

Shigella Isolates From the Global Enteric Multicenter Study Inform Vaccine Development

Sofie Livio,¹ Nancy A. Strockbine,^{2,3} Sandra Panchalingam,¹ Sharon M. Tennant,¹ Eileen M. Barry,¹ Mark E. Marohn,¹ Martin Antonio,⁴ Anowar Hossain,⁵ Inacio Mandomando,⁶ John B. Ochieng,⁷ Joseph O. Oundo,⁷ Shahida Qureshi,⁸ Thandavarayan Ramamurthy,⁹ Boubou Tamboura,¹⁰ Richard A. Adegbola,^{4,a} Mohammed Jahangir Hossain,⁴ Debasish Saha,^{4,b} Sunil Sen,¹ Abu Syed Golam Faruque,⁵ Pedro L. Alonso,⁶ Robert F. Breiman,^{7,11,c} Anita K. M. Zaidi,^{8,d} Dipika Sur,^{9,12} Samba O. Sow,¹⁰ Lynette Y. Berkeley,^{1,13} Ciara E. O'Reilly,³ Eric D. Mintz,³ Kousick Biswas,¹⁴ Dani Cohen,¹⁵ Tamer H. Farag,^{1,d} Dilruba Nasrin,¹ Yukun Wu,¹ William C. Blackwelder,¹ Karen L. Kotloff,¹ James P. Nataro,^{1,e} and Myron M. Levine¹

¹Center for Vaccine Development, University of Maryland School of Medicine, Baltimore; ²*Escherichia* and *Shigella* Reference Unit, Enteric Diseases Laboratory Branch, and ³Division of Foodborne, Waterborne and Environmental Diseases, Centers for Disease Control and Prevention, Atlanta, Georgia; ⁴Medical Research Council Unit (United Kingdom), Fajara, The Gambia; ⁵International Centre for Diarrhoeal Disease Research, Mohakhali, Dhaka, Bangladesh; ⁶Centro de Investigação em Saúde de Manhiça, Manhiça, Mozambique and the Centre de Recerca en Salut Internacional de Barcelona, Hospital Clinic/Universitat de Barcelona, Spain; ⁷Kenya Medical Research Institute/Centers for Disease Control and Prevention, Kisumu, Kenya; ⁸Department of Paediatrics and Child Health, The Aga Khan University, Karachi, Pakistan; ⁹National Institute of Cholera and Enteric Diseases, Kolkata, India; ¹⁰Centre pour le Développement des Vaccins du Mali, Bamako, Mali; ¹¹Global Disease Detection Division, Kenya Office of the US Centers for Disease Control and Prevention, Nairobi, Kenya; ¹²Program for Appropriate Technology in Health (PATH), New Delhi, India; ¹³US Food and Drug Administration, Rockville; ¹⁴Department of Veterans Affairs, Cooperative Studies Program Coordinating Center, Perry Point, Maryland; and ¹⁵Department of Epidemiology and Preventive Medicine, School of Public Health, Sackler Faculty of Medicine, Tel Aviv University, Ramat Aviv, Israel

(See the Editorial Commentary by Van de Verg and Venkatesan on pages 942–3.)

Background. *Shigella*, a major diarrheal disease pathogen worldwide, is the target of vaccine development. The Global Enteric Multicenter Study (GEMS) investigated burden and etiology of moderate-to-severe diarrheal disease in children aged <60 months and matched controls without diarrhea during 3 years at 4 sites in Africa and 3 in Asia. *Shigella* was 1 of the 4 most common pathogens across sites and age strata. GEMS *Shigella* serotypes are reviewed to guide vaccine development.

Methods. Subjects' stool specimens/rectal swabs were transported to site laboratories in transport media and plated onto xylose lysine desoxycholate and MacConkey agar. Suspect *Shigella* colonies were identified by biochemical tests and agglutination with antisera. *Shigella* isolates were shipped to the GEMS Reference Laboratory (Baltimore, MD) for confirmation and serotyping of *S. flexneri*; one-third of isolates were sent to the Centers for Disease Control and Prevention for quality control.

Results. *Shigella dysenteriae* and *S. boydii* accounted for 5.0% and 5.4%, respectively, of 1130 *Shigella* case isolates; *S. flexneri* comprised 65.9% and *S. sonnei* 23.7%. Five serotypes/subserotypes comprised 89.4% of *S. flexneri*, including *S. flexneri* 2a, *S. flexneri* 6, *S. flexneri* 3a, *S. flexneri* 2b, and *S. flexneri* 1b.

