (4o):** To a solution of **7j** (0.25 g, 0.73 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (5 mL) was added EDCI (0.17 g, 0.88 mmol) and DMAP (0.11 g,

0.88 mmol). To the reaction was added 4-methyl-3-(morpholinosulfonyl)aniline (0.19 g, 0.73 mmol). The reaction mixture was stirred at room temperature for 12 h and then treated with 10% HCl. The organic layer was collected, and the aqueous layer was extracted with EtOAc. The combined organic solution was dried over Na<sub>2</sub>SO<sub>4</sub>, evaporated under a vacuum. The resulting yellow oil was solidified by MeOH to afford **4o** as a white solid (0.31 g, 74% yield), mp 190.1 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.04 (s, 1H), 7.72 (s, 1H), 7.54 (s, 1H), 7.48 (t, 2H), 7.34 (s, 1H), 7.28 (m, 3H), 7.20 (d, 1H), 6.94 (t, 1H), 6.88 (s, 1H), 3.82 (s, 3H), 3.80 (s, 3H), 3.74 (t, 4H), 3.17 (t, 4H), 2.57 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 163.2, 161.6, 160.0, 151.4, 137.7, 137.4, 135.7, 134.9, 134.5, 131.7, 131.6, 125.0, 123.1, 122.1, 122.0, 116.9, 116.2, 115.5, 115.4, 67.8, 58.1, 56.9, 56.7, 46.9, 21.7. Anal. Calcd for C<sub>30</sub>H<sub>32</sub>N<sub>4</sub>O<sub>5</sub>S · ½ H<sub>2</sub>O: C, 61.51; H, 5.50; N, 9.56. Found: C, 61.50; H, 5.28; N, 9.39.

**4.1.53. 1-Isopropyl-N-(4-methyl-3-(morpholinosulfonyl)phenyl)-3-(4-(trifluoromethyl)phenyl)-1H-pyrazole-4-carboxamide (4p):** To a solution of **7k** (0.2 g, 0.67 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (5 mL) was added EDCI (0.15 g, 0.81 mmol) and DMAP (0.08 g, 0.81 mmol). To the reaction was added 4-methyl-3-(morpholinosulfonyl)aniline (0.17 g, 0.67 mmol). The reaction mixture was stirred at room temperature for 12 h, quenched with water and then treated with 10% HCl. The organic layer was collected, and the aqueous layer was extracted with EtOAc. The combined organic solution was dried over Na<sub>2</sub>SO<sub>4</sub>, evaporated under a vacuum. The resulting yellow oil was purified by flash column chromatography (H/EtOAc 1:1) to afford **4p** as a yellow solid (0.18 g, 51% yield), mp 192.2 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.12 (s, 1H), 8.09 (s, 1H), 7.87 (d, 2H), 7.81 (d, 2H), 7.65 (d, 2H), 7.24 (d, 1H), 4.53 (m, 1H), 3.64 (t, 4H), 3.07 (t, 4H), 2.54 (s, 3H), 1.53 (s, 6H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 161.5, 148.8, 138.6, 136.2, 134.9, 133.7, 133.3, 130.4, 129.3, 125.7, 119.4, 66.2, 66.1, 45.3, 45.2, 22.5, 20.1, 20.0. Anal. Calcd for C<sub>25</sub>H<sub>27</sub>F<sub>3</sub>N<sub>4</sub>O<sub>4</sub>S · ½ H<sub>2</sub>O: C, 55.03; H, 4.98; N, 10.44. Found: C, 55.23; H, 5.03; N, 10.26.

**4.1.54. 1-(3-Methoxybenzyl)-N-(4-methyl-3-(morpholinosulfonyl)phenyl)-3-(4-(trifluoromethyl)phenyl)-1H-pyrazole-4-carboxamide (4q):** To a solution of **7l** (0.06 g, 0.16 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (2 mL) was added EDCI (0.04 g, 0.019 mmol) and DMAP (0.02 g, 0.019 mmol). To the reaction was added 4-methyl-3-(morpholinosulfonyl)aniline (0.04 g, 0.16 mmol). The reaction mixture was stirred at room temperature for 12 h, quenched with water and then treated with 10% HCl. The organic layer was collected and the aqueous layer was extracted with EtOAc. The combined organic solution was dried over Na<sub>2</sub>SO<sub>4</sub>, evaporated under a vacuum. The resulting yellow oil was purified by flash column chromatography (H/EtOAc 1:1) to afford **4q** as a yellow solid (0.022 g, 61% yield), mp 208.2 °C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 7.98 (s, 1H), 7.89 (d, 2H), 7.81 (s, 1H), 7.74 (d, 2H), 7.62 (d, 1H), 7.47 (s, 1H), 7.27 (t, 1H), 7.26 (t, 1H), 6.94 (m, 3H), 5.34 (s, 2H), 3.82 (s, 3H), 3.72 (t, 4H), 3.15 (t, 4H), 2.58 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 161.5, 148.8, 138.6, 136.2, 134.9, 133.7, 133.3, 130.4, 129.3, 125.7, 119.4, 66.2, 66.1, 45.3, 45.2, 22.5, 20.1, 20.0. Anal. Calcd for C<sub>30</sub>H<sub>29</sub>F<sub>3</sub>N<sub>4</sub>O<sub>5</sub>S · ½ H<sub>2</sub>O: C, 57.77; H, 4.68; N, 8.98. Found: C, 58.08; H, 4.64; N, 8.64.

**4.1.55. 1-Isopropyl-N-methyl-N-(4-methyl-3-(morpholinosulfonyl)phenyl)-3-(p-tolyl)-1H-pyrazole-4-carboxamide (8a):** A mixture of **4f** (0.1 g, 0.21 mmol), NaH (60% dispersion in oil) (0.018 g, 0.46 mmol) and anhydrous DMF (10 mL) was stirred at room

temperature for 30 min. The reaction mixture was cooled to 0 °C, and iodomethane (0.1 mL, 1.47 mmol) was added to it. The resulting mixture was stirred at room temperature under an atmosphere of N<sub>2</sub> and monitored by TLC. The reaction was stopped, water (30 mL) was added to the reaction mixture, and the aqueous layer was extracted with ethyl acetate (3 × 15 mL). The combined organic layers were washed with water and brine (3 × 10 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated. The resulting yellow oil was purified by flash column chromatography (H/EtOAc 1:1) to afford **8a** as a clear oil (0.07 g, 70% yield). <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 7.34 (s, 1H), 7.15 (d, 2H), 7.09 (t, 2H), 6.94 (d, 2H), 6.87 (d, 1H), 4.26 (m, 1H), 3.67 (t, 4H), 3.27 (s, 3H), 3.05 (t, 4H), 2.55 (s, 3H), 2.41 (s, 3H), 1.31 (d, 6H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 166.5, 144.0, 140.8, 136.8, 134.4, 131.6, 130.8, 130.7, 130.4, 129.6, 129.3, 128.5, 127.4, 67.6, 51.5, 46.6, 39.0, 22.8, 22.7, 21.6. HRMS (ESI); Calcd for C<sub>26</sub>H<sub>32</sub>N<sub>4</sub>O<sub>4</sub>S (M+1): 497.2217. Found: 497.2215.

**4.1.56. 3-Isopropyl-1-(3-methoxybenzyl)-N-methyl-N-(4-methyl-3-**

**(morpholinosulfonyl)phenyl)-1H-pyrazole-4-carboxamide (8b):** A mixture of **4m** (0.3 g, 0.58 mmol), NaH (60% dispersion in oil) (0.046 g, 1.27 mmol) and anhydrous DMF (15 mL) was stirred at room temperature for 30 min. The reaction mixture was cooled to 0 °C, and iodomethane (0.25 mL, 4.09 mmol) was added to it. The resulting mixture was stirred at room temperature under an atmosphere of N<sub>2</sub> and monitored by TLC. The reaction was stopped, water (50 mL) was added to the reaction mixture and the aqueous layer was extracted with ethyl acetate (3 × 25 mL). The combined organic layers were washed with water and brine (3 × 10 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated. The resulting yellow oil was purified by flash column chromatography (H/EtOAc 1:1) to afford **8b** as a clear oil (0.27 g, 87% yield). <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 7.45 (s, 1H), 7.18 (dd, 3H), 6.78 (d, 1H), 6.52 (d, 2H), 6.50 (d, 1H), 4.95 (s, 2H), 3.71 (s, 3H), 3.63 (t, 4H), 3.34 (s, 3H), 3.31 (m, 1H), 2.82 (t, 4H), 2.53 (s, 3H), 1.23 (d, 6H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 166.5, 144.0, 140.8, 136.8, 134.4, 131.6, 130.8, 130.7, 130.4, 129.6, 129.3, 128.5, 127.4, 67.6, 51.5, 46.6, 39.0, 22.8, 22.7, 21.6. HRMS (ESI); Calcd for C<sub>27</sub>H<sub>34</sub>N<sub>4</sub>O<sub>5</sub>S (M+1): 527.2323. Found: 527.2321.

