

Published in final edited form as:

Obesity (Silver Spring). 2011 March ; 19(3): 662–666. doi:10.1038/oby.2010.180.

MC4R Variant Is Associated With BMI but Not Response to Resistance Training in Young Females

Funda E. Orkunoglu-Suer¹, Brennan T Harmon¹, Heather Gordish-Dressman¹, Priscilla M. Clarkson², Paul D. Thompson³, Theodore J. Angelopoulos⁴, Paul M. Gordon⁵, Monica J. Hubal¹, Niall M. Moyna⁶, Linda S. Pescatello⁷, Paul S. Visich⁸, Robert F. Zoeller⁹, Eric P. Hoffman¹, and Joseph M. Devaney¹

¹Research Center for Genetic Medicine, Children's National Medical Center, Washington, DC, USA ²Department of Kinesiology, University of Massachusetts, Amherst, Massachusetts, USA ³Division of Cardiology, Henry Low Heart Center, Hartford Hospital, Hartford, Connecticut, USA ⁴Center for Lifestyle Medicine, Department of Health Professions, University of Central Florida, Orlando, Florida, USA ⁵Laboratory for Physical Activity and Exercise Intervention Research, University of Michigan, Ann Arbor, Michigan, USA ⁶School of Health and Human Performance, Dublin City University, Dublin, Ireland ⁷Department of Kinesiology, Human Performance Laboratory, University of Connecticut, Storrs, Connecticut, USA ⁸Human Performance Laboratory, Central Michigan University, Mount Pleasant, Michigan, USA ⁹Department of Exercise Science and Health Promotion, Florida Atlantic University, Davie, Florida, USA

Abstract

Recently, a genome-wide association study (GWAS) that identified eight single-nucleotide polymorphisms (SNPs) associated with BMI highlighted a possible neuronal influence on the development of obesity. We hypothesized these SNPs would govern the response of BMI and subcutaneous fat to resistance training in young individuals (age = 24 years). We genotyped the eight GWAS-identified SNPs in the article by Willer *et al.* in a cohort ($n = 796$) that undertook a 12-week resistance-training program. Females with a copy of the rare allele (C) for rs17782313 (*MC4R*) had significantly higher BMIs (CC/CT: $n = 174$; 24.70 ± 0.33 kg/m², TT: $n = 278$; 23.41 ± 0.26 kg/m², $P = 0.002$), and the SNP explained 1.9% of overall variation in BMI. Males with a copy of the rare allele (T) for rs6548238 (*TMEM18*) had lower amounts of subcutaneous fat pretraining (CT/TT: $n = 65$; $156,534 \pm 7,415$ mm³, CC: $n = 136$; $177,825 \pm 5,139$ mm³, $P = 0.019$) and males with a copy of the rare allele (A) for rs9939609 (*FTO*) lost a significant amount of subcutaneous fat with exercise (AT/AA: $n = 83$; $-798.35 \pm 2,624.30$ mm³, TT: $n = 47$; $9,435.23 \pm 3,494.44$ mm³, $P = 0.021$). Females with a copy of the G allele for a missense variant in the *SH2B1* (rs7498665) was associated with less change of subcutaneous fat volume with exercise

© 2010 The Obesity Society

Correspondence: Funda E. Orkunoglu-Suer (fsuer@cnmcresearch.org) or Joseph M. Devaney (jdevaney@cnmcresearch.org).

SUPPLEMENTARY MATERIAL

Supplementary material is linked to the online version of the paper at <http://www.nature.com/oby>

All authors contributed to the final critical revision of the manuscript.

Disclosure

The authors declared no conflict of interest.

(AG/GG: $n = 191$; $9,813 \pm 2,250 \text{ mm}^3$ vs. AA: $n = 126$; $770 \pm 2,772 \text{ mm}^3$; $P = 0.011$). These data support the original finding that there is an association between measures of obesity and a variant near the *MC4R* gene and extends these results to a younger population and implicates *FTO*, *TMEM18*, and *SH2B1* polymorphisms in subcutaneous fat regulation.

The World Health Organization estimates that there are one billion adults who are overweight (BMI $>25 \text{ kg/m}^2$), with 300 million of these individuals clinically obese (BMI $>30 \text{ kg/m}^2$). These statistics suggest that Westernized societies are yielding to the global epidemic of obesity (1). Science has progressed to the point where we have begun to understand the biology behind obesity and illustrated the enormous role that genetic–environment interactions ($G \times E$) play in the obese state.