Conclusions. A broad-spectrum *Shigella* vaccine must protect against *S. sonnei* and 15 *S. flexneri* serotypes/subserotypes. A quadrivalent vaccine with O antigens from *S. sonnei*, *S. flexneri* 2a, *S. flexneri* 3a, and *S. flexneri* 6 can provide broad direct coverage against these most common serotypes and indirect coverage against all but 1 (rare) remaining subserotype through shared *S. flexneri* group antigens.

Keywords. serotyping; *Shigella*; shigellosis; vaccines.

Received 15 January 2014; accepted 5 May 2014; electronically published 23 June 2014.

^aPresent affiliation: GlaxoSmithKline Vaccines, Wavre, Belgium.

^bPresent affiliation: Center for International Health, University of Otago, Dunedin, New Zealand.

^cPresent affiliation: Global Health Institute, Emory University, Atlanta, Georgia.

^dPresent affiliation: Bill and Melinda Gates Foundation, Seattle, Washington.

^ePresent affiliation: Department of Pediatrics, University of Virginia School of Medicine, Charlottesville.

Correspondence: Myron M. Levine, MD, DTPH, Center for Vaccine Development, University of Maryland School of Medicine, 685 West Baltimore St., Baltimore, MD 21201 (mlevine@medicine.umaryland.edu).

Clinical Infectious Diseases 2014;59(7):933–41

© The Author 2014. Published by Oxford University Press on behalf of the Infectious Diseases Society of America. This is an Open Access article distributed under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs licence (<http://creativecommons.org/licenses/by-nc-nd/3.0/>), which permits non-commercial reproduction and distribution of the work, in any medium, provided the original work is not altered or transformed in any way, and that the work properly cited. For commercial re-use, please contact journals.permissions@oup.com.

DOI: 10.1093/cid/ciu468

The Global Enteric Multicenter Study (GEMS) of the burden and etiology of moderate-to-severe diarrheal illness (MSD) in children aged <5 years performed over 3 years at 4 sites in sub-Saharan Africa (Basse, The Gambia; Bamako, Mali; Siaya County, Kenya; Manhica, Mozambique) and 3 in South Asia (Karachi, Pakistan; Kolkata, India; Mirzapur, Bangladesh) established *Shigella* as 1 of 4 top pathogens [1]. The increased diagnostic yield observed when stool specimens are examined using gel-based or quantitative real-time polymerase chain reaction (PCR) suggests that the burden of disease may be greater than estimated using standard cultures [2, 3]. Although pediatric morbidity from shigellosis remains substantial, mortality has diminished, in part, because of the virtual disappearance worldwide of the highly virulent Shiga toxin-producing *S. dysenteriae* 1 serotype and because World Health Organization guidelines recommend antibiotic treatment for clinical dysentery (diarrhea with gross blood). Regrettably, *Shigella* relentlessly acquires resistance to antibiotics that were previously effective in diminishing disease severity and duration and pathogen excretion [2, 4].

Based on clinical severity, disease burden, and emergence of antimicrobial resistance, *Shigella* is a prime target for vaccine development [2, 4–6]. The 4 species (also called groups or subgroups) of *Shigella* encompass 50 serotypes and subserotypes that include the following: *S. dysenteriae* (15 serotypes); *S. flexneri* (15 serotypes and subserotypes, including *S. flexneri* 1a, 1b, 2a, 2b, 3a, 3b, 4a, 4b, 5a, 5b, 6, X, and Y and 2 new subserotypes 7a and 7b, previously referred to as *Shigella* provisional 88–893, Y394, or “*S. flexneri* 1c” [7]); *S. boydii* (19 serotypes); and *S. sonnei* (1 serotype). The distinct serotypes/subserotypes are defined by conformational epitopes of their O polysaccharide antigens [6]. Challenge/rechallenge studies in nonhuman primates [8] and volunteers [9–11], epidemiological field studies [12], and seroepidemiological studies [13, 14] indicate that clinical infection with wild type *Shigella* strains bestows approximately 75% subgroup-specific (and usually subserotype-specific) immunity. Live oral vaccines [15–18] and O-polysaccharide-protein conjugate vaccines [19, 20] that have conferred protection in randomized controlled field trials corroborate the importance of immune responses to *Shigella* O antigens. Most *Shigella* vaccines in clinical development are based on eliciting protection against multiple epidemiologically important serotypes. Accordingly, to rationally guide vaccine formulation, it is imperative to have robust data on the distribution of *Shigella* serotypes associated with shigellosis. GEMS serotype data provide such information from the geographic areas where 80% of deaths due to diarrheal disease among young children occur [1, 21].

