

Generation of more effective cancer vaccines

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Cancer vaccines represent a promising therapeutic approach for which prime time is imminent. However, clinical efficacy must be improved in order for cancer vaccines to become a valid alternative or complement to traditional cancer treatments. Considerable efforts have been undertaken so far to better understand the fundamental requirements for clinically-effective cancer vaccines. Recent data emphasize that important requirements, among others, are (1) the use of multi-epitope immunogens, possibly deriving from different tumor antigens; (2) the selection of effective adjuvants; (3) the association of cancer vaccines with agents able to counteract the regulatory milieu present in the tumor microenvironment; and (4) the need to choose the definitive formulation and regimen of a vaccine after accurate preliminary tests comparing different antigen formulations. The first requirement deals with issues related to HLA restriction of tumor antigen presentation, as well as usefulness of tumor antigen spreading and counteraction of immune escape phenomena, linked to tumor antigen down-modulation, for an effective anti-cancer immune response. The second point underscores the necessity of optimal activation of innate immunity to achieve an efficient adaptive anti-cancer immune response. The third point focuses on the importance to inhibit subsets of regulatory cells. The last requirement stresses the concept that the regimen and formulation of the vaccine impacts profoundly on cancer vaccine efficacy. A new generation of cancer vaccines, provided with

both immunological and clinical efficacy, will hopefully soon address these requirements.

The approval by the Food and Drug Administration of Sipuleucel-T (Provenge®) for the treatment of advanced prostate cancer provided a boost to the development of cancer vaccines.¹ For the first time the therapeutic potential of cancer vaccines has been officially recognized. However, those working in the field recognize that substantial improvements are required to make cancer vaccines a viable alternative or complement to traditional anti-cancer therapies. Although the rate of vaccine-specific immunological responses is often elevated, the rate of clinical responses is generally low.^{2–4} One of the reasons for these unsatisfactory results could be the inappropriate use of follow-up criteria adopted for conventional cancer therapy. Indeed, RECIST criteria may not be suitable for immunotherapy since they are mainly based on the evaluation of the treatment's eradication potential applied, for instance, to cytolytic therapies.^{5,6} Effective cancer vaccines usually are not directly cytotoxic, leading to inflammation of the tumor rather than immediate necrosis. Hence, an immunotherapy-sensitive lesion could show stable or even increased size for long time, thus failing to show amelioration when RECIST criteria are used to assess disease progression. This could cause premature withdrawal from the immunotherapy protocol, thus precluding potentially positive responses to treatment from being discovered. For this reasons modified RECIST criteria have been proposed for immunotherapies, and

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future analyses will enable an understanding of whether vaccination can improve the rate of successful treatments.⁷

Apart from these considerations, we must be aware that optimal schedules for cancer vaccine protocols must be identified, which is the real challenge. In fact, several aspects must be taken into consideration in the setting of an optimal vaccine regimen. Indeed, the first point of discussion is the choice of the immunizing antigen. A plethora of tumor antigens has been identified—but how to choose among them? An attempt to clarify the issue by the National Cancer Institute categorizes each tumor antigen on the basis of its capacity to fulfill criteria such as therapeutic function, immunogenicity, oncogenicity, specificity, expression level, stem cell expression, number of patients with antigen-positive cancers, number of epitopes, and cellular location of expression.⁸ What emerges from this proposal is that the ideal tumor antigen does not exist and hardly one will be identified with such characteristics. Therefore, before selecting a tumor antigen, an answer must be provided to these questions: (1) Is the vaccine antigen specific to a single tumor type, or is it common to many types of cancer? (2) Is it a surface antigen? (3) Is the candidate tumor antigen necessary for tumor growth and survival, or not? (4) Can several tumor antigens be associated with each other? In addition, tumor-specific antigens^{9,10} need to selectively induce immune responses against tumors while sparing normal tissues.

