

## Video Article

# Optimized Protocols for *Mycobacterium leprae* Strain Management: Frozen Stock Preservation and Maintenance in Athymic Nude Mice

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## Abstract

Leprosy, caused by *Mycobacterium leprae*, is an important infectious disease that is still endemic in many countries around the world, including Brazil. There are currently no known methods for growing *M. leprae* *in vitro*, presenting a major obstacle in the study of this pathogen in the laboratory. Therefore, the maintenance and growth of *M. leprae* strains are preferably performed in athymic nude mice (NU-*Foxn1*<sup>nu</sup>). The laboratory conditions for using mice are readily available, easy to perform, and allow standardization and development of protocols for achieving reproducible results. In the present report, we describe a simple protocol for purification of bacilli from nude mouse footpads using trypsin, which yields a suspension with minimum cell debris and with high bacterial viability index, as determined by fluorescent microscopy. A modification to the standard method for bacillary counting by Ziehl-Neelsen staining and light microscopy is also demonstrated. Additionally, we describe a protocol for freezing and thawing bacillary stocks as an alternative protocol for maintenance and storage of *M. leprae* strains.

## Video Link

The video component of this article can be found at <http://www.jove.com/video/50620/>

## Introduction

Leprosy, a disease caused by *Mycobacterium leprae*, is an important public health problem in many countries around the world<sup>1,2</sup>. Despite being known as an infectious disease that affects both skin and peripheral nerves, there are still several gaps in knowledge about the mechanisms involved in the complex immunopathogenesis of the disease.

Among the challenging characteristics of *M. leprae* that hinder its study are its inability to grow in artificial culture media and its relatively long doubling time (approximately 14 days)<sup>3,4</sup>. Most experimental procedures that use *M. leprae* are set up with bacilli purified from skin lesions of leprosy patients or from experimental animal models, such as armadillos and several strains of mice<sup>5,6</sup>.

For many years, researchers have depended on purification of *M. leprae* from skin lesions of multibacillary leprosy patients for use in experimental procedures. Several laboratory conditions for the *in vitro* cultivation or maintenance of live *M. leprae* have been attempted, but to date, animal models have proved to be most suitable for research purposes. In the 1960s and 1970s, researchers began using mice and armadillos for detection and assessment of *M. leprae* viability, monitoring bacterial growth in response to anti-*M. leprae* drugs, and in the growth and maintenance of mycobacterial strains<sup>2,4</sup>.

The animal models have some limitations, especially in armadillos and nonhuman primates, including ethical concerns, cost of maintenance, special infrastructure needed for animal maintenance, and poor reproducibility of results and final yields. Currently, mice are the preferred model animal for leprosy research. The laboratory conditions for using mice are readily available, easy to perform, and allow standardized protocols<sup>7,8,9</sup>. Nude mice have been used in the maintenance of *M. leprae* strains because, even with low viable bacillary loads, the T-cell deficient immune response leads to exuberant granuloma formation and good bacillary multiplication. Thus, this animal model has gained broad acceptance within the leprosy research community.

In the present report, we describe a simple protocol for enzymatic purification of bacilli from nude mouse footpads, intended for preparation of a *M. leprae* suspension with minimal cell debris and with high bacterial viability index. The viability is determined by fluorescent microscopy, which can be rapidly performed in the laboratory. We also demonstrate a modification to the standard method for bacillary counting by cold Ziehl-

Neelsen staining and light microscopy. Additionally, we present an alternative protocol for maintenance and storage of *M. leprae* strains, allowing freezing and thawing of bacillary stocks.

