

Published in final edited form as:

Cancer Epidemiol Biomarkers Prev. 2011 October ; 20(10): 2222–2231. doi:
10.1158/1055-9965.EPI-11-0569.

Confirmation of 5p12 as a susceptibility locus for progesterone-receptor-positive, lower grade breast cancer

A full list of authors and affiliations appears at the end of the article.

Abstract

Background—The single nucleotide polymorphism 5p12-rs10941679 has been found to be associated with risk of breast cancer, particularly estrogen receptor (ER)-positive disease. We aimed to further explore this association overall, and by tumor histopathology, in the Breast Cancer Association Consortium.

Methods—Data were combined from 37 studies, including 40,972 invasive cases, 1,398 cases of ductal carcinoma in situ (DCIS) and 46,334 controls, all of white European ancestry, as well as 3,007 invasive cases and 2,337 controls of Asian ancestry. Associations overall and by tumor invasiveness and histopathology were assessed using logistic regression.

Results—For white Europeans, the per-allele odds ratio (OR) associated with 5p12-rs10941679 was 1.11 (95% confidence interval [CI] =1.08–1.14, $P=7\times 10^{-18}$) for invasive breast cancer and 1.10 (95% CI=1.01–1.21, $P=0.03$) for DCIS. For Asian women, the estimated OR for invasive disease was similar (OR=1.07, 95% CI=0.99–1.15, $P=0.09$). Further analyses suggested that the association in white Europeans was largely limited to progesterone receptor (PR)-positive disease (per-allele OR=1.16, 95% CI=1.12–1.20, $P=1\times 10^{-18}$ versus OR=1.03, 95% CI=0.99–1.07, $P=0.2$ for PR-negative disease; P -heterogeneity= 2×10^{-7}); heterogeneity by estrogen receptor status was not observed ($P=0.2$) once PR status was accounted for. The association was also stronger for lower-grade tumors (per-allele OR [95% CI]=1.20 [1.14–1.25], 1.13 [1.09–1.16] and 1.04 [0.99–1.08] for grade 1, 2 and 3/4, respectively; P -trend= 5×10^{-7}).

Conclusion—5p12 is a breast cancer susceptibility locus for PR-positive, lower grade breast cancer.

Impact—Multi-centre fine-mapping studies of this region are needed as a first step to identifying the causal variant or variants.

Keywords

Breast cancer; SNP; susceptibility; disease subtypes

Corresponding author: Roger L. Milne, Genetic & Molecular Epidemiology Group, Spanish National Cancer Research Centre (CNIO), Melchor Fernández Almagro 3, 28029 Madrid, Spain (rmilne@cnio.es).

*Contributed equally to this work

Conflicts of interest: None to declare

Introduction

Genome-wide association studies (GWAS) have identified several single-nucleotide polymorphisms (SNPs) associated with breast cancer risk. Key to these findings, and to the more precise estimation of the associated relative risks, has been their replication in large independent case-control series. SNPs in or close to the genes *LSP1*, *MAP3K1*, *FGFR2*, *TOX3*, *MRPS30*, *COX 11*, *MRPS30*, and *SLC4A7*, and in chromosomal regions 8p24 and 2q35 have all been replicated in the Breast Cancer Association Consortium (BCAC) (1–4)

An Icelandic GWAS and replication study of 5,028 breast cancer cases and 32,090 controls revealed multiple signals in the 5p12 chromosomal region (5). The most strongly associated SNP was rs10941679, the minor G allele being associated with an estimated per-allele odds ratio (OR) of 1.19 (95% confidence interval [CI]=1.11–1.26, $P=2.9\times 10^{-11}$). The authors reported that the increased risk was particularly marked for estrogen receptor (ER)-positive disease (N=2,726 cases, per-allele OR=1.27, 95%CI=1.19–1.35, $P=2.5\times 10^{-12}$; P -heterogeneity=0.004) (5). In addition, our previous breast cancer GWAS reported evidence of an association with another SNP on 5p12, rs981782 ($P=9\times 10^{-6}$), based on data from 23,408 cases and 24,636 controls from 21 BCAC studies (2). The Icelandic-led study evaluated both these SNPs in multivariable models and observed that only 5p12-rs10941679 was independently associated with breast cancer risk (5). This SNP was not among those genotyped in the first phase of our previous GWAS (2).

The aims of the present study were to estimate the relative risk of breast cancer associated with 5p12-rs10941679 in a much larger case-control series comprising studies participating in the BCAC, to evaluate its independence of any association with 5p12-rs981782, and to assess associations by disease subtypes defined by histopathology.

Materials and Methods

Thirty-seven studies from Europe, North America, Australia, and Asia participated in genotyping for the present study via the BCAC, contributing a total of 41,243 invasive breast cancer cases, 1,406 cases of ductal carcinoma in-situ (DCIS) and 46,621 controls of white European origin and 3,082 invasive cases and 2,402 controls of Asian origin. All 37 studies genotyped 5p12-rs10941679. Seven studies also genotyped 5p12-rs981782 in a total of 8,247 invasive cases and 10,363 controls, all of European origin. Fourteen of the 37 participating studies had already genotyped 5p12-rs981782 in white Europeans as part of the previously published study (2). Descriptions of studies are provided in Supplemental Table 1 and final sample sizes are presented in Supplemental Table 2.

All studies provided information on disease status and self-reported race/ethnicity for all subjects and age at diagnosis for cases. All but five studies (BIGGS, FBSC, HUBCS, RBCS and UCIBCS) also provided age at data collection for controls. “Selected cases” were a subset of 7,128 invasive cases selected for inclusion or oversampled by nine studies because they had bilateral breast cancer, a family history of breast cancer and/or other characteristics that suggested they were at increased genetic risk (all cases from BBSC, FBSC, GC-HBOC, kConFab/AOCS, MBCSG, NC-BCFR, OFBCR and RBCS and 211 cases from CNIO-BCS,

see Supplemental Table 1). ER and progesterone receptor (PR) status were provided for a subset of cases, respectively, as were human epidermal growth factor receptor 2 (HER2) status and other histological features including axillary node status and tumor grade, size, and morphology (see Supplemental Table 3). This histopathology information was generally abstracted from medical reports.

Subjects who reported having ethnicity other than white European were excluded, with the exception of those from the three Asian studies, for which only subjects of Asian origin were included. Only subjects from studies that genotyped at least 30 cases of DCIS were included in the analyses of risk of DCIS. All subjects gave written informed consent, where applicable, and each study was approved by the relevant local institutional review boards.

Most studies carried out genotyping using Taqman nuclease assay (Taqman®), with reagents designed by Applied Biosystems as Assays-by-Design™ and genotyping performed using the ABI PRISM 7900HT, 7700 or 7500 Sequence Detection Systems according to manufacturer's instructions. Three studies used Sequenom's MassARRAY system and iPLEX technology (Sequenom, San Diego, CA, USA), with oligonucleotides design carried out according to the guidelines of Sequenom and performed using MassARRAY Assay Design software (version 3.1). The method used by each study is identified in Supplemental Table 2. All studies complied with BCAC genotyping quality control (QC) standards by including at least 2% of samples in duplicate and a common set of 93 CEPH DNAs used by the HapMap Consortium (HAPMAPPT01, Coriell Institute for Medical Research, Cambden, NJ).

Statistical Methods

Departure from Hardy-Weinberg equilibrium (HWE) was tested for in controls from each centre using Pearson's χ^2 test (1df). The association of each SNP with breast cancer risk was assessed by estimating genotype-specific and per-allele ORs using logistic regression, adjusted for study. The exclusion of studies for which the age of controls was not known and the additional adjustment for age (in 5-year categories and as a continuous covariate) made no substantial difference to the results. Between-study heterogeneity in ORs was assessed using a likelihood ratio test (LRT) comparing the model with interaction terms for the per-allele log-OR by study to the model with no interaction terms. Differences in ORs by ethnicity (white European, Asian) and age (<40, 40–49, 50–59, 60–69, 70 years) were evaluated using a similar LRT, the latter modeled as a linear trend by fitting the median age for each of the defined categories.