MATERIALS AND METHODS

Conducted over 3 years, GEMS was an age-stratified, matched case-control study of MSD among children aged 0–59 months

residing in censused populations and seeking care at medical facilities serving 7 sites in sub-Saharan Africa and South Asia. Rationale for the GEMS and detailed clinical, epidemiologic, and microbiological methods have been published elsewhere [21–24].

Epidemiological and Clinical Methods

The University of Maryland, Baltimore Institutional Review Board and ethics committees at each field site approved the protocol. A censused population provided the sampling frame at each study site where sentinel hospitals or health centers serving the population enrolled cases from 3 age strata: infants (0–11 months), toddlers (12–23 months), and young children (24–59 months) [1, 24]. Age-eligible children from the censused population visiting the centers with diarrhea (≥ 3 loose stools in the previous 24 hours) were examined for eligibility. To be included, the child’s diarrheal episode had to be new (onset after ≥ 7 diarrhea-free days), acute (duration <7 days), and had to meet at least 1 of the following criteria defining MSD: clinical evidence of moderate-to-severe dehydration (sunken eyes, loss of skin turgor, or initiation of intravenous fluids based on clinical judgment); dysentery; or clinical judgment that the child with diarrheal illness needed to be hospitalized. For each MSD case, 1–3 (occasionally 4) controls without diarrhea, randomly selected from each site’s census database and matched by age, gender, and residential community, were enrolled within 14 days of the matched index case.

Upon enrollment, each case and matched control provided a stool specimen (≥ 3 grams) that, within 1 hour of passage, was stored cold until delivered to the laboratory. If antibiotics were to be administered to patients before stool was produced, 2 rectal swabs were obtained for bacterial culture pending passage of whole stool for the remaining assays.

Bacteriologic Methods

Stool samples/rectal swabs were introduced into Cary–Blair and buffered glycerol saline (BGS) transport media, the latter to enhance yield of *Shigella* [22, 25]; inoculation onto solid media occurred within 18 hours. To isolate *Shigella*, the BGS swab was plated onto MacConkey and xylose lysine desoxycholate agar. After incubation at 37°C, suspicious colonies were subjected to biochemical tests [22]. *Shigella* isolates at the field sites were serotyped with polyvalent group A, B, C, and D antisera (Denka Seiken Co., Ltd., Tokyo, Japan or Reagensia AB, Solna, Sweden) and shipped to the GEMS Reference Laboratory at the Center for Vaccine Development (CVD) for confirmation and identification of individual *S. flexneri* serotypes/subserotypes and *S. dysenteriae* 1. One-third of isolates serotyped at CVD were sent to the Centers for Disease Control and Prevention (CDC) for serotype confirmation.

Chromosomal genes encoding *Shigella* enterotoxin 1 (ShET1) [26, 27] were amplified by PCR using the following primers:

set1AForward: 5'- CAG CGT CTT TCA GCG ACA GTG
TTT -3'
set1AReverse: 5'- AGC ATG ATA CTC AAC AGC CAG
ACC -3'
set1BForward: 5'- ATA CTG GCT CCT GTC ATT CAC
GGT -3'
set1BReverse: 5'- GGA AGT GAC AGG GCA TTT GTG
GAT -3' [28, 29].

Statistical Methods

Distributions of species were compared by χ^2 test with 3 degrees of freedom. Individual species and subserotypes were compared using χ^2 test with no continuity correction or 2-sided Fisher exact test. $P \leq .05$ was considered statistically significant.

RESULTS

We investigated 1130 *Shigella* isolates from cases (1120 from the matched case-control dataset [1] and 10 from MSD cases for whom there was no control) and 219 isolates from controls without diarrhea; 11 other isolates from the sites (9 cases and 2 controls) were not sent to CVD. The distribution of *Shigella* species among case isolates is shown in Table 1. Only 5.0% of case isolates (N = 56) were *S. dysenteriae* (none were *S. dysenteriae* 1) and 5.4% (N = 61) were *S. boydii*. Overall, 89.6% of case isolates were *S. flexneri* (N = 745; 65.9%) or *S. sonnei* (N = 268; 23.7%). Four serotypes/subserotypes, *S. flexneri* 2a, *S. flexneri* 2b, *S. flexneri* 3a, and *S. flexneri* 6, comprised 581 of the 745 *S. flexneri* isolates (78.0%; Table 1); inclusion of *S. flexneri* 1b raises the total to 89.4% of all *S. flexneri*.