## Protocol

### 1. Preparation of *Mycobacterium leprae* Suspension

- Before euthanizing the athymic nude mouse (NU-*Foxn1*<sup>nu</sup>), prepare the trypsin solution and culture media (Protocol of the reagents - Protocols 1, 2, and 3). Euthanize the nude mice 4-5 months post inoculation with *M. leprae* according to the AVMA Guidelines for the Euthanasia of Animals: 2013 Edition<sup>10</sup> (the experiments and use of images of animals were approved and conducted in accordance with the guidelines of the Animal Care Committee of Faculdade de Odontologia de Bauru, USP). Inside a biological safety cabinet used for handling animals, clean the entire mouse with 70% ethanol alcohol.
- Hold the rear paw using hemostatic forceps distal to the tarsal joint. Cut the paw off with scissors below the forceps and immerse the paw in 2% iodine for 20 min. Then, repeat the procedure for the other rear paw.
- Transfer the paws to a clean biological safety cabinet for further work. Take paws out of the 2% iodine, dry them with sterile gauze and then collect the two footpads using number 22 or 23 scalpel blade, removing all soft tissue (dermis, epidermis, tendons, and nerves) close to the bone. Remove the metatarsals bones and the 1st and 5th phalanges. Place the material in a sterile Petri dish and remove the epidermis, by scraping it off with the scalpel blade. Cut the tissue into small pieces with scissors and transfer them to a round bottom tube. Weight the tissue and add 1 ml of Hanks Balanced Salt Solution (HBSS). Keep the tube on ice to avoid warming up of the sample.
- Add another 1 ml of HBSS and homogenize the sample using a tissue homogenizer.
  - First homogenization cycle: 3 pulses of 15 sec at speed 4 (14,450 rpm).
  - Transfer the supernatant to a sterile 50 ml conical tube, passing it through a cell strainer to eliminate the remaining debris. Allow the solution to pass by gravity.
  - Add an additional 2 ml of HBSS, and repeat the homogenization cycle (Section 1.4.1), and pass the samples through the cell strainer again.
  - Rinse the strainer by adding HBSS to bring the volume to 9 ml. Discard the cell strainer.
- Thaw an aliquot of 0.5% trypsin solution. Add 1 ml of trypsin to the 9 ml of cell suspension to obtain a final concentration 0.05% trypsin. Discard remaining thawed trypsin. Incubate for 60 min in a 37 °C water bath. After incubation, bring the volume up to 40 ml by adding sterile saline to dilute the trypsin.
- Centrifuge at 1,700 x g for 30 min at 4°C.
- Discard the supernatant by carefully inverting the tube. Resuspend the pellet by tapping the tube and then add 1 ml of sterile saline. Transfer the suspension to a new conical tube measuring the final volume of the suspension (to calculate the yield/gram of tissue).
- Homogenize the bacillary suspension using a 1 ml insulin syringe with a 26 G needle while transferring the suspension to a new tube.
- Perform microbiological control of the suspension by placing 2 drops (50  $\mu$ l) of it in each medium: 7H9 and Lowenstein-Jensen medium and incubate at 37 °C for 30 days, for detection of contaminating mycobacteria. Similarly, inoculate a Brain Heart Infusion (BHI) medium and incubate for 24 hr at 37 °C, for the detection of contaminating aerobic bacteria.
- Aliquot the suspension into tubes for staining: 35  $\mu$ l for Cold Ziehl-Neelsen staining (Protocol 2), and 200  $\mu$ l for viability determination (Protocol 3).
- After Ziehl-Neelsen staining (ZN), calculate the number of acid fast bacilli/ml (AFB/ml). For nude mice inoculation, prepare a 1 x 10<sup>8</sup> AFB/ml suspension; make sure to have enough volume to inoculate 30  $\mu$ l/footpad/animal (Protocol 5), and keep the suspension cold until inoculation. For freezing, prepare a 1 x 10<sup>7</sup> AFB/ml suspension (Protocol 4).