ORs specific to disease subtypes defined by ER, PR and HER2 status (positive, negative), by combinations of these markers, and by axillary node status (none, 1 affected), tumor grade (1, 2, 3), tumor size (<10, 11–20, >20mm) and tumor morphology (ductal, lobular), were estimated for white Europeans using polytomous logistic regression with control status as the reference outcome. Heterogeneity in the OR by subtypes was tested for by applying polytomous logistic regression to cases-only, treating the number of minor alleles as the outcome and restricting, for each explanatory variable, the beta coefficient for the comparison of 2 to 0 minor alleles to be double that for the comparison of 1 to 0 minor alleles. This is equivalent to modeling a log-additive per-allele OR and allows multiple

tumor markers to be modeled simultaneously. Linear trends were tested for grade by fitting values 1, 2, and 3, and for size by fitting the median value, for the defined categories, respectively. Enrichment of the risk allele in “selected cases” was assessed using the likelihood ratio test comparing polytomous logistic regression models with and without the per-allele OR constrained to be equal for selected and unselected cases, relative to controls (1df). We minimised bias in the estimation of OR by repeating all analyses after excluding “selected cases”. All statistical tests were two-sided. The term “genome-wide” statistically significant is taken to imply $p < 10^{-7}$; otherwise “statistically significant” implies $p < 0.05$. All analyses were carried out using Stata: Release 10 (6).

Meta-analyses were carried out using *metan* command based on log-transformed OR estimates and their 95% CI from different reports.

Results

Minimum genotype concordance of 98% for duplicate samples and 95% for the CEPH samples was observed in all studies, as were minimum genotype calls of 97% for both SNPs in cases and controls. Evidence of departure from HWE was observed for 5p12-rs10941679 in controls from two studies (FBCS [$P = 0.02$] and HABCS [$P = 0.005$], Supplemental Table 2); for both studies, cluster plots were double-checked visually and determined to be of high quality, and all their genotype data were therefore included in the final analysis.

Thirty-four studies successfully genotyped 5p12-rs10941679 in a total of 40,972 invasive breast cancer cases (7,037 of which were selected for increased genetic risk), 1,398 DCIS cases, and 46,334 controls of white European origin. Genotypes were also obtained for this SNP for 3,007 cases and 2,337 controls of Asian origin recruited by three studies. A total of 8,213 invasive cases and 10,340 controls were successfully genotyped for 5p12-rs981782 by seven studies. Previously published data for this SNP were included from 14 studies to obtain genotypes for both SNPs in 5p12 for 23,548 invasive cases and 28,142 controls, all of white European origin. Genotype counts by study are provided in Supplemental Table 4.

The minor G allele of 5p12-rs10941679 was less frequent in white European women (26%) than in Asian women (49%). Estimated ORs, for invasive breast cancer and DCIS, and by ethnicity, are presented in Table 1. The genotype-specific OR estimates were consistent with a log-additive model for white Europeans. The per-G-allele OR estimate was 1.12 (95% CI=1.10–1.15, $P = 6 \times 10^{-24}$), and there was no evidence of heterogeneity in the OR among studies overall ($P = 0.1$) or among studies of Asians ($P = 0.3$), studies of white Europeans that included “selected cases” ($P = 0.3$) or studies of white Europeans without “selected cases” ($P = 0.2$) (Figure 1). This association was maintained at genome-wide statistical significance when SNP 5p12-rs981782 was included as a covariate in the logistic regression model (OR=1.11, 95% CI=1.09–1.15, $P = 6 \times 10^{-13}$). For 5p12-rs10941679 the estimated per-G-allele OR for DCIS was similar to that for invasive disease (OR=1.10, 95% CI=1.01–1.21, $P = 0.03$). The per-G-allele OR for invasive breast cancer did not appear to be different for Asian women (P -heterogeneity=0.3), but the estimate was lower and not statistically significant (1.07; 95% CI=0.99–1.15, $P = 0.09$). All further analyses were based on white European women only.

We observed weak evidence that the G allele of 5p12-rs10941679 was enriched in “selected cases” ($P=0.06$). The estimated per-allele OR was 1.18 (95%CI=1.11–1.24, $P=1\times 10^{-8}$) when comparing cases selected for increased genetic risk to controls from the same studies and 1.11 (95%CI=1.08–1.14, $P=7\times 10^{-18}$) when they were excluded. There was evidence of an increase in the per-allele OR associated with 5p12-rs10941679 with age ($P=0.01$), with estimates of 1.10 (95%CI=1.01–1.19), 1.08 (95%CI=1.02–1.15), 1.14 (95%CI=1.09–1.19), 1.12 (1.07–1.17) and 1.16 (95%CI=1.06–1.26) for white European women aged <40, 40–49, 50–59, 60–69 and 70, respectively. This trend was also observed when age was modeled in years ($P=0.02$), with an estimated interaction OR of 1.02 (95%CI=1.00–1.04) per G allele, per 10-year increase in age. The same trend was observed after excluding “selected cases” (interaction OR=1.02, 95%CI=1.00–1.05, $P=0.03$).

Results from analyses by breast cancer subtypes defined by histopathological features are presented in Tables 2 and 3. The number of cases from each study for which this information was available is presented in Supplemental Table 3. We observed strong evidence that the per-allele OR associated with 5p12-rs10941679 differed by ER status, PR status and ER/PR combined (all $P<10^{-5}$, Table 2). When ER and PR status were modeled together in the case-only analysis only the association with PR status was maintained ($P=6\times 10^{-4}$, compared to $P=0.2$ for ER status). The OR estimates by combined ER/PR status were consistent with the heterogeneity being present by PR rather than ER status. Furthermore, heterogeneity in the OR by PR status was observed when only cases with ER-positive disease were considered ($P=0.005$), but not by ER status when only cases with PR-positive disease were considered ($P=0.8$). After excluding “selected cases”, the per-G-allele OR estimates were 1.03 (95%CI=0.99–1.07, $P=0.2$) for PR-negative disease and 1.16 (95%CI=1.12–1.20, $P=1\times 10^{-18}$) for PR-positive disease. We observed no further heterogeneity by HER2 status ($P=0.3$). The trend of increasing per-allele OR with age remained apparent when only the risk of PR-positive disease was considered (interaction OR=1.03, 95%CI=1.00–1.06, $P=0.05$).

There was also clear evidence that 5p12-rs10941679 was more strongly associated with the risk of lower-grade breast cancer ($P=5\times 10^{-7}$, Table 3). When grade and PR status were modeled together in a case-only analysis, evidence of heterogeneity was observed for both tumor characteristics ($P<0.002$). After excluding “selected cases”, the per-G-allele OR estimates were 1.20 (95%CI=1.14–1.25), OR=1.13 (95%CI=1.09–1.16) and OR=1.04 (95%CI=0.99–1.08) for grade 1, 2 and 3/4 disease, respectively). Further restriction to PR-positive cases gave consistent results [OR=1.22 [95%CI=1.14–1.30], OR=1.15 [95%CI=1.10–1.20] and OR=1.07 [95%CI=1.00–1.15], respectively). We also assessed the association of 5p12-rs10941679 with risk of different grades of DCIS and, while based on limited data (N=210, 102 and 124 cases with grade 1, 2 and 3 DCIS, respectively), the per-allele OR estimates were consistent with the trend observed for invasive disease (OR=1.33 [95%CI=0.88–2.02], OR=1.08 [95%CI=0.78–1.49] and OR=0.98 [95%CI=0.72–1.33] for grade 1, 2 and 3 DCIS, respectively). There was no evidence of heterogeneity in per-G-allele OR by nodal status, tumor size or tumor type.