Recently we have witnessed the attempt to develop personalized cancer vaccines, and some groups are applying genomic and proteomic approaches to the search for unique single-tumor-specific antigens.^{11,12} Theoretically, the more tumor-restricted the antigen, the more specific the immune response will be, thereby creating the conditions for high specificity of the response and greater avoidance of side effects related to collateral damage of healthy tissues by the vaccine-induced immune response. While appealing, this approach would result in the proliferation of numbers of cancer vaccines that exceed the types of cancer. An extreme characterization of this new trend will be a personalized cancer vaccine unique for each patient. Is this an effort we can afford? If we think to cancer vaccines as a therapy of the future, possibly adopted in any country and at any latitude, is it realistic to imagine a widespread diffusion of such an onerous (technically and economically) approach? An alternative could be the choice as immunogens of universal tumor-associated antigens (TAA, e.g., telomerase, survivin),^{13,14} expressed by the majority of cancers. In this case, the same vaccine could be applied to the treatment of several tumor types. Telomerase-based vaccines are examples demonstrating feasibility and efficacy of this approach.^{15–21} But are the two strategies really alternatives to each other?

Evidence suggests that antigen spreading may occur in effective vaccine-induced immune responses against cancer.^{22,23}

Hence, a scenario can be envisaged in which an initial anti-tumor immune response (in any kind of tumor) could be induced by a cancer vaccine based on a universal TAA, and a subsequent tumor-specific boost could be provided by immunization against a tumor-specific antigen. The immunization against the universal TAA would induce an initial immune response leading to *in situ* inflammation and recruitment of lymphocytes specific to a broad spectrum of tumor antigen specificities; the immunization against the tumor-specific antigen(s) would allow the expansion and affinity selection of (more) strictly tumor-specific T-cell clones. This dual-faced strategy also would have the advantage of targeting multiple epitopes/antigens at the same time. This is important since one of the most effective mechanisms of tumor immune escape is down-modulation of tumor antigen.²⁴ Indeed, the greater the number (and type) of target antigens, the more difficult it will be for the tumor to escape immune surveillance through antigen modulation. Accordingly, vaccines have been developed including multiple epitopes of tumor antigens.^{11,15,25–27} However, the inclusion of multiple epitopes in a vaccine does not protect against the risk of tumor immune escape via antigen down-modulation. We reason that the biological characteristics of the immunizing molecule may have an impact on the immunogenicity of the cancer vaccine. For instance, tumor antigens likely to offer greater immunogenicity could be (1) those strictly related to the

Table 1. Some options and requirements relevant for the setting of a cancer vaccine

Issue	Options	Suggested requirements
Tumor antigen(s)	(1) Tumor-specific (2) Universal (3) Both	(1) Involved in pathways of tumor growth or survival (2) Surface antigen
Immunizing epitope(s)	(1) Single (2) Multiple	(1) Restricted by both HLA class I and II molecules (2) Restricted by multiple HLA alleles
Activation of innate immunity	(1) One adjuvant (2) Multiple adjuvants	Activation of multiple pathways and functions
Inhibition of regulatory cells	(1) Biological agents (2) Chemotherapy	Inhibition of multiple cell types and functions
Regimen and formulation	(1) Routes of administration (2) Schedules (3) Antigen formulation (cell lysate, protein, peptide, DNA, RNA) (4) Co-administration of cytokines	Selection through preclinical testing

mechanisms of tumor growth/survival, and (2) those expressed at the cell surface. The former type of antigens would tend not to be down-modulated due to the importance in the economy of the tumor growth; the efficacy of immunotherapies targeting protein tyrosine phosphatases supports this consideration.²⁸ The second type of tumor antigen could allow the combined targeting of the same molecule by vaccine-induced T cells and by specific cytotoxic/neutralizing antibodies. This combined but little-explored approach²⁹ could prove beneficial against immune escape due to antigen downregulation and/or an impairment of the antigen presentation machinery.²⁴

An important issue in formulating a multi-epitope cancer vaccine is selection of the immunogenic epitopes. Two major rules must be taken into consideration: (1) the need to activate both CD4⁺ and CD8⁺ tumor-specific T lymphocytes in order to achieve efficient tumor rejection;^{30–33} and (2) the opportunity to conform to the widest possible array of HLA haplotypes in order to make the vaccine suitable for patients with the greatest genetic assortment. Several vaccination procedures allow to fulfill such requirements, e.g., those using as immunogens either whole molecules (as in the case of DNA vaccines, RNA vaccines, or vaccines consisting of tumor lysate-pulsed dendritic cells [DC])³⁴ or multi-peptide vaccines containing promiscuous peptides (i.e., peptides binding to various HLA molecules).²⁶ As for peptide vaccines, recent data from this and other laboratories corroborate the concept that multi-epitope vaccines may be both immunologically and clinically effective.^{11,15,35}