### 2. Cold Ziehl-Neelsen Staining

- Before starting the staining, prepare the serum/phenol solution and the ZN staining solutions (Protocol of the reagents - Protocols 4 and 5). Draw three circles (internal diameter 10 mm) on three glass slides using an immunohistochemistry pen. Identify the slides: (1) normal or undiluted, (2) 1:10, and (3) 1:100 using a number 2 pencil (Note: some graphite fades off after ZN staining).
- Dilute the bacillary suspension (Step 1.10) to 1:10 and 1:100 serial dilutions, *i.e.* add 5  $\mu$ l of undiluted suspension and 45  $\mu$ l of saline, then take 5  $\mu$ l of 1:10 and add 45  $\mu$ l of saline.
- Add 5  $\mu$ l of the serum/phenol solution and 10  $\mu$ l of bacillary suspension per circle. Homogenize and spread evenly around the area of the circle with the handle of a disposable loop. Wait for the suspension to dry on a leveled table.
- After drying, fix the smear by passing the slide 3 times over the blue flame of a Bunsen burner (around 20 sec total)<sup>11</sup>.
- For staining, cover the entire surface of the slide with filtered Ziehl-Neelsen carbofuchsin (about 5 ml) for 20 min.
- Rinse the slide in running water (slow flow).
- Cover the slide with a 10% acid alcohol solution for about 20 sec.
- Wash the slide again in running water.
- Cover the slide with a methylene blue solution for 5 min.
- Rinse the slide in running water and let it dry at room temperature.
- Count 20 fields/circle (total 60 fields/slide) using a 100X oil immersion objective. The calculation of acid fast bacilli/ml (AFB/ml) is done as follows, according to the description in the Laboratory Techniques for Leprosy<sup>12</sup>:  

$$\text{AFB/field} = \text{total number of bacilli found in the 3 circles (60 fields) divided by 60.}$$

$$\text{AFB/ml} = \text{AFB/field} \times \text{constant (area of the circle} \times 100 \text{ divided by area of the objective lens aperture), and if counting a diluted suspension, multiply the number of AFB/ml by the dilution factor (10 or 100).}$$

### 3. Viability Determination

1. Before starting, prepare the autoclaved *M. leprae* suspension (Protocol of the reagents - Protocol 7). Use the appropriate kit (see table of reagents and equipment) for qualitative determination of *M. leprae* viability in the suspension. Dilute the solutions as follows (all procedures should be performed under dim light): dilute solution A by a factor of 10 in saline (1  $\mu$ l stock plus 9  $\mu$ l saline), dilute solution B 20 by a factor of 10 in saline (1  $\mu$ l stock plus 19  $\mu$ l saline).
2. Add 3.6  $\mu$ l of diluted solution A and 6  $\mu$ l of diluted solution B to 200  $\mu$ l of the bacillary suspension to be tested (step 1.10). A previously autoclaved *M. leprae* suspension is routinely used as a negative control for viability staining.
3. Incubate the suspensions for 15 min, at room temperature in the dark.
4. Centrifuge the tube at 10,600 x g for 5 min at 4 °C.
5. Discard the supernatant and resuspend the pellet in 15  $\mu$ l of 10% glycerol. Apply 8  $\mu$ l to the surface of a clean glass slide and cover it with a small cover slip. Analyze the slide using a fluorescent microscope, containing the appropriate filters (see Molecular Probes recommendations). Evaluate the results by comparing Syto 9 (Sy) staining to propidium iodide (PI) staining.  
 Note: Sy stain penetrates 100% of both dead and alive bacteria, and PI stain penetrates only bacteria with damaged membranes. The autoclaved suspension (negative control) may form clumps of bacteria due to the presence of cellular debris. The semiquantitative scale used for live/dead evaluation varies from 0 to 2+; 0 indicates that less than 30% of the cells are positive for PI stain; 1+ means between 30-50% of the cells are positive for PI stain; 2+ means more than 50% of the cells are positive for PI stain. A score of 0 is considered most adequate for use.