There have been numerous genome-wide association studies (GWAS) in the past 2 years that have discovered loci for BMI (2–6). Previously, we examined the effect of one of these loci (*INSIG2*; rs7566605) on the interaction between resistance training and subcutaneous fat levels in healthy college-aged individuals (age = 24 years) (7). We found that the *INSIG2* polymorphism underlies variation in subcutaneous adiposity in young adult females and suppresses the positive effects of resistance training in young men.

In this study, eight newly revealed GWAS loci (*NEGR1* (rs2815752), *MTCH2* (rs10838738), *TMEM18* (rs6548238), *GNPDA2* (rs10938397), *SH2B1* (rs7498665), *MC4R* (rs17782313), *FTO* (rs9939609), and *KCTD15* (rs11084753)) that have been associated with increased BMI in individuals aged 43–65 years were examined for associations with BMI in a younger population (4). We hypothesized that these loci would show a stronger effect in young people due to removal of interactions with age. In addition, we wanted to examine the BMI-associated loci for a potential role in influencing changes in BMI and subcutaneous fat in response to resistance training.

Methods

Subjects

The 796 individuals included in this report were white college-aged adults living within the United States and Ireland who participated in a supervised resistance-training program as part of the Functional SNPs Associated with Muscle Size and Strength (FAMUSS) study (8) (clinical characteristics are reported in Table 1). The study protocol was approved by an ethics committee at Children’s National Medical Center (institutional review board protocol #2449) and at all testing sites.

Measures of obesity

Anthropometric measures were collected using a protocol standardized across all study centers. Height and weight were measured using a calibrated wall-mounted stadiometer and scale, respectively. BMI was calculated as weight in kilogram divided by height in meters squared (kg/m^2). In addition to BMI, magnetic resonance imaging measurements of subcutaneous fat were used as another measure of adiposity (7). We were not able to obtain

magnetic resonance images for all FAMUSS study participants (see Table 1); therefore, data from only 546 of the 796 total subjects were used for analysis of subcutaneous fat volume.

Exercise training program

A 12-week resistance-training program was performed with the nondominant arm (trained arm). The protocol has been described elsewhere (7–9).

Genotyping

Genomic DNA was isolated from peripheral blood lymphocytes with the Gentra Puregene DNA extraction kit (Qiagen, Valencia, CA) and genotyping was completed using Taqman assays from ABI (Foster City, CA) according to the manufacturer's instructions (see Table 2 for assay IDs). The end point fluorescent readings were performed on an ABI 7900HT Sequence Detection System (SDS V 2.3 software; Applied Biosystems, Foster City, CA).

Statistical analyses

Hardy–Weinberg equilibrium was tested for each single-nucleotide polymorphism (SNP) using a one-degree of freedom χ^2 -test. Baseline BMI and subcutaneous fat volume were analyzed as continuous quantitative traits. Due to large gender differences in baseline values, all analyses were performed separately for males and females. Bivariate correlation analyses of BMI and fat volume showed statistically significant correlations with age and baseline weight; therefore, genotype–phenotype associations were assessed using analysis of covariance, with BMI adjusted for age and fat volume adjusted for age and body weight. All significant associations from the main analysis of covariance model were subjected to pairwise statistical tests among the three genotype groups. Linear tests were performed between each of the genotype groups to determine which groups were significantly different from one another. Linear regression analysis, including likelihood ratio tests between full (containing genotype and covariates) and constrained (containing covariates only) models were performed to estimate the proportion of variance in volumetric measurements that was attributable to genotype. Analyses used a nominal *P* value of 0.05 as significant. The resulting *P* values from these tests were adjusted for multiple comparisons using the Sidak *post hoc* test. To address to some extent the problem of multiple testing, we compared our unadjusted *P* values to a significance level of 0.006 (0.05/8 SNPs) for BMI, subcutaneous fat, and changes in BMI and subcutaneous fat with unilateral-resistance training on the nondominant arm.

Results

We studied participants in the FAMUSS study of young adults (average age 24 years), enrolled into a unilateral upper arm 12-week supervised resistance-training program. The demographics of our population of European-descent (white) individuals are provided in Table 1. We have divided the study population into two groups (BMI cohort and volumetric fat cohort) because we were not able to measure subcutaneous fat volumes from the magnetic resonance images of some subjects.