**4.1.57. 1-(3-Methoxybenzyl)-3-(3-methoxyphenyl)-N-methyl-N-(4-methyl-3-**

**(morpholinosulfonyl)phenyl)-1H-pyrazole-4-carboxamide (8c):** A mixture of **4o** (0.1 g, 0.17 mmol), NaH (60% dispersion in oil) (0.02 g, 0.51 mmol) and anhydrous DMF (10 mL) was stirred at room temperature for 30 min. The reaction mixture was cooled to 0 °C, and iodomethane (0.075 mL, 1.21 mmol) was added to it. The resulting mixture was stirred at room temperature under an atmosphere of N<sub>2</sub> and monitored by TLC. The reaction was stopped, water (50 mL) was added into the reaction mixture and the aqueous layer was extracted with ethyl acetate (3 × 25 mL). The combined organic layers were washed with water and brine (3 × 10 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated. The resulting yellow oil was purified by flash column chromatography (H/EtOAc 1:1) to afford **8c** as a clear oil (0.08 g, 87% yield). <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 7.45 (s, 1H), 7.21 (t, 2H), 7.14 (t, 1H), 6.85 (d, 2H), 6.80 (t, 3H), 6.72 (d, 2H), 6.70 (s, 1H), 5.12 (s, 2H), 3.72 (s, 6H), 3.58 (t, 4H), 3.26 (s, 3H), 2.84 (t, 4H), 2.41 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 164.3, 159.0, 158.6, 148.3, 140.8, 135.5, 134.4, 133.9, 132.6, 131.6, 131.3, 129.0, 128.3, 127.0, 119.3, 118.8, 113.3, 112.8, 111.3, 65.2, 54.3, 54.2, 44.0, 19.0. HRMS (ESI); Calcd for C<sub>31</sub>H<sub>34</sub>N<sub>4</sub>O<sub>6</sub>S (M+1): 591.2272. Found: 591.2270.

**4.1.58. *N*-((1-Isopropyl-3-(*p*-tolyl)-1*H*-pyrazol-4-yl)methyl)-4-methyl-3-**

**(morpholinosulfonyl)aniline (9a):** To a solution of **4f** (0.1 g, 0.21 mmol) in anhydrous THF (5 mL), 1 N borane-THF complex (7 mL) was added at 0 °C under N<sub>2</sub>. The reaction mixture was heated to reflux for 3 h. After it was cooled to 0 °C, 1N methanolic HCl (3 mL) was added dropwise to quench excess borane reagent and MeOH (5 mL) was added. The reaction mixture was heated to reflux for 2 h. The solvent was evaporated at reduced pressure. The oily residue was dissolved in EtOAc and neutralized with 2N NaOH. The resulting mixture was extracted with EtOAc (3 × 15 mL). The combined extracts were washed with 0.5 N KOH, dried over Na<sub>2</sub>SO<sub>4</sub>, and evaporated *in vacuo*. The resulting yellow oil was purified by flash column chromatography (H/EtOAc 1:1) to afford **9a** as colorless oil (0.08 g, 81% yield). <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 7.55 (d, 2H), 7.47 (s, 1H), 7.20 (d, 2H), 7.14 (s, 1H), 7.10 (d, 1H), 6.72 (d, 1H), 4.52 (m, 1H), 4.27 (s, 2H), 3.71 (t, 4H), 3.12 (t, 4H), 2.51 (s, 3H), 2.36 (s, 3H), 1.53 (d, 6H), <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 150.5, 147.6, 138.8, 136.7, 135.1, 130.7, 128.8, 128.5, 127.6, 118.7, 115.7, 69.0, 46.7, 33.1, 24.1, 24.0. HRMS (ESI); Calcd for C<sub>25</sub>H<sub>32</sub>N<sub>4</sub>O<sub>3</sub>S (M+1): 469.6116. Found: 469.2715.

**4.1.59. *N*-((3-Isopropyl-1-(3-methoxybenzyl)-1*H*-pyrazol-4-yl)methyl)-4-methyl-3-**

**(morpholinosulfonyl)aniline (9b):** To a solution of **4m** (0.1 g, 0.19 mmol) in anhydrous THF (5 mL), 1 N borane-THF complex (7 mL) was added at 0 °C under N<sub>2</sub>. The reaction mixture was heated to reflux for 3 h. After it was cooled to 0 °C, 1N methanolic HCl (3 mL) was added dropwise to quench excess borane reagent and additional MeOH was added. The reaction mixture was heated to reflux for 3 h. The solvent was evaporated at reduced pressure. The oily residue was dissolved in EtOAc and neutralized with 2N NaOH. The resulting mixture was extracted with EtOAc (3 × 15 mL). The combined extracts were washed with 0.5 N KOH, dried over Na<sub>2</sub>SO<sub>4</sub>, and evaporated *in vacuo*. The resulting yellow oil was purified by flash column chromatography (H/EtOAc 1:1) to afford **9b** as colorless oil (0.07 g, 77% yield). <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 7.22 (d, 2H), 7.21 (s, 1H), 7.12 (s, 1H), 7.10 (d, 1H), 6.83 (d, 1H), 6.77 (d, 1H), 6.72 (t, 2H), 5.18 (s, 2H), 4.11 (t, 2H), 3.76 (s, 3H), 3.69 (t, 4H), 3.11 (t, 4H), 3.05 (m, 1H), 2.48 (s, 3H), 1.26 (d, 6H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 161.3, 157.7, 147.6, 139.6, 136.4, 135.1, 131.2, 130.5, 127.1, 121.3, 118.5, 116.6, 115.6, 114.8, 114.7, 67.7, 57.1, 56.6, 46.7, 39.8, 26.5, 24.0, 23.6, 21.1. HRMS (ESI); Calcd for C<sub>26</sub>H<sub>34</sub>N<sub>4</sub>O<sub>4</sub>S (M+1): 499.6376. Found: 499.3676.

**4.1.60. *N*-((1-(3-Methoxybenzyl)-3-(3-methoxyphenyl)-1*H*-pyrazol-4-yl)methyl)-4-**

**methyl-3-(morpholinosulfonyl)aniline (9c):** To a solution of **4o** (0.1 g, 0.17 mmol) in anhydrous THF (5 mL), 1 N borane-THF complex (8 mL) was added at 0 °C under N<sub>2</sub>. The reaction mixture was heated to reflux for 3 h. After it was cooled to 0 °C, 1N methanolic HCl (3 mL) was added dropwise to quench excess borane reagent and additional MeOH was added. The reaction mixture was heated to reflux for 2 h. The solvent was evaporated at reduced pressure. The oily residue was dissolved in EtOAc and neutralized with 2N NaOH. The resulting mixture was extracted with EtOAc (3 × 15 mL). The combined extracts were washed with 0.5 N KOH, dried over Na<sub>2</sub>SO<sub>4</sub>, and evaporated *in vacuo*. The resulting yellow oil was purified by flash column chromatography (H/EtOAc 1:1) to afford **9-c** as colorless oil (0.065 g, 72% yield). <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 7.38 (s, 1H), 7.32 (m, 5H), 7.18 (d, 1H), 7.11 (s, 1H), 6.89 (t, 2H), 6.81 (d, 2H), 6.69 (t, 1H), 5.28 (s, 2H), 4.26 (s, 2H), 3.81 (s, 3H), 3.76

(s, 3H), 3.71 (t, 4H), 3.11 (t, 4H), 2.49 (s, 3H).  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta$  161.3, 161.2, 151.1, 147.3, 139.1, 136.4, 135.1, 131.7, 131.3, 127.4, 121.5, 118.6, 117.7, 115.7, 115.3, 115.1, 116.2, 114.8, 114.1, 67.7, 57.5, 56.6, 46.7, 40.5, 21.2. HRMS (ESI); Calcd for  $\text{C}_{30}\text{H}_{34}\text{N}_4\text{O}_5\text{S}$  (M+Na): 565.6798. Found: 585.2798.

## 4.2. Biology

**4.2.1. Materials**—DMSO and charcoal/dextran-treated FBS were obtained from Fisher Scientific (Pittsburgh, PA). GW4064 was purchased from R&D Systems, Inc. (Minneapolis, MN); LCA and rifampicin were purchased from Sigma-Aldrich (St. Louis, MO); TO901317 was obtained from Cayman Chemical Company (Ann Arbor, MI); Z26476908 (E16) was purchased from Enamine LLC (Monmouth Junction, NJ). Black polypropylene 384-well plates were purchased from Matrical Bioscience (Spokane, WA). Black 384-well tissue culture-treated clear bottom plates were obtained from Corning Incorporated (Corning, NY). White tissue culture-treated 384-well plates were purchased from PerkinElmer (Waltham, MA). GST-hFXR-LBD, Tb-anti-GST antibody, coregulator buffer G, 1 M DTT, GeneBLazer FXR-UAS-bla HEK 293T cells, DMEM, phenol red-free DMEM, dialyzed FBS, non-essential amino acids, sodium pyruvate, HEPES, hygromycin B, zeocin, penicillin/ streptomycin, G418 and CCF4-AM substrate were purchased from Invitrogen (Carlsbad, CA).