### 4. Freezing/thawing *M. leprae* Suspension

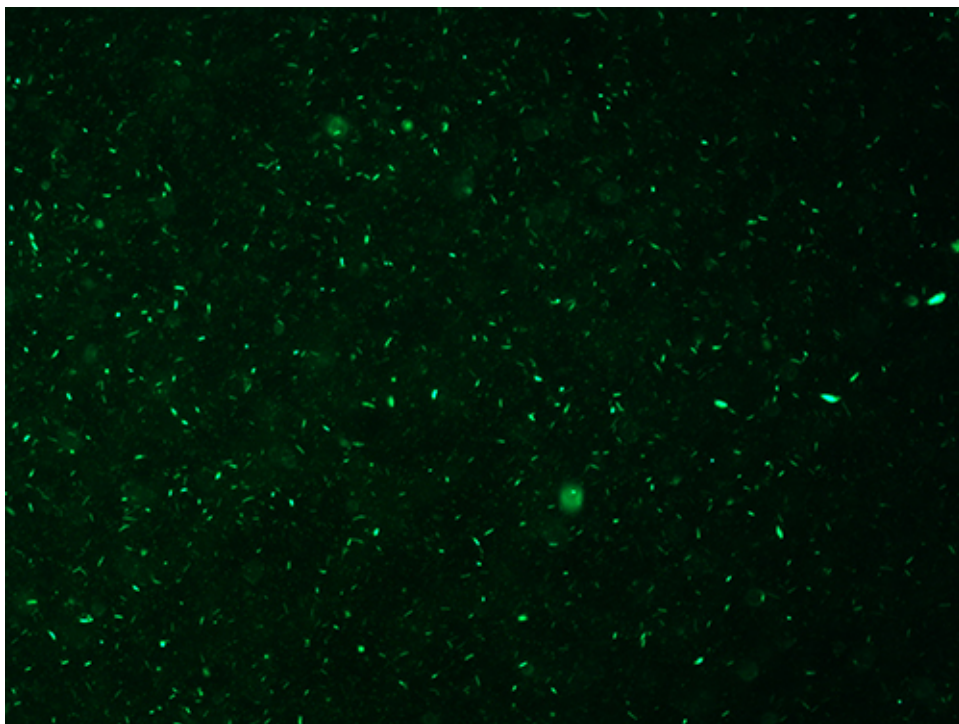
1. Before starting, prepare the freezing medium (Protocol of the reagents - Protocol 6). For freezing, use the steps described below:
  1. Add 150  $\mu$ l of the suspension containing  $1 \times 10^7$  AFB/ml to a sterile cryovial along with 1 ml of freezing medium.
  2. Place the cryovial in a freezing container and store the container at -80 °C for 24 hr.
  3. Transfer the frozen vial to a storage box and store it at -80 °C.
2. For thawing, use the steps described below:
  1. Remove the cryovial with the frozen suspension from -80 °C and place it in a 37 °C water bath to start thawing the suspension.
  2. Pour the suspension into a tube containing 20 ml of sterile saline. Mix the suspension gently until thawing is complete.
  3. Centrifuge the suspension for 30 min, at 1,700 x g, at 4 °C.
  4. Discard the supernatant and add enough sterile saline to reach the desired volume. Homogenize the suspension by passing it through a syringe (step 1.8).
  5. For ZN staining, viability determination and inoculation, follow Protocols 2, 3 and 5.