The weight gains in the university-based population studied are well documented (10). After the 12-week study period, females gained $0.87 \pm 3.21\%$ kg, (pre/postexercise: $65.23 \pm 12.81/65.75 \pm 12.80$ kg, $P < 0.0001$ and males gained $0.55 \pm 2.69\%$ kg of weight (pre/postexercise: $79.81 \pm 15.94/80.22 \pm 15.89$ kg, $P = 0.003$). Therefore females added 0.9% to their BMI and males added 0.7%. Both the trained and untrained arms in females and males as a whole gained subcutaneous fat volume after 12 weeks of resistance training, with the gains significant in females (3%, $P = 0.005$; 2.8%, $P = 0.13$ females and males, respectively).

We tested eight GWAS loci previously associated with BMI in older adults (*NEGR1* (rs2815752), *MTCH2* (rs10838738), *TMEM18* (rs6548238), *GNPDA2* (rs10938397), *SH2B1* (rs7498665), *MC4R* (rs17782313), *FTO* (rs9939609), and *KCTD15* (rs11084753)) (4). All eight variants were in Hardy–Weinberg equilibrium in our population of European-descent individuals. The allele frequencies for the SNPs are provided in Table 3. The common allele frequencies for our population (whites) were very similar to the allele frequencies for the HapMap CEPH population (Utah residents with ancestry from northern and western Europe).

Each locus was then tested for association with BMI and subcutaneous fat volume, and the change in these variables in response to resistance training (see Supplementary Table S1). The allele frequencies for the eight SNPs and the alleles associated with BMI in the Willer *et al.* GWAS (4) are provided in Table 3. One of the eight SNPs we tested was found to be associated with BMI in our young cohort. Specifically, the *MC4R* rs17782313 SNP was associated with BMI in females using a dominant model (CC/CT: $n = 174$; 24.70 ± 0.33 kg/m², TT: $n = 278$; 23.41 ± 0.26 kg/m², $P = 0.003$). This SNP explained 1.9% ($P = 0.002$) of the phenotype (Figure 1). There were no significant associations with BMI response to upper arm 12-week resistance training. *NEGR1* (rs2815752), *MTCH2* (rs10838738), *GNPDA2* (rs10938397), and *KCTD15* (rs11084753) SNPs did not show any associations with BMI, subcutaneous fat, and their response to resistance training in our young cohort.

Males with a copy of the rare allele (T) for rs6548238 (*TMEM18* gene) had lower amounts of subcutaneous fat pretraining (CT/TT: $n = 65$; $156,534 \pm 7,415$ mm³, CC: $n = 136$; $177,825 \pm 5,139$ mm³, $P = 0.019$). This SNP explained 0.12% of phenotype.

Two of the SNPs that we examined were associated with changes in subcutaneous fat with resistance training. Females with a copy of the G allele for a missense variant in the *SH2B1* gene (rs7498665) that changes a Thr to Ala at position 484 was associated with less of a difference in subcutaneous fat volume using a dominant model after the 12-week training period (AG/GG: $n = 191$; $9,813 \pm 2,250$ mm³ vs. AA: $n = 126$; $770 \pm 2,772$ mm³; $P = 0.011$). This SNP explained 2.0% of the phenotype in women. The *FTO* rs9939609 SNP was associated with difference in subcutaneous fat volume in men using a dominant model (AT/AA: $n = 83$; $-798.35 \pm 2,624.30$ mm³ vs. TT: $n = 47$; $9,435.23 \pm 3,494.44$ mm³; $P = 0.02$). This SNP explained 4.1% of the phenotype in men. However, adjustment for multiple testing showed only the baseline BMI in females associated with allele (C) for rs17782313 (*MC4R*) to remain statistically significant.

Discussion

The current obesity epidemic is related to decreased habitual physical activity levels, changes in dietary intake, and genetic predisposition to obesity (11). In this study, we hypothesized that GWAS loci associated with BMI in older populations may have a stronger genetic effect on BMI in a younger, healthy cohort. However, our results show that only one of the eight GWAS-identified SNPs was associated with BMI in our young population. Specifically, we ascertained that a variant near the *MC4R* gene (rs17782313) is associated with BMI values in young women. This work confirms one of the GWAS results from Willer *et al.* (4) for BMI and extends the findings to a younger population. The gender specificity observed for this variant warrants further investigation.

