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Trends in *Clostridium difficile* Disease: Epidemiology and Intervention

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Abstract

Clostridium difficile is the most common cause of nosocomial infectious diarrhea. The incidence of *C difficile* infection (CDI) is increasing in both inpatients and outpatients, and outbreaks caused by a hypervirulent strain of *C difficile* are resulting in more severe disease. Moreover, community-associated CDI is occurring in persons who lack the traditional risk factors, which include antibiotic use, advanced age, and severe underlying disease. The clinical severity of CDI ranges from a mild, self-limited diarrheal illness to a fulminant, life-threatening colitis. Enzyme-linked immunosorbent assay is the most common laboratory method used for detection of *C difficile* toxins and can confirm the diagnosis within several hours. The choice of treatment should be based on disease severity. Oral metronidazole is generally regarded as the treatment of choice for mild to moderate CDI, while oral vancomycin is recommended for severe disease. Timely surgical intervention is important in patients who have severe complicated CDI.

Keywords

Clostridium difficile; Pseudomembranous colitis

Clostridium difficile is a spore-forming, anaerobic, gram-positive bacillus. Although the bacterium was discovered in 1935,¹ it was not associated with pseudomembranous colitis until 1977.^{2,3} *C difficile* is now recognized as the most common cause of nosocomial infectious diarrhea.⁴ It is responsible for up to 25% of cases of antibiotic-associated diarrhea,⁵ up to 75% of cases of antibiotic-associated colitis, and greater than 90% of cases of antibiotic-associated pseudomembranous colitis.⁶

This review provides an overview of *C difficile* infection (CDI), including a discussion of epidemiology, risk factors, clinical presentation, diagnosis, treatment, and preventative measures.

EPIDEMIOLOGY

C difficile is primarily a nosocomial pathogen. The prevalence of asymptomatic colonization in healthy adults is only 3%⁷; however, the prevalence in long-term care facilities has been reported to be up to 50%.^{8,9} Colonized persons can serve as a reservoir for infection by

contaminating the environment with *C difficile* spores.^{10, 11} The organism can be isolated from patient rooms several months after an infected patient has been discharged.¹² Health care workers are thought to be the primary source of transmission in health care facilities, spreading the organism on their hands or medical equipment after coming into contact with patients with symptomatic infection.¹¹

The risk of becoming colonized with *C difficile* is directly proportional to the length of hospitalization,¹⁰ with a median time from admission to acquisition of about 2 weeks. Length of stay is likely to be a surrogate marker for the risk of exposure to other patients with *C difficile* infection (CDI).^{13, 14}

The occurrence of symptomatic disease from *C. difficile* varies widely between hospitals, but the overall incidence is estimated to be approximately 1% of all inpatients in acute care facilities.¹⁵ Although numerous strains exist within a single center, outbreaks typically are linked to a single strain.¹⁵ The risk of symptomatic disease is higher in newly exposed and infected patients, possibly because those who are already colonized have pre-existing immunity to *C difficile* toxins.¹⁶ After infection, the patient's immune response to *C difficile* toxins appears to play an important role in determining whether the person becomes asymptotically colonized or whether disease develops. Persons who mount high antibody titers to toxin A are less likely to have diarrhea than are patients with a poor antibody response.¹⁷

The incidence and severity of CDI among hospitalized patients are increasing worldwide. The number of patients discharged from US hospitals with diagnosed CDI nearly doubled from 31 per 100,000 population in 1996 to 61 per 100,000 population in 2003.¹⁸ Since 2000, many CDI epidemics in the United States, Canada, United Kingdom, and Europe have been associated with the North American pulsed-field gel electrophoresis type 1 (NAP1) strain of *C difficile*.^{15, 19, 20} CDI outbreaks associated with the NAP1 strain are more severe, leading to more colectomies²¹ and an attributable mortality of 16.7%.²²

A deletion in the negative regulator of toxin production, *tcdC*, is thought to be the reason that the NAP1 strain produces greater than 15 times more toxin A and B than previously identified strains.²⁰ This strain is highly resistant to fluoroquinolones, and outbreaks attributed to this strain are strongly associated with the use of fluoroquinolones, but other antibiotics are also implicated.^{21,22}