### 5. Mouse Inoculation

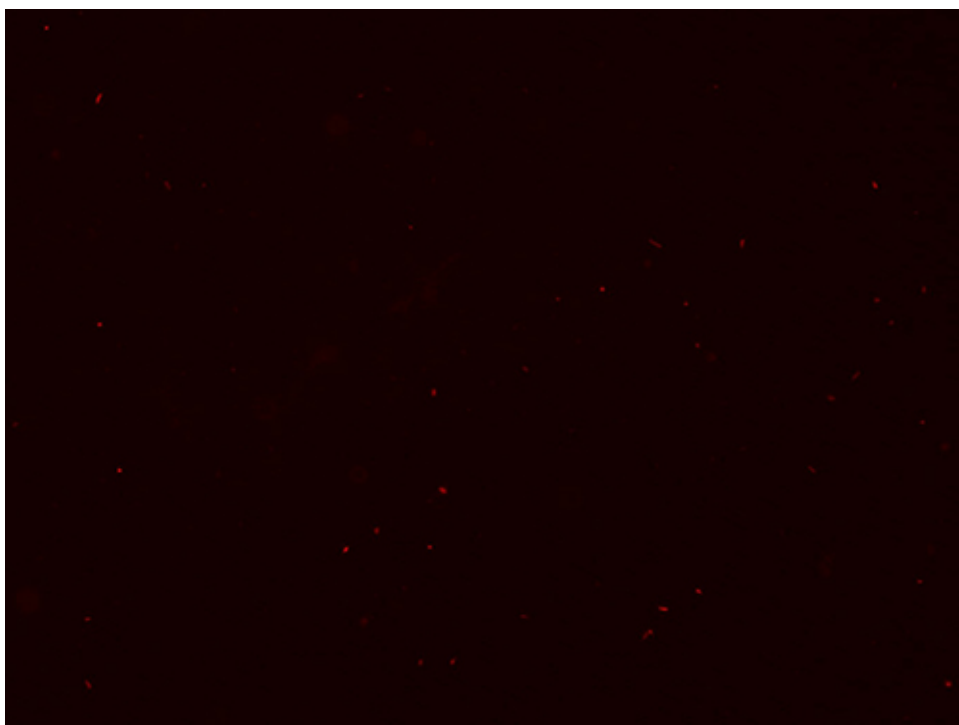
1. Two people are needed for this procedure, one to restrain the mouse and another to administer the inoculum. The animal to be inoculated should be handled according to ethical guidelines and regulations and all procedures must be approved by the institutional animal care and use committee. Restrain the nude mouse by the scruff of the neck with the paws facing up.
2. Homogenize the suspension by passing it through a 26 G needle. With a 1 ml syringe, aspirate enough bacillary suspension to inoculate two footpads (30  $\mu$ l/footpad).
3. Hold the rear paw, clean the footpad with 70% ethanol alcohol prior to injection and introduce the needle intradermally with the bevel of the needle pointed up, from the proximal to the distal side of the footpad. Inject 30  $\mu$ l of the suspension. The footpad injections may be performed in anesthetized mice.
4. Wait 5 sec before withdrawing the needle from the skin to avoid reflux of the inoculum. Mice should be housed on soft bedding post inoculation, and ambulation and signs of self-mutilation should be monitored.

### Representative Results

The success of the protocol can be assessed in three ways. First, evaluation of the quality of the inoculum is determined by the amount of cellular debris and the resulting percentage of viable bacilli in the final suspension (See Protocol 3). As shown in **Figures 1** and **2**, the final bacillary suspension contained very little cell debris and high viability (score 0+); note that most bacilli stained with the Syto 9 green fluorescent stain (stain penetrates 100% of both dead and alive bacteria, **Figure 1**) and only a few bacilli stained with the red PI fluorescent stain (stain penetrates only bacteria with damaged membranes, **Figure 2**).



**Figure 1. Syto 9 staining of *M. leprae* suspension.** Green fluorescence demonstrates the presence of alive and dead *M. leprae*. 400X.



**Figure 2. PI staining of *M. leprae* suspension.** Red fluorescence showing the small number of dead *M. leprae*. 400X.

Second, an inoculum with high viability will impact on the multiplication of bacilli in the footpads of animals after 4-5 months post inoculation, with development of obvious macroscopic lesion, as shown in the video (Protocol 1). The third way to assess the success of the protocol is by evaluating the survival and multiplication of AFB in mouse footpads inoculated with bacilli that had been frozen for different periods (See Protocol 4).