We found the rare allele for rs6548238 in the *TMEM18* gene to be associated with lower values of subcutaneous fat volume in the upper arm before the 12-week training period in males. This same allele (T) shows an association with lower values of BMI in 91,469 individuals as part of a large GWAS (4,5). The *TMEM18* gene is involved in the modulation of neural stem cell migration to glial cells (12). In addition, this gene is expressed at high levels in the brain and hypothalamus and points to a neuronal component as part of the development of obesity (4).

The young adult cohort studied completed a 12-week supervised unilateral resistance-training program of the upper nondominant arm. After completion of the resistance training, both males and females showed an increase in BMI and subcutaneous fat volume (see Table 1). We hypothesized that the alleles associated with higher BMI values in the GWAS from Willer *et al.* (4) would be associated with stunted changes in BMI and subcutaneous fat volume in response to resistance training. The response of subcutaneous fat to resistance training has been varied, with multiple studies showing a reduction in subcutaneous fat as measured by magnetic resonance imaging (13–15) and one study showing no change in subcutaneous fat (16) with resistance training. We discovered that males with a copy of the rare allele (A) for the *FTO* variant (rs9939609) lost a significant amount of subcutaneous fat in the upper arm with resistance training. This result is in agreement with a article by Mitchell *et al.* (17) that showed women with two copies of the rare allele for the rs8050136 SNP (in complete linkage disequilibrium with rs9939609) lost significantly more weight than carriers of the common allele with aerobic training (cycle ergometers or treadmill). However, another study showed that carriers of the common allele for rs8050136 lost three times greater percent body fat with aerobic exercise (18). Both of these studies used a different form of exercise (aerobic exercise) than the FAMUSS study (resistance training). In addition, both studies used different durations for their aerobic exercise program, with the Mitchell *et al.* (17) study using 6 months (3–4 sessions a week) and the Rankinen *et al.* (18) study using a 20-week program (3 sessions a week). Our data show that there is an interaction between the *FTO* rs9939609 SNP and resistance training in men. However, this association needs to be explored further and tested in other cohorts using different exercise modalities.

In addition, we discovered an interaction between a missense mutation in the *SH2B1* gene (rs7498665; Thr484Ala) with resistance training in women. Women with two copies of the

common allele (A allele; Thr) did not gain as much subcutaneous fat as women with a copy of the rare risk allele (G) after the unilateral- resistance training. The common allele has been shown to be associated with a lower value of BMI in 91,469 individuals as part of a large GWAS (4,5). Mice with a disruption of the *Sh2b1* gene resulted in metabolic disorders including hyperlipidemia, leptin resistance, hyperphagia, obesity, hyperglycemia, insulin resistance, and glucose intolerance (19). This gene is thought to regulate energy balance (19) and may interact with resistance training to promote weight homeostasis (20).

There are some limitations to our study. Only one arm was trained for the 12-week time period; therefore, individuals did not see any loss in weight or BMI, which has been shown for whole body resistance training (13–15). In addition, we do not have a replication group that provides insurance against errors and bias that can affect an individual study's results (21). Finally, three of the associations are not significant after correction for multiple testing but the results have biological relevance because of previous associations for BMI values (4,5). In addition, the statistical community is divided on correction methods for multiple testing as the Bonferonni correction may be too harsh and thus increasing false negatives (22). The results would need to be validated in additional populations (21).

In summary, we found that a variant near the *MC4R* gene (rs17782313) is associated with BMI in young females and a variant in the *TMEM18* gene (rs6548238) to be associated with subcutaneous fat volume in males. In addition, we found that the *FTO* SNP that has been associated with exercise response (17,18,23) has an effect on subcutaneous fat loss following resistance training in males and, a missense variant in the *SH2B1* gene was associated with a greater gain in subcutaneous fat following resistance training in women. These findings emphasize that genetics can be utilized in combination with resistance training to lower subcutaneous fat levels and may ultimately provide the public with genetic information that can influence exercise choice.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

This study was supported by a grant from the National Institutes of Health 3R01AR055100-06. F.E.O.-S. designed the study, genotyped the samples, analyzed data, and drafted the manuscript. B.T.H. assisted in genotyping. H.G.D. ran the statistics, analyzed and interpreted data and drafted the manuscript. P.M.C., M.J.H., P.D.T., T.J.A., P.M.G., N.M.M., L.S.P., P.S.V., and R.F.Z. recruited the subjects, collected the clinical data, and supervised exercise trainings. J.M.D. and E.P.H. designed the study and drafted the manuscript.

