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## Oligoarticular and polarticular JIA: epidemiology and pathogenesis

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### Summary

Juvenile idiopathic arthritis (JIA) refers to a group of chronic childhood arthropathies, currently classified into subtypes primarily on the basis of clinical features. Research has focused on the hypothesis that these subtypes arise through distinct etiologic pathways. In this Review, we discuss four subtypes of JIA: persistent oligoarticular, extended oligoarticular, rheumatoid-factor positive polyarticular and rheumatoid-factor-negative polyarticular. These subtypes differ in prevalence between ethnic groups and are associated with different HLA alleles. Non-HLA genetic risk factors have also been identified, some of which reveal further molecular differences between these subtypes, while others suggest mechanistic overlap. Investigations of immunophenotypes also provide insights into subtype differences: adaptive immunity appears to have a prominent role in both polyarticular and oligoarticular JIA, and the more-limited arthritis observed in persistent oligoarticular JIA as compared with extended oligoarticular JIA may reflect more-potent immunoregulatory T-cell activity in the former. Tumor necrosis factor seems to be a key mediator of both polyarticular and oligoarticular JIA, especially in the extended oligoarticular subtype, although elevated levels of other cytokines also are observed. Limited data on monocytes, dendritic cells, B cells, natural killer T cells, and neutrophils suggest that the contributions of these cells differ across subtypes of JIA. Within each subtype, however, common pathways appear to drive joint damage.

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## INTRODUCTION

Juvenile idiopathic arthritis (JIA) is a term that collectively refers to a group of chronic childhood arthropathies, which together constitute the most common rheumatic condition in children. The latest (2001) International League of Associations for Rheumatology (ILAR) criteria define seven subtypes of JIA: systemic JIA (sJIA), oligoarticular JIA, rheumatoid factor (RF)-negative (RF<sup>-</sup>) polyarticular JIA, RF-positive (RF<sup>+</sup>) polyarticular JIA, enthesitis-related arthritis (ERA), psoriatic arthritis and undifferentiated JIA.(1) The ILAR classification includes persistent and extended oligoarthritis as subcategories of oligoarticular JIA, but not as distinct subtypes. Discussion regarding the robustness of this classification scheme continues, however.(2–5) This Review focuses on developments regarding the epidemiology, genetics and immunopathogenesis of the oligoarticular and polyarticular subtypes. Progress in the understanding of sJIA, which appears to be an autoinflammatory rather than an autoimmune disease,(6) and of ERA, which is a clinically well-defined subtype typically associated with HLA-B27,(7) while of interest, are beyond the scope of this discussion.

Oligoarthritis is defined as arthritis affecting 4 or fewer joints during the first 6 months after disease onset, excluding children with enthesitis and boys after age 6 with HLA-B27\*<sup>\*</sup>. Oligoarthritis is subdivided into persistent and extended forms, based on the number of joints affected after the first 6 months; that is, 1–4 and more than 4 joints, respectively. RF<sup>+</sup> and RF<sup>-</sup> polyarthritis both affect 5 or more joints during the first 6 months of the disease; RF<sup>+</sup> disease is defined by two or more positive RF tests.(1) Table 1 summarizes the main clinical features associated with these four subtypes. Clinical differences imply distinct pathogenetic pathways for these subtypes, although current treatment approaches do not distinguish extended-oligoarticular, RF<sup>+</sup> polyarticular and RF<sup>-</sup> polyarticular disease. Our understanding of the molecular basis of JIA subtypes is still unfolding, as discussed in this Review. Of note, the interpretation of some studies is confounded by changing schemes for JIA classification (Box 1) and by analyses that combine subtypes to increase patient numbers.

## EPIDEMIOLOGY

The reported incidence and prevalence of JIA vary widely, in part because JIA is a heterogeneous condition that is identified clinically without specific diagnostic tests. Available numbers are likely to underestimate the true incidence and prevalence of JIA, because of under-diagnosis and because most studies are clinic- rather than community-based.(8–10) In a comprehensive survey of data published as of 2002, Manners and Bower(11)concluded that the incidence of chronic arthritis of childhood ranges from 0.008 to 0.226 per 1000 children, and that the prevalence ranges from 0.07 to 4.01 per 1000 children worldwide. Recent US and Canadian studies report the incidence of JIA to be 0.041 to 0.061 per 1000 children.(12–14). The best estimate of JIA prevalence in the US uses data recorded in the 2001–2004 National Ambulatory and National Hospital Ambulatory Medical

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\*In older classification schemes (see text box), the term pauciarticular disease was used, and this subtype included subjects with ERA or extended oligoarticular disease.

Care Survey. Based on ICD-9-CM (International Classification of Diseases, Ninth Revision, Clinical Modification) codes, approximately 294,000 US children aged 0–17 years (95% CI 188,000–400,000) are affected by the broadly defined “arthritis or other (similar) rheumatic conditions”.(15)

The major JIA subtypes vary by sex ratio and age at onset.(7) Girls are overrepresented in groups of patients with oligoarticular JIA and polyarticular JIA. Age at onset for oligoarticular JIA peaks at 2–4 years; by contrast, age at onset of polyarticular JIA has a biphasic pattern, with peaks at 1–4 years and 6–12 years of age, with RF<sup>+</sup> disease contributing disproportionately to the second peak.

In North America and Europe, the relative frequencies of JIA subtypes are 10–30% RF<sup>-</sup> polyarticular, 5–10% RF<sup>+</sup> polyarticular, and 30–60% oligoarticular.(7) Studies of multi-ethnic communities from North America and from Europe suggest that, with the exception of RF<sup>+</sup> polyarticular JIA, European ancestry is an important predisposing factor for JIA, especially for extended oligoarticular JIA and antinuclear-antibody (ANA)-associated JIA with uveitis.(16, 17) Conversely, patients of African American or Native American descent are more likely to have RF<sup>+</sup> polyarticular JIA than children of European descent (16, 18, 19). Studies from different geographic areas are difficult to compare, but it seems that in Asia the overall prevalence of JIA is lower than in Europe and North America (20–22) and that ERA is particularly common.(20–23) The prevalence of uveitis in oligoarticular and polyarticular JIA subtypes is seemingly highest in Scandinavia and lowest in East Asia and India.(24) Black South African patients, like African Americans, have a higher rate of RF<sup>+</sup> polyarticular JIA and lower rate of arthritis with uveitis compared to children of European descent.(25)

Environmental influences are hypothesized to contribute to JIA pathogenesis, but specific agents have not been identified. In some reports, JIA is more prevalent in rural than urban settings.(26, 27) A single study conducted in Manitoba, Canada in 1975–1992 correlated cyclic increases in JIA incidence with the prevalence of mycoplasma infection.(14) Introduction of measles–mumps–rubella vaccination in Finland did not change the annual incidence of JIA, which argues against these infections as triggers of JIA.(28) An increasing incidence of JIA seems to be a worldwide trend, although the incidence of chronic pediatric arthritis in Rochester, Minnesota reportedly decreased from 0.150 cases per 1000 children (1960–1969) to 0.078 per 1000 children (1980–1993).(29, 30) Further work is needed to confirm incidence trends and to identify factors that contribute to these patterns.

Overall, epidemiological data corroborate clinical evidence that JIA subtypes represent discrete conditions. Data showing the influence of ethnicity on subtype susceptibility imply a role for genetics in JIA risk. To reveal more-detailed hypotheses on the pathogenesis of these subtypes, however, we must turn to studies of immunogenetics and immunophenotyping.

## **PATHOGENESIS**

The presence of associations between HLA class II alleles and susceptibility to both oligoarticular and polyarticular JIA implicates CD4<sup>+</sup> T cells in these subtypes. The specific

class II alleles associated, however, differ between the subtypes, which strongly suggests differences in molecular mechanisms. Indeed, as knowledge of CD4<sup>+</sup> T cell subsets has grown, models of their involvement in oligoarticular and polyarticular JIA have been refined and support the idea that mechanistic differences exist. Available evidence also argues that antigen-driven autoimmunity is insufficient to explain all the features of oligoarticular and polyarticular JIA,(31) and implicate the involvement of both innate and adaptive immune responses. These issues are the focus of the rest of this Review.