Further to our previous report of a possible association with breast cancer risk of variant rs981782, also in 5p12, we genotyped this SNP in an additional 8,213 invasive cases and

10,340 controls from studies participating in the BCAC and found no evidence of association (per-G-allele OR=0.97, 95%CI=0.93–1.02, $P=0.2$, Table 4). When we added in data for participants genotyped as part of the previous study (2), to give a combined total of 23,548 invasive cases and 28,142 controls of white European origin, the per-G-allele OR estimate was 0.95 (95%CI=0.92–0.97, $P=2\times 10^{-5}$). The MAF for controls genotyped using Taqman, iPlex and the GWAS SNP-array (2) were similar (0.47, 0.45 and 0.50, respectively). This possible association was attenuated, but still nominally statistically significant, when 5p12-rs10941679 was also included in the logistic regression model (OR=0.97, 95%CI=0.94–0.99, $P=0.007$). The per-allele OR estimate for each SNP adjusted for the other was unchanged after excluding “selected cases”. The estimated linkage disequilibrium coefficient (r^2) between the two SNPs was 0.04, both across all white European subjects genotyped ($N=52,506$) and for controls only ($N=28,142$).

Discussion

This analysis of 40,972 invasive cases and 46,334 controls from 37 studies definitively confirms the 5p12-rs10941679 as a breast cancer susceptibility locus in white European women. After excluding cases selected for increased genetic susceptibility, we estimated an OR for white European women of 1.11 per G allele (95%CI=1.08–1.14). This association was independent of a possible association with 5p12-rs981782 and appeared to be stronger for older women.

The estimated OR for Asian women (3,007 cases and 2,337 controls) was consistent with that for European women, but the confidence limits were too wide to demonstrate a definite association (OR=1.07, 95%CI=0.99–1.15). The Shanghai Breast Cancer Study also estimated a per-G-allele OR of 1.07 (95%CI=0.99–1.15) using data from 2,950 cases and 2,986 controls (7). Assessed together, these two large studies suggest that 5p12-rs10941679 is associated with increased breast cancer risk in Asian women (meta-analysis OR=1.07 (95%CI=1.02–1.13, $P=0.01$), although even larger studies will be required to precisely estimate the corresponding OR.

The initial paper on 5p12-rs10941679, based on 6,145 cases and 33,016 controls, reported a larger per-G-allele OR for breast cancer in general of 1.19 (95%CI=1.11–1.28) (5). This larger initial estimate probably reflects “winner’s curse” whereby the estimated OR in the study discovering the association tends to be overestimated (8). Stacey et al. (5) also reported that the increased risk associated with 5p12-rs10941679 was limited to ER-positive disease and consistent results have been observed in other studies (7, 9). We replicated this finding, but our larger sample size enabled us to carry out a more detailed analysis that revealed a stronger association with PR-positive disease (per-G-allele OR=1.16, 95%CI=1.12–1.20). The heterogeneity in the OR by ER status appeared to be driven by the correlation between ER and PR status. Furthermore, we have established that the association is also stronger for lower grade tumors in general, and lower grade PR-positive tumors specifically. Stacey et al. (5), also noted a possibly larger OR for lower grade tumors, but not after ER-status was taken into account. This multifaceted heterogeneity has also been observed for 10q26-rs2981582, which appears to be associated with increased risk of lower grade ER-positive tumors (10).

In our previous GWAS, we observed that the minor G allele of 5p12-rs981782 in the same region was associated with reduced risk of breast cancer. The estimated per-G-allele OR for white European women was 0.94 (95%CI=0.91–0.97, $P=9\times 10^{-6}$), based on data from 20,649 cases of invasive breast cancer and 22,578 controls from 19 studies participating in the replication stages (2 and 3) (2). We did not replicate this association in the present study of 8,247 cases and 10,363 controls, but our result (OR=0.97, 95%CI=0.93–1.02, $P=0.2$) was not inconsistent with the existence of a modest protective effect. Reeves et al. (11) studied 5p12-rs981782 in 10,306 cases and 10,309 controls from the Million Women Study and estimated a per-G-allele OR of 0.94 (95%CI=0.91–0.98, $P=0.003$). Stacey et al. (5) also evaluated this SNP in 5,028 cases and 32,090 controls and reported the corresponding OR to be 0.96 (95%CI=0.92–1.01, $P=0.1$). The Cancer Genetic Markers of Susceptibility (CGEMS) project studied SNP 5p12-rs4866929, which is in high linkage disequilibrium with 5p12-rs981782 ($r^2=0.96$), in a total of 5,692 cases and 5,576 controls and obtained an OR estimate of 0.97 (95%CI=0.91–1.03, $P=0.4$) (3, 9). When combined together in a meta-analysis, these results for 5p12-rs981782 (excluding those from CGEMS) give an estimated OR of 0.95, 95%CI=0.93–0.97, $P=5\times 10^{-8}$, suggesting that this SNP is associated with a modest reduction in breast cancer risk.

It is less clear whether the association between 5p12-rs981782 and breast cancer risk is independent of the effect marked by 5p12-rs10941679. In contrast to the findings of Stacey et al. (5), in the present study (23,548 cases and 28,142 controls) the evidence of association for 5p12-rs981782 remained after adjusting for 5p12-rs10941679 ($P=0.007$), although the effect was slightly attenuated. Furthermore, the correlation between these two SNPs was too weak ($r^2=0.04$, $N=52,506$) for the association seen with 5p12-rs981782 to be due to confounding by 5p12-rs10941679. However, it remains possible that these two associations are driven by a variant correlated with both SNPs. Stacey et al. (5) found that a third SNP nearby, 5p12-rs4415084 was associated with breast cancer risk, independently of 5p12-rs10941679 ($r^2=0.51$ between the two SNPs). Multi-centre fine-mapping studies of this region are needed to obtain the sample sizes required to identify the causal variant or variants behind these multiple small signals.

As discussed by Stacey et al. (5), 5p12-rs10941679 resides between two genes: *MRPS30*, which encodes the smaller 28S subunit of the mitochondrial ribosome and has pro-apoptotic properties (12), and *FGF10*, a fibroblast growth factor which is amplified in around 10% of breast cancers (13), although there is a recombination hotspot separating this SNP from *FGF10*. Therefore, as for the vast majority of low-penetrance breast cancer susceptibility loci identified by GWAS, an implicated gene or functional mechanism remains to be elucidated.

A potential limitation of our study was that information on tumor histopathology was not available for all cases and, where it was, these data were predominantly abstracted from medical records, rather than being obtained through a standardised pathology review. Thus some misclassification may have been present and this may have attenuated some associations with tumor phenotype. However the strong association with PR and ER status suggests that this effect was minimal, at least for the features examined here.

In conclusion, 5p12-rs10941679 is a confirmed marker of breast cancer susceptibility. In white European women it is associated with increased risk of lower-grade, PR-positive tumors. These findings suggest that commonly collected tumor characteristics other than ER status can be used to identify subtypes of breast cancer with distinct etiology, and that large collaborative studies are required to identify the genetic contributions specific to each of these.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Authors