No national surveillance system is in place in the United States for tracking community-associated CDI; however, the incidence is estimated to be 8 to 12 cases per 100,000 population. Surveillance data from Connecticut and Philadelphia show incidences of 6.9 and 7.6 community-associated CDI cases per 100,000 population, respectively.^{23, 24} In Philadelphia this corresponded to one case of community-associated CDI per 5549 outpatient antibiotic prescriptions, although only 76% of these patients had received antibiotics in the preceding 3 months.²³ Furthermore, severe cases of community-associated CDI have occurred in relatively young and healthy persons who lack the traditional risk factors for CDI.²³

This increase in community-associated CDI has raised the concern that *C difficile* may be a zoonotic or food-borne pathogen. Although a definitive link between the food supply and CDI remains unproven, the spore form of *C difficile* is capable of surviving the standard cooking process and could potentially infect humans if consumed during a meal.²⁵ CDI has been described in many animals,^{26–29} including house pets,³⁰ and *C difficile* or its toxins have been isolated from the stool of livestock and poultry,^{31–33} causing concerns about the contamination of meat and dairy products. Added support for a possible food-borne association comes from a study in which *C difficile* was cultured from 12 of 60 (20%) retail ground beef samples during a 10-month period in Canada.³⁴ One quarter of the isolates were identical to non-NAP1 isolates that were also found in humans with CDI.

Further support of a possible zoonotic link comes from the recent outbreaks of severe CDI in the Netherlands. The predominant strain of *C. difficile* infecting pigs and cattle in this area is a ribotype 078 that contains a toxin pattern similar to that of the NAP1(ribotype 027) strain. The number of infections in humans secondary to the ribotype 078 strain has been increasing rapidly in the Netherlands, from 3% in 2005 to 13% in 2008.³⁵ Patients infected with this strain of *C. difficile* were more likely to be younger and have community-acquired disease. Severity of the illness in patients infected with ribotype 078 is equivalent to that in patients infected with the NAP1 strain. Analysis of these strains isolated from both pigs and humans revealed overlapping antimicrobial susceptibilities and numerous other similarities indicating a very close genetic relationship.^{35, 36}

PATHOGENESIS

C difficile causes symptoms by producing exotoxins in the intestinal tract. Toxin A causes mucosal damage via an intense neutrophilic infiltrate that leads to inflammatory diarrhea. Toxin B is a very potent cytotoxin but appears to be less enterotoxic than toxin A.³⁷ Direct interaction between the toxins and surface receptors in the colonic mucosa result in actin filament degradation that causes necrosis and sloughing of cellular debris into the colonic lumen. Both toxins and other surface proteins further induce an inflammatory response by triggering cytokine release from monocytes and dendritic cells.

Exudation of inflammatory cells and proteins from the resulting ulcers causes the visible yellow plaques that form the characteristic pseudomembrane. The number of lesions appears to be dose-dependent, with greater amounts of toxin resulting in a more confluent pseudomembrane.³⁷

RISK FACTORS

The development of CDI depends on both an interruption in the usual host flora and acquisition of the organism. Risk factors most consistently identified in the literature include antibiotic use, advanced age, and severe underlying disease.³⁸ The most important risk factor in hospitalized patients is antibiotic exposure.³⁹ A history of antibiotic exposure is found in more than 90% of inpatients with CDI.⁷ Clindamycin, ampicillin, and cephalosporins were most frequently implicated before the NAP1 epidemic.^{39, 40} Fluoroquinolones were implicated in the most recent epidemic of CDI in hospitalized

patients.²² Longer duration of antimicrobial administration and the use of multiple antimicrobials also have been associated with an increased risk.²²

Aging is a risk factor for CDI. Incidence increases with each decade of life, with the greatest increase seen in patients older than 65 years.³⁸ Colonization rates in the elderly are up to 100 times greater than rates in young adults and adolescents, and disease rates are 20-fold higher.^{15, 41} Institutionalization, longer hospital stays, and comorbid conditions and infections that necessitate frequent treatment with antibiotics are contributing factors.