An essential requirement for achieving effective anti-cancer immune responses is the optimal activation of innate immunity. In fact, the immune system generates effector responses when it “senses” danger signals.³⁶ Pattern recognition receptors are devoted to this function; when stimulated they induce the activation of genomic pathways leading to complex integrated effector functions.³⁷ Adoptive immune responses are strictly dependent on adequate activation of innate immunity.³⁸ Hence, the selection of the appropriate adjuvant is as important as that of

the immunizing tumor antigen. Again, a wide choice of adjuvants is now available.³⁹ Each of them has specific activities, but none has the capacity to mediate alone the complex network of activating signals that pathogens have. This provides the rationale for the use of multiple adjuvants, preferably those with complementary functions.⁴⁰ Accordingly, in the GX301 vaccine we associated Montanide ISA-51 and Imiquimod. The first adjuvant generates a water-in-oil emulsion, which protects vaccine peptides from tissue proteases and favors uptake by antigen presenting cells. Moreover, it induces IFN γ release by innate immunity cells, favoring the expression of HLA molecules by tumor cells.⁴¹ The second adjuvant is a potent activator of Toll-like receptors 7 and 8, inducing strong activation of DCs.⁴² Collectively, these two adjuvants exert complementary functions fostering simultaneous uptake and presentation of vaccine peptides. It is of interest that by associating the multi-epitope and the dual adjuvant strategies, we achieved 100% of cancer-specific immune responses in a series of highly advanced prostate or renal cancer patients.¹⁵ Indeed, future studies are needed to explore the feasibility and efficacy of other adjuvant associations in order to select the most effective ones.

Several other aspects deserve mention, including the importance of the route, tools and schedule of vaccine administration, the selection of the appropriate antigen-presenting cell, the usefulness of cytokine co-administration, and the opportunity to associate cancer vaccines and chemotherapy drugs. However, all these points have been exhaustively reviewed recently,^{10,43–46} and will not be analyzed here.

A final issue has fundamental relevance to effective cancer vaccines: the inhibition of tumor-dependent regulatory functions. The therapeutic efficacy of Ipilimumab supports this point.⁴⁷ We are aware that several regulatory cell subsets co-exist within tumor microenvironment, e.g., CD4⁺ and CD8⁺ T regulatory lymphocytes, myeloid derived suppressor cells, and tumor-associated macrophages.^{48,49} Hence, a combination of inhibitors that counteract different regulatory pathways is likely to enhance the immunogenicity

of cancer vaccines. Thus, in an experimental model of melanoma, we consistently achieved 100% protection from tumor growth when the vaccine was co-administered with a neutralizing anti-IL10 antibody,⁵⁰ given that IL10 secretion is a hallmark of different subtypes of cells with regulatory function.

In conclusion, an ideal cancer vaccine must integrate many options with respect to its composition and the several requirements discussed above (Table 1). Furthermore every possible vaccine combination should be tested at the preclinical level to select the combination that best fulfills the requirements desired for clinical efficacy. It is no surprise that the same tumor antigen may yield different clinical results depending on the immunization setting.⁵⁰

Collectively, the multitude of data obtained so far indicate that the right approach for generating more effective cancer vaccines is combinatorial. In fact, only by combining different (biological and traditional) agents will it be possible to fulfill the numerous requirements enabling cancer vaccinology to move from empiricism to a well-structured science, and cancer vaccines to move from promise to reality.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