### Genetic factors

Data from family and twin studies were among the first to imply that susceptibility to JIA is inherited (reviewed elsewhere(32)). For example, siblings of probands are 15-fold to 30-fold more likely than the general population to develop JIA.(33)

All JIA subtypes are probably complex genetic traits, as they lack single-gene, Mendelian patterns of inheritance. Inherited risk factors for both disease susceptibility and disease severity have been reported (Tables 2–4). The highly polymorphic HLA genes confer the strongest genetic effects in JIA, and, to a large extent, HLA allelic associations confirm clinically defined subtypes, including differences between persistent and extended oligoarthritis (Table 2). Initial findings on HLA genetic risk from association and linkage studies have been confirmed by genome wide association studies (GWASs) (34, 35). HLA effects are observed within an age-window-of-effect, and gender also influences the age at which some of the associated alleles have their effect (36). Associations of HLA-A, HLA-B and HLA-DR with oligoarthritis are observed in girls but not boys, which raises the possibility of further heterogeneity in disease pathogenesis.(37) Shared HLA allelic associations imply a mechanistic relationship between extended oligoarticular JIA and RF<sup>-</sup> polyarticular JIA; however, this relationship might be modified by the effects of differences in the associated HLA-DP alleles for these subtypes. Evidence suggests that different alleles confer susceptibility in different ethnic groups, as is the case for other MHC-associated diseases.(38–40) A caveat of interpreting genetic studies that use pre-ILAR classification criteria is that the patient groups are possibly heterogeneous; for example, the pauciarticular subtype could include HLA-B27<sup>+</sup> cases.(4)

HLA class II has been estimated to account for ~20% of the sibling recurrence risk in oligoarticular and polyarticular JIA, which implies a role for non-HLA genes.(41) Approximately 100 non-HLA genes have been investigated for an association with JIA; however, many studies do not have the statistical power to detect moderate or mild gene effects, particularly after stratification for JIA subtype. Nevertheless, associations with some non-HLA genes have been confirmed (Table 3). Some of these genes, such as *PTPN22* and *IL2RA* (also known as *CD25*), are associated with multiple autoimmune diseases, (42) an observation that is consistent with the occurrence of various autoimmune disorders in families of JIA probands.(43) A 2009 GWAS found evidence of an association between *VTCN1*, a gene that encodes a negative regulator of T-cell-mediated responses, and JIA susceptibility.(35) Other candidate non-HLA genes are reportedly associated with risk of both oligoarticular and polyarticular JIA subtypes, including genes that encode regulators of innate immunity such as the proinflammatory cytokine macrophage migration inhibitory

factor (MIF). By contrast, some studies reveal subtype-specific genetic effects. A GWAS found linkage of different chromosomal regions to oligoarticular and polyarticular JIA subtypes, and distinction between early-onset and late-onset disease.(34) Another study found *TNFAIP3* and *C12orf30* variants associated with oligoarticular JIA, and a *STAT4* variant associated with polyarticular JIA.(44) The unique association between oligoarticular JIA and *IL-2RA/CD25* (42) which is highly expressed on regulatory T cells, suggests the possibility that these cells have a particular role in this subtype. Some, but not all, cytokine gene associations distinguish the subtypes (Table 4). The association of a low-expression-level allele of osteopontin with persistent oligoarticular JIA is consistent with the more-limited disease observed in this subtype as compared with extended oligoarticular JIA, and further argues for a pathogenic distinction between the two forms of oligoarticular JIA (45). Overall, observed genetic associations lend further support to the idea that the oligoarticular and polyarticular subtypes have different etiologies, but also argue that certain immunological pathways, such as MIF-driven inflammation, could contribute to pathology across subtypes.

### Immune-related factors

**T cells and autoantigens**—Associations of HLA alleles with polyarticular and oligoarticular JIA imply a role for adaptive immunity, and especially T cells, in the pathogenesis of these subtypes. A number of lines of cellular evidence support this possibility, the oldest being the presence of T-cell and plasma-cell infiltrates in affected joints. However, there are some differences in T-cell phenotype in oligoarticular compared to polyarticular JIA, providing clues to subtype-specific mechanisms.

One aspect of T-cell biology that might have a role in JIA is impaired thymic function. Accelerated loss and premature aging of naïve CD4<sup>+</sup> T cells, the latter detected as telomeric erosion, have been reported in oligoarticular and polyarticular JIA, independent of treatment.(46) A study of all JIA subtypes, however, found no difference in thymic function between patients with JIA and controls.(47) Difficulty discriminating between recent thymic emigrants and long-lived peripheral naïve T cells could be one of the reasons for these discordant results; markers that better distinguish these two T-cell populations may clarify the role of thymic function in JIA.(48)

Evidence of T cell activation is a feature of patients with oligoarticular and polyarticular JIA, although critical or initiating antigens have not been determined. Activated CD4<sup>+</sup> T cells are found at increased levels in circulation(49) in comparison to healthy, age-matched children and in the synovium, with the CD4:CD8 ratio being higher in polyarticular JIA joint tissue compared to oligoarticular JIA.(50) Selected CD4<sup>+</sup> clones not abundant in circulation are expanded within the joints in oligoarticular patients, (51) but precipitating antigens for these expansions have not been identified. Evidence for auto-reactivity of circulating T cells includes responses to the joint-related proteins aggrecan and fibrillin, the latter primarily in patients with polyarticular JIA.(52) Several heat shock proteins (HSPs) are also potential autoantigens in JIA. Highly conserved across species, HSPs are produced in response to cell stress. Circulating T cells from oligoarticular and polyarticular JIA patients, but not controls, react to human HSP60.(53, 54) Human HSP70 is expressed on

synovial cell membranes in all JIA subtypes and might act as an autoantigen. Alternatively, ligation of HSP70 and autoantigens may promote autoantigen processing and presentation. (55)

Autoreactive CD8<sup>+</sup> T cells have also been implicated in the pathogenesis of polyarticular and oligoarticular JIA. Activated CD8<sup>+</sup> T cells are found in circulation(49) and in the synovium,(50) with a suggestion of higher levels of activation in the oligoarticular JIA compared to the polyarticular subtype. Consistent with re-distribution of these cells to the joints, absolute numbers of circulating CD8<sup>+</sup>CD45RA<sup>+</sup> (naïve) T cells are decreased in oligoarticular JIA in comparison to healthy, age-matched controls, even after clinical remission.(56) Several studies have addressed potential autoantigenic specificities of CD8<sup>+</sup> T cells. Peptides from self-HLA proteins (HLA DRB1\*1101, DRB1\*0801 and DRP1\*0201) with homology to Epstein–Barr-virus-derived peptides trigger cytotoxicity and proinflammatory cytokine production from peripheral blood CD8<sup>+</sup> T cells of patients with oligoarticular JIA.(57) In a study of patients primarily with polyarticular JIA, peripheral blood cytotoxic T cells lysed autologous synovial cells and keratinocytes *in vitro*.(58) Mechanisms of loss of CD4<sup>+</sup> and CD8<sup>+</sup> T-cell tolerance to autoantigens could include molecular mimicry (that is, cross-reactive responses to self-peptides that resemble pathogen-derived peptides) and inflammation-induced epitope spreading.

T cells in inflamed synovia of patients with polyarticular and oligoarticular JIA are predominantly activated memory type 1 T helper (Th1) cells, which are IFN $\gamma$ <sup>+</sup> and CD45RO<sup>+</sup>.(59) Evidence suggests that these cells home to the joint where they receive further activation by Th1- polarizing stimuli. Synovial fluid from patients with polyarticular JIA has been found to contain interleukin (IL)-12, a key inducer of Th1 cells. (60) The CC-chemokine receptor (CCR) 7, which controls the migration of memory T cells to inflamed tissues, is expressed on a subset of synovial fluid CD4<sup>+</sup> T cells in patients with oligoarticular and polyarticular JIA.(61) Levels of CC-chemokine ligand (CCL) 21, the ligand of CCR7, are elevated in synovial fluid of polyarticular and oligoarticular JIA patients in comparison with blood levels. CCR7<sup>+</sup> synovial CD4<sup>+</sup> T cells have higher expression levels of the Th1-biased CXC-chemokine receptor (CXCR) 3 than do CCR7<sup>-</sup> cells, and the CCR7<sup>-</sup> population is more enriched with the Th-1-biased chemokine receptor CCR5<sup>+</sup> as compared to CCR7<sup>+</sup> cells.(59) Synovial cells express transcripts encoding the ligand for CXCR3, CXC-chemokine ligand (CXCL) 10 (also known as IP-10), and the ligands for CCR5, CCL3 (also known as MIP-1 $\alpha$ ) and CCL5 (also known as RANTES).(61–63) One model that accounts for these observations is that CCR7<sup>+</sup>CXCR3<sup>+</sup> cells might be recruited directly to the synovium, where they may become effector cells that downregulate CCR7 and upregulate CCR5; CCR5, in turn, may be the main receptor used for migration within the synovial tissue.(61) Furthermore, CCL5 may enhance T-cell production of IFN $\gamma$ .(64) While effector T cells in the synovial fluid of both polyarticular and oligoarticular JIA patients express higher IFN $\gamma$ :IL-4 ratios than their peripheral blood counterparts,(65) IL-4 is more prominent in the synovial compartments of patients with oligoarticular JIA compared with polyarticular JIA. This pattern is consistent with the more-limited arthritis associated with this subtype, and suggests a deviation from Th1 towards Th2-biased phenotypes.(66) In polyarticular JIA, even peripheral blood mononuclear cells have elevated levels of genes induced by IFN $\gamma$ .(67)