Roger L. Milne^{1,3,*}, Ellen L. Goode^{5,*}, Montserrat García-Closas⁷, Fergus J. Couch⁶, Gianluca Severi^{8,3}, Rebecca Hein⁹, Zachary Fredericksen⁵, Núria Malats¹, M. Pilar Zamora¹³, Jose Ignacio Arias Pérez¹⁴, Javier Benítez², Thilo Dörk¹⁵, Peter Schürmann¹⁵, Johann H. Karstens¹⁶, Peter Hillemanns¹⁵, Angela Cox¹⁷, Ian W. Brock¹⁷, Graeme Elliot^{17,19}, Simon S. Cross¹⁸, Sheila Seal²⁰, Clare Turnbull²⁰, Anthony Renwick²⁰, Nazneen Rahman²⁰, Chen-Yang Shen²¹, Jyh-Cherng Yu²², Chiun-Sheng Huang²³, Ming-Feng Hou²⁴, Børge G. Nordestgaard²⁵, Stig E. Bojesen²⁵, Charlotte Lanng²⁶, Grethe Grenaker Alnæs²⁷, Vessela Kristensen^{27,28}, Anne-Lise Børrensen-Dale^{27,28}, John L. Hopper³, Gillian S. Dite³, Carmel Apicella³, Melissa C. Southey⁴, Diether Lambrechts²⁹, Betül T. Yesilyurt²⁹, Giuseppe Floris³⁰, Karin Leunen³⁰, Suleeporn Sangrajang³¹, Valerie Gaborieau³², Paul Brennan³², James McKay³², Jenny Chang-Claude^{9,33}, Shan Wang-Gohrke³³, Paolo Radice^{34,36}, Paolo Peterlongo^{34,36}, Siranoush Manoukian³⁵, Monica Barile³⁷, Graham G. Giles^{8,3}, Laura Baglietto^{8,3}, Esther M. John³⁸, Alexander Miron³⁹, Stephen J. Chanock⁴⁰, Jolanta Lissowska⁴¹, Mark E. Sherman⁴⁰, Jonine D. Figueroa⁴⁰, Natalia V. Bogdanova^{15,16}, Natalia N. Antonenkova⁴², Iosif V. Zalutsky⁴², Yuri I. Rogov⁴², Peter A. Fasching^{43,44}, Christian M. Bayer⁴³, Arif B. Ekici⁴⁵, Matthias W. Beckmann⁴³, Hermann Brenner¹⁰, Heiko Müller¹⁰, Volker Arndt¹⁰, Christa Stegmaier⁴⁶, Irene L. Andrulis^{47,48,50}, Julia A. Knight^{49,51}, Gord Glendon⁴⁷, Anna Marie Mulligan^{52,53}, Arto Mannermaa^{54,55,56}, Vesa Kataja^{54,55,57}, Veli-Matti Kosma^{54,55,56}, Jaana M. Hartikainen^{54,55,56}, Alfons Meindl⁵⁸, Joerg Heil⁵⁹, Claus R. Bartram⁶⁰, Rita K. Schmutzler⁶², Gilles D. Thomas⁴⁰, Robert N. Hoover⁴⁰, Olivia Fletcher⁶³, Lorna J. Gibson⁶⁴, Isabel dos Santos Silva⁶⁴, Julian Peto⁶⁴, Stefan Nickels⁹, Dieter Flesch-Janys⁶⁵, Hoda Anton-Culver⁶⁶, Argyrios Ziogas⁶⁶, Elinor Sawyer⁶⁷, Ian Tomlinson⁶⁸, Michael Kerin⁶⁹, Nicola Miller⁶⁹, Marjanka K. Schmidt⁷⁰, Annegien Broeks⁷⁰, Laura J. Van 't Veer⁷⁰, Rob A.E.M. Tollenaar⁷¹, Paul D.P. Pharoah^{72,73,74}, Alison M. Dunning^{72,74}, Karen A. Pooley^{73,74}, Frederik Marme^{59,61}, Andreas Schneeweiss^{59,61}, Christof Sohn⁵⁹, Barbara Burwinkel^{11,59}, Anna Jakubowska⁷⁵, Jan Lubinski⁷⁵, Katarzyna Jaworska^{75,76}, Katarzyna Durda⁷⁵, Daehee Kang⁷⁷, Keun-Young Yoo⁷⁷, Dong-Young Noh⁷⁷, Sei-Hyun Ahn⁷⁸, David J. Hunter^{79,80}, Susan E. Hankinson^{80,81}, Peter Kraft^{79,80}, Sara Lindstrom^{79,80}, Xiaoqing Chen⁸², Jonathan Beesley⁸², Ute Hamann¹², Volker Harth⁸³, Christina Justenhoven^{84,85}, Robert Winqvist^{86,87}, Katri

Pyhkäs^{86,87}, Arja Jukkola-Vuorinen⁸⁸, Mervi Grip⁸⁹, Maartje Hooning⁹⁰, Antoinette Hollestelle⁹¹, Rogier A. Oldenburg⁹², Madeleine Tilanus-Linthorst⁹³, Elza Khusnutdinova⁹⁴, Marina Bermisheva⁹⁴, Darya Prokofieva⁹⁴, Albina Farahtdinova⁹⁴, Janet E. Olson⁵, Xianshu Wang⁶, Manjeet K. Humphreys⁷⁴, Qin Wang⁷⁴, Georgia Chenevix-Trench⁸², and Douglas F. Easton^{73,74} **for the GENICA Network for the kConFab Investigators and the AOCs Group**

Affiliations

¹Genetic and Molecular Epidemiology Group, Spanish National Cancer Research Centre (CNIO), Madrid, Spain ²Human Genetics Group, Spanish National Cancer Research Centre (CNIO), Madrid, Spain ³Centre for Molecular, Environmental, Genetic, and Analytic Epidemiology, The University of Melbourne, Australia ⁴Department of Pathology, The University of Melbourne, Australia ⁵Department of Health Sciences Research, Mayo Clinic, Rochester, MN, USA ⁶Department of Laboratory Medicine and Pathology, Mayo Clinic, Rochester, MN, USA ⁷Sections of Epidemiology and Genetics, Institute of Cancer Research and Breakthrough Breast Cancer Research Centre, London, UK ⁸Cancer Epidemiology Centre, The Cancer Council Victoria, Melbourne, Australia ⁹Division of Cancer Epidemiology, German Cancer Research Center (DKFZ), Heidelberg, Germany ¹⁰Division of Clinical Epidemiology and Aging Research, German Cancer Research Center (DKFZ), Heidelberg, Germany ¹¹Molecular Epidemiology Unit, German Cancer Research Center (DKFZ), Heidelberg, Germany ¹²Molecular Genetics of Breast Cancer, German Cancer Research Center (DKFZ), Heidelberg, Germany ¹³Servicio de Oncología Médica, Hospital Universitario La Paz, Madrid, Spain ¹⁴Servicio de Cirugía General y Especialidades, Hospital Monte Naranco, Oviedo, Spain ¹⁵Department of Obstetrics and Gynaecology, Hannover Medical School, Hannover, Germany ¹⁶Department of Radiation Oncology, Hannover Medical School, Hannover, Germany ¹⁷Institute for Cancer Studies, Department of Oncology, University of Sheffield, UK ¹⁸Academic Unit of Pathology, Department of Neuroscience, University of Sheffield, UK ¹⁹University of Manchester, Manchester, UK ²⁰Section of Cancer Genetics, Institute of Cancer Research, Sutton, Surrey, UK ²¹Institute of Biomedical Sciences, Academia Sinica, and Taiwan Biobank, Taipei, Taiwan ²²Department of Surgery, Tri-Service General Hospital, Taipei, Taiwan ²³Department of Surgery, National Taiwan University Hospital, Taipei, Taiwan ²⁴Cancer Center and Department of Surgery, Kaohsiung Medical University Chung-Ho Memorial Hospital, Kaohsiung, Taiwan ²⁵Copenhagen General Population Study and Department of Clinical Biochemistry, Copenhagen University Hospital, Copenhagen, Denmark ²⁶Department of Breast Surgery, Herlev Hospital, Copenhagen University Hospital, Copenhagen, Denmark ²⁷Department of Genetics, Institute for Cancer Research, Oslo University Hospital, Radiumhospitalet, Oslo, Norway ²⁸Faculty of Medicine (Faculty Division Ahus), UiO, Norway ²⁹Vesalius Research Center (VRC), VIB, Leuven, Belgium ³⁰Multidisciplinary Breast Center, University Hospital Gasthuisberg, Leuven, Belgium ³¹National Cancer Institute, Bangkok, Thailand ³²International Agency for Research on Cancer, Lyon, France ³³Department of Obstetrics and Gynecology, University of Ulm, Ulm, Germany