References

1. Kantoff PW, Higano CS, Shore ND, Berger ER, Small EJ, Penson DF, Redfern CH, Ferrari AC, Dreicer R, Sims RB, et al.; IMPACT Study Investigators. Sipuleucel-T immunotherapy for castration-resistant prostate cancer. *N Engl J Med* 2010; 363:411–22; PMID:20818862; <http://dx.doi.org/10.1056/NEJMoa1001294>
2. Parmiani G, Castelli C, Dalerba P, Mortarini R, Rivoltini L, Marincola FM, Anichini A. Cancer immunotherapy with peptide-based vaccines: what have we achieved? Where are we going? *J Natl Cancer Inst* 2002; 94:805–18; PMID:12048268; <http://dx.doi.org/10.1093/jnci/94.11.805>
3. Rosenberg SA, Yang JC, Restifo NP. Cancer immunotherapy: moving beyond current vaccines. *Nat Med* 2004; 10:909–15; PMID:15340416; <http://dx.doi.org/10.1038/nm1100>
4. Yang JC. Melanoma vaccines. *Cancer J* 2011; 17:277–82; PMID:21952276; <http://dx.doi.org/10.1097/PPO.0b13e3182325f72>

5. Therasse P, Arbuick SG, Eisenhauer EA, Wanders J, Kaplan RS, Rubinstein L, Verweij J, Van Glabbeke M, van Oosterom AT, Christian MC, et al. New guidelines to evaluate the response to treatment in solid tumors. European Organization for Research and Treatment of Cancer, National Cancer Institute of the United States, National Cancer Institute of Canada. *J Natl Cancer Inst* 2000; 92:205-16; PMID:10655437; <http://dx.doi.org/10.1093/jnci/92.3.205>
6. Scher HI, Halabi S, Tannock I, Morris M, Sternberg CN, Carducci MA, Eisenberger MA, Higano C, Bubley GJ, Dreicer R, et al.; Prostate Cancer Clinical Trials Working Group. Design and end points of clinical trials for patients with progressive prostate cancer and castrate levels of testosterone: recommendations of the Prostate Cancer Clinical Trials Working Group. *J Clin Oncol* 2008; 26:1148-59; PMID:18309951; <http://dx.doi.org/10.1200/JCO.2007.12.4487>
7. Wolchok JD, Hoos A, O'Day S, Weber JS, Hamid O, Lebbé C, Maio M, Binder M, Bohnsack O, Nichol G, et al. Guidelines for the evaluation of immune therapy activity in solid tumors: immune-related response criteria. *Clin Cancer Res* 2009; 15:7412-20; PMID:19934295; <http://dx.doi.org/10.1158/1078-0432.CCR-09-1624>
8. Cheever MA, Allison JP, Ferris AS, Finn OJ, Hastings BM, Hecht TT, Mellman I, Prindiville SA, Viner JL, Weiner LM, et al. The prioritization of cancer antigens: a national cancer institute pilot project for the acceleration of translational research. *Clin Cancer Res* 2009; 15:5323-37; PMID:19723653; <http://dx.doi.org/10.1158/1078-0432.CCR-09-0737>
9. Parmiani G, Castelli C, Rivoltini L, Casati C, Tully GA, Novellino L, Patuzzo A, Tosi D, Anichini A, Santinami M. Immunotherapy of melanoma. *Semin Cancer Biol* 2003; 13:391-400; PMID:15001157; <http://dx.doi.org/10.1016/j.semcancer.2003.09.001>
10. Aly HA. Cancer therapy and vaccination. *J Immunol Methods* 2012; 382:1-23; PMID:22658969; <http://dx.doi.org/10.1016/j.jim.2012.05.014>
11. Walter S, Weinschenk T, Stenzl A, Zdrojowy R, Pluzanska A, Szczylik C, Staehler M, Brugger W, Dietrich PY, Mendrzyk R, et al. Multipetide immune response to cancer vaccine IMA901 after single-dose cyclophosphamide associates with longer patient survival. *Nat Med* 2012; 18:1254-61; PMID:22842478; <http://dx.doi.org/10.1038/nm.2883>
12. Haen SP, Rammensee HG. The repertoire of human tumor-associated epitopes—identification and selection of antigens and their application in clinical trials. *Curr Opin Immunol* 2013; 25:277-83; PMID:23619309; <http://dx.doi.org/10.1016/j.coi.2013.03.007>
13. Beatty GL, Vonderheide RH. Telomerase as a universal tumor antigen for cancer vaccines. *Expert Rev Vaccines* 2008; 7:881-7; PMID:18767939; <http://dx.doi.org/10.1586/14760584.7.7.881>
14. Andersen MH, Svane IM, Becker JC, Straten PT. The universal character of the tumor-associated antigen survivin. *Clin Cancer Res* 2007; 13:5991-4; PMID:17947459; <http://dx.doi.org/10.1158/1078-0432.CCR-07-0686>