A new paradigm suggests that a key inflammatory CD4<sup>+</sup> T-cell subset produces IL-17, and that the development of this subset is negatively regulated by Th1 cells.(68) The inflamed joints of all JIA subtypes are enriched in IL-17-producing T cells, and high levels of IL-17, in excess of serum levels, are detected in synovial fluid in polyarticular JIA.(69, 70) IL-17 potently induces synoviocyte production of IL-6, matrix metalloproteinases (MMPs) 1 and 3, and IL-8 (which is chemotactic for neutrophils), all of which have been implicated in joint damage.(70)

The extent of synovial inflammation is probably influenced by the balance between inflammatory cells and regulatory CD4<sup>+</sup> T cells (Treg cells), which are detected as CD25<sup>hi</sup>CD127<sup>lo</sup>FOXP3<sup>+</sup> cells.(71) In active synovitis, Treg cell function is apparently insufficient. One reason may be high synovial levels of IL-7 and IL-15, which are found in polyarticular and oligoarticular JIA and are associated with the induction of T-cell proliferation and abrogation of Treg-cell-mediated suppression of immune activation.(72, 73) In addition, the frequency of Th17 cells is inversely related to the frequency of FOXP3<sup>+</sup> Treg cells in the synovium.(69)

Evidence particularly implicates the activity of these anti-inflammatory T cells as a disease-limiting mechanism in oligoarticular JIA. Compared with the circulation, there are more CD4<sup>+</sup>CD25<sup>+</sup> Treg cells in oligoarticular JIA synovial fluid, and these have higher suppressive capacity than peripheral cells.(74) Higher numbers of circulating CD4<sup>+</sup>CD25<sup>hi</sup> Tregs are observed in persistent compared to extended oligoarticular JIA,(74) polyarticular JIA or sJIA.(75) A regulatory mechanism may involve reactivity to HSP: in individuals with persistent oligoarticular JIA, an HSP dnaJ epitope stimulates synovial mononuclear cells to expand T cells with suppressive activity (CD4<sup>+</sup>CD25<sup>+</sup>CTLA4<sup>+</sup> T cells).(76) T-cell reactivity to human HSP60 is associated with remission in oligoarticular JIA; the protective mechanism seems to involve induction of IL-10 production by CD30<sup>+</sup> T cells.(77, 78) In summary, it seems that proinflammatory T cells are recruited to the joint in both oligoarticular and polyarticular JIA, where they drive inflammation, but this activity is perhaps better controlled by regulatory mechanisms in persistent oligoarticular disease than in extended oligoarticular disease.

**Antigen-presenting cells: dendritic cells and B cells**—T-cell activation is dependent on antigen-presenting cells, the most potent of which are dendritic cells (DCs). DCs are thought to participate in the initiation, maintenance and/or regulation of inflammation in JIA, although few studies provide direct evidence for this. In oligoarticular and polyarticular JIA patients, conventional DCs (cDCs) and IFN $\alpha$ -producing DCs (plasmacytoid dendritic cells [pDCs]) are enriched in synovial fluid compared with the circulation.(79) One study found synovial fluid DCs to be more numerous in polyarticular JIA than in oligoarticular JIA and found lower numbers of circulating DCs to be associated with a reduced response to treatment. (80) Within synovial tissue, pDCs are found at lymphoid follicle-like structures, while cDCs are mostly found at the lining and sublining layers, which suggests that DC subtypes mediate distinct processes in the joint.(79) cDCs expressing RANK (receptor activator for nuclear factor  $\kappa$  B) are found in the joints of patients with oligoarticular and polyarticular JIA.(81) RANK-mediated activation of DCs leads to increased survival of these cells and production of inflammatory cytokines.(82)

Further investigation of DC migration to and function in the joint microenvironment is warranted.

The major autoantibody specificities found in JIA (ANA, RF, and anti-cyclic citrullinated peptide [CCP] antibody) have not been directly implicated in disease development, although they serve as biomarkers of clinical subsets. ANA is associated with uveitis in oligoarticular JIA, and IgM RF and/or anti-CCP antibodies are associated with erosive disease in polyarticular JIA.(83) Ravelli *et al.* have proposed that ANA<sup>+</sup> patients form an homogenous group, regardless of the course of arthritis; that is, ANA<sup>+</sup> oligoarticular JIA patients share clinical features with ANA<sup>+</sup> polyarticular JIA patients but not ANA<sup>-</sup> polyJIA patients.(17) Microarray data from Griffin *et al.* also indicate that ANA<sup>+</sup> polyarticular JIA patients with early-onset disease might represent a subgroup separate from other patients with RF<sup>-</sup> and RF<sup>+</sup> polyarticular JIA.(84)

Analyses of B-cell populations in JIA are few. Expansions of CD5<sup>+</sup> B cells, a minor B-cell subset, are observed in oligoarticular and polyarticular JIA (as well as Rheumatoid Arthritis).(49, 85) CD5<sup>+</sup> B cells typically produce antibodies against bacterial wall components, but are also a source of low-affinity multispecific antibodies. Whether CD5<sup>+</sup> B cells produce disease-related autoantibodies in JIA is not known. Given the efficacy of rituximab for RA, and the renewed interest in B cells as pathogenic effectors,(86) more studies of B cells in JIA are likely to be forthcoming.

**Monocyte/macrophages and proinflammatory cytokines**—Elevated levels of proinflammatory cytokines are found in the circulation and synovial compartments in patients with JIA. Serum levels of tumor necrosis factor (TNF) are elevated in oligoarticular and polyarticular JIA,(87) as are serum levels of IL-6, but the highest IL-6 levels are found in sJIA.(87–90) Circulating IL-1 $\beta$  can be elevated in oligoarticular and polyarticular JIA, (89) although this finding is not consistent.(87) In active arthritis, levels of proinflammatory cytokines are generally higher in synovial fluid than in circulation, mostly likely due to local production. For example, complexes of IL-6 and a soluble form of the IL-6 receptor, which bind to surface gp130 on chondrocytes and fibroblasts, are thought to trigger further IL-6 production by these cells.(91)

On the basis of subtype-specific responses to anti-cytokine therapies, particular cytokines seem to be primary effectors in JIA subtypes. For polyarticular JIA and extended oligoarticular JIA, blockade of TNF has been efficacious, arguing for a crucial role for TNF in these subsets.(92) TNF is inhibited endogenously by its soluble receptors, and its damaging effects in polyarticular JIA might be attributable to a reduced ratio of soluble TNF receptor to TNF. (93) A recent study of transcriptional profiles of peripheral blood mononuclear cells from patients with newly diagnosed polyarticular JIA identified a subgroup with elevated expression of monocyte-associated genes, including genes that encode the monocyte markers CD64 and CD14. This subgroup included primarily RF<sup>+</sup> individuals, which raises the possibility that monocyte-driven inflammation is of particular importance in these patients.(84)



A key observation in all JIA subtypes is that, while levels of inflammatory cytokines are higher during active disease, levels are abnormal during clinical remission,(90) which suggests that remission represents a state of compensated inflammation that is overwhelmed during disease flare.

**Innate immune dysfunction in JIA subtypes**—Although HLA associations and other immunophenotypic data argue strongly for dysregulated adaptive immune responses in oligoarticular and polyarticular JIA, recent evidence also suggests an important role for innate immune activation in these subtypes. Levels of protein S100A8 and S100A9 (or their heterodimeric form S100A8/S100A9) and S100A12, which are secreted products of activated neutrophils and monocytes, are increased in the serum in oligoarticular and polyarticular JIA, and their concentration correlates positively with degree of inflammation.(94) These molecules belong to the damage-associated molecular pattern proteins (DAMPs), which act as endogenous danger signals and activate immune cells and vascular endothelium.(95) Direct cytotoxic effects of the S100 proteins may also contribute to tissue damage.(95) In patients with polyarticular and oligoarticular JIA, protein S100A8/S100A9 levels correlate positively with disease activity and increase before onset of clinical flare, which suggests they act early in the inflammatory cascade associated with periods of disease activity.(96) Levels of protein S100A12 also correlate positively with, and may be early mediators of, disease activity for oligoarticular JIA and polyarticular JIA.(97)

Although neutrophils are the most abundant cell population in JIA synovial fluid, few studies have concentrated on this cell type. Gene expression analysis suggests neutrophil abnormalities exist in polyarticular JIA patients. Expression of more than 700 genes, including IFN $\gamma$ -related and IL-8-related genes, differs in patients with polyarticular JIA compared with healthy controls and some differences persist after response to therapy(98) or even during drug-free clinical remission.(99) The the NADPH cycle is dysregulated in neutrophils from patients with active and inactive RF- polyarticular JIA, which suggests an intrinsic neutrophil defect in these patients.(31)

**Synovial inflammation and tissue damage**—Within each of the oligoarticular and polyarticular JIA subtypes, a subset of patients suffers significant joint damage: the RF<sup>+</sup>/anti-CCP<sup>+</sup> group in polyarticular JIA, and patients with the extended oligoarticular phenotype of oligoarticular JIA.(100, 101) The processes mediating joint damage appear to be shared across JIA subtypes. The hallmarks of inflamed synovium are increased vascularization and endothelial activation. Factors such as vascular endothelial growth factor, osteopontin, and reactive oxygen species induce the formation of new vasculature, thereby facilitating an influx of inflammatory cells. Inflammatory cells subsequently drive the activation of RANK-expressing osteoclasts and proteases, especially MMPs, which increases the ratio of MMPs to their inhibitors, tissue inhibitor of metalloproteinases (TIMPs). Cartilage and bone destruction ensues, enhanced by a reduced capacity for tissue repair (for example, due to increased osteoblast apoptosis) in the setting of inflammation. (102–108), (109)