All chemicals were initially solubilized in DMSO as 10 mM stocks and then diluted in corresponding assay buffer or medium to indicated concentrations. All experiments were repeated in triplicate.

**4.2.2. FXR TR-FRET binding assay**—The FXR TR-FRET binding assay was performed as previously described<sup>23</sup> with modifications. Briefly, in a black polypropylene 384-well plate, DMSO, 10  $\mu\text{M}$  GW4064, and titrations of GW4064, lithocholic acid or the synthesized chemicals were mixed with 10 nM GST-hFXR-LBD, 1.5 nM Tb-anti-GST antibody, and 10 nM DY246 in a 20  $\mu\text{L}$ /well of coregulator buffer G supplemented with 10 mM DTT. The final DMSO concentration was 0.4% for all wells. The plate was spun down after a brief shake and incubated at room temperature for 20 min. The TR-FRET emission signals at 520 nm and 490 nm for each well were determined using a PHERAstar plate reader (BMG LABTECH, Cary, NC) and an excitation wavelength of 337 nm, 100  $\mu\text{s}$  delay time, and 200  $\mu\text{s}$  integration time. The 520 nm/490 nm ratio from each well was then calculated. The DMSO group and 10  $\mu\text{M}$  GW4064 group served as negative (0% inhibition) and positive (100% inhibition) controls, respectively. The % inhibition of tested chemicals at given concentration was calculated based on negative and positive controls using equation 1.

$$\% \text{Inhibition} = 100\% - 100\% \times \frac{\text{Chemical}_{520\text{nm}/490\text{nm}} - 10\mu\text{M GW4064}_{520\text{nm}/490\text{nm}}}{\text{DMSO}_{520\text{nm}/490\text{nm}} - 10\mu\text{M GW4064}_{520\text{nm}/490\text{nm}}} \quad \text{Equation 1}$$

For those chemicals with a maximal % inhibition > 50% and activities that behaved in a dose-dependent manner, the inhibitory activities at different concentrations were fit into a

one-site competitive binding equation with GraphPad PRISM 6.01 (GraphPad Software, Inc., La Jolla, CA) to derive IC<sub>50</sub> values.

**4.2.3. FXR cell-based transactivation assay**—FXR transactivation assays were performed using GeneBLazer FXR-UAS-bla HEK 293T cells (Invitrogen). Cells were maintained according to the manufacturer's instructions. Briefly, cells were grown in DMEM supplemented with 10% dialyzed FBS, 0.1 mM non-essential amino acids, 25 mM HEPES, 100 µg/mL of hygromycin B, 100 µg/ml of zeocin, 100 units/mL penicillin and 100 µg/mL streptomycin in regular tissue culture flasks. Upon transactivation assay, DMSO, 400 nM GW4064, 10 µM GW4064, and titrations of GW4064, LCA or chemicals without 400 nM GW4064 (for FXR agonistic activity determination) or with 400 nM GW4064 (for FXR antagonistic activity determination) were mixed with cells (20,000 cells/well in a 30 µL) in assay medium (phenol red-free DMEM supplemented with 2% charcoal/dextran-treated FBS, 0.1 mM non-essential amino acids, 1 mM sodium pyruvate, 100 units/mL penicillin and 100 µg/mL streptomycin) in black 384-well tissue culture-treated clear bottom plates (Corning Incorporated, Corning, NY). As a background control, the wells in column 24 of each plate were filled with 30 µL assay medium along with 0.5% DMSO. The final DMSO concentration was 0.5% in each assay well. Plates were incubated (16 h, cell culture incubator at 37°C), after which 6 µl/well of loading solution with CCF4-AM substrate was added. Plates were then incubated in the dark for 90 min at room temperature before measuring the fluorescent emissions at 460 nm and 535 nm (using excitation at 400 nm) with an Envision plate reader with bottom read capacity. After background signal subtraction for individual emission channels, emission signals at 460 and 535 nm from each well were used to determine the ratio of 460 nm/535 nm. For agonist assays, the DMSO group and 10 µM GW4064 group served as negative (0% activation) and positive (100% activation) controls, respectively. The % activation was calculated using equation 2.

$$\%Activation = 100\% \times \frac{Chemical_{460nm/535nm} - DMSO_{460nm/535nm}}{10\mu MGW4064_{460nm/535nm} - DMSO_{460nm/535nm}} \quad \text{Equation 2}$$

For antagonistic assays in which 400 nM GW4064 was included in the assay conditions, the DMSO group and 400 nM GW4064 group served as positive (100% inhibition) and negative (0% inhibition) controls, respectively. The % inhibition was calculated using equation 3.

$$\%Inhibition = 100\% - 100\% \times \frac{(Chemical + 400nMGW4064)_{460nm/535nm} - DMSO_{460nm/535nm}}{400nMGW4064_{460nm/535nm} - DMSO_{460nm/535nm}} \quad \text{Equation 3}$$

For those chemicals with a maximal % inhibition > 50% and activities that behaved in a dose-dependent manner, the inhibitory activities at different concentrations were fit into a sigmoidal dose-response equation with GraphPad PRISM 6.01 to derive IC<sub>50</sub> values.

**4.2.4. Cell-based cytotoxicity assay**—Cytotoxicity assays were performed side-by-side with the FXR agonistic assay using GeneBLazer FXR-UAS-bla HEK 293T cells and the CellTiter-Glo® Luminescent Cell Viability Assay reagent (Promega). Briefly, cells were

maintained in DMEM supplemented with 10% dialyzed FBS, 0.1 mM non-essential amino acids, 25 mM HEPES, 100 µg/mL of hygromycin B, 100 µg/mL of zeocin, 100 units/mL penicillin and 100 µg/mL streptomycin in regular tissue culture flasks. For the cytotoxicity assay, DMSO, titrations of GW4064, LCA or of chemicals were mixed with cells (20,000 cells/well in a 30 µL) in assay medium (phenol red-free DMEM supplemented with 2% charcoal/dextran-treated FBS, 0.1 mM non-essential amino acids, 1 mM sodium pyruvate, 100 units/mL penicillin and 100 µg/mL streptomycin) in black 384-well tissue culture-treated clear bottom plates. As a positive control, the wells in column 24 of each plate were filled with 30 µL assay medium along with 0.5% DMSO. The final DMSO concentration was 0.5% in each assay well. After a 16 h incubation in a cell culture incubator at 37°C, the cell plates were cooled at room temperature for 20 min and then CellTiter-Glo® reagent 20 µL/well was added followed by an incubation for 20 min in the dark to allow the development of optimal luminescence signal. The luminescence signal from each well was collected with an Envision plate reader equipped with Ultra-sensitive Luminescence detector. The DMSO group and no cell group served as negative (0% activation) and positive (100% activation) controls, respectively. The % inhibition was calculated using equation 4 below.

$$\%Inhibition = 100\% - 100\% \times \frac{Signal_{compound} - Signal_{No\ Cell}}{Signal_{DMSO} - Signal_{No\ Cell}} \quad \text{Equation 4}$$

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## ABBREVIATIONS USED

<b>HTS</b>	High-Throughput Screen
<b>FXR</b>	farnesoid X receptor
<b>NRs</b>	nuclear receptors
<b>Cyp7A1</b>	cytochrome 7A1
<b>BAs</b>	bile acids
<b>BSEP</b>	bile salt export pump
<b>SAR</b>	structure-activity relationship
<b>LBD</b>	ligand-binding domain

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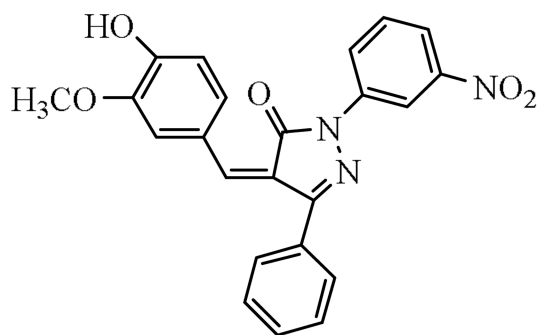
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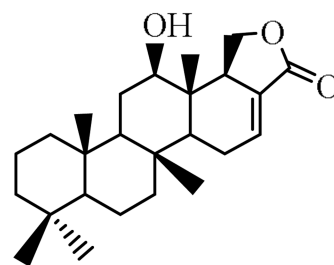