REFERENCES

1. Obesity: preventing and managing the global epidemic. Report of a WHO consultation. World Health Organ Tech Rep Ser. 2000; 894:i–xii. 1–253. [PubMed: 11234459]
2. Frayling TM, Timpson NJ, Weedon MN, et al. A common variant in the *FTO* gene is associated with body mass index and predisposes to childhood and adult obesity. *Science*. 2007; 316:889–894. [PubMed: 17434869]
3. Loos RJ, Lindgren CM, Li S, et al. Common variants near *MC4R* are associated with fat mass, weight and risk of obesity. *Nat Genet*. 2008; 40:768–775. [PubMed: 18454148]

4. Willer CJ, Speliotes EK, Loos RJ, et al. Six new loci associated with body mass index highlight a neuronal influence on body weight regulation. *Nat Genet.* 2009; 41:25–34. [PubMed: 19079261]
5. Thorleifsson G, Walters GB, Gudbjartsson DF, et al. Genome-wide association yields new sequence variants at seven loci that associate with measures of obesity. *Nat Genet.* 2009; 41:18–24. [PubMed: 19079260]
6. Herbert A, Gerry NP, McQueen MB, et al. A common genetic variant is associated with adult and childhood obesity. *Science.* 2006; 312:279–283. [PubMed: 16614226]
7. Orkunoglu-Suer FE, Gordish-Dressman H, Clarkson PM, et al. INSIG2 gene polymorphism is associated with increased subcutaneous fat in women and poor response to resistance training in men. *BMC Med Genet.* 2008; 9:117. [PubMed: 19105843]
8. Thompson PD, Moyna N, Seip R, et al. Functional polymorphisms associated with human muscle size and strength. *Med Sci Sports Exerc.* 2004; 36:1132–1139. [PubMed: 15235316]
9. Kostek MA, Pescatello LS, Seip RL, et al. Subcutaneous fat alterations resulting from an upper-body resistance training program. *Med Sci Sports Exerc.* 2007; 39:1177–1185. [PubMed: 17596787]
10. Cluskey M, Grobe D. College weight gain and behavior transitions: male and female differences. *J Am Diet Assoc.* 2009; 109:325–329. [PubMed: 19167962]
11. Campbell WW, Crim MC, Young VR, Evans WJ. Increased energy requirements and changes in body composition with resistance training in older adults. *Am J Clin Nutr.* 1994; 60:167–175. [PubMed: 8030593]
12. Jurvansuu J, Zhao Y, Leung DS, et al. Transmembrane protein 18 enhances the tropism of neural stem cells for glioma cells. *Cancer Res.* 2008; 68:4614–4622. [PubMed: 18559506]
13. Wilmore JH. Alterations in strength, body composition and anthropometric measurements consequent to a 10-week weight training program. *Med Sci Sports.* 1974; 6:133–138. [PubMed: 4461973]
14. Treuth MS, Ryan AS, Pratley RE, et al. Effects of strength training on total and regional body composition in older men. *J Appl Physiol.* 1994; 77:614–620. [PubMed: 8002507]
15. Staron RS, Leonardi MJ, Karaondo DL, et al. Strength and skeletal muscle adaptations in heavy-resistance-trained women after detraining and retraining. *J Appl Physiol.* 1991; 70:631–640. [PubMed: 1827108]
16. Häkkinen K, Pakarinen A, Kraemer WJ, Newton RU, Alen M. Basal concentrations and acute responses of serum hormones and strength development during heavy resistance training in middle-aged and elderly men and women. *J Gerontol A Biol Sci Med Sci.* 2000; 55:B95–B105. [PubMed: 10737684]
17. Mitchell JA, Church TS, Rankinen T, et al. FTO genotype and the weight loss benefits of moderate intensity exercise. *Obesity (Silver Spring).* 2010; 18:641–643. [PubMed: 19798072]
18. Rankinen T, Rice T, Teran-Garcia M, Rao DC, Bouchard C. FTO genotype is associated with exercise training-induced changes in body composition. *Obesity (Silver Spring).* 2010; 18:322–326. [PubMed: 19543202]
19. Ren D, Zhou Y, Morris D, et al. Neuronal SH2B1 is essential for controlling energy and glucose homeostasis. *J Clin Invest.* 2007; 117:397–406. [PubMed: 17235396]
20. Poirier P, Després JP. Exercise in weight management of obesity. *Cardiol Clin.* 2001; 19:459–470. [PubMed: 11570117]
21. Hattersley AT, McCarthy MI. What makes a good genetic association study? *Lancet.* 2005; 366:1315–1323. [PubMed: 16214603]
22. Rice TK, Schork NJ, Rao DC. Methods for handling multiple testing. *Adv Genet.* 2008; 60:293–308. [PubMed: 18358325]
23. Andreasen CH, Stender-Petersen KL, Mogensen MS, et al. Low physical activity accentuates the effect of the FTO rs9939609 polymorphism on body fat accumulation. *Diabetes.* 2008; 57:95–101. [PubMed: 17942823]