## CONCLUSIONS

Using clinical phenotypes, epidemiological data and HLA immunogenetics as a foundation, the ILAR criteria for JIA subtypes were defined by consensus conference and have been used globally for clinical and translational research and for the development of practice patterns and treatment algorithms. Implied in this subtyping is the hypothesis that different forms of chronic arthritis in childhood arise through distinct etiopathological pathways. Evidence reviewed here lends support to the general idea of differences in pathogenesis between subtypes. However, some results suggest areas of mechanistic overlap, even between oligoarticular and polyarticular JIA, for example, contributions of Th1 cells, MIF-driven innate immune activation and S100 proteins to joint inflammation. Other results imply further heterogeneity than is captured in the current classification scheme. Based on epidemiological and new molecular data, the ANA+ arthritis subgroup may prove to be mechanistically distinct. The subcategories of persistent and extended oligoarticular JIA may eventually be recognized as independent subtypes, with a defining role for CD4+ CD25+ T reg cells in controlling inflammation in the persistent form. However, much remains to be done to generate full molecular pictures of these conditions. Fortunately, the collaborative research effort that is probably necessary to achieve this goal is starting: an international team of investigators will soon begin working towards developing a biological classification system for JIA (ILAR II, personal communication, R. Yeung).

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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## References

1. Petty RE, Southwood TR, Manners P, Baum J, Glass DN, Goldenberg J, et al. International League of Associations for Rheumatology classification of juvenile idiopathic arthritis: second revision, Edmonton 2001. *The Journal of rheumatology*. 2004; 31(2):390–392. [PubMed: 14760812]
2. Manners P, Lesslie J, Speldewinde D, Tunbridge D. Classification of juvenile idiopathic arthritis: should family history be included in the criteria? *The Journal of rheumatology*. 2003; 30(8):1857–1863. [PubMed: 12913947]
3. Tsitsami E, Bozzola E, Magni-Manzoni S, Viola S, Pistorio A, Ruperto N, et al. Positive family history of psoriasis does not affect the clinical expression and course of juvenile idiopathic arthritis patients with oligoarthritis. *Arthritis and rheumatism*. 2003; 49(4):488–493. [PubMed: 12910554]
4. Berntson L, Damgrd M, Andersson-Gre B, Herlin T, Nielsen S, Nordal E, et al. HLA-B27 predicts a more extended disease with increasing age at onset in boys with juvenile idiopathic arthritis. *The Journal of rheumatology*. 2008; 35(10):2055–2061. [PubMed: 18785306]

5. Stoll M, Lio P, Sundel R, Nigrovic P. Comparison of Vancouver and International League of Associations for rheumatology classification criteria for juvenile psoriatic arthritis. *Arthritis and rheumatism*. 2008; 59(1):51–58. [PubMed: 18163407]
6. Hayem F. Is Still's disease an autoinflammatory syndrome? *Joint, bone, spine*. 2009; 76(1):7–9. [PubMed: 19084456]
7. Ravelli A, Martini A. Juvenile idiopathic arthritis. *The Lancet*. 2007; 369(9563):767–778.
8. Manners PJ. Delay in diagnosing juvenile arthritis. *Medical journal of Australia*. 1999; 171(7):367–369. [PubMed: 10590726]
9. Manners PJ, Diepeveen DA. Prevalence of juvenile chronic arthritis in a population of 12-year-old children in urban Australia. *Pediatrics*. 1996; 98(1):84–90. [PubMed: 8668417]
10. Mielants H, Veys EM, Maertens M, Goemaere S, De Clercq L, Castro S, et al. Prevalence of inflammatory rheumatic diseases in an adolescent urban student population, age 12 to 18, in Belgium. *Clinical and experimental rheumatology*. 1993; 11(5):563–567. [PubMed: 8275595]
11. Manners PJ, Bower C. Worldwide prevalence of juvenile arthritis why does it vary so much? *The Journal of rheumatology*. 2002; 29(7):1520–1530. [PubMed: 12136914]
12. Malleson PN, Fung MY, Rosenberg AM. The incidence of pediatric rheumatic diseases: results from the Canadian Pediatric Rheumatology Association Disease Registry. *The Journal of rheumatology*. 1996; 23(11):1981–1987. [PubMed: 8923379]
13. Denardo BA, Tucker LB, Miller LC, Szer IS, Schaller JG. Demography of a regional pediatric rheumatology patient population. Affiliated Children's Arthritis Centers of New England. *The Journal of rheumatology*. 1994; 21(8):1553–1561. [PubMed: 7983664]
14. Oen K, Fast M, Postl B. Epidemiology of juvenile rheumatoid arthritis in Manitoba, Canada, 1975–92: cycles in incidence. *The Journal of rheumatology*. 1995; 22(4):745–750. [PubMed: 7791175]
15. Sacks JJ, Helmick CG, Luo Y-H, Ilowite NT, Bowyer S. Prevalence of and annual ambulatory health care visits for pediatric arthritis and other rheumatologic conditions in the United States in 2001–2004. *Arthritis and rheumatism*. 2007; 57(8):1439–1445. [PubMed: 18050185]
16. Saurenmann RK, Rose JB, Tyrrell P, Feldman BM, Laxer RM, Schneider R, et al. Epidemiology of juvenile idiopathic arthritis in a multiethnic cohort: ethnicity as a risk factor. *Arthritis and rheumatism*. 2007; 56(6):1974–1984. [PubMed: 17530723]
17. Ravelli A, Felici E, Magni-Manzoni S, Pistorio A, Novarini C, Bozzola E, et al. Patients with antinuclear antibody-positive juvenile idiopathic arthritis constitute a homogeneous subgroup irrespective of the course of joint disease. *Arthritis and rheumatism*. 2005; 52(3):826–832. [PubMed: 15751057]
18. Schwartz MM, Simpson P, Kerr KL, Jarvis JN. Juvenile rheumatoid arthritis in African Americans. *The Journal of rheumatology*. 1997; 24(9):1826–1829. [PubMed: 9292811]
19. Rosenberg AM, Petty RE, Oen KG, Schroeder ML. Rheumatic diseases in Western Canadian Indian children. *The Journal of rheumatology*. 1982; 9(4):589–592. [PubMed: 6982335]
20. Wu CJ, Huang JL, Yang MH, Yan DC, Ou LS, Ho HH. Clinical characteristics of juvenile rheumatoid arthritis in Taiwan. *Journal of microbiology, immunology and infection*. 2001; 34(3):211–214.
21. See Y, Koh ET, Boey ML. One hundred and seventy cases of childhood-onset rheumatological disease in Singapore. *Annals of the Academy of Medicine, Singapore*. 1998; 27(4):496–502.
22. Fujikawa S, Okuni M. A nationwide surveillance study of rheumatic diseases among Japanese children. *Acta paediatrica Japonica*. 1997; 39(2):242–244. [PubMed: 9141265]
23. Sawhney S, Magalhaes C. Paediatric rheumatology--a global perspective. *Best Practice & Research Clinical Rheumatology*. 2006; 20(2):201–221. [PubMed: 16546053]
24. Carvounis PE, Herman DC, Cha S, Burke JP. Incidence and outcomes of uveitis in juvenile rheumatoid arthritis, a synthesis of the literature. *Graefe's archive for clinical and experimental ophthalmology*. 2006; 244(3):281–290.
25. Haffejee IE, Raga J, Coovadia HM. Juvenile chronic arthritis in black and Indian South African children. *South African Medical Journal*. 1984; 65(13):510–514. [PubMed: 6710254]
26. Kurahara DK, Grandinetti A, Fujii LLA, Tokuda AA, Galario JA, Han MJ, et al. Visiting consultant clinics to study prevalence rates of juvenile rheumatoid arthritis and childhood systemic