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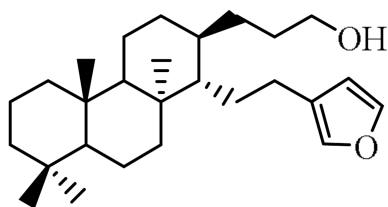
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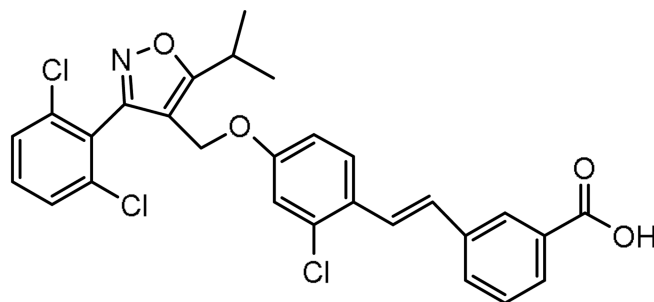
**1,3,4-Trisubstituted-pyrazolone Derivative**



**Scalarane Sesterterpene**



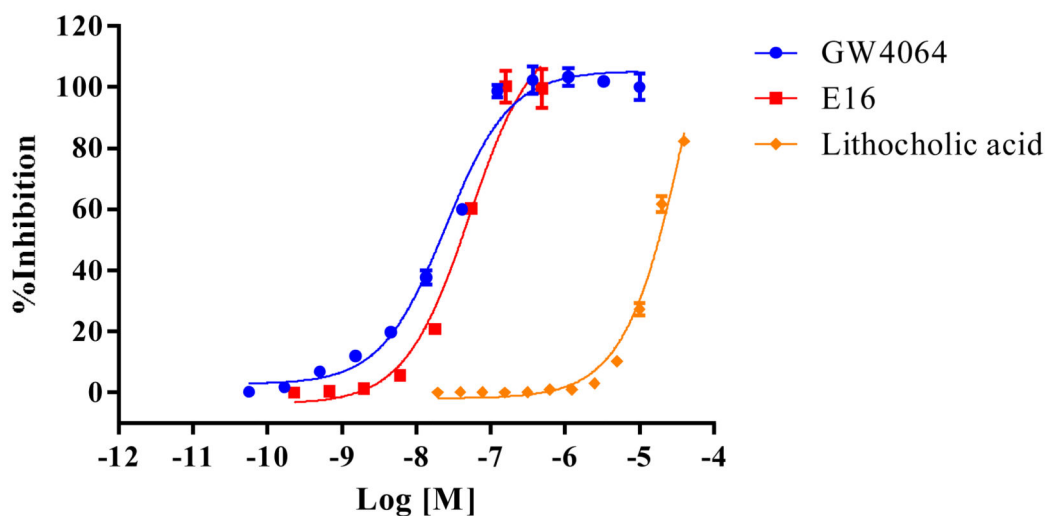
**Suvanine**



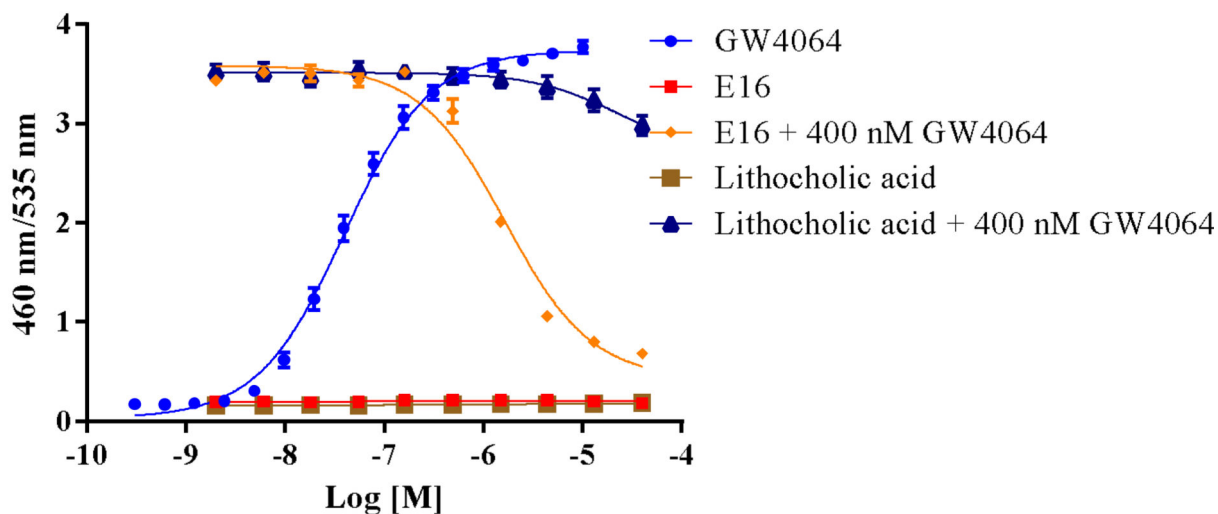
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**Figure 1.**  
Chemical structures of examples of FXR ligands.

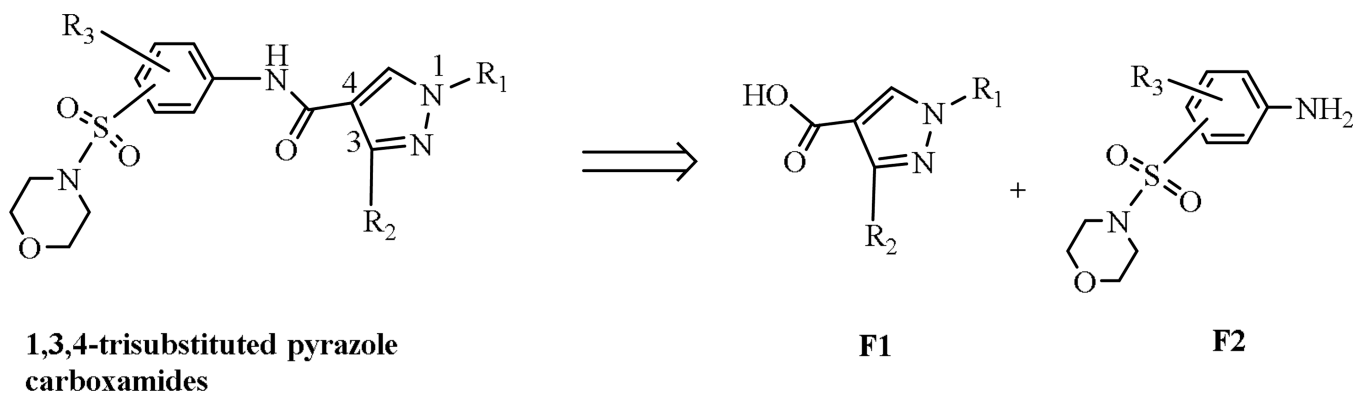
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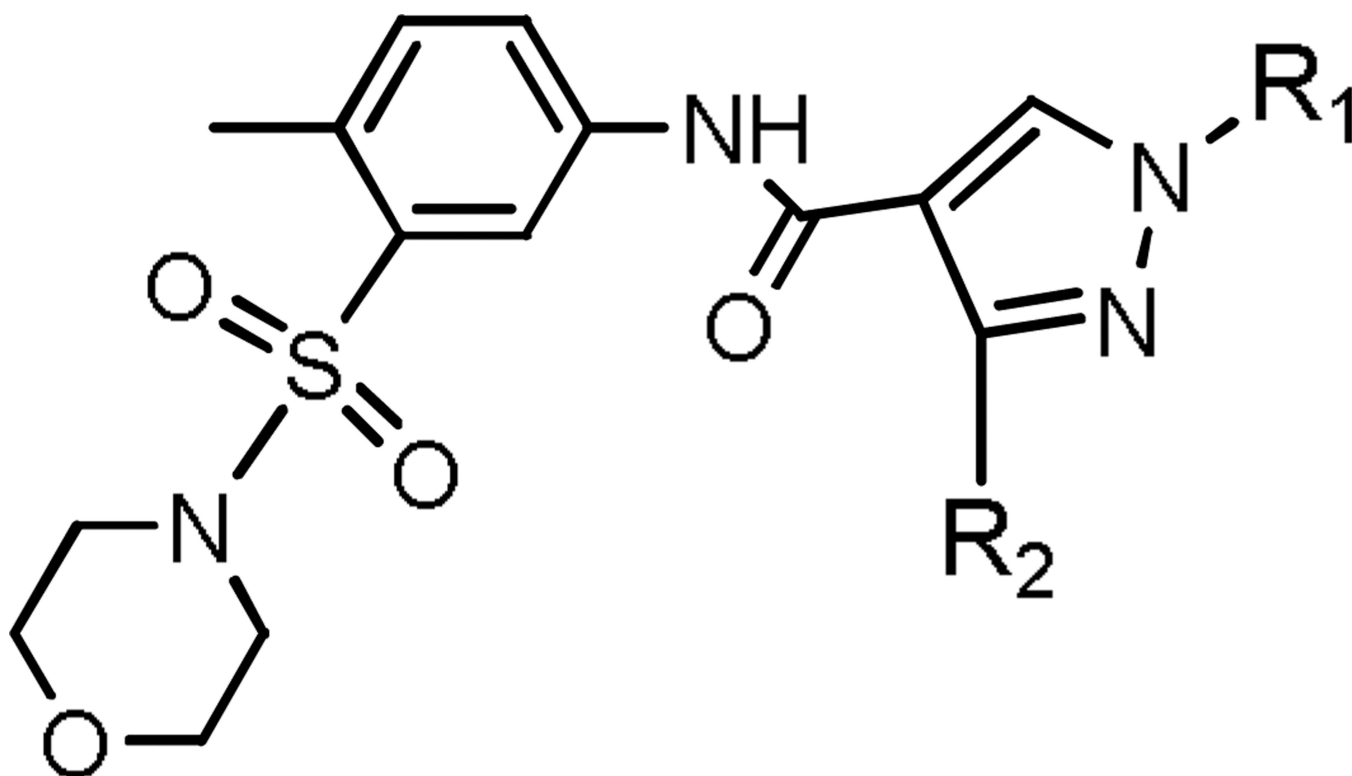
B