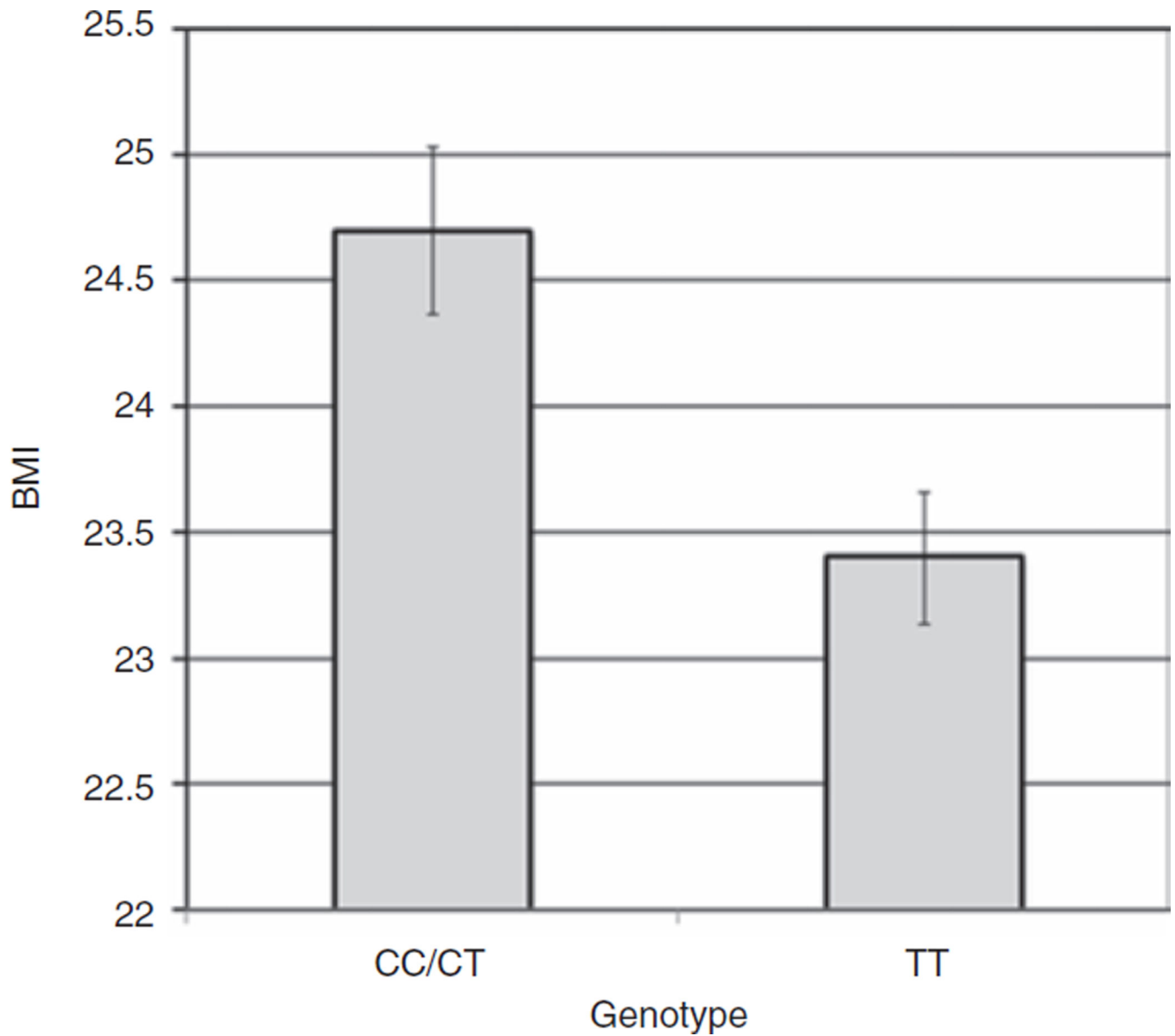


Figure 1.