Experiment (animal)	Number of AFB/ml frozen at day 0	Viability score/day 0	Freezing period (days)	Number of AFB inoculated after freezing	Viability score after freezing	Number of AFB/ml recovered after 7 months
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1 (1)	$1.0 \times 10^7$	0+	60	$1.7 \times 10^5$	1+	$2.3 \times 10^8$
1 (2)	$1.0 \times 10^7$	0+	60	$1.7 \times 10^5$	1+	$3.8 \times 10^7$
1 (3)	$1.0 \times 10^7$	0+	60	$1.7 \times 10^5$	1+	$8.5 \times 10^7$
1 (4)	$1.0 \times 10^7$	0+	60	$1.7 \times 10^5$	1+	$4.6 \times 10^7$
2 (1)	$1.0 \times 10^7$	0+	15	$4.2 \times 10^5$	1+	$1.7 \times 10^7$
2 (2)	$1.0 \times 10^7$	0+	15	$4.2 \times 10^5$	1+	$4.8 \times 10^6$
2 (3)	$1.0 \times 10^7$	0+	15	$4.2 \times 10^5$	1+	$8.6 \times 10^6$
3 (1)	$1.0 \times 10^7$	0+	15	$1.7 \times 10^5$	1+	$1.2 \times 10^7$
3 (2)	$1.0 \times 10^7$	0+	15	$1.7 \times 10^5$	1+	$1.8 \times 10^7$
3 (3)	$1.0 \times 10^7$	0+	15	$1.7 \times 10^5$	1+	$1.8 \times 10^7$
3 (4)	$1.0 \times 10^7$	0+	15	$1.7 \times 10^5$	1+	$2.6 \times 10^7$
3 (5)	$1.0 \times 10^7$	0+	15	$1.7 \times 10^5$	1+	$3.7 \times 10^6$

**Table 1. Results of *M. leprae* multiplication using post freezing suspensions.**

**Table 1** depicts results of *M. leprae* growth using suspensions after freezing. The viability score of the suspensions after thawing was 1+. The freezing protocol was carried out in three independent experiments to test different freezing periods. In both experiments with inocula frozen for 15 or 60 days, the outcome was similar, propagation after freezing yielded 10-1,000 times increase in the number of AFB recovered from each footpad after 7 months of inoculation (**Table 1**). Therefore, the freezing of *M. leprae* suspensions in 7H9 medium supplemented with OADC (oleic acid-albumin-dextrose-catalase) resulted in maintenance of viability.

Three inocula were used to evaluate *M. leprae* multiplication after two different freezing periods (15 and 60 days). After 7 months, bacilli were recovered from footpads of inoculated mice and counted after Ziehl-Neelsen staining. Semiquantitative viability scale: 0 means absent up to 30% of PI stained cells; 1+ means between 30-50% of PI stained cells; 2+ means above 50% of PI stained cells. AFB: acid fast bacilli.

## Discussion

A detailed description of a well-illustrated, successful protocol for propagation of *M. leprae* is greatly needed. Our study demonstrates that the protocol of inoculum preparation by filtration and trypsin digestion allows the inocula to be obtained with very little cellular debris and with high viability of bacilli (score 0+). Sodium hydroxide has been used to disaggregate the tissue for purification of bacilli<sup>6</sup>. Studies performed in our laboratory using sodium hydroxide for purification of *M. leprae* resulted in formation of clumps of bacilli, hampering the homogenization of the suspension for viability determination and animal inoculation (data not shown).

Potential problems encountered with the inoculum preparation by filtration and trypsin digestion include large amount of cellular debris and contamination of the inoculum with bacterial or fungal agents. In case large amounts of cellular debris are observed after purification, either the trypsin is no longer active or there is an excessive amount of biological material. Enzymatic activity of the trypsin stock solution must be evaluated. If excessive amount of initial biological material is suspected, the material should be divided into aliquots and the protocol should be carried out in separate batches. To avoid contamination of the inoculum with bacterial or fungal agents, care must be taken to process the material under aseptic conditions. If fungal and/or bacterial contamination are detected the suspension should be discarded.