**Figure 2.**

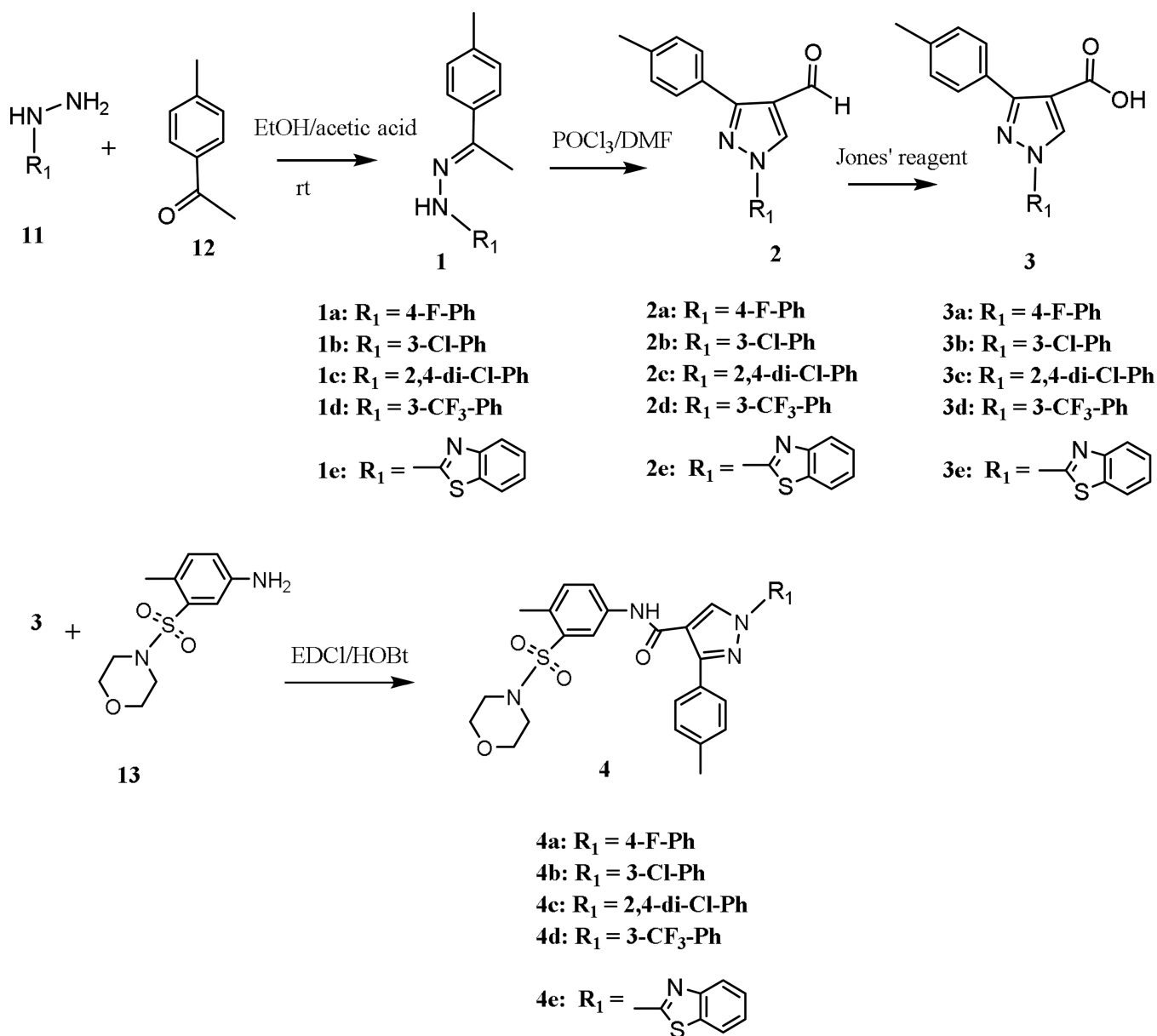
(A) FXR binding activities of GW4064, E16, and lithocholic acid in an FXR TR-FRET binding assay. (B) FXR agonistic activities (GW4064, E16, or lithocholic acid alone) and antagonistic activities (E16 and lithocholic acid in the presence of 400 nM GW4064) in a cell-based FXR transactivation assay. GW4064 and lithocholic acid were used as agonistic and antagonistic controls, respectively.



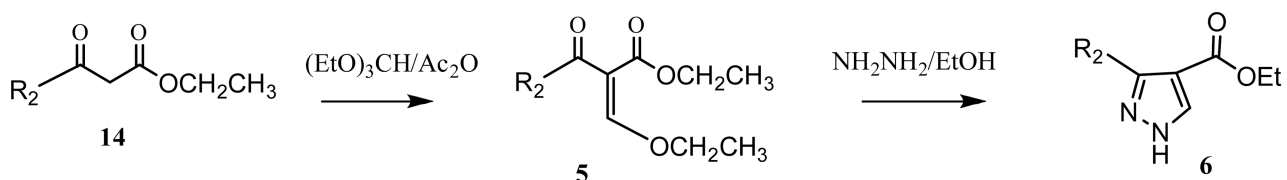
**Figure 3.**  
The retro-analysis of 1,3,4-trisubstituted pyrazole carboxamides.



**Figure 4.**  
A focused two-component small array of pyrazoles designed for optimization.

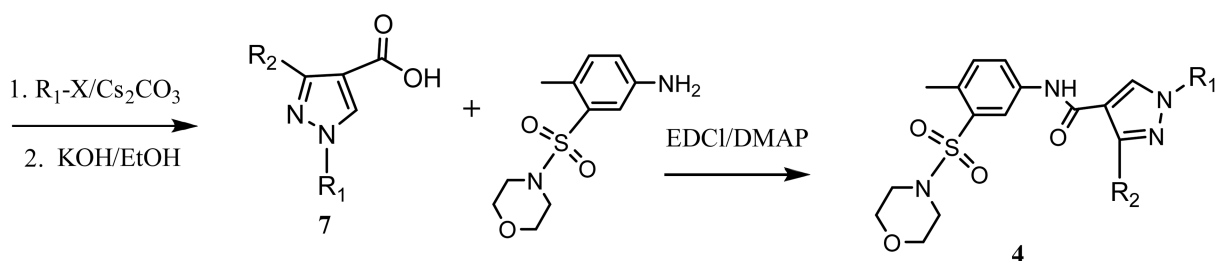


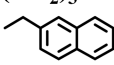
**Figure 5.** Initial exploration of pyrazole core via Vilsmeier-Haack reaction by the introduction of the first diversity element ( $R_1$ ).