³⁴Unit of Molecular Bases of Genetic Risk and Genetic Testing, Department of Preventive and Predictive Medicine, Fondazione IRCCS Istituto Nazionale Tumori (INT), Milan, Italy ³⁵Unit of Medical Genetics, Department of Preventive and Predictive Medicine, Fondazione IRCCS Istituto Nazionale Tumori (INT), Milan, Italy ³⁶IFOM, Fondazione Istituto FIRC di Oncologia Molecolare, Milan, Italy ³⁷Division of Cancer Prevention and Genetics, Istituto Europeo di Oncologia (IEO), Milan, Italy ³⁸Cancer Prevention Institute of California, Fremont and Stanford University School of Medicine, Stanford, California, USA ³⁹Dana-Farber Cancer Institute, Boston, Massachusetts, USA ⁴⁰Division of Cancer Epidemiology and Genetics, National Cancer Institute, Rockville, MD, USA ⁴¹Department of Cancer Epidemiology and Prevention, M. Sklodowska-Curie Memorial Cancer Center & Institute of Oncology, Warsaw, Poland ⁴²N.N. Alexandrov Research Institute of Oncology and Medical Radiology, Minsk, Belarus ⁴³University Breast Center Franconia, Department of Gynecology and Obstetrics, University Hospital Erlangen, Comprehensive Cancer Center Erlangen-Nuremberg, Erlangen, Germany ⁴⁴David Geffen School of Medicine, Department of Medicine Division of Hematology and Oncology, University of California at Los Angeles, CA, USA ⁴⁵Institute of Human Genetics, Friedrich Alexander University Erlangen-Nuremberg, Erlangen, Germany ⁴⁶Saarland Cancer Registry, Saarbrücken, Germany ⁴⁷Ontario Cancer Genetics Network, Cancer Care Ontario, Toronto, ON, Canada ⁴⁸Fred A. Litwin Center for Cancer Genetics, Mount Sinai Hospital, Toronto, ON, Canada ⁴⁹Samuel Lunenfeld Research Institute, Mount Sinai Hospital, Toronto, ON, Canada ⁵⁰Department of Molecular Genetics, University of Toronto, Toronto, ON, Canada ⁵¹Division of Epidemiology, Dalla Lana School of Public Health, University of Toronto, Toronto, ON, Canada ⁵²Department of Laboratory Medicine and Pathobiology, University of Toronto, Toronto, ON, Canada ⁵³Department of Laboratory Medicine and Keenan Research Centre of the Li Ka Shing Knowledge Institute, St Michael's Hospital, Toronto, ON, Canada ⁵⁴School of Medicine, Institute of Clinical Medicine, Pathology and Forensic Medicine, University of Eastern Finland, Kuopio, Finland ⁵⁵Biocenter Kuopio, Kuopio, Finland ⁵⁶Department of Clinical Pathology, Kuopio University Hospital, Kuopio, Finland ⁵⁷Department of Oncology, Kuopio University Hospital, Kuopio, Finland ⁵⁸Division of Gynecology and Obstetrics, Klinikum rechts der Isar at the Technical University Munich, Munich, Germany ⁵⁹Department of Obstetrics and Gynecology, University of Heidelberg, Heidelberg, Germany ⁶⁰Institute of Human Genetics, University of Heidelberg, Heidelberg, Germany ⁶¹National Center for Tumor Diseases, University of Heidelberg, Heidelberg, Germany ⁶²Division of Molecular Gyneco-Oncology, Department of Gynaecology and Obstetrics, Center of Molecular Medicine Cologne (CMMC), University Hospital of Cologne, Cologne, Germany ⁶³Breakthrough Breast Cancer Research Centre, The Institute of Cancer Research, London, UK ⁶⁴London School of Hygiene and Tropical Medicine, London, UK ⁶⁵Department of Cancer Epidemiology/Clinical Cancer Registry and Institute for Medical Biometrics and Epidemiology, University Clinic Hamburg-Eppendorf, Hamburg, Germany ⁶⁶Department of Epidemiology, University of California Irvine, Irvine, California, USA ⁶⁷Division of Cancer Studies, NIHR Comprehensive

Biomedical Research Centre, Guy's & St. Thomas' NHS Foundation Trust in partnership with King's College London, London, UK ⁶⁸Wellcome Trust Centre for Human Genetics and Oxford Biomedical Research Centre, University of Oxford, UK ⁶⁹Clinical Science Institute, University Hospital Galway, Galway, Ireland ⁷⁰Netherlands Cancer Institute, Antoni van Leeuwenhoek hospital, Amsterdam, The Netherlands ⁷¹Leiden University Medical Center, Leiden, The Netherlands ⁷²Department of Oncology, University of Cambridge, Cambridge, UK ⁷³Department of Public Health and Primary Care, University of Cambridge, Cambridge, UK ⁷⁴Centre for Cancer Genetic Epidemiology, University of Cambridge, Cambridge, UK ⁷⁵Department of Genetics and Pathology, Pomeranian Medical University, Szczecin, Poland ⁷⁶Postgraduate School of Molecular Medicine, Warsaw Medical University, Warsaw, Poland ⁷⁷Seoul National University College of Medicine, Seoul, Korea ⁷⁸Ulsan University College of Medicine, Seoul, Korea ⁷⁹Program in Molecular and Genetic Epidemiology, Harvard School of Public Health, Boston, MA, USA ⁸⁰Department of Epidemiology, Harvard School of Public Health, Boston, MA, USA ⁸¹Channing Laboratory, Department of Medicine, Brigham and Women's Hospital and Harvard Medical School, Boston, MA, USA ⁸²Queensland Institute of Medical Research, Brisbane, Australia ⁸³Institute for Prevention and Occupational Medicine of the German Social Accident Insurance (IPA), Bochum, Germany ⁸⁴Dr. Margarete Fischer-Bosch-Institute of Clinical Pharmacology, Stuttgart, Germany ⁸⁵University of Tübingen, Tübingen, Germany ⁸⁶Laboratory of Cancer Genetics, Department of Clinical Genetics, Oulu University Hospital, University of Oulu, Oulu, Finland ⁸⁷Biocenter Oulu, Oulu University Hospital, University of Oulu, Oulu, Finland ⁸⁸Department of Oncology, Oulu University Hospital, University of Oulu, Oulu, Finland ⁸⁹Department of Surgery, Oulu University Hospital, University of Oulu, Oulu, Finland ⁹⁰Department of Medical Oncology, Family Cancer Clinic, Erasmus University Medical Center, Rotterdam, The Netherlands ⁹¹Department of Medical Oncology, Josephine Nefkens Institute, Erasmus University Medical Center, Rotterdam, The Netherlands ⁹²Department of Clinical Genetics, Family Cancer Clinic, Erasmus University Medical Center, Rotterdam, The Netherlands ⁹³Department of Surgical Oncology, Family Cancer Clinic, Erasmus University Medical Center, Rotterdam, The Netherlands ⁹⁴Institute of Biochemistry and Genetics, Ufa Scientific Center of Russian Academy of Sciences, Ufa, Russia

Acknowledgments

We thank all the individuals who took part in these studies and all the researchers, clinicians, technicians and administrative staff who have enabled this work to be carried out. In particular, we thank: Charo Alonso, Tais Moreno, Guillermo Pita, Primitiva Menendez, Anna González-Neira, Michael Bremer, Sue Higham, Helen Cramp, Dan Connley, The Wellcome Trust Case Control Consortium (see the WTCCC website for a full list of contributing investigators), Maggie Angelakos, Judi Maskiell, Gillian Dite, Gilian Peuteman, Dominiek Smeets, Thomas Van Brussel, Kathleen Corthouts, Ursula Eilber, Tanya Koehler, Marco Pierotti, Bernard Peissel, Daniela Zaffaroni, Bernardo Bonanni, Loris Bernard, the personnel of the Cancer Genetics Testing laboratory at the IFOM-IEO campus, Louise Brinton, Neonila Szeszenia-Dabrowska, Beata Peplonska, Witold Zatonski, Pei Chao, Michael Stagner, Hartwig Ziegler, Sonja Wolf, Volker Hermann, Teresa Selander, Nayana Weerasooriya, Eija Myöhänen, Helena Kemiläinen, Bernd Frank, Eileen Williams, Elaine Ryder-Mills, Kara Sargus, Tracy Slanger, Elke Mutschelknauss, Ramona Salazar, S. Behrens, R. Birr, W. Busch, U. Eilber, B. Kaspereit, N. Knese, K. Smit, Irene Masunaka, Niall McInerney, Gabrielle Colleran, Andrew Rowan, Angela Jones, Richard van Hien, Sten