Immunosuppression from HIV infection or from chemotherapy or transplants also puts patients at higher risk for CDI. A retrospective analysis of 44,778 patients over 10 years revealed that CDI was the cause of diarrhea in more than 5% of HIV-positive patients.⁴² The incidence in solid organ and hematopoietic stem cell transplant recipients ranged from 1% to 31%.^{43–45}

Gastric acid suppression also may increase the risk of CDI. The acidic environment of the stomach has been shown to be fatal to vegetative *C difficile*⁴⁵; however this effect is nullified once gastric pH reaches 5.⁴⁶ The increasing use of proton-pump inhibitors (PPIs) has also been correlated with the increasing incidence of CDI.⁴⁷ Several studies have shown that CDI is more than twice as likely to develop in hospitalized patients prescribed PPIs than in other patients.^{14, 15, 48} The use of PPIs also has been shown to alter the normal flora, which may enhance the ability of *C difficile* to colonize the GI tract.⁴⁹

Enteral feeding has been shown to increase the risk of *C difficile* acquisition from 8% to 20% and the risk of CDI from 1% to 9%.⁵⁰ Several factors associated with tube feeding are thought to increase the risk of infection, including contamination of the formula during preparation or the equipment by handling or alteration of the normal colonic flora.^{51, 52} Although any feeding tube appears to increase the incidence of CDI, post-pyloric tubes confer the greatest risk.⁵⁰

CLINICAL MANIFESTATIONS

The clinical severity of CDI ranges from a mild, self-limited diarrheal illness to a fulminant, life-threatening colitis. Symptoms can develop within the first several days after antibacterial treatment is initiated or be delayed until up to 10 weeks after cessation of antibiotics.^{53, 54} Low-grade fever and cramping abdominal pain often accompany the diarrhea of CDI. The disease progresses to fulminant colitis in 1% to 8% of patients.⁵⁵

Mild to moderate CDI consists of diarrhea with abdominal cramping, but not systemic symptoms. Severe disease involves profuse diarrhea, abdominal pain, abdominal distention, leukocytosis, and systemic symptoms such as fever. Severe disease with complications includes paralytic ileus, colonic dilatation with systemic toxicity (toxic megacolon), or other immediate life-threatening conditions. In severe CDI, ileus or toxic megacolon may cause a paradoxical decrease in the volume of diarrhea.⁵³

CDI may also have a variety of unusual or unexpected presentations. If patients have ileus but not the diarrhea typically associated with severe CDI, clinicians will need to rely on

supportive examination and laboratory findings. Striking leukocytosis, often greater than 30,000 cells/uL, is a common supportive laboratory finding in patients with CDI that frequently precedes organ dysfunction.^{56, 57} Other unusual manifestations of CDI may include perforation of the small or large bowel,⁵⁸ protein-losing enteropathy,⁵⁹ and a variety of extracolonic manifestations such as visceral abscesses, reactive arthritis, and post-traumatic soft tissue infections.⁶⁰ These otherwise unexplained findings should prompt the clinician to consider CDI to prevent the high morbidity and mortality that results from undiagnosed disease.

The differential diagnosis of CDI includes both non-infectious and other infectious causes of antibiotic-associated diarrhea. Antibiotics may directly cause diarrhea through osmotic factors or stimulation of intestinal motility. Many other medications that are commonly co-administered with antibiotics in hospitalized patients, such as PPIs, should also be considered as potential causes of diarrhea.⁶¹ Other causes of non-infectious colitis may be confused with CDI, especially in patients with inflammatory bowel disease since there is an increase in *C. difficile* colonization in this group that may result in misleading diagnostic study results.⁶² Other less common infectious causes of antibiotic-associated diarrhea include *Staphylococcus aureus*, especially if methicillin-resistant, and antibiotic-resistant gram negative bacilli, such as *Klebsiella oxytoca*.^{63, 64}

DIAGNOSIS

The diagnosis of CDI is most frequently established by confirming the presence of *C. difficile* or one of its toxins in the stool of a symptomatic patient. The clinical laboratory gold standard is the cytotoxicity cell assay,⁶⁵ but cost and the 48-hour delay in results have caused this test to be replaced by the enzyme-linked immunosorbent assay (ELISA) in most US laboratories.⁶⁶ ELISA is the most common diagnostic laboratory method used in the United States for detection of *C. difficile* toxins. Growth in culture does not differentiate toxigenic from non-toxigenic strains of *C. difficile*. This is important because only toxigenic strains of *C. difficile* can cause CDI, and therefore stool culture false-positive rate can exceed 10%.⁶⁵

Toxin detecting ELISA can confirm the diagnosis within several hours and is relatively inexpensive. The major disadvantage is that its sensitivity is 70% to 90%, whereas the sensitivity of the cytotoxicity cell assay is greater than 90%.^{67, 68} Most currently available ELISAs detect both toxin A and toxin B; however older assays that only detect toxin A have a higher false-negative rate. There appears to be little value in repeating the ELISA after an initial negative test as few convert to positive⁶⁹ and there is a precipitous decline in the positive predictive value of the test.