15. Fenoglio D, Traverso P, Parodi A, Tomasello L, Negrini S, Kalli F, Battaglia F, Ferrera F, Sciallero S, Murdaca G, et al. A multi-peptide, dual-adjunct telomerase vaccine (GX301) is highly immunogenic in patients with prostate and renal cancer. *Cancer Immunol Immunother* 2013; 62:1041-52; PMID:23591981; <http://dx.doi.org/10.1007/s00262-013-1415-9>
16. Brunsvig PF, Kyte JA, Kersten C, Sundström S, Møller M, Nyakas M, Hansen GL, Gaudernack G, Aamdal S. Telomerase peptide vaccination in NSCLC: a phase II trial in stage III patients vaccinated after chemoradiotherapy and an 8-year update on a phase I/II trial. *Clin Cancer Res* 2011; 17:6847-57; PMID:21918169; <http://dx.doi.org/10.1158/1078-0432.CCR-11-1385>
17. Hunger RE, Kernland Lang K, Markowski CJ, Trachsel S, Möller M, Eriksen JA, Rasmussen AM, Braathen LR, Gaudernack G. Vaccination of patients with cutaneous melanoma with telomerase-specific peptides. *Cancer Immunol Immunother* 2011; 60:1553-64; PMID:21681371; <http://dx.doi.org/10.1007/s00262-011-1061-z>
18. Rittig SM, Haentschel M, Weimer KJ, Heine A, Müller MR, Brugger W, Horger MS, Maksimovic O, Stenzl A, Hoerr I, et al. Intradermal vaccinations with RNA coding for TAA generate CD8+ and CD4+ immune responses and induce clinical benefit in vaccinated patients. *Mol Ther* 2011; 19:990-9; PMID:21189474; <http://dx.doi.org/10.1038/mt.2010.289>
19. Berntsen A, Trepiakos R, Wenandy L, Geertsen PF, thor Straten P, Andersen MH, Pedersen AE, Claesson MH, Lorentzen T, Johansen JS, et al. Therapeutic dendritic cell vaccination of patients with metastatic renal cell carcinoma: a clinical phase 1/2 trial. *J Immunother* 2008; 31:771-80; PMID:18779742; <http://dx.doi.org/10.1097/CJL.0b013e3181833818>
20. Cortez-Gonzalez X, Zanetti M. Telomerase immuno-unity from bench to bedside: round one. *J Transl Med* 2007; 5:12; PMID:17324292; <http://dx.doi.org/10.1186/1479-5876-5-12>
21. Su Z, Dannull J, Heiser A, Yancey D, Pruitt S, Madden J, Coleman D, Niedzwiecki D, Gilboa E, Vieweg J. Immunological and clinical responses in metastatic renal cancer patients vaccinated with tumor RNA-transfected dendritic cells. *Cancer Res* 2003; 63:2127-33; PMID:12727829
22. Corbière V, Chapiro J, Stroobant V, Ma W, Lurquin C, Lethé B, van Baren N, Van den Eynde BJ, Boon T, Coulie PG. Antigen spreading contributes to MAGE vaccination-induced regression of melanoma metastases. *Cancer Res* 2011; 71:1253-62; PMID:21216894; <http://dx.doi.org/10.1158/0008-5472.CAN.10-2693>
23. Inderberg-Suso EM, Trachsel S, Lislerud K, Rasmussen AM, Gaudernack G. Widespread CD4+ T-cell reactivity to novel hTERT epitopes following vaccination of cancer patients with a single hTERT peptide GV1001. *Oncoimmunology* 2012; 1:670-86; PMID:22934259; <http://dx.doi.org/10.4161/onci.20426>
24. Rivoltini L, Carrabba M, Huber V, Castelli C, Novellino L, Dalerba P, Mortarini R, Arancia G, Anichini A, Fais S, et al. Immunity to cancer: attack and escape in T lymphocyte-tumor cell interaction. *Immunol Rev* 2002; 188:97-113; PMID:12445284; <http://dx.doi.org/10.1034/j.1600-065X.2002.18809.x>
25. Slingluff CL Jr., Lee S, Zhao F, Chianese-Bullock KA, Olson WC, Butterfield LH, Whiteside TL, Leming PD, Kirkwood JM. A Randomized Phase II Trial of Multi-epitope Vaccination with Melanoma Peptides for Cytotoxic T-Cells and Helper T-Cells for Patients with Metastatic Melanoma (E1602). *Clin Cancer Res* 2013; In press; <http://dx.doi.org/10.1158/1078-0432.CCR-13-0002>
26. Yamada A, Sasada T, Noguchi M, Itoh K. Next-generation peptide vaccines for advanced cancer. *Cancer Sci* 2013; 104:15-21; PMID:23107418; <http://dx.doi.org/10.1111/cas.12050>
27. Phuphanich S, Wheeler CJ, Rudnick JD, Mazer M, Wang H, Nuño MA, Richardson JE, Fan X, Ji J, Chu RM, et al. Phase I trial of a multi-epitope-pulsed dendritic cell vaccine for patients with newly diagnosed glioblastoma. *Cancer Immunol Immunother* 2013; 62:125-35; PMID:22847020; <http://dx.doi.org/10.1007/s00262-012-1319-0>