Cornelissen, Linde Braaf, Flora van Leeuwen (NKI-AVL), Bas Bueno-de-Mesquita (RIVM; for the release of control samples), the SEARCH and EPIC teams, Heather Thorne, Eveline Niedermayr, the AOCS Management Group (D Bowtell, G Chenevix-Trench, A deFazio, D Gertig, A Green, P Webb), the ACS Management Group (A Green, P Parsons, N Hayward, P Webb, D Whiteman), The GENICA network: Dr. Margarete Fischer-Bosch-Institute of Clinical Pharmacology, Stuttgart, and University of Tübingen, Germany; [CJ, Hiltrud Brauch], Department of Internal Medicine, Evangelische Kliniken Bonn gGmbH, Johanniter Krankenhaus, Bonn, Germany [Yon-Dschun Ko, Christian Baisch], Institute of Pathology, University of Bonn, Bonn, Germany [Hand-Peter Fischer], Molecular Genetics of Breast Cancer, Deutsches Krebsforschungszentrum (DKFZ), Heidelberg, Germany [UH] and Institute for Prevention and Occupational Medicine of the German Social Accident Insurance (IPA), Bochum, Germany [Thomas Bruening, Beate Pesch, Sylvia Rabstein, Anne Spickenheuer, VH], Meeri Otsukka, Kari Mononen, Petra Bos, Jannet Blom, Ellen Crepin, Elisabeth Huijskens, Annette Heemskerk and the Erasmus MC Family Cancer Clinic.

Grant support

Part of this work was supported by the European Community's Seventh Framework Programme under grant agreement number 223175 (grant number HEALTH-F2-2009-223175) (COGS). The **BCAC** is funded by CR-UK (C1287/A10118 and C1287/A12014). Meetings of the BCAC have been funded by the European Union COST programme (BM0606). D.F.E. is a Principal Research Fellow of CR-UK. The **ABCS** was supported by the Dutch Cancer Society [grants NKI 2001-2423; 2007-3839] and the Dutch National Genomics Initiative. The **ABCFS**, **NC-BCFR** and **OFBCR** work was supported by the United States National Cancer Institute, National Institutes of Health (NIH) under RFA-CA-06-503 and through cooperative agreements with members of the Breast Cancer Family Registry (BCFR) and Principal Investigators, including Cancer Care Ontario (U01 CA69467), Northern California Cancer Center (U01 CA69417), University of Melbourne (U01 CA69638). Samples from the NC-BCFR were processed and distributed by the Coriell Institute for Medical Research. The content of this manuscript does not necessarily reflect the views or policies of the National Cancer Institute or any of the collaborating centers in the BCFR, nor does mention of trade names, commercial products, or organizations imply endorsement by the US Government or the BCFR. The **ABCFS** was also supported by the National Health and Medical Research Council of Australia, the New South Wales Cancer Council, the Victorian Health Promotion Foundation (Australia) and the Victorian Breast Cancer Research Consortium. J.L.H. is a National Health and Medical Research Council (NHMRC) Australia Fellow and a Victorian Breast Cancer Research Consortium Group Leader. M.C.S. is a NHMRC Senior Research Fellow and a Victorian Breast Cancer Research Consortium Group Leader. Financial support for the **AOCS** was provided by the United States Army Medical Research and Materiel Command [DAMD17-01-1-0729], the Cancer Council of Tasmania and Cancer Foundation of Western Australia and the NHMRC [199600]. G.C.T. and P.W. are supported by the NHMRC. The work of the **BBCC** was partly funded by ELAN-Fond of the University Hospital of Erlangen. The **BBCS** is funded by Cancer Research UK and Breakthrough Breast Cancer and acknowledges NHS funding to the NIHR Biomedical Research Centre, and the National Cancer Research Network (NCRN). The **BSUCH** study was supported by the Dietmar-Hopp Foundation, the Helmholtz Society and the German Cancer Research Center (DKFZ). The **CGPS** was supported by the Chief Physician Johan Boserup and Lise Boserup Fund, the Danish Medical Research Council and Herlev Hospital. The **CNIO-BCS** was supported by the Genome Spain Foundation, the Red Temática de Investigación Cooperativa en Cáncer and grants from the Asociación Española Contra el Cáncer and the Fondo de Investigación Sanitario (PI081583 and PI081120). The **ESTHER** study was supported by a grant from the Baden Württemberg Ministry of Science, Research and Arts. Additional cases were recruited in the context of the VERDI study, which was supported by a grant from the German Cancer Aid (Deutsche Krebshilfe). The **FBCS** is supported by funds from Cancer Research UK (C8620/A8372 and C8620/A8857), a US Military Acquisition (ACQ) Activity, Era of Hope Award (W81XWH-05-1-0204) and the Institute of Cancer Research (UK). C.T. is funded by a Medical Research Council (UK) Clinical Research Fellowship. The **FBCS** acknowledges NHS funding to the Royal Marsden / Institute of Cancer Research NIHR Specialist Cancer Biomedical Research Centre. The **GC-HBOC** was supported by Deutsche Krebshilfe [107054], the Dietmar-Hopp Foundation, the Helmholtz society and the German Cancer Research Centre (DKFZ). The **GENICA** was funded by the Federal Ministry of Education and Research (BMBF) Germany grants 01KW9975/5, 01KW9976/8, 01KW9977/0 and 01KW0114, the Robert Bosch Foundation, Stuttgart, Deutsches Krebsforschungszentrum (DKFZ), Heidelberg, Institute for Prevention and Occupational Medicine of the German Social Accident Insurance (IPA), Bochum, as well as the Department of Internal Medicine, Evangelische Kliniken Bonn gGmbH, Johanniter Krankenhaus, Bonn, Germany. The **GESBC** was supported by the Deutsche Krebshilfe e. V. [70492] and genotyping in part by the state of Baden-Württemberg through the Medical Faculty of the University of Ulm [P.685]. The **HABCS** study was supported by an intramural grant from Hannover Medical School. The **HMBCS** was supported by short-term fellowships from the German Academic Exchange Program [to N.B.], and the Friends of Hannover Medical School [to N.B.]. The **HUBCS** was supported by a grant from the German Federal Ministry of Research and Education (RUS08/017). The **KBCP** was financially supported by the special Government Funding (EVO) of Kuopio University Hospital grants, Cancer Fund of North Savo, the Finnish Cancer Organizations, The Academy of Finland and by the strategic funding of the University of Eastern Finland. **kConFab** is supported by grants from the National Breast Cancer Foundation, the NHMRC, the Queensland Cancer Fund, the Cancer Councils of New South Wales, Victoria, Tasmania and South Australia and the Cancer Foundation of Western Australia. The kConFab Clinical Follow Up Study was funded by the NHMRC [145684, 288704, 454508]. **LMBC** is supported by the 'Stichting tegen Kanker' (232-2008 and