- lupus erythematosus across dispersed geographic areas. *The Journal of rheumatology*. 2007; 34(2): 425–429. [PubMed: 17295431]
27. Mauldin J, Cameron HD, Jeanotte D, Solomon G, Jarvis JN. Chronic arthritis in children and adolescents in two Indian health service user populations. *BMC musculoskeletal disorders*. 2004; 5:30. [PubMed: 15333136]
  28. Kaipiainen-Seppänen O, Savolainen A. Incidence of chronic juvenile rheumatic diseases in Finland during 1980–1990. *Clinical and experimental rheumatology*. 1996; 14(4):441–444. [PubMed: 8871846]
  29. Towner SR, Michet CJ, O'Fallon WM, Nelson AM. The epidemiology of juvenile arthritis in Rochester, Minnesota 1960–1979. *Arthritis and rheumatism*. 1983; 26(10):1208–1213. [PubMed: 6626278]
  30. Peterson LS, Mason T, Nelson AM, O'Fallon WM, Gabriel SE. Juvenile rheumatoid arthritis in Rochester, Minnesota 1960–1993. Is the epidemiology changing? *Arthritis and rheumatism*. 1996; 39(8):1385–1390. [PubMed: 8702448]
  31. Jarvis JN, Jiang K, Petty HR, Centola M. Neutrophils: the forgotten cell in JIA disease pathogenesis. *Pediatric rheumatology online journal*. 2007; 5:13. [PubMed: 17567896]
  32. Prahalad S, Glass DN. A comprehensive review of the genetics of juvenile idiopathic arthritis. *Pediatric rheumatology online journal*. 2008; 6:11. [PubMed: 18644131]
  33. Prahalad S, O'Brien E, Fraser AM, Kerber RA, Mineau GP, Pratt D, et al. Familial aggregation of juvenile idiopathic arthritis. *Arthritis and rheumatism*. 2004; 50(12):4022–4027. [PubMed: 15593218]
  34. Thompson S, Moroldo M, Guyer L, Ryan M, Tombragel E, Shear E, et al. A genome-wide scan for juvenile rheumatoid arthritis in affected sibpair families provides evidence of linkage. *Arthritis and rheumatism*. 2004; 50(9):2920–2930. [PubMed: 15457461]
  35. Hinks A, Barton A, Shephard N, Eyre S, Bowes J, Cargill M, et al. Identification of a novel susceptibility locus for juvenile idiopathic arthritis by genome-wide association analysis. *Arthritis and rheumatism*. 2009; 60(1):258–263. [PubMed: 19116933]
  36. Murray KJ, Moroldo MB, Donnelly P, Prahalad S, Passo MH, Giannini EH, et al. Age-specific effects of juvenile rheumatoid arthritis-associated HLA alleles. *Arthritis and rheumatism*. 1999; 42(9):1843–1853. [PubMed: 10513798]
  37. Zeggini E, Donn R, Ollier WER, Thomson W. Evidence for linkage of HLA loci in juvenile idiopathic oligoarthritis: independent effects of HLA-A and HLA-DRB1. *Arthritis and rheumatism*. 2002; 46(10):2716–2720. [PubMed: 12384931]
  38. Garavito G, Yunis EJ, Egea E, Ramirez LA, Malagon C, Iglesias A, et al. HLA-DRB1 alleles and HLA-DRB1 shared epitopes are markers for juvenile rheumatoid arthritis subgroups in Colombian mestizos. *Human immunology*. 2004; 65(4):359–365. [PubMed: 15120191]
  39. Huang JL, Yeh CC, Shaw CK, Yao TC, Chen LC, Lee TD, et al. HLA-DRB1 genotyping in patients with juvenile idiopathic arthritis in Taiwan. *European journal of immunogenetics*. 2004; 31(4):185–188. [PubMed: 15265024]
  40. Alsaied K, Haider MZ, Sharma PN, Ayoub EM. The prevalence of human leukocyte antigen (HLA) DR/DQ/DP alleles in Kuwaiti children with oligoarticular juvenile idiopathic arthritis. *Rheumatology international*. 2006; 26(3):224–228. [PubMed: 15703957]
  41. Prahalad S, Ryan MH, Shear ES, Thompson SD, Giannini EH, Glass DN. Juvenile rheumatoid arthritis: linkage to HLA demonstrated by allele sharing in affected sibpairs. *Arthritis and rheumatism*. 2000; 43(10):2335–2338. [PubMed: 11037894]
  42. Hinks A, Ke X, Barton A, Eyre S, Bowes J, Worthington J, et al. Association of the IL2RA/CD25 gene with juvenile idiopathic arthritis. *Arthritis and rheumatism*. 2009; 60(1):251–257. [PubMed: 19116909]
  43. Prahalad S, Shear ES, Thompson SD, Giannini EH, Glass DN. Increased prevalence of familial autoimmunity in simplex and multiplex families with juvenile rheumatoid arthritis. *Arthritis and rheumatism*. 2002; 46(7):1851–1856. [PubMed: 12124869]
  44. Prahalad S, Hansen S, Whiting A, Guthery S, Clifford B, McNally B, et al. Variants in TNFAIP3, STAT4, and C12orf30 loci associated with multiple autoimmune diseases are also associated with

- juvenile idiopathic arthritis. *Arthritis and rheumatism*. 2009; 60(7):2124–2130. [PubMed: 19565500]
45. Marciano R, Giacomelli F, Divizia MT, Gattorno M, Felici E, Pistorio A, et al. A polymorphic variant inside the osteopontin gene shows association with disease course in oligoarticular juvenile idiopathic arthritis. *Annals of the rheumatic diseases*. 2006; 65(5):662–665. [PubMed: 16150788]
  46. Prelog M, Schwarzenbrunner N, Sailer-Hock M, Kern H, Klein-Franke A, Auserlechner MJ, et al. Premature aging of the immune system in children with juvenile idiopathic arthritis. *Arthritis and rheumatism*. 2008; 58(7):2153–2162. [PubMed: 18576332]
  47. Lorenzi AR, Morgan TA, Anderson A, Catterall J, Patterson AM, Foster HE, et al. Thymic function in juvenile idiopathic arthritis. *Annals of the rheumatic diseases*. 2009; 68(6):983–990. [PubMed: 18628282]
  48. Haines C, Giffon T, Lu L-S, Lu X, Tessier-Lavigne M, Ross D, et al. Human CD4+ T cell recent thymic emigrants are identified by protein tyrosine kinase 7 and have reduced immune function. *The Journal of experimental medicine*. 2009; 206(2):275–285. [PubMed: 19171767]
  49. Wouters CHP, Ceuppens JL, Stevens EAM. Different circulating lymphocyte profiles in patients with different subtypes of juvenile idiopathic arthritis. *Clinical and experimental rheumatology*. 2002; 20(2):239–248. [PubMed: 12051407]
  50. Murray KJ, Luyrink L, Grom AA, Passo MH, Emery H, Witte D, et al. Immunohistological characteristics of T cell infiltrates in different forms of childhood onset chronic arthritis. *The Journal of rheumatology*. 1996; 23(12):2116–2124. [PubMed: 8970050]
  51. Wedderburn LR, Patel A, Varsani H, Woo P. Divergence in the degree of clonal expansions in inflammatory T cell subpopulations mirrors HLA-associated risk alleles in genetically and clinically distinct subtypes of childhood arthritis. *International immunology*. 2001; 13(12):1541–1550. [PubMed: 11717195]
  52. Kamphuis S, Hrafnkelsdottir Kn, Klein MR, de Jager W, Haverkamp MH, van Bilsen JHM, et al. Novel self-epitopes derived from aggrecan, fibrillin, and matrix metalloproteinase-3 drive distinct autoreactive T-cell responses in juvenile idiopathic arthritis and in health. *Arthritis research & therapy*. 2006; 8(6):R178. [PubMed: 17129378]
  53. Sedlackova L, Velek J, Vavrincova P, Hromadnikova I. Peripheral blood mononuclear cell responses to heat shock proteins and their derived synthetic peptides in juvenile idiopathic arthritis patients. *Inflammation research*. 2006; 55(4):153–159. [PubMed: 16807696]
  54. De Graeff-Meeder ER, van der Zee R, Rijkers GT, Schuurman HJ, Kuis W, Bijlsma JW, et al. Recognition of human 60 kD heat shock protein by mononuclear cells from patients with juvenile chronic arthritis. *The Lancet*. 1991; 337(8754):1368–1372.
  55. Nguyen TTH, Gehrman M, Zlacka D, Sosna A, Vavrincova P, Multhoff G, et al. Heat shock protein 70 membrane expression on fibroblast-like synovial cells derived from synovial tissue of patients with rheumatoid and juvenile idiopathic arthritis. *Scandinavian journal of rheumatology*. 2006; 35(6):447–453. [PubMed: 17343252]
  56. Prelog M, Schwarzenbrunner N, Tengge E, Sailer-Hock M, Kern H, Zimmerhackl LB, et al. Quantitative alterations of CD8(+) T cells in juvenile idiopathic arthritis patients in remission. *Clinical rheumatology*. 2008
  57. Massa M, Mazzoli F, Pignatti P, De Benedetti F, Passalia M, Viola S, et al. Proinflammatory responses to self HLA epitopes are triggered by molecular mimicry to Epstein-Barr virus proteins in oligoarticular juvenile idiopathic arthritis. *Arthritis and rheumatism*. 2002; 46(10):2721–2729. [PubMed: 12384932]
  58. Stechova K, Vavrincova P, Chudoba D, Frantlova M, Reitzova H, Sosna A, et al. In vitro autoreactivity against skin and synovial cells in patients with juvenile idiopathic and rheumatoid arthritis. *Pathobiology*. 2002; 70(2):76–82. [PubMed: 12476032]
  59. Wedderburn LR, Robinson N, Patel A, Varsani H, Woo P. Selective recruitment of polarized T cells expressing CCR5 and CXCR3 to the inflamed joints of children with juvenile idiopathic arthritis. *Arthritis and rheumatism*. 2000; 43(4):765–774. [PubMed: 10765921]
  60. Saxena N, Aggarwal A, Misra R. Elevated concentrations of monocyte derived cytokines in synovial fluid of children with enthesitis related arthritis and polyarticular types of juvenile idiopathic arthritis. *The Journal of rheumatology*. 2005; 32(7):1349–1353. [PubMed: 15996076]