ELISAs that detect *C. difficile* glutamate dehydrogenase (GDH) are widely available and inexpensive.⁷⁰ The sensitivity of the assay exceeds 95% with a negative predictive value of over 99%;⁷¹ however the positive predictive value is only about 50%.⁷⁰ GDH is constitutively produced by all strains of *C. difficile*, so this test cannot differentiate between toxigenic and non-toxigenic strains.⁷² To improve the specificity of the GDH assay, a second confirmatory test that identifies *C. difficile* toxins must also be used. A 2-step

algorithm consisting of the GDH assay followed by a confirmatory cytotoxicity assay on positive samples has superior sensitivity and specificity to toxin detecting ELISAs, but increases turnaround time for positive results by several days.⁷⁰

Polymerase chain reaction (PCR) shows promise as a sensitive and rapid method for diagnosing CDI.^{73, 74} Real-time PCR detection of the toxin B gene also has proved promising. In a study of 1368 stool specimens, the sensitivity was 93% compared with 78% for the cytotoxicity cell assay and 73% for a toxin-detecting EIA.⁷⁵ CT scanning may reveal patterns that suggest CDI and support the diagnosis in uncertain cases. The most common finding is diffuse or segmental thickening of the bowel wall more than 4 mm.⁷⁶ Other supportive findings include colonic distention, pericolonic stranding, colonic fold effacement, and nodular fold thickening.^{76, 77} The sensitivity of CT is approximately 50%.

Endoscopy is generally not necessary to establish the diagnosis and may increase the incidence of perforation in patients with severe disease.⁷⁸ Sigmoidoscopy or colonoscopy may be used if the ELISA finding is negative but clinical suspicion remains high or the diagnosis cannot be delayed.⁷⁸

TREATMENT

The choice of treatment should be based on disease severity (Figure). Cessation of the causative antibiotic is important, although this may not always be feasible. If the patient has a concomitant infection that requires antibiotic therapy, it is reasonable to change the antibiotic to a more narrow-spectrum alternative or use antimicrobials that are less often associated with CDI.

Oral metronidazole, 500 mg tid or 250 mg qid for 10 to 14 days, is generally regarded as the drug of choice for mild to moderate CDI. This recommendation is based on the significant cost savings and comparable efficacy compared with oral vancomycin for mild to moderate CDI.^{79–81} Oral vancomycin, 125 mg qid for 10 to 14 days, is becoming the preferred first-line agent for patients with severe CDI, based on studies that showed significantly improved cure rates compared with metronidazole in these patients.^{81, 82}

When severe CDI is complicated by ileus, the efficacy of oral or nasogastric vancomycin may be compromised by an inability of the drug to reach the site of infection. Although this approach has not been studied, increasing the vancomycin dose to 250–500mg every six hours may theoretically improve the chance that adequate drug concentrations are achieved in the colon. Intracolonic administration of vancomycin is another possible strategy in the management of patients with ileus and the route is supported by several case reports when used in conjunction with other treatments.^{83–86}

Timely surgical intervention is important in patients with severe complicated CDI. Even before the emergence of the NAP1 strain of *C difficile*, it was noted that mortality was reduced if surgery was done within 48 hours of a failure to respond to medical therapy.⁸⁷ Because rapid progression to death is associated with the hypervirulent strain of *C difficile*, surgical consultation is especially urgent in these patients.