Mirzapur, Bangladesh, where shigellosis exhibits a striking seasonal peak [30], contributed the most *Shigella* cases. Thus, it was important to compare Bangladesh serotype data with data from the other 6 GEMS sites (Table 1). The overall species distributions in Bangladesh and the other sites combined were significantly different ($P = .015$), but the absolute differences for individual species were modest. Percentages of the 2 most prevalent species, *S. flexneri* and *S. sonnei*, were similar and not significantly different in Bangladesh vs the combined other sites. The percentages of *S. flexneri* subserotypes were significantly different only for *S. flexneri* 1b, *S. flexneri* 2b, *S. flexneri* 6, and *S. flexneri* X. The proportion of isolates that were *S. flexneri* plus *S. sonnei* by year of the study at all sites, Bangladesh, and the 6 sites other than Bangladesh is shown in Table 2. Remarkably, there was little variation.

Among 219 isolates from control subjects (Table 1), there was less representation of *S. flexneri* (N = 115; 52.5%) compared with

cases, where 65.9% of all isolates were *S. flexneri* ($P < .001$). *Shigella flexneri* 2a, *S. flexneri* 2b, *S. flexneri* 3a, and *S. flexneri* 6 accounted for 78.0% of *S. flexneri* case isolates vs 71.3% of control *S. flexneri* isolates. *Shigella flexneri* 2a ranked first in frequency followed by *S. flexneri* 6 among cases; among controls, *S. flexneri* 6 ranked first followed by *S. flexneri* 2a.

There was excellent concordance between CVD and CDC results in serotyping quality control. The 46 *S. dysenteriae* isolates sent by CVD were confirmed by CDC as *S. dysenteriae* serotypes 2 (N = 17), 3 (N = 10), 4 (N = 12), 8 (N = 1), 9 (N = 1), and 12 (N = 5). CDC confirmed 76 of 77 putative *S. boydii* that they serotyped as *S. boydii* 1 (N = 11), 2 (N = 16), 3 (N = 1), 4 (N = 16), 5 (N = 1), 7 (N = 1), 8 (N = 4), 10 (N = 9), 11 (N = 1), 12 (N = 4), 14 (N = 3), 15 (N = 2), 18 (N = 3), and 20 (N = 5); the remaining isolate was *Escherichia albertii*, which is known to share O antigens with *S. boydii* 13 [31]. Of 37 putative *S. sonnei* isolates, 36 were confirmed and 1 was found by CDC to be *S. dysenteriae*-like provisional serotype 96–3162 serotype. CDC confirmed CVD's subserotyping of 146 of the 147 *S. flexneri* sent, the exception being a strain identified by CVD as *S. flexneri* 1b but shown by CDC to be *S. dysenteriae* 3 (strain mix-up). Finally, 22 *Shigella* isolates sent by CVD were deemed untypable with available reagents. CDC identified 21 of the 22; 20 were *S. dysenteriae*-like provisional serotype 96–3162 and 1 was *S. boydii*-like provisional serotype 2009C-3081.

ShET1, a classic enterotoxin consisting of 1 enzymatically active A subunit linked to 5 binding B subunits, is encoded by *setAB* [26, 32] located within chromosomal *Shigella* pathogenicity island 1 [26, 32], originally identified in *S. flexneri* 2a. Notably, 85 of 86 GEMS *S. flexneri* 2a tested positive for ShET1, as did all 33 *S. flexneri* 2b strains tested. Only 4 of 134 (3.0%) isolates of other *S. flexneri* serotypes were positive, including 2/2 *S. flexneri* 5b, 1/1 *S. flexneri* Y, and 1/22 *S. flexneri* 3a.

DISCUSSION

Natural immunity to *Shigella* is largely based on immune responses to O antigens. Follow-up of a Chilean pediatric cohort where *S. sonnei*, *S. flexneri* 2a, and *S. flexneri* 6 were the most prevalent serotypes (79% of cases) indicated that an initial episode of shigellosis conferred approximately 75% protection against subsequent shigellosis due to the same serotype but did not significantly cross protect against illness caused by the other predominant serotypes [12]. Seroepidemiological studies from Israel corroborate the importance of preexistent O antibody (denoting prior exposure) in lowering the risk of *S. sonnei* and *S. flexneri* 2a disease [13, 14]. Accordingly, developers of vaccines to prevent shigellosis have designed serotype-based vaccines [4, 6], some of which have conferred significant protection [9, 15–20, 33].

Table 1. Species and Serotype Distribution by Site of 1130 *Shigella* Isolates From Children Aged <60 Months With Moderate to Severe Diarrhea in the Global Enteric Multicenter Study and of 219 Isolates From Control Children Without Diarrhea

Serogroup, Serotype, or Subserotype	Cases									Controls
	All 7 GEMS Sites	6 GEMS Sites Other Than