61. Gattorno M, Prigione I, Morandi F, Gregorio A, Chiesa S, Ferlito F, et al. Phenotypic and functional characterisation of CCR7+ and CCR7- CD4+ memory T cells homing to the joints in juvenile idiopathic arthritis. *Arthritis research & therapy*. 2005; 7(2):R256–R267. [PubMed: 15743472]
62. Martini G, Zulian F, Calabrese F, Bortoli M, Facco M, Cabrelle A, et al. CXCR3/CXCL10 expression in the synovium of children with juvenile idiopathic arthritis. *Arthritis research & therapy*. 2005; 7(2):R241–R249. [PubMed: 15743470]
63. Pharoah DS, Varsani H, Tatham RW, Newton KR, de Jager W, Prakken BJ, et al. Expression of the inflammatory chemokines CCL5, CCL3 and CXCL10 in juvenile idiopathic arthritis, and demonstration of CCL5 production by an atypical subset of CD8+ T cells. *Arthritis research & therapy*. 2006; 8(2):R50. [PubMed: 16507178]
64. Appay V, Dunbar PR, Cerundolo V, McMichael A, Czaplowski L, Rowland-Jones S. RANTES activates antigen-specific cytotoxic T lymphocytes in a mitogen-like manner through cell surface aggregation. *International immunology*. 2000; 12(8):1173–1182. [PubMed: 10917892]
65. Chiesa S, Prigione I, Morandi F, Buoncompagni A, Picco P, Bocca P, et al. Cytokine flexibility of early and differentiated memory T helper cells in juvenile idiopathic arthritis. *The Journal of rheumatology*. 2004; 31(10):2048–2054. [PubMed: 15468374]
66. Murray KJ, Grom AA, Thompson SD, Liewen D, Passo MH, Glass DN. Contrasting cytokine profiles in the synovium of different forms of juvenile rheumatoid arthritis and juvenile spondyloarthritis: prominence of interleukin 4 in restricted disease. *J Rheumatology*. 1998; 25(7):1388–1398.
67. Jarvis JN, Dozmorov I, Jiang K, Frank MB, Szodoray P, Alex P, et al. Novel approaches to gene expression analysis of active polyarticular juvenile rheumatoid arthritis. *Arthritis research & therapy*. 2004; 6(1):R15–R32. [PubMed: 14979934]
68. Harrington LE, Hatton RD, Mangan PR, Turner H, Murphy TL, Murphy KM, et al. Interleukin 17-producing CD4+ effector T cells develop via a lineage distinct from the T helper type 1 and 2 lineages. *Nature immunology*. 2005; 6(11):1123–1132. [PubMed: 16200070]
69. Nistala K, Moncrieffe H, Newton KR, Varsani H, Hunter P, Wedderburn LR. Interleukin-17-producing T cells are enriched in the joints of children with arthritis, but have a reciprocal relationship to regulatory T cell numbers. *Arthritis and rheumatism*. 2008; 58(3):875–887. [PubMed: 18311821]
70. Agarwal S, Misra R, Aggarwal A. Interleukin 17 levels are increased in juvenile idiopathic arthritis synovial fluid and induce synovial fibroblasts to produce proinflammatory cytokines and matrix metalloproteinases. *The Journal of rheumatology*. 2008; 35(3):515–519. [PubMed: 18203309]
71. Milojevic D, Nguyen KD, Wara D, Mellins ED. Regulatory T cells and their role in rheumatic diseases: a potential target for novel therapeutic development. *Pediatric rheumatology online journal*. 2008; 6:20. [PubMed: 19046457]
72. Smolewska E, Brozik H, Smolewski P, Darzynkiewicz Z, Stanczyk J. Regulation of peripheral blood and synovial fluid lymphocyte apoptosis in juvenile idiopathic arthritis. *Scandinavian journal of rheumatology*. 2004; 33(1):7–12. [PubMed: 15124936]
73. Ruprecht CR, Gattorno M, Ferlito F, Gregorio A, Martini A, Lanzavecchia A, et al. Coexpression of CD25 and CD27 identifies FoxP3+ regulatory T cells in inflamed synovia. *The Journal of experimental medicine*. 2005; 201(11):1793–1803. [PubMed: 15939793]
74. de Kleer IM, Wedderburn LR, Taams LS, Patel A, Varsani H, Klein M, et al. CD4+CD25bright regulatory T cells actively regulate inflammation in the joints of patients with the remitting form of juvenile idiopathic arthritis. *The journal of immunology*. 2004; 172(10):6435–6443. [PubMed: 15128835]
75. Wei C-M, Lee J-H, Wang L-C, Yang Y-H, Chang L-Y, Chiang B-L. Frequency and phenotypic analysis of CD4+CD25+ regulatory T cells in children with juvenile idiopathic arthritis. *Journal of microbiology, immunology and infection*. 2008; 41(1):78–87.
76. Massa M, Passalia M, Manzoni SM, Campanelli R, Ciardelli L, Yung GP, et al. Differential recognition of heat-shock protein dnaJ-derived epitopes by effector and Treg cells leads to modulation of inflammation in juvenile idiopathic arthritis. *Arthritis and rheumatism*. 2007; 56(5):1648–1657. [PubMed: 17469159]

77. de Kleer IM, Kamphuis SM, Rijkers GT, Scholtens L, Gordon G, De Jager W, et al. The spontaneous remission of juvenile idiopathic arthritis is characterized by CD30+ T cells directed to human heat-shock protein 60 capable of producing the regulatory cytokine interleukin-10. *Arthritis and rheumatism*. 2003; 48(7):2001–2010. [PubMed: 12847694]
78. Kamphuis S, Kuis W, de Jager W, Teklenburg G, Massa M, Gordon G, et al. Tolerogenic immune responses to novel T-cell epitopes from heat-shock protein 60 in juvenile idiopathic arthritis. *Lancet*. 2005; 366(9479):50–56. [PubMed: 15993233]
79. Gattorno M, Chicha L, Gregorio A, Ferlito F, Rossi F, Jarrossay D, et al. Distinct expression pattern of IFN-alpha and TNF-alpha in juvenile idiopathic arthritis synovial tissue. *Rheumatology*. 2007; 46(4):657–665. [PubMed: 17085467]
80. Smolewska E, Stanczyk J, Brozik H, Biernacka-Zielinska M, Cebula B, Robak T, et al. Distribution and clinical significance of blood dendritic cells in children with juvenile idiopathic arthritis. *Annals of the rheumatic diseases*. 2008; 67(6):762–768. [PubMed: 17965117]
81. Varsani H, Patel A, van Kooyk Y, Woo P, Wedderburn LR. Synovial dendritic cells in juvenile idiopathic arthritis (JIA) express receptor activator of NF-kappaB (RANK). *Rheumatology*. 2003; 42(4):583–590. [PubMed: 12649407]
82. Anderson DM, Maraskovsky E, Billingsley WL, Dougall WC, Tometsko ME, Roux ER, et al. A homologue of the TNF receptor and its ligand enhance T-cell growth and dendritic-cell function. *Nature*. 1997; 390(6656):175–179. [PubMed: 9367155]
83. van Rossum M, van Soesbergen R, de Kort S, ten Cate R, Zwinderman AH, de Jong B, et al. Anti-cyclic citrullinated peptide (anti-CCP) antibodies in children with juvenile idiopathic arthritis. *The Journal of rheumatology*. 2003; 30(4):825–828. [PubMed: 12672206]
84. Griffin T, Barnes M, Ilowite N, Olson J, Sherry D, Gottlieb B, et al. Gene expression signatures in polyarticular juvenile idiopathic arthritis demonstrate disease heterogeneity and offer a molecular classification of disease subsets. *Arthritis and rheumatism*. 2009; 60(7):2113–2123. [PubMed: 19565504]
85. Taniguchi O, Miyajima H, Hirano T, Noguchi M, Ueda A, Hashimoto H, et al. The Leu-1 B-cell subpopulation in patients with rheumatoid arthritis. *Journal of clinical immunology*. 1987; 7(6):441–448. [PubMed: 3500961]
86. Looney RJ. B cell-targeted therapy for rheumatoid arthritis: an update on the evidence. *Drugs*. 2006; 66(5):625–639. [PubMed: 16620141]
87. Mangge H, Kenzian H, Gallistl S, Neuwirth G, Liebmann P, Kaulfersch W, et al. Serum cytokines in juvenile rheumatoid arthritis. Correlation with conventional inflammation parameters and clinical subtypes. *Arthritis and rheumatism*. 1995; 38(2):211–220. [PubMed: 7848311]
88. Yilmaz M, Kendirli SG, Altintas D, Bingol G, Antmen B. Cytokine levels in serum of patients with juvenile rheumatoid arthritis. *Clinical rheumatology*. 2001; 20(1):30–35. [PubMed: 11254238]
89. Lotito APN, Campa A, Silva CAA, Kiss MHB, Mello SBV. Interleukin 18 as a marker of disease activity and severity in patients with juvenile idiopathic arthritis. *The Journal of rheumatology*. 2007; 34(4):823–830. [PubMed: 17343316]
90. de Jager W, Hoppenreijns EPAH, Wulfraat NM, Wedderburn LR, Kuis W, Prakken BJ. Blood and synovial fluid cytokine signatures in patients with juvenile idiopathic arthritis: a cross-sectional study. *Annals of the rheumatic diseases*. 2007; 66(5):589–598. [PubMed: 17170049]
91. Peake NJ, Khawaja K, Myers A, Nowell MA, Jones SA, Rowan AD, et al. Interleukin-6 signalling in juvenile idiopathic arthritis is limited by proteolytically cleaved soluble interleukin-6 receptor. *Rheumatology*. 2006; 45(12):1485–1489. [PubMed: 16690760]
92. Prince FH, Twilt M, Ten Cate R, Van Rossum MA, Armbrust W, Hoppenreijns EP, et al. Long-term follow-up on effectiveness and safety of etanercept in JIA: the Dutch national register. *Annals of the rheumatic diseases*. 2008
93. Rooney M, Varsani H, Martin K, Lombard PR, Dayer JM, Woo P. Tumour necrosis factor alpha and its soluble receptors in juvenile chronic arthritis. *Rheumatology*. 2000; 39(4):432–438. [PubMed: 10817778]
94. Foell D, Wittkowski H, Roth J. Mechanisms of disease: a 'DAMP' view of inflammatory arthritis. *Nature clinical practice rheumatology*. 2007; 3(7):382–390.