Patients who are elderly or immunocompetent or have leukocytosis or elevated lactate levels benefit most by emergency colectomy.⁸⁸ Postoperative mortality is increased in patients with very severe underlying illness, mental status changes, protracted poor response to medical treatment, or hypotension requiring treatment with vasopressors before colectomy.⁸⁹

In addition to vancomycin and metronidazole, possible antibiotic therapy includes bacitracin, rifampin, rifaximin, nitazoxanide, fusidic acid, and teicoplanin. Symptomatic cure rates with bacitracin are similar to those of vancomycin, but bacteriological cure rates are inferior.⁹⁰ The efficacy of rifaximin, nitazoxanide, and fusidic acid appears to be identical to that of metronidazole and vancomycin.⁹⁰

Teicoplanin was found to be superior to vancomycin and metronidazole at reducing toxin levels in the stool, but was not statistically superior in reducing symptoms or relapse rates.⁹⁰ Although teicoplanin has shown some potential benefit in comparison with vancomycin, it is not available in the United States and is much more expensive than other commonly used medications. As a result of the equal efficacy in clinical cure, limited availability, and greater experience with metronidazole and vancomycin, these other antibiotics are rarely used.

Probiotics such *Saccharomyces* or *Lactobacillus* tablets have been studied as adjunctive therapy because they theoretically restore nonpathogenic flora to the GI tract, inhibit *C difficile* toxin production, and stimulate the host immune system. Several randomized controlled trials have been conducted but none have shown that the addition of a probiotic to vancomycin or metronidazole therapy improved the course of disease.^{91–94} Adverse effects of probiotics are rare in immunocompetent patients; however, probiotics should be avoided in patients who are immunocompromised or critically ill or have central venous catheters, because organisms in the probiotic preparation may cause bloodstream infections.^{95, 96}

Cholestyramine and colestipol bind the toxins of *C difficile* in vitro; however, the efficacy of these agents in acute disease has not been demonstrated.^{97–99} Furthermore, because these agents bind vancomycin, concomitant administration of these drugs may result in subtherapeutic fecal concentrations of vancomycin.¹⁰⁰

Several case studies and series suggest that toxin-binding resins may help prevent relapses, but further study is needed before such medications can be recommended in the management of CDI.^{39, 101, 102} Avoidance of medications that may exacerbate CDI is also important. Although the evidence is anecdotal, antiperistaltic agents, including narcotics, may contribute to the development of toxic megacolon and, thus, generally should not be administered to patients with CDI.¹⁰³

Treatment failure and relapse

Factors associated with metronidazole failure include a serum albumin level of 2.5 g/dL or less, previous or current stay in the ICU, and continuation of the causative antibiotic.^{104, 105} Although antibiotic resistance does not appear to be a significant cause of metronidazole

failure, switching to oral vancomycin is a common practice and may provide symptomatic relief.^{106, 107}

CDI recurs in up to 35% of patients who respond to therapy with metronidazole or vancomycin,^{108, 109} and more half of these patients will have additional recurrences.¹¹⁰ Associated risk factors are similar to those for disease acquisition and include antibiotic use, older age, gastric acid-suppressive therapy, prolonged hospital stay, nursing home residence, and the presence of comorbidities.⁶ More than half of relapses appear to be the result of reinfection with a different strain of *C difficile* rather than reactivation of spores remaining in the colon.¹⁰⁸

The first relapse should be treated similarly to the initial episode, with severity of the disease guiding the choice of medication.¹⁰⁸ Management of multiple relapses has not been definitively studied, but prolonged tapering and pulsed dosing of oral vancomycin have been used successfully (Table).^{111, 112}

Several studies suggest that rifaximin, which has in vitro activity against *C difficile*, may be effective in treating recurrent disease.^{113–115} In a case series of 8 women who each had at least 4 previous episodes of CDI and who received a 2-week course of rifaximin after vancomycin therapy, 7 remained symptom-free after a mean follow-up of 233 days.¹¹⁵ After receiving a second course of rifaximin, the eighth patient had no further relapses. Similar results were generated in another case series involving 6 symptomatic patients who had at least 1 previous episode of CDI.¹¹³ Five patients had no recurrence after a mean follow-up of 310 days. Interestingly, the patient that continued to have recurrent disease was culture-negative for *C difficile*. The emergence of rifaximin resistance during therapy has been noted in several studies and may be a potential deterrent to routine use.^{113, 115}