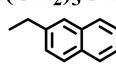
5a:  $R_2 = 4\text{-CH}_3\text{-Ph}$   
 5b:  $R_2 = \text{CH}(\text{CH}_3)_2$   
 5c:  $R_2 = 3\text{-OCH}_3\text{-Ph}$   
 5d:  $R_2 = 4\text{-CF}_3\text{-Ph}$   
 5e:  $R_2 = (\text{CH}_2)_2\text{-OCH}_3$

6a:  $R_2 = 4\text{-CH}_3\text{-Ph}$   
 6b:  $R_2 = \text{CH}(\text{CH}_3)_2$   
 6c:  $R_2 = 3\text{-OCH}_3\text{-Ph}$   
 6d:  $R_2 = 4\text{-CF}_3\text{-Ph}$   
 6e:  $R_2 = (\text{CH}_2)_2\text{-OCH}_3$



7a:  $R_1 = \text{CH}(\text{CH}_3)_2$ ,  $R_2 = 4\text{-CH}_3\text{-Ph}$   
 7b:  $R_1 = (\text{CH}_2)_3\text{CH}_3$ ,  $R_2 = 4\text{-CH}_3\text{-Ph}$   
 7c:  $R_1 =$  ,  $R_2 = 4\text{-CH}_3\text{-Ph}$

7d:  $R_1 = \text{CH}_2\text{CH}(\text{CH}_2)_2$ ,  $R_2 = 4\text{-CH}_3\text{-Ph}$   
 7e:  $R_1 = \text{CH}_2\text{-3-OCH}_3\text{-Ph}$ ,  $R_2 = 4\text{-CH}_3\text{-Ph}$   
 7f:  $R_1 = \text{CH}_2\text{-3-OCF}_3\text{-Ph}$ ,  $R_2 = 4\text{-CH}_3\text{-Ph}$   
 7g:  $R_1 = (\text{CH}_2)_4\text{OH}$ ,  $R_2 = 4\text{-CH}_3\text{-Ph}$   
 7h:  $R_1 = \text{CH}_2\text{-3-OCH}_3\text{-Ph}$ ,  $R_2 = \text{CH}(\text{CH}_3)_2$   
 7i:  $R_1 = \text{CH}(\text{CH}_3)_2$ ,  $R_2 = 3\text{-OCH}_3\text{-Ph}$   
 7j:  $R_1 = \text{CH}_2\text{-3-OCH}_3\text{-Ph}$ ,  $R_2 = 3\text{-OCH}_3\text{-Ph}$   
 7k:  $R_1 = \text{CH}(\text{CH}_3)_2$ ,  $R_2 = 4\text{-CF}_3\text{-Ph}$   
 7l:  $R_1 = \text{CH}_2\text{-3-OCH}_3\text{-Ph}$ ,  $R_2 = 4\text{-CF}_3\text{-Ph}$   
 7m:  $R_1 = \text{CH}(\text{CH}_3)_2$ ,  $R_2 = (\text{CH}_2)_2\text{-OCH}_3$   
 7n:  $R_1 = \text{CH}_2\text{-3-OCH}_3\text{-Ph}$ ,  $R_2 = (\text{CH}_2)_2\text{-OCH}_3$

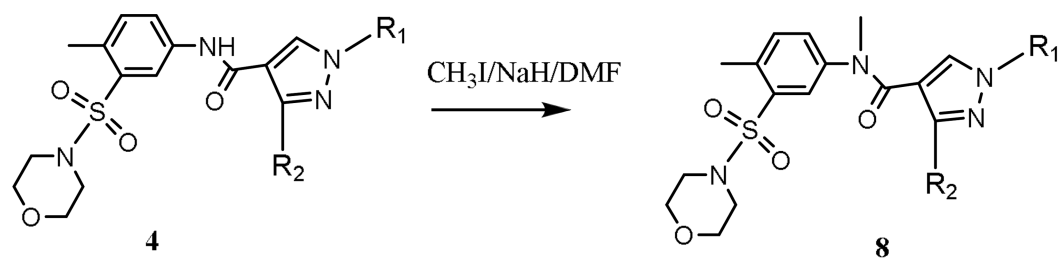
4f:  $R_1 = \text{CH}(\text{CH}_3)_2$ ,  $R_2 = 4\text{-CH}_3\text{-Ph}$   
 4g:  $R_1 = (\text{CH}_2)_3\text{CH}_3$ ,  $R_2 = 4\text{-CH}_3\text{-Ph}$   
 4h:  $R_1 =$  ,  $R_2 = 4\text{-CH}_3\text{-Ph}$

4i:  $R_1 = \text{CH}_2\text{CH}(\text{CH}_2)_2$ ,  $R_2 = 4\text{-CH}_3\text{-Ph}$   
 4j:  $R_1 = \text{CH}_2\text{-3-OCH}_3\text{-Ph}$ ,  $R_2 = 4\text{-CH}_3\text{-Ph}$   
 4k:  $R_1 = \text{CH}_2\text{-3-OCF}_3\text{-Ph}$ ,  $R_2 = 4\text{-CH}_3\text{-Ph}$   
 4l:  $R_1 = (\text{CH}_2)_4\text{OH}$ ,  $R_2 = 4\text{-CH}_3\text{-Ph}$   
 4m:  $R_1 = \text{CH}_2\text{-3-OCH}_3\text{-Ph}$ ,  $R_2 = \text{CH}(\text{CH}_3)_2$   
 4n:  $R_1 = \text{CH}(\text{CH}_3)_2$ ,  $R_2 = 3\text{-OCH}_3\text{-Ph}$   
 4o:  $R_1 = \text{CH}_2\text{-3-OCH}_3\text{-Ph}$ ,  $R_2 = 3\text{-OCH}_3\text{-Ph}$   
 4p:  $R_1 = \text{CH}(\text{CH}_3)_2$ ,  $R_2 = 4\text{-CF}_3\text{-Ph}$   
 4q:  $R_1 = \text{CH}_2\text{-3-OCH}_3\text{-Ph}$ ,  $R_2 = 4\text{-CF}_3\text{-Ph}$   
 4r:  $R_1 = \text{CH}(\text{CH}_3)_2$ ,  $R_2 = (\text{CH}_2)_2\text{-OCH}_3$   
 4s:  $R_1 = \text{CH}_2\text{-3-OCH}_3\text{-Ph}$ ,  $R_2 = (\text{CH}_2)_2\text{-OCH}_3$

**Figure 6.**

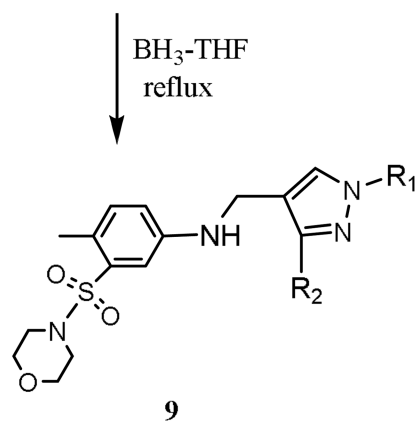
Alternative exploration of the pyrazole core to introduce  $R_1$  and  $R_2$  as substituted  $N^1$ - $C^3$ -substituted pyrazoles of **4f-4s**.