95. Foell D, Wittkowski H, Vogl T, Roth J. S100 proteins expressed in phagocytes: a novel group of damage-associated molecular pattern molecules. *Journal of leukocyte biology*. 2007; 81(1):28–37. [PubMed: 16943388]
96. Schulze zur Wiesch A, Foell D, Frosch M, Vogl T, Sorg C, Roth J. Myeloid related proteins MRP8/MRP14 may predict disease flares in juvenile idiopathic arthritis. *Clinical and experimental rheumatology*. 2004; 22(3):368–373. [PubMed: 15144135]
97. Foell D, Wittkowski H, Hammerschmidt I, Wulffraat N, Schmeling H, Frosch M, et al. Monitoring neutrophil activation in juvenile rheumatoid arthritis by S100A12 serum concentrations. *Arthritis and rheumatism*. 2004; 50(4):1286–1295. [PubMed: 15077313]
98. Jarvis JN, Petty HR, Tang Y, Frank MB, Tessier PA, Dozmorov I, et al. Evidence for chronic, peripheral activation of neutrophils in polyarticular juvenile rheumatoid arthritis. *Arthritis research & therapy*. 2006; 8(5):R154. [PubMed: 17002793]
99. Jarvis J, Jiang K, Frank M, Knowlton N, Aggarwal A, Wallace C, et al. Gene expression profiling in neutrophils from children with polyarticular juvenile idiopathic arthritis. *Arthritis and rheumatism*. 2009; 60(5):1488–1495. [PubMed: 19404961]
100. Sandborg C, Holmes TH, Lee T, Biederman K, Bloch DA, Emery H, et al. Candidate early predictors for progression to joint damage in systemic juvenile idiopathic arthritis. *The Journal of rheumatology*. 2006; 33(11):2322–2329. [PubMed: 16960920]
101. Singh-Grewal D, Schneider R, Bayer N, Feldman BM. Predictors of disease course and remission in systemic juvenile idiopathic arthritis: significance of early clinical and laboratory features. *Arthritis and rheumatism*. 2006; 54(5):1595–1601. [PubMed: 16645998]
102. Urakami T, Manki A, Inoue T, Oda M, Tanaka H, Morishima T. Clinical significance of decreased serum concentration of cartilage oligomeric matrix protein in systemic juvenile idiopathic arthritis. *The Journal of rheumatology*. 2006; 33(5):996–1000. [PubMed: 16652429]
103. Nakajima S, Naruto T, Miyamae T, Imagawa T, Mori M, Nishimaki S, et al. Improvement of reduced serum cartilage oligomeric matrix protein levels in systemic juvenile idiopathic arthritis patients treated with the anti-interleukin-6 receptor monoclonal antibody tocilizumab. *Modern rheumatology*. 2008
104. Sarma PK, Misra R, Aggarwal A. Elevated serum receptor activator of NFkappaB ligand (RANKL), osteoprotegerin (OPG), matrix metalloproteinase (MMP)3, and ProMMP1 in patients with juvenile idiopathic arthritis. *Clinical rheumatology*. 2008; 27(3):289–294. [PubMed: 17703334]
105. Gattorno M, Vignola S, Falcini F, Sabatini F, Buoncompagni A, Simonini G, et al. Serum and synovial fluid concentrations of matrix metalloproteinases 3 and its tissue inhibitor 1 in juvenile idiopathic arthritides. *The Journal of rheumatology*. 2002; 29(4):826–831. [PubMed: 11950028]
106. Peake NJ, Khawaja K, Myers A, Jones D, Cawston TE, Rowan AD, et al. Levels of matrix metalloproteinase (MMP)-1 in paired sera and synovial fluids of juvenile idiopathic arthritis patients: relationship to inflammatory activity, MMP-3 and tissue inhibitor of metalloproteinases-1 in a longitudinal study. *Rheumatology*. 2005; 44(11):1383–1389. [PubMed: 16049050]
107. Spelling P, Bonfá E, Caparbo VF, Pereira RMR. Osteoprotegerin/RANKL system imbalance in active polyarticular-onset juvenile idiopathic arthritis: a bone damage biomarker? *Scandinavian journal of rheumatology*. 2008; 37(6):439–444. [PubMed: 18802807]
108. Masi L, Simonini G, Piscitelli E, Del Monte F, Giani T, Cimaz R, et al. Osteoprotegerin (OPG)/RANK-L system in juvenile idiopathic arthritis: is there a potential modulating role for OPG/RANK-L in bone injury? *The Journal of rheumatology*. 2004; 31(5):986–991. [PubMed: 15124262]
109. Caparbo VF, Prada F, Silva CAA, Regio PL, Pereira RMR. Serum from children with polyarticular juvenile idiopathic arthritis (pJIA) inhibits differentiation, mineralization and may increase apoptosis of human osteoblasts "in vitro". *Clinical rheumatology*. 2009; 28(1):71–77. [PubMed: 18685881]



### Key points

- Epidemiologic, genetic and immunophenotypic evidence suggests that oligoarticular (persistent and extended) and polyarticular (RF+ and RF-) JIA subtypes are distinct etiopathological conditions.
- Reported incidence and prevalence of JIA vary widely, due partly to lack of specific diagnostic tests and to rarity of community-based studies.
- Inherited susceptibility factors (both HLA and non-HLA) generally differ among JIA subtypes, although some overlap in the latter suggests shared immunological pathways.
- Evidence supports a key role for T-cell-driven inflammation in oligoarticular and polyarticular JIA and suggests a contribution of regulatory T cells to limiting disease in persistent oligoarticular JIA.
- Dysregulation of cytokines, especially tumor necrosis factor, contribute to oligoarticular and polyarticular JIA pathogenesis, and persistent cytokine abnormalities argue that clinical remission is a state of compensated inflammation.

Table 1

Clinical features of JIA subtypes in white patients from the USA and Europe

Feature	Oligoarticular JIA		Polyarticular JIA	
	Overall	Persistent	Extended	RF <sup>+</sup>
Age at onset	Peak at age 2–4 years	—	—	Late childhood or adolescence
Female:male ratio	3:1	—	—	5.7:1
Number of joints affected	1–4 joints in the first 6 months	1–4 joints throughout the disease course	5 or more joints after first 6 months	5 or more joints
Pattern of arthritis	Affects large joints asymmetrically in the first 6 months (commonly knees and ankles)	Pattern maintained throughout the disease	After 6 months small joints also affected. Presence of ankle and/or wrist involvement within the first 6 months is more common in this subcategory. Erosive disease.	Symmetric polyarthritis in large and small joints, similar to adult rheumatoid arthritis
Frequency	30–60% of all JIA	50% of all oligoarticular JIA *	50% of all oligoarticular JIA *	5–10% of all JIA
Uveitis <sup>‡</sup>	17–26% <sup>§</sup>	14–26% <sup>§</sup>	17–36% <sup>§,  </sup>	0–2%
Systemic symptoms (e.g. fever, weight loss, anemia)	Absent	—	—	Present
Anti-nuclear antibody	75–85%	70–80%	80–95%	~ 55%
Rheumatoid factor	Absent	—	—	Present <sup>¶</sup>
Anti-CCP antibody <sup>#</sup>	0–6%	0–9%	0–6%	~ 55%
Treatment <sup>109</sup>	—	NSAIDs, intra-articular corticosteroids	NSAIDs, methotrexate, TNF inhibitors <sup>**†††</sup>	NSAIDs, methotrexate, TNF inhibitors <sup>**</sup>

\* 4–6 years after disease onset.