196-2010). The **MARIE** study was supported by the Deutsche Krebshilfe e.V. [70-2892-BR I], the Hamburg Cancer Society, the German Cancer Research Center and the genotype work in part by the Federal Ministry of Education and Research (BMBF) Germany [01KH0402]. **MBCSG** was supported by grants from Ministero della Salute (Extraordinary National Cancer Program 2006 “Alleanza contro il Cancro”, and “Progetto Tumori Femminili” to PR), Ministero dell’Università e Ricerca (RBLAO3-BETH to PR), Fondazione Italiana per la Ricerca sul Cancro (Special Project “Hereditary tumors”), Associazione Italiana per la Ricerca sul Cancro (4017 to PP) and by funds from Italian citizens who allocated the 5/1000 share of their tax payment in support of the Fondazione IRCCS Istituto Nazionale Tumori, according to Italian laws (INT-Institutional strategic projects “5×1000”). The **MCBCS** was supported by the NIH grants [CA122340, CA128978] and a Specialized Program of Research Excellence (SPORE) in Breast Cancer [CA116201]. **MCCS** cohort recruitment was funded by VicHealth and Cancer Council Victoria. The MCCS was further supported by Australian NHMRC grants 209057, 251553 and 504711 and by infrastructure provided by Cancer Council Victoria. SEE ABCFS. The **NBCS** was supported by grants from the Norwegian Research Council, 155218/V40, 175240/S10 to ALBD, FUGE-NFR 181600/V11 to VNK and a Swizz Bridge Award to ALBD. The **NHS** was funded by NIH grant CA87969. The **OBCS** was supported by research grants from the Finnish Cancer Foundation, the Sigrid Juselius Foundation, the Academy of Finland, the University of Oulu, and the Oulu University Hospital. The **PBCS** was funded by Intramural Research Funds of the National Cancer Institute, Department of Health and Human Services, USA. The **RBCS** was funded by the Dutch Cancer Society (DDHK 2004-3124, DDHK 2009-4318). The **SBCS** was supported by Yorkshire Cancer Research and the Breast Cancer Campaign. **SEARCH** is funded by programme grants from Cancer Research UK [C490/A10124, C8197/A10123, C1287/A10118]. The **SEBCS** was supported by the Korea Health 21 R&D Project [AO30001], Ministry of Health and Welfare, Republic of Korea. The **SZBCS** was supported by Grant PBZ_KBN_122/P05/2004; Katarzyna Jaworska is a fellow of International PhD program, Postgraduate School of Molecular Medicine, Warsaw Medical University, supported by the Polish Foundation of Science. The **TBCS** was funded by The National Cancer Institute Thailand. The **TWBCS** is supported by the Taiwan Biobank project of the Institute of Biomedical Sciences, Academia Sinica, Taiwan. The **UCIBCS** component of this research was supported by the NIH [CA58860, CA92044] and the Lon V Smith Foundation [LVS39420]. ES is supported by NIHR Comprehensive Biomedical Research Centre, Guy’s & St. Thomas’ NHS Foundation Trust in partnership with King’s College London, United Kingdom. IT is supported by the Oxford Biomedical Research Centre.

References

- Ahmed S, Thomas G, Ghossaini M, Healey CS, Humphreys MK, Platte R, et al. Newly discovered breast cancer susceptibility loci on 3p24 and 17q23. *Nat Genet.* 2009; 41:585–90. [PubMed: 19330027]
- Easton DF, Pooley KA, Dunning AM, Pharoah PD, Thompson D, Ballinger DG, et al. Genome-wide association study identifies novel breast cancer susceptibility loci. *Nature.* 2007; 447:1087–93. [PubMed: 17529967]
- Hunter DJ, Kraft P, Jacobs KB, Cox DG, Yeager M, Hankinson SE, et al. A genome-wide association study identifies alleles in FGFR2 associated with risk of sporadic postmenopausal breast cancer. *Nat Genet.* 2007; 39:870–4. [PubMed: 17529973]
- Stacey SN, Manolescu A, Sulem P, Rafnar T, Gudmundsson J, Gudjonsson SA, et al. Common variants on chromosomes 2q35 and 16q12 confer susceptibility to estrogen receptor-positive breast cancer. *Nat Genet.* 2007; 39:865–9. [PubMed: 17529974]
- Stacey SN, Manolescu A, Sulem P, Thorlacius S, Gudjonsson SA, Jonsson GF, et al. Common variants on chromosome 5p12 confer susceptibility to estrogen receptor-positive breast cancer. *Nat Genet.* 2008; 40:703–6. [PubMed: 18438407]
- StataCorp. Stata Statistical Software: Release 10.0. College Station, Texas (USA): Stata Corporation LP; 2007.
- Long J, Shu XO, Cai Q, Gao YT, Zheng Y, Li G, et al. Evaluation of breast cancer susceptibility loci in Chinese women. *Cancer Epidemiol Biomarkers Prev.* 2010; 19:2357–65. [PubMed: 20699374]
- Kraft P. Curses--winner’s and otherwise--in genetic epidemiology. *Epidemiology.* 2008; 19:649–51. discussion 657–8. [PubMed: 18703928]
- Thomas G, Jacobs KB, Kraft P, Yeager M, Wacholder S, Cox DG, et al. A multistage genome-wide association study in breast cancer identifies two new risk alleles at 1p11.2 and 14q24. 1 (RAD51L1). *Nat Genet.* 2009; 41:579–84. [PubMed: 19330030]
- Broeks A, Schmidt MK, Sherman ME, Couch FJ, Hopper JL, Dite GS, et al. Low penetrance breast cancer susceptibility loci are associated with specific breast tumor subtypes: Findings from the Breast Cancer Association Consortium. *Human Molecular Genetics.* in press.

11. Reeves GK, Travis RC, Green J, Bull D, Tipper S, Baker K, et al. Incidence of breast cancer and its subtypes in relation to individual and multiple low-penetrance genetic susceptibility loci. *JAMA*. 2010; 304:426–34. [PubMed: 20664043]
12. Cavdar Koc E, Burkhart W, Blackburn K, Moseley A, Spremulli LL. The small subunit of the mammalian mitochondrial ribosome. Identification of the full complement of ribosomal proteins present. *J Biol Chem*. 2001; 276:19363–74. [PubMed: 11279123]
13. Theodorou V, Boer M, Weigelt B, Jonkers J, van der Valk M, Hilkens J. Fgf10 is an oncogene activated by MMTV insertional mutagenesis in mouse mammary tumors and overexpressed in a subset of human breast carcinomas. *Oncogene*. 2004; 23:6047–55. [PubMed: 15208658]