Effect of genotype for the *MC4R* SNP rs17782313 on BMI levels in females. Females with a copy of the C allele for the *MC4R* SNP (rs17782313) showed higher values for BMI (CC/CT: $N = 174$; 24.70 ± 0.33 kg/m² vs. TT: $N = 278$; 23.41 ± 0.26 kg/m²; $P = 0.003$) and this accounted for 1.9% of the BMI phenotype ($P = 0.002$).

Table 1
 Demographic characteristics of the Functional SNPs Associated with Muscle Size and Strength cohort

Characteristic	BMI cohort				Volumetric fat cohort			
	Females		Males		Females		Males	
	N	Mean ± s.d.	N	Mean ± s.d.	N	Mean ± s.d.	N	Mean ± s.d.
Age (years)	475	23.12 ± 5.48	321	23.72 ± 5.47	335	22.81 ± 5.17	211	23.91 ± 5.66
Height (cm)	478	165.02 ± 6.80	323	177.97 ± 6.88	338	165.02 ± 7.00	212	178.19 ± 6.64
Pre-exercise weight (kg)	478	65.23 ± 12.81	323	79.77 ± 15.98	338	64.53 ± 12.17	212	78.19 ± 14.44
Postexercise weight (kg)	479	65.75 ± 12.79	323	80.22 ± 15.89	338	65.05 ± 12.29	213	78.73 ± 14.62
Pre-exercise BMI (kg/m ²)	478	23.95 ± 4.53	323	25.15 ± 4.70	338	23.68 ± 4.18	212	24.57 ± 4.02
Postexercise BMI (kg/m ²)	479	24.14 ± 4.51	323	25.30 ± 4.65	338	23.86 ± 4.25	213	24.76 ± 4.00
Difference in BMI (kg/m ²)	478	0.19 ± 0.87	322	0.13 ± 0.80	338	0.19 ± 0.82	212	0.18 ± 0.82
Pre-exercise subcutaneous fat volume (mm ³)					338	256,784 ± 114,086	213	170,126 ± 88,886
Postexercise subcutaneous fat volume (mm ³)					337	263,082 ± 116,803	210	174,163 ± 93,611
Difference in subcutaneous fat volume (mm ³)					337	6,181 ± 30,619	210	3,996 ± 23,857

Table 2

Genotyping assays for GWAS-discovered SNPs

dBSNP ID	Assay ID ^a
rs2815752	C__26668839_10
rs10838738	C___432493_10
rs6548238	C__29311887_10
rs10938397	C___1594245_10
rs7498665	C__25999166_10
rs17782313	C__32667060_10
rs9939609	C__30090620_10
rs11084753	C__31497814_10

GWAS, genome-wide association study; SNP, single-nucleotide polymorphism.

^aFrom Applied Biosystems website.

Table 3

Genotype frequencies of SNPs for participants in FAMUSS

dBSNP ID	Alleles (+/-)	Effect Allele ^a	Frequency in FAMUSS (+)	Published frequency for CEU (+) ^b	Allele (+) (%)	Heteros (%)	Allele (-) (%)
rs2815752	A/G	A	0.639	0.637	40.1	47.5	12.4
rs10838738	G/A	G	0.663	0.637	43.7	45.3	11.0
rs6548238	C/T	C	0.818	0.850	67.1	29.4	3.5
rs10938397	A/G	G	0.571	0.554	31.1	51.9	17.0
rs7498665	A/G	G	0.612	0.619	37.7	47.0	15.3
rs17782313	T/C	T	0.781	0.735	61.1	33.9	5.0
rs9939609	T/A	A	0.602	0.540	35.1	50.1	14.8
rs11084753	G/A	G	0.667	0.690	44.6	44.1	11.4

CEU, European Caucasian; FAMUSS, Functional SNPs Associated with Muscle Size and Strength; SNP, single-nucleotide polymorphism.

^a According to Willer *et al.* (4).^b Utah residents with northern and western European ancestry from the CEPH collection used in HAPMAP.