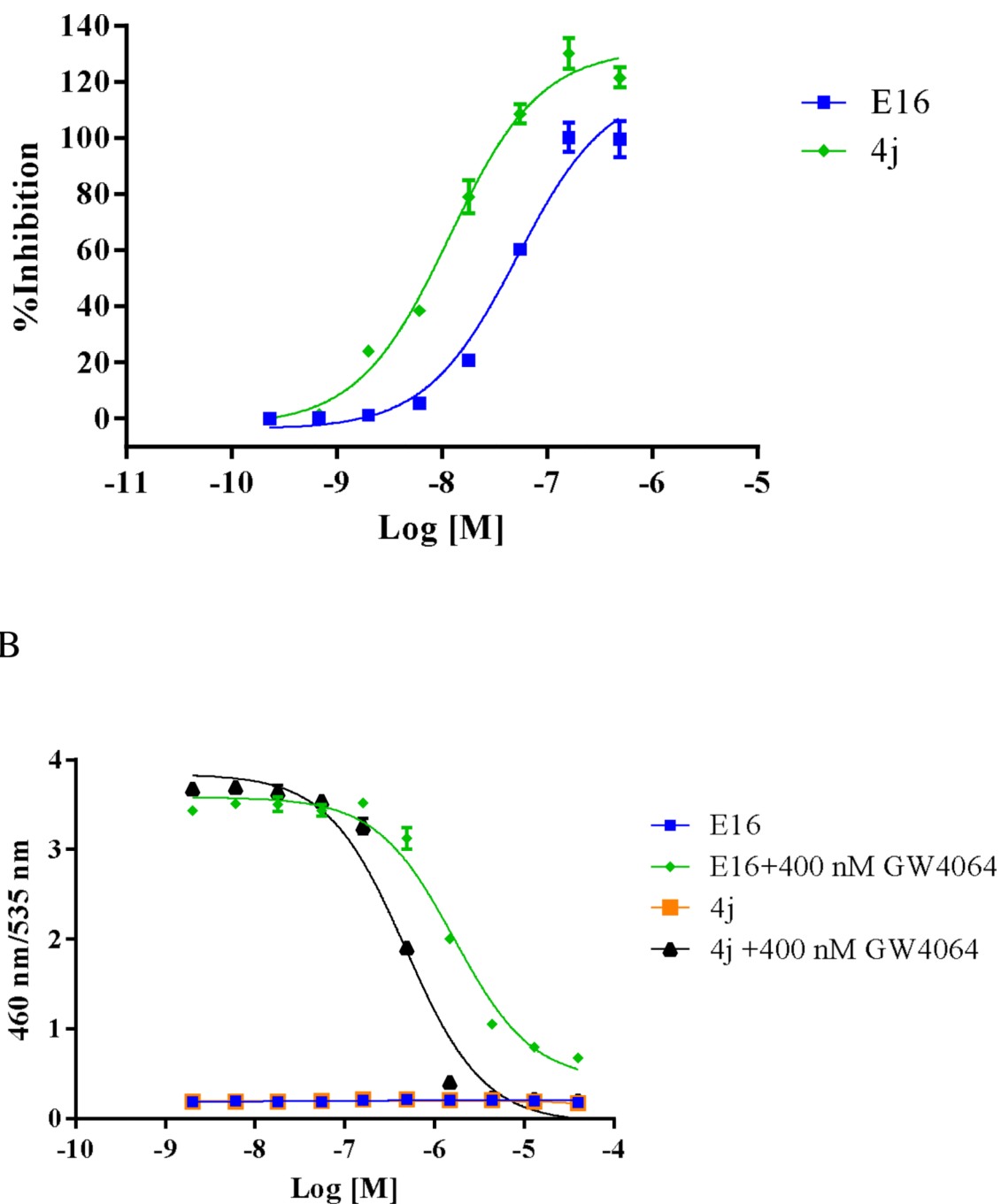
**4f:**  $\text{R}_1 = \text{CH}(\text{CH}_3)_2$ ,  $\text{R}_2 = p\text{-CH}_3\text{-Ph}$   
**4m:**  $\text{R}_1 = \text{CH}_2\text{-3-OCH}_3\text{-Ph}$ ,  $\text{R}_2 = \text{CH}(\text{CH}_3)_2$   
**4o:**  $\text{R}_1 = \text{CH}_2\text{-3-OCH}_3\text{-Ph}$ ,  $\text{R}_2 = 3\text{-OCH}_3\text{-Ph}$

**8a:**  $\text{R}_1 = \text{CH}(\text{CH}_3)_2$ ,  $\text{R}_2 = p\text{-CH}_3\text{-Ph}$   
**8b:**  $\text{R}_1 = \text{CH}_2\text{-3-OCH}_3\text{-Ph}$ ,  $\text{R}_2 = \text{CH}(\text{CH}_3)_2$   
**8c:**  $\text{R}_1 = \text{CH}_2\text{-3-OCH}_3\text{-Ph}$ ,  $\text{R}_2 = 3\text{-OCH}_3\text{-Ph}$



**9a:**  $\text{R}_1 = \text{CH}(\text{CH}_3)_2$ ,  $\text{R}_2 = p\text{-CH}_3\text{-Ph}$   
**9b:**  $\text{R}_1 = \text{CH}_2\text{-3-OCH}_3\text{-Ph}$ ,  $\text{R}_2 = \text{CH}(\text{CH}_3)_2$   
**9c:**  $\text{R}_1 = \text{CH}_2\text{-3-OCH}_3\text{-Ph}$ ,  $\text{R}_2 = 3\text{-OCH}_3\text{-Ph}$

**Figure 7.**  
*N*-methylation and amide reduction of **4f**, **4m**, and **4o** gave corresponding compounds **8** and **9**.



**Figure 8.** (A) FXR binding activities of E16 and 4j in a FXR TR-FRET binding assay. (B) Agonistic (E16 and 4j) and antagonistic activities (E16 and 4j in an FXR cell-based transactivation assay).

**Table 1**

Structures of the 16 substituted pyrazole-4-carboxamides (from Enamine Small Molecule Compound Libraries).

<b>E 1</b>	<b>E2</b>	<b>E3</b>	<b>E4</b>
<b>E5</b>	<b>E6</b>	<b>E7</b>	<b>E8</b>
<b>E9</b>	<b>E10</b>	<b>E11</b>	<b>E12</b>
<b>E13</b>	<b>E14</b>	<b>E15</b>	<b>E16</b>

Table 2

Biological activities of the 16 substituted pyrazole-4-carboxamides as determined by FXR TR-FRET binding, cell-based FXR antagonistic and cytotoxic assays. None of the 16 compounds showed FXR agonistic activity (data not shown).

Compound	FXR TR-FRET Binding		GB FXR Cell Anta.		GB FXR Cell Cyto	
	%Inh.	IC <sub>50</sub>	%Inh.	IC <sub>50</sub>	%Inh.	IC <sub>50</sub>
E1	21.2±0.6 (164 nM)	NA	34.7±1.2 (40 μM)	NA	NT	NA
E2	70.6±4.2 (13.3 μM)	3.3±0.6 μM	99.0±3.7 (40 μM)	11.8±0.6 μM	47.4±1.4 (40 μM)	NA
E3	11.6±0.2 (54.8 nM)	NA	27.0±0.3 (40 μM)	NA	NT	NA
E4	83.8±1.4 (13.3 μM)	8.3±0.9 μM	80.3±0.1 (40 μM)	9.4±0.9 μM	27.4±1.2 (40 μM)	NA
E5	99.8±6.1 (13.3 μM)	996±67 nM	101.1±2.7 (40 μM)	11.8±1.1 μM	51.3±1.7 (40 μM)	NA
E6	12.7±0.9 (493 nM)	NA	18.6±0.4 (40 μM)	NA	7.4±0.3 (40 μM)	NA
E7	44.4±1.9 (493 nM)	NA	54.9±2.1 (40 μM)	NA	7.1±0.3 (40 μM)	NA
E8	53.3±0.6 (493 nM)	NA	42.6±0.9 (40 μM)	NA	16.3±0.2 (40 μM)	NA
E9	90.4±4.0 (4.44 μM)	1.3±0.2 μM	82.3±2.6 (40 μM)	10.8±0.4 μM	NT	NA
E10	85.1±0.9 (13.3 μM)	6.6±0.7 μM	66.9±3.0 (40 μM)	29.3±1.7 μM	NT	NA
E11	36.9±1.5 (54.8 nM)	NA	38.8±1.7 (40 μM)	NA	NT	NA
E12	17.8±0.2 (54.8 nM)	NA	20.6±0.3 (40 μM)	NA	NT	NA
E13	27.5±0.9 (493 nM)	NA	32.8±1.1 (40 μM)	NA	5.4±0.2 (40 μM)	NA
E14	18.8±1.2 (13.3 μM)	NA	16.1±0.5 (40 μM)	NA	27.9±0.7 (40 μM)	NA
E15	23.0±0.5 (1.48 μM)	NA	24.0±0.9 (40 μM)	NA	10.3±0.3 (40 μM)	NA
E16	100.2±5.2 (164 nM)	47.0±5.1 nM	77.2±2.2 (40 μM)	2.62±0.13 μM	17.9±0.5 (40 μM)	NA

NA: not applicable

NT: Not toxic (toxicity less than 5% of control)

GB FXR Cell Anta: GeneBLAzer FXR cell-based antagonistic assay

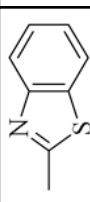
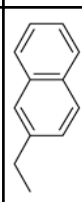
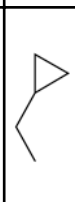
GB FXR Cell Cyto: Cytotoxicity assay performed in the GeneBLAzer FXR cells

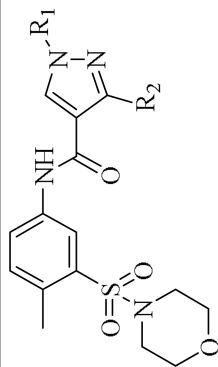
<sup>a</sup>IC<sub>50</sub> values represent the means ± SE of at least three independent experiments derived as described in the EXPERIMENTAL SECTION







%Inh: %Inhibition is presented as the means ± SE of at least three independent experiments calculated as described in the EXPERIMENTAL SECTION. The value shown in the table represents the maximum percent inhibition observed, and its lowest corresponding compound concentration is noted in the parentheses.

Table 3

The effect of varying the substitution pattern of the 1,3,4-substituted-pyrazole-4-carboxamide derivatives on their biological activities.