PREVENTION

The prevention of CDI depends both on eliminating the spread of the organism and reducing the risk of infection in individual patients. Reducing *C difficile* transmission within institutions entails strict hand hygiene and appropriate contact precautions, such as wearing a gown and gloves when entering an infected patient's room.^{116–119} *C difficile* spores are resistant to standard disinfectants and can contaminate dry surfaces for months.¹³ Bleach diluted 1:10 with water is sporicidal in 10 minutes.¹²⁰ It is not clear whether routine environmental decontamination with a sporicidal agent is necessary, although it may be of benefit during disease outbreaks.^{121, 122}

Antimicrobial stewardship programs and formulary restrictions are important in reducing the patient's risk of infection after exposure to *C difficile*. A 54% reduction in antibiotic use after introduction of an antimicrobial stewardship program during the nosocomial CDI outbreak of the NAP1 strain in Quebec was associated with a 60% decrease in the incidence of CDI.¹²³ Similar results were reported with an antimicrobial stewardship program that reduced use of broad-spectrum antibiotics while leaving overall antibiotic use unchanged.¹²⁴

Specific restrictions of cephalosporins and clindamycin have led to statistically significant decreases in the incidence of CDI at many centers.^{125–128} Because CDI outbreaks caused by

the NAP1 strain are increasingly being associated with fluoroquinolones, restriction on fluoroquinolones use may prove beneficial.

SUMMARY

The incidence and severity of CDI in both the inpatient and outpatient settings are increasing worldwide. Although traditional risk factors for CDI continue to apply in nosocomial disease, severe community-associated CDI has begun to appear in previously healthy persons. The continued search for a deeper understanding of the epidemiology of CDI has become even more important now that a hypervirulent strain of *C difficile*—NAP1—has emerged.

Treatment recommendations are evolving. Vancomycin is now the drug of choice for treatment of severe disease. Additional therapies, however, are needed to stem the increasing morbidity and mortality associated with CDI. Attention to infection control practices, including hand hygiene and contact precautions, in combination with antimicrobial control programs, have proved beneficial in controlling nosocomial outbreaks and should be used to reduce the rising incidence of CDI.

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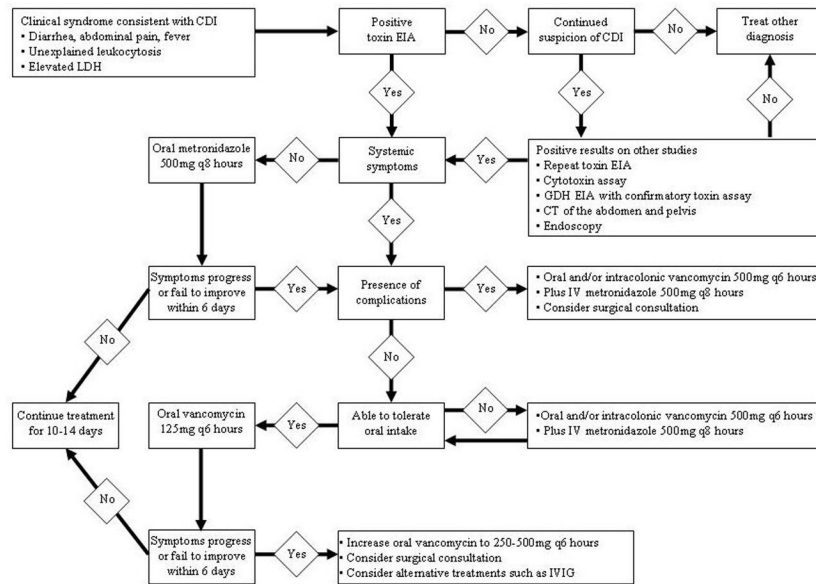


Figure 1.
Diagnosis and Treatment of CDI

Table 1

Possible Treatment Options for Multiple Recurrences of CDI

Week #	Treatment Phase		Tapering Phase			
	1	2	3	4	5	6
Vancomycin Taper						
Vancomycin Dose	125mg q6 hours	125mg q6 hours	125mg q8 hours	125mg q12 hours	125mg q24 hours	125mg q48 hours
Vancomycin Pulse						
Vancomycin Dose	125mg q6 hours	125mg q6 hours	125mg qMWF	125mg qMWF	125mg qMWF	125mg qMWF
Rifaximin Chaser						
Vancomycin Dose	125mg q6 hours	125mg q6 hours	---	---	---	---
Rifaximin Dose	---	---	200-400mg Q12	200-400mg Q12	---	---