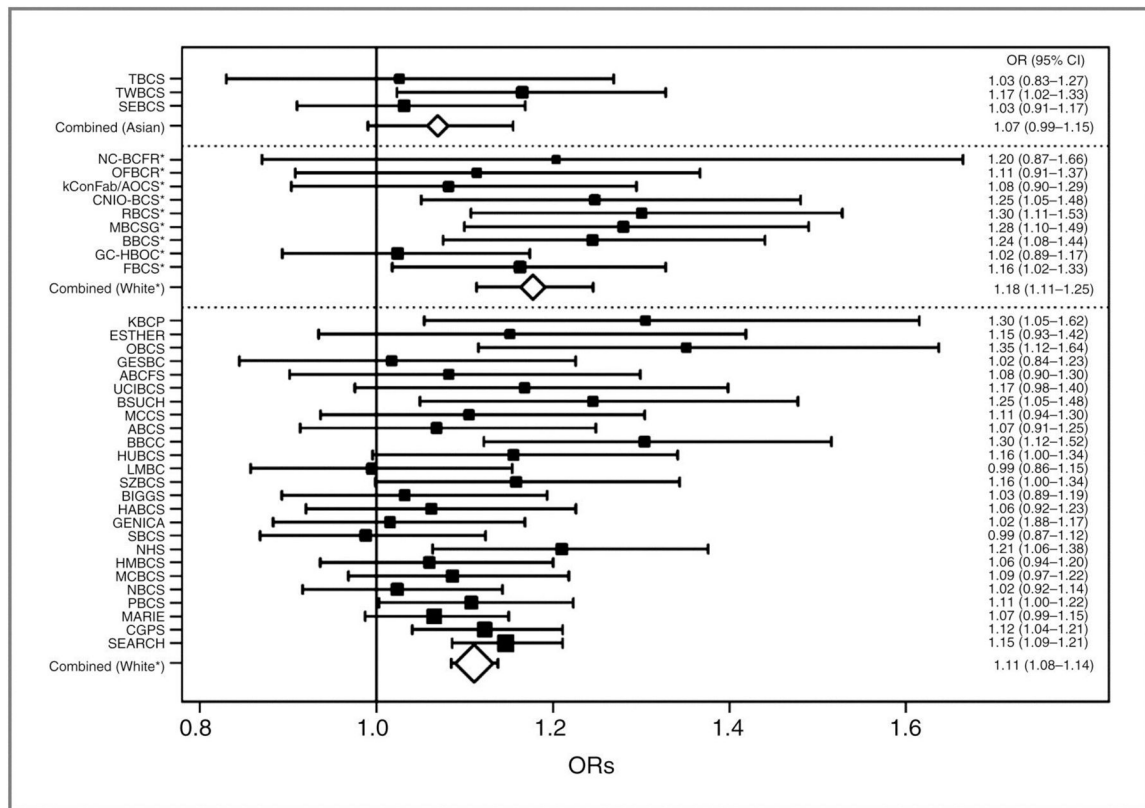


Figure 1. Association of 5p12-rs10941679 with risk of invasive breast cancer, by study
 Per-allele odds ratios (OR) and 95% confidence intervals (CI), with studies grouped into those that recruited Asian women and white European women. The latter group is further divided into those including “selected cases” likely to be at higher genetic risk (marked by an asterisk) and those with unselected cases. The area of the box/diamond is inversely proportional to the standard error of the log OR estimate.

Table 1

SNP 5p12-rs10941679 and risk of breast cancer

Group/invasiveness/genotype	Controls, N (%)	Cases, N (%)	OR (95%CI)	P
White European women				
<u>Invasive disease</u> (34 studies)				
AA	25,622 (55)	21,241 (52)	1.00	
AG	17,668 (38)	16,671 (41)	1.14 (1.11–1.17)	9×10 ⁻¹⁹
GG	3,044 (6.6)	3,060 (7.5)	1.22 (1.15–1.29)	2×10 ⁻¹²
per G-allele			1.12 (1.10–1.15)	6×10 ⁻²⁴
Adjusted for 5p12-rs981782*				
AA	15,648 (56)	12,243 (52)	1.00	
AG	10,684 (38)	9,598 (41)	1.14 (1.10–1.18)	2×10 ⁻¹¹
GG	1,810 (6.4)	1,707 (7.3)	1.19 (1.11–1.28)	3×10 ⁻⁶
per G-allele			1.11 (1.09–1.15)	6×10 ⁻¹³
White European women				
<u>DCIS</u> (15 studies)				
AA	14,187 (55)	726 (52)	1.00	
AG	9,733 (38)	578 (41)	1.17 (1.04–1.31)	0.009
GG	1,662 (6.5)	94 (6.7)	1.10 (0.88–1.38)	0.4
per G-allele			1.10 (1.01–1.21)	0.03
Asian women				
<u>Invasive disease</u> (3 studies)				
AA	612 (26)	765 (25)	1.00	
AG	1,155 (49)	1,460 (49)	1.03 (0.90–1.18)	0.6
GG	570 (24)	782 (26)	1.14 (0.98–1.33)	0.09
per G-allele			1.07 (0.99–1.15)	0.09

OR, odds ratio; CI, confidence interval; DCIS, ductal carcinoma in situ

* SNP 5p12-981782 included under a codominant model (2df), based on data on both variants available from 19 studies, including previously published data on 5p12-rs198782 (2).

Table 2

SNP 5p12-rs10941679 and risk of invasive breast cancer for white Europeans, by ER, PR and HER2 status

	N Participants		Log-additive Model		
	Cases	Controls	OR (95%CI)	P	Case-only P-heterogeneity
<u>ER Status (29 studies)</u>					
ER+	20,044	41,436	1.15 (1.12–1.19)	7×10 ⁻²⁴	
ER-	6,201	41,436	1.04 (0.99–1.08)	0.1	4×10 ⁻⁶
<u>PR Status (28 studies)</u>					
PR+	14,557	40,577	1.16 (1.12–1.19)	8×10 ⁻²⁰	
PR-	7,744	40,577	1.04 (1.00–1.08)	0.08	5×10 ⁻⁷
<u>ER/PR Status (28 studies)</u>					
ER+/PR+	13,517	40,577	1.16 (1.12–1.20)	3×10 ⁻¹⁹	
ER+/PR-	3,104	40,577	1.06 (1.00–1.13)	0.04	
ER-/PR+	960	40,577	1.13 (1.02–1.26)	0.02	
ER-/PR-	4,558	40,577	1.02 (0.97–1.07)	0.5	6×10 ⁻⁶
<u>ER, PR and HER2 Status (16 studies)</u>					
(ER+ or PR+) & HER2-	8,078	25,726	1.15 (1.10–1.20)	2×10 ⁻¹¹	
(ER+ or PR+) & HER2+	1,188	25,726	1.11 (1.02–1.22)	0.02	
ER-/PR-/HER2-	1,641	25,726	1.04 (0.95–1.12)	0.4	
ER-/PR-/HER2+	750	25,726	0.93 (0.83–1.05)	0.3	

OR, odds ratio; CI, confidence interval; ER, estrogen receptor; PR, progesterone receptor; HER2, human epidermal growth factor receptor 2

Table 3

SNP 5p12-rs10941679 and risk of invasive breast cancer for white Europeans, by pathology characteristics

N Participants		Log-additive Model		
Cases	Controls	OR (95%CI)	P	Case-only P-heterogeneity*
<u>Axillary node status (27 studies)</u>				
Negative	16,566	39,148	1.12 (1.09–1.15)	3×10 ⁻¹³
Positive	9,762	39,148	1.13 (1.09–1.17)	1×10 ⁻¹⁰ 0.9
<u>Grade (26 studies)</u>				
1	5,032	38,601	1.20 (1.14–1.25)	1×10 ⁻¹³
2	12,204	38,601	1.13 (1.09–1.17)	1×10 ⁻¹²
3/4	7,802	38,601	1.04 (1.00–1.08)	0.05 5×10 ⁻⁷
<u>Size (19 studies)</u>				
10mm	3,707	28,102	1.16 (1.09–1.22)	3×10 ⁻⁷
11–20mm	7,964	28,102	1.11 (1.07–1.16)	1×10 ⁻⁷
>20mm	6,823	28,102	1.11 (1.07–1.16)	1×10 ⁻⁶ 0.4
<u>Morphology (26 studies)</u>				
Ductal	20,283	38,447	1.12 (1.09–1.15)	2×10 ⁻¹⁵
Lobular	3,829	38,447	1.07 (1.02–1.13)	0.01 0.1

OR, odds ratio; CI, confidence interval

* For size and grade, a linear trend was tested by fitting the median values for each category as a continuous variable.

Table 4

SNP 5p12-rs981782 and risk of invasive breast cancer in white European women

Group/genotype	Controls, N (%)	Cases, N (%)	OR (95%CI)	P
Subjects genotyped for the present study (7 studies)				
AA	2,845 (28)	2,310 (28)	1.00	
AG	5,162 (50)	4,048 (49)	0.95 (0.89–1.02)	0.2
GG	2,333 (23)	1,855 (23)	0.94 (0.87–1.03)	0.2
per G-allele			0.97 (0.93–1.01)	0.2
Combined analysis* (19 studies)				
AA	7,815 (28)	6,890 (29)	1.00	
AG	14,056 (50)	11,626 (49)	0.93 (0.90–0.97)	0.001
GG	6,271 (22)	5,032 (21)	0.90 (0.86–0.95)	3×10 ⁻⁵
per G-allele			0.95 (0.92–0.97)	2×10 ⁻⁵
Adjusted for rs10941679* (19 studies)				
AA	7,815 (28)	6,890 (29)	1.00	
AG	14,056 (50)	11,626 (49)	0.95 (0.91–0.99)	0.02
GG	6,271 (22)	5,032 (21)	0.93 (0.89–0.98)	0.009
per G-allele			0.97 (0.94–0.99)	0.007

OR, odds ratio; CI, confidence interval

* based on white European subjects genotyped for 5p12-rs10941679 and 5p12-rs981782, including previously published data on the latter (2)