A limitation of our protocol is the subjectivity of the viability evaluation using the described semi-quantitative method. Viability assessed semi-quantitatively is more practical, although less precise than the published quantitative method<sup>6</sup>. Viability score of 0+ and 1+ are satisfactory for maintenance of propagation and freezing of *M. leprae*. Lahiri *et al.* have already shown that nude mice inoculated with 80-90% viable inoculum, result in footpads suitable for harvesting (high viability bacilli) at 4-5 months of inoculation. Therefore, early infection (around 4 months) is the best harvesting time. For harvesting of frozen inocula, the mice in the present protocol were maintained inoculated for longer periods (7 months) to guarantee growth curves. A critical step to ensure adequate viability is the use of fresh bacilli suspensions, preferably within 24 hr after collection of biological material from the host and processing. Moreover, quality of the reagents, freshly prepared diluted trypsin and viability staining solutions, are necessary to guarantee reproducible results.

Another limitation of this protocol is that the final *M. leprae* suspension is not free of host DNA, RNA, protein, *etc.* Therefore, other purification steps must be added to obtain a *M. leprae* suspension free of host cell components.

A method for maintaining viable bacilli by freezing whole tissue specimens of *M. leprae* lesions has been reported<sup>9</sup>. However, the study by Portaels *et al.* demonstrated significant loss of viability, ranging between 65-97% after freezing and thawing of *M. leprae* infected tissue specimens obtained from armadillo<sup>9</sup>. Our protocol demonstrated that the viability index observed in *M. leprae* suspensions after freezing and thawing dropped when compared to the aliquot that had not been frozen (**Table 1**). Indeed, freezing the *M. leprae* suspension in freezing media yielded viability ranging from 50-70%, with viability score 1+, while viability score 0+ was obtained in the unfrozen suspension. Nonetheless, the multiplication of *M. leprae* was satisfactory after 7 months post inoculation of nude mice (**Table 1**). The inoculation of nude mice with reconstituted samples maintained frozen for 60 days resulted in mean 100 times increase in the number of bacilli compared to the initial inoculum. It appears that freezing the *M. leprae* suspension in freezing media, instead of infected tissue specimens, is more efficient. A critical

step of our protocol is the slow freezing of the AFB in a freezing container, necessary to maintain the bacilli viable, as demonstrated by Colston and Hilson<sup>8</sup>. Future experiments will be conducted to assess the viability of bacilli after longer freezing periods.

In summary, because *M. leprae* does not grow *in vitro*, our protocol allows for a fast and easy alternative for maintenance of viable inoculum, and the successful freezing step makes possible the maintenance of strains without continuous passage in animals, thus enabling the establishment of a bank of defined strains.

This section contains instructions for preparing reagents to perform this protocol.

### 1. Trypsin

Trypsin	0.5 g
Distilled water	up to 100 ml

Filter sterilize. Store at -20 °C.

### 2. 7H9

7H9 broth base	4.7 g
40% glycerol stock	5 ml
Distilled water	up to 900 ml

Mix the base with water then add the glycerol while stirring. Autoclave at 121 °C for 20 min to sterilize. Store at 4 °C.

### 3. Brain heart infusion (BHI)

BHI	37 g
Distilled water	up to 1,000 ml

Autoclave at 121 °C for 15 min to sterilize. Store at 4 °C.

### 4. Phenol serum

#### 4.1) 5% phenol

Phenol	5 ml
Distilled water	up to 100 ml

#### 4.2) Serum phenol

fetal bovine serum	2 ml
5% phenol	98 ml

Store at 4 °C.

### 5. Solutions for cold Ziehl-Neelsen Stain

#### 5.1) CarboFuchsin

Fuchsin	1 g
Phenol crystals fused to 60 °C	5 ml
Pure ethyl alcohol	10 ml
Distilled water	up to 100 ml

Filter before each use.

#### 5.2) Methylene Blue Base

Methylene Blue	3 g
95% ethyl alcohol	up to 200 ml

#### 5.3) Alcohol acid

70% alcohol	990 ml
chloridric acid	10 ml

## 6. Medium for freezing:

OADC	10 ml
Glycerol	20 ml
7H9 medium	up to 100 ml

Autoclave glycerol before use and sterilize OADC by filtration.

## 7. Autoclaved *M. leprae* suspension

Autoclave at 121 °C for 20 min. Store at -20 °C.

## Disclosures

No conflicts of interest declared.

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