Compds	R <sub>1</sub>	R <sub>2</sub>	FXR TR-FRET Binding		GB FXR Cell Anta.		GB FXR Cell Cyto	
			%Inh.	IC <sub>50</sub> <sup>a</sup>	%Inh.	IC <sub>50</sub> <sup>a</sup>	%Inh.	IC <sub>50</sub>
GW4064			100±4.3 (10 µM)	19.8±2.5 nM	NA	NA	NT	NA
Lithocholic Acid			82.3±0.9 (40 µM)	18.3±1.2	16.1±0.5 (40 µM)	NA	NT	NA
E16	Ph	4-CH <sub>3</sub> -Ph	100.2±5.2 (164 nM)	47.0±5.1 nM	77.2±2.2 (40 µM)	2.62±0.13 µM	17.9±0.5 (40 µM)	NA
4a	4-F-Ph	4-CH <sub>3</sub> -Ph	95.0±1.2 (493 nM)	24.3±0.6 nM	72.0±2.2 (40 µM)	2.49±0.0,06 µM	18.5±1.0 (40 µM)	
4b	3-Cl-Ph	4-CH <sub>3</sub> -Ph	99.3±1.1 (164 nM)	10.4±0.9 nM	60.6±0.9 (40 µM)	5.91±0.15 µM	NT	NA
4c	2,4-Di-Cl-Ph	4-CH <sub>3</sub> -Ph	119.8±7.8 (164 nM)	14.1±0.3 nM	89.9±1.9 (40 µM)	1.24±0.12 µM	NT	NA
4d	3-CF <sub>3</sub> -Ph	4-CH <sub>3</sub> -Ph	77.5±4.3 (164 nM)	23.9±2.1 nM	54.8±1.3 (40 µM)	22.29±1.14µM	7.3±0.2 (40 µM)	NA
4e		4-CH <sub>3</sub> -Ph	101.8±2.8 (164 nM)	17.7±0.4 nM	79.1±2.4 (40 µM)	7.91±1.16 µM	11.5±0.2 (13.3 µM)	NA
4f	(CH <sub>3</sub> ) <sub>2</sub> CH	4-CH <sub>3</sub> -Ph	130.6±3.8 (4.4 µM)	49.1±9.9 nM	99.1±4.4 (40 µM)	1.69±0.18µM	NT	NA
4g	CH <sub>3</sub> (CH <sub>2</sub> ) <sub>3</sub>	4-CH <sub>3</sub> -Ph	130.1±0.7 (1.5 µM)	52.5±4.1 nM	97.9±8.1 (20 µM)	11.51±0.07 µM	10.3±0.3 (4.4 µM)	NA
4h		4-CH <sub>3</sub> -Ph	97.8±1.2 (164 nM)	10.2±1.6 nM	84.4±4.1 (40 µM)	844.1±10.3 nM	7.9±0.3 (4.4 µM)	NA
4i		4-CH <sub>3</sub> -Ph	105.0±4.9 (1.5 µM)	76.9±2.7 nM	100.1±3.2 (40 µM)	848.8±69.0 nM	NT	NA



Compds	R <sub>1</sub>	R <sub>2</sub>	FXR TR-FRET Binding		GB FXR Cell Anta.		GB FXR Cell Cyto	
			%Inh.	IC <sub>50</sub> <sup>a</sup>	%Inh.	IC <sub>50</sub> <sup>a</sup>	%Inh.	IC <sub>50</sub>
4j		4-CH <sub>3</sub> -Ph	130.1±5.4 (164 nM)	7.5±0.8 nM	98.9±4.3 (40 μM)	468.5±8.4 nM	NT	NA
4k		4-CH <sub>3</sub> -Ph	99.2±6.4 (164 nM)	18.4±1.2 nM	93.4±3.9 (40 μM)	515.0±25.4 nM	NT	NA
4l	HO(CH <sub>2</sub> ) <sub>4</sub>	4-CH <sub>3</sub> -Ph	89.3±0.1 (493 nM)	130.3±16 nM	97.6±3.7 (40 μM)	2.61 ±0.29 μM	5.9±0.2 (40 μM)	NA
4m		CH(CH <sub>3</sub> ) <sub>2</sub>	100.6±5.6 (4.4 μM)	199.4±21.2 nM	99.2±2.7 (40 μM)	3.11 ±0.58 μM	NT	NA
4n	(CH <sub>3</sub> ) <sub>2</sub> CH	3-CH <sub>3</sub> O-Ph	149.8±6.9 (13.3 μM)	159.7±22.6 nM	97.8±1.5 (40 μM)	3.49 ±0.64 μM	10.8±0.2 (4.4 μM)	
4o		3-CH <sub>3</sub> O-Ph	114.1±3.3 (493 nM)	42.8±0.5 nM	89.4±4.2 (40 μM)	1.24 ±0.09 μM	NT	NA
4p	(CH <sub>3</sub> ) <sub>2</sub> CH	4-CF <sub>3</sub> -Ph	133.2±5.3 (4.4 μM)	77.5±3.7 nM	100.0±3.7 (40 μM)	1.95 ±0.28 μM	19.5±1.1 (40 μM)	NA
4q		4-CF <sub>3</sub> -Ph	109.1±3.6 (164 nM)	30.9±3.6 nM	97.6±0.9 (40 μM)	592.7±3.0 nM	NT	NA
4r	(CH <sub>3</sub> ) <sub>2</sub> CH	CH <sub>3</sub> O(CH <sub>2</sub> ) <sub>2</sub>	98.2±3.8 (40 μM)	3.42±0.19 μM	62.5±0.8 (40 μM)	29.58 ±1.75 μM	NT	NA
4s		CH <sub>3</sub> O(CH <sub>2</sub> ) <sub>2</sub>	113.5±0.8 (13.3 μM)	389.5±9.4 nM	95.9±2.4 (40 μM)	11.1 ±0.64 μM	NT	NA

NA: not applicable

NT: Not toxic (toxicity less than 5%)

GB FXR Cell Antia: GeneBL/Azer FXR cell-based antagonistic assay



GB FXR Cell Cyto: Cytotoxicity assay performed in the GeneBL/Azer FXR cells

<sup>4</sup>IC<sub>50</sub> values represent the means  $\pm$  SE of at least three independent experiments derived as described in the EXPERIMENTAL SECTION

% Inh: % Inhibition is presented as the means  $\pm$  SE of at least three independent experiments calculated as described in the EXPERIMENTAL SECTION. The value shown in the table represents the maximum percent inhibition observed, and its lowest corresponding compound concentration is noted in the parentheses.

Table 4

The effect of varying the substitution pattern of the 1,3,4-substituted-pyrazole-4-carboxamide derivatives on their biological activities (*N*-methylation).

Cmpd	R <sub>1</sub>	R <sub>2</sub>	FXR TR-FRET Binding		GB FXR Cell Anta.		GB FXR Cell Cyto	
			%Inh.	IC <sub>50</sub>	%Inh.	IC <sub>50</sub>	%Inh.	IC <sub>50</sub>
8a	(CH <sub>3</sub> ) <sub>2</sub> CH	4-CH <sub>3</sub> -Ph	132.9±6.2 (40 μM)	489.4±21.2 nM	86.4±2.8 (40 μM)	16.16±0.28 μM	15.0±0.7 (40 μM)	NA
8b		(CH <sub>3</sub> ) <sub>2</sub> CH	80.3±5.2 (40 μM)	5.33±0.24 μM	2.4±0.1 (20 μM)	NA	13.5±0.4 (40 μM)	NA
8c		3-CH <sub>3</sub> -O-Ph	69.7±3.5 (40 μM)	3.81±0.20 μM	23.8±0.4 (40 μM)	NA	19.7±0.4 (40 μM)	NA

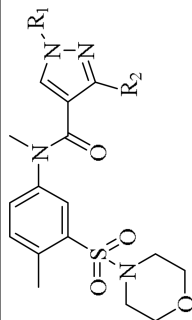
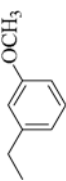
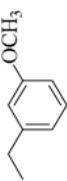




Table 5

The effect of varying the substitution pattern of the 1,3,4-substituted-pyrazole-4-carboxamide derivatives on their biological activities (amide reduction).

Cmpd	R <sub>1</sub>	R <sub>2</sub>	FXR TR-FRET Binding		GB FXR Cell Anta.		GB FXR Cell Cyto	
			%Inh.	IC <sub>50</sub>	%Inh.	IC <sub>50</sub>	%Inh.	IC <sub>50</sub>
9a	(CH <sub>3</sub> ) <sub>2</sub> CH	4-CH <sub>3</sub> -Ph	99.5±2.6 (13.3 μM)	2.04±0.02 μM	36.8±0.3 (40 μM)	NA	10.5±0.3 (40 μM)	NA
9b		(CH <sub>3</sub> ) <sub>2</sub> CH	72.5±2.7 (4.4 μM)	2.31±0.32 μM	64.8±1.9 (40 μM)	27.83 ±1.54 μM	13.9±0.6 (40 μM)	NA
9c		3-CH <sub>3</sub> O-Ph	39.8±1.8 (1.48 μM)	NA	3.1±0.1 (40 μM)	NA	NT	NA