28. Nunes-Xavier CE, Martín-Pérez J, Elson A, Pulido R. Protein tyrosine phosphatases as novel targets in breast cancer therapy. *Biochim Biophys Acta* 2013; 1836:211-26; PMID:23756181
29. Disis ML, Wallace DR, Gooley TA, Dang Y, Slota M, Lu H, Coveler AL, Childs JS, Higgins DM, Fintak PA, et al. Concurrent trastuzumab and HER2/neu-specific vaccination in patients with metastatic breast cancer. *J Clin Oncol* 2009; 27:4685-92; PMID:19720923; <http://dx.doi.org/10.1200/JCO.2008.20.6789>
30. Pardoll DM, Topalian SL. The role of CD4+ T cell responses in antitumor immunity. *Curr Opin Immunol* 1998; 10:588-94; PMID:9794842; [http://dx.doi.org/10.1016/S0952-7915\(98\)80228-8](http://dx.doi.org/10.1016/S0952-7915(98)80228-8)
31. Mumberg D, Monach PA, Wanderling S, Philip M, Toledano AY, Schreiber RD, Schreiber H. CD4(+) T cells eliminate MHC class II-negative cancer cells in vivo by indirect effects of IFN-gamma. *Proc Natl Acad Sci U S A* 1999; 96:8633-8; PMID:10411927; <http://dx.doi.org/10.1073/pnas.96.15.8633>
32. Hung K, Hayashi R, Lafond-Walker A, Lowenstein C, Pardoll D, Levitsky H. The central role of CD4(+) T cells in the antitumor immune response. *J Exp Med* 1998; 188:2357-68; PMID:9858522; <http://dx.doi.org/10.1084/jem.188.12.2357>
33. Ossendorf F, Mengedé E, Camps M, Filius R, Melief CJ. Specific T helper cell requirement for optimal induction of cytotoxic T lymphocytes against major histocompatibility complex class II negative tumors. *J Exp Med* 1998; 187:693-702; PMID:9480979; <http://dx.doi.org/10.1084/jem.187.5.693>
34. Vergari M, Intrivici C, Huen NY, Schlom J, Tsang KY. Strategies for cancer vaccine development. *J Biomed Biotechnol* 2010; 2010:1; PMID:20706612; <http://dx.doi.org/10.1155/2010/596432>
35. Kenter GG, Welters MJ, Valentijn AR, Lowik MJ, Berends-van der Meer DM, Vloon AP, Essahsah F, Fathers LM, Offringa R, Drijfhout JW, et al. Vaccination against HPV-16 oncoproteins for vulvar intraepithelial neoplasia. *N Engl J Med* 2009; 361:1838-47; PMID:19890126; <http://dx.doi.org/10.1056/NEJMoa0810097>
36. Gallucci S, Matzinger P. Danger signals: SOS to the immune system. *Curr Opin Immunol* 2001; 13:114-9; PMID:1154927; [http://dx.doi.org/10.1016/S0952-7915\(00\)00191-6](http://dx.doi.org/10.1016/S0952-7915(00)00191-6)
37. Kumar H, Kawai T, Akira S. Pathogen recognition by the innate immune system. *Int Rev Immunol* 2011; 30:16-34; PMID:21235323; <http://dx.doi.org/10.3109/08830185.2010.529976>
38. Akira S. Innate immunity and adjuvants. *Philos Trans R Soc Lond B Biol Sci* 2011; 366:2748-55; PMID:21893536; <http://dx.doi.org/10.1098/rstb.2011.0106>
39. Dubensky TW Jr., Reed SG. Adjuvants for cancer vaccines. *Semin Immunol* 2010; 22:155-61; PMID:20488726; <http://dx.doi.org/10.1016/j.smim.2010.04.007>
40. Chiang CL, Kandalafte LE, Coukos G. Adjuvants for enhancing the immunogenicity of whole tumor cell vaccines. *Int Rev Immunol* 2011; 30:150-82; PMID:21557641; <http://dx.doi.org/10.3109/08830185.2011.572210>
41. Aucouturier J, Dupuis L, Deville S, Ascarateil S, Ganne V. Montanide ISA 720 and 51: a new generation of water in oil emulsions as adjuvants for human vaccines. *Expert Rev Vaccines* 2002; 1:111-8; PMID:12908518; <http://dx.doi.org/10.1586/14760584.1.1.111>
42. Schön MP, Schön M. Imiquimod: mode of action. *Br J Dermatol* 2007; 157(Suppl 2):8-13; PMID:18067624; <http://dx.doi.org/10.1111/j.1365-2133.2007.08265.x>
43. Bal SM, Ding Z, van Riet E, Jiskoot W, Bouwstra JA. Advances in transcutaneous vaccine delivery: do all ways lead to Rome? *J Control Release* 2010; 148:266-82; PMID:20869998; <http://dx.doi.org/10.1016/j.jconrel.2010.09.018>
44. Palucka K, Banchereau J, Mellman I. Designing vaccines based on biology of human dendritic cell subsets. *Immunity* 2010; 33:464-78; PMID:21029958; <http://dx.doi.org/10.1016/j.immuni.2010.10.007>