‡ Risk factors for uveitis include ANA<sup>+</sup>, young age at diagnosis, female sex and oligoarticular JIA subtype.

§ Higher percentage reported in Finland.<sup>24</sup>

|| Uveitis generally develops before extension of arthritis.<sup>110,111</sup>

¶ RF detected in two or more tests at least 3 months apart during the first 6 months of the disease

# Using a synthetic CCP variant, Low *et al.* found CCP<sup>+</sup> for in 60% of patients with oligoarticular JIA and 93% of those with polyarticular RF<sup>+</sup> JIA.<sup>112</sup>

\*\* Patients refractory to this treatment can be administered other medications including a second anti-TNF agent, sulfasalazine, CTLA4-Ig, rituximab, and leflunomide; no consensus exists on the order of introduction of these medications.

†† Evidence suggests that extended oligoarticular JIA responds to a similar or perhaps greater extent than polyarticular JIA to methotrexate and anti-TNF agents, and comparably to CTLA4-Ig (D. Lovell, personal communication, 113–116)

Abbreviations: +, positive; –, negative; CCP, cyclic citrullinated peptide; JIA, juvenile idiopathic arthritis; RF, rheumatoid factor; TNF, tumor necrosis factor; NSAIDs, Non-steroidal anti-inflammatory drugs.

**Table 2**

HLA alleles associated with oligoarticular and polyarticular JIA in white populations

Clinical subtype of JIA	Susceptibility alleles	Protective alleles
Oligoarticular (both persistent and extended) [AU: OK?OK] <sup>32,117,118</sup>	A2, DRB1*08, DRB1*11, DPB1*0201, DQA1*04, DQB1*04	DRB1*04, DRB1*07
Persistent oligoarticular <sup>117,118</sup>	DRB1*13	DRB1*04
Extended oligoarticular <sup>117</sup>	DRB1*01	DRB1*04
RF <sup>-</sup> polyarticular <sup>32,117,118</sup>	DRB1*08, DQA1*04, DPB1*03	DRB1*04, DRB1*07
RF <sup>+</sup> polyarticular <sup>32,117,118</sup>	DRB1*04, DQA1*03	DQA1*02
Uveitis <sup>117,119,120</sup>	DRB1*11, DRB1*13	DRB1*01 (Zulian et al, 2002)

Abbreviations: JIA, juvenile idiopathic arthritis; RF, rheumatoid factor

**Table 3**

Associations of non-HLA genes with JIA susceptibility independently replicated in white populations

Gene	Subtypes of JIA tested	Subtypes with significant findings	Function of gene product
<i>IL2RA/CD25</i> <sup>42</sup>	All ILAR subtypes	persistent oligoarticular JIA	α chain of the high-affinity IL-2 receptor
<i>MIF</i> <sup>*121–123</sup>	All ILAR subtypes	All ILAR subtypes <sup>§</sup>	Pro-inflammatory cytokine; inhibits macrophage migration
<i>PTPN22</i> <sup>124–126</sup>	All ILAR subtypes except 'other arthritis'	Oligoarticular, polyarticular	Protein tyrosine phosphatase May inhibit the T-cell receptor signaling pathway Predisposing allele is a gain-of-function mutant
<i>SLC11A1</i> (also known as <i>NRAMP1</i> ) <sup>127,128</sup>	OligoarticularJIA, polyarticular JIA	Oligoarticular, polyarticular	Transport of divalent cations in endosomes of macrophages, neutrophils and dendritic cells
<i>TNF</i> <sup>‡129–131</sup>	All ILAR subtypes	Oligoarticular, polyarticular	Pro-inflammatory cytokine
<i>VTCN1</i> (also known as <i>B7H4</i> ) <sup>//5</sup>	Persistent and extended oligoarticular JIA, RF <sup>-</sup> and RF <sup>+</sup> polyarticular-JIA	Oligoarticular (persistent and extended), Polyarticular (RF <sup>-</sup> and RF <sup>+</sup> )	Negatively regulates T-cell-mediated immune response
<i>WISP3</i> <sup>132</sup>	All ILAR subtypes except other arthritis	Polyarticular-course JIA (including extended oligoarticularJIA, RF <sup>-</sup> and RF <sup>+</sup> polyarticular JIA)	Involved in postnatal skeletal growth and cartilage homeostasis

Abbreviations: IL, interleukin; ILAR, International League Against Rheumatism; JIA, juvenile idiopathic arthritis; MIF, macrophage migration inhibitory factor; RF, rheumatoid factor; PTPN22, protein tyrosine phosphatase, non-receptor type 22 (lymphoid); SLC11A1, solute carrier family 11 (proton-coupled divalent metal ion transporters), member 1; NRAMP1, Natural resistance-associated macrophage protein 1; TNF, tumor necrosis factor; VTCN1, V-set domain containing T cell activation inhibitor 1; WISP3, WNT1 inducible signaling pathway protein 3.

\* Also associated with poor outcome in oligoarticular JIA, polyarticular JIA and systemic JIA<sup>133,134</sup>.

‡ Also associated with poor outcome and poor response to treatment.<sup>135–137</sup>

§ Other arthritis included in one study only

// Results of genome-wide association study

**Table 4**

Other non-HLA genes associated with JIA - single reports\*

Gene	Subtypes of JIA tested	Significant associations	Function of gene product
IL-1 family of genes <sup>138-141</sup>	All ILAR subtypes	Susceptibility for oligoarticular JIA ( <i>IL1A</i> , <i>IL1RN</i> ), JIA ( <i>IL1B</i> ) <sup>‡</sup> sJIA ( <i>IL1A</i> , <i>ILF10</i> , <i>IL1RN</i> , <i>IL1R2</i> )	<i>IL1A</i> , <i>IL1B</i> pro-inflammatory cytokines <i>IL1RN</i> IL-1 receptor antagonist <i>IL1R2</i> receptor for IL-1 $\alpha$ , IL-1 $\beta$ and IL-1RN
<i>IL6</i> <sup>§142,143</sup>	Oligoarticular JIA, polyarticular JIA, sJIA	Susceptibility for sJIA	Immunoregulatory cytokine
<i>IL10</i> <sup>144,145</sup>	All ILAR subtypes except other arthritis	Disease course of extended (as opposed to persistent) oligoarticular JIA// Susceptibility for sJIA	Immunoregulator cytokine Inhibits the synthesis of a number of cytokines
<i>IL15</i> <sup>146</sup>	ANA <sup>+</sup> JIA	Susceptibility for ANA <sup>+</sup> JIA	Structurally similar to IL-2 Involved in T and NK cell homeostasis and controls CD8 T cell memory expansion together with IL-2
<i>IL18</i> <sup>¶147</sup>	Oligoarticular JIA, polyarticular JIA, sJIA	Susceptibility for oligoarticular JIA	Belongs to the IL-1 family Pro-inflammatory cytokine
<i>SPP1</i> <sup>148</sup>	Oligoarticular JIA	Disease course of persistent oligoarticular <sup>#</sup>	Osteopontin Immunomodulator Involved in bone remodeling
<i>STAT4</i> <sup>44</sup>	All ILAR subtypes	Susceptibility for RF <sup>-</sup> and RF <sup>+</sup> polyarticular JIA	Essential mediator of signals from IL-12 receptor
<i>TNFAIP3</i> <sup>44</sup>	All ILAR subtypes	Susceptibility for and protective effects in oligoarticular JIA	Modulates inflammatory response by regulating NF- $\kappa$ B signaling

\* All studies, except as noted, conducted in white populations.

<sup>‡</sup> Association with JIA as a whole, without specificity to particular subtypes<sup>§</sup> Two studies showed an association between *IL6* and systemic JIA but with different ages of disease onset (< and > 5 years of age).// Genotype associated with lower IL-10 production.<sup>144,149</sup><sup>¶</sup> Association found in Japanese population; polymorphism association with higher IL-18 production in sJIA.<sup>#</sup> Polymorphism associated with low expression of osteopontin

Abbreviations: +, positive; -, negative; ANA, anti-nuclear antibody; IL, interleukin, ILAR, International League Against Rheumatism; JIA, juvenile idiopathic arthritis; NF- $\kappa$ B, nuclear factor NF; sJIA, systemic JIA; IL2RA: interleukin 2 receptor, alpha; ANA, anti-nuclear antibody; SPP1, secreted phosphoprotein 1; Stat4, signal transducer and activator of transcription 4; TNFAIP3, tumor necrosis factor, alpha-induced protein 3