45. Church SE, Jensen SM, Twitty CG, Bahjat K, Hu HM, Urba WJ, Fox BA. Multiple vaccinations: friend or foe. *Cancer J* 2011; 17:379-96; PMID:21952289; <http://dx.doi.org/10.1097/PPO.0b013e3182346320>
46. Arens R, van Hall T, van der Burg SH, Ossendorp F, Melief CJ. Prospects of combinatorial synthetic peptide vaccine-based immunotherapy against cancer. *Semin Immunol* 2013; 00024-9; PMID:23706598
47. Sharma P, Wagner K, Wolchok JD, Allison JP. Novel cancer immunotherapy agents with survival benefit: recent successes and next steps. *Nat Rev Cancer* 2011; 11:805-12; PMID:22020206; <http://dx.doi.org/10.1038/nrc3153>
48. Ostrand-Rosenberg S. Immune surveillance: a balance between protumor and antitumor immunity. *Curr Opin Genet Dev* 2008; 18:11-8; PMID:18308558; <http://dx.doi.org/10.1016/j.gde.2007.12.007>
49. Filaci G, Fenoglio D, Fravega M, Ansaldo G, Borgonovo G, Traverso P, Villaggio B, Ferrera A, Kunkl A, Rizzi M, et al. CD8+ CD28- T regulatory lymphocytes inhibiting T cell proliferative and cytotoxic functions infiltrate human cancers. *J Immunol* 2007; 179:4323-34; PMID:17878327
50. Kalli F, Machiorlatti R, Battaglia F, Parodi A, Conteduca G, Ferrera F, Proietti M, Tardito S, Sanguineti M, Millo E, et al. Comparative analysis of cancer vaccine settings for the selection of an effective protocol in mice. *J Transl Med* 2013; 11:120; PMID:23663506; <http://dx.doi.org/10.1186/1479-5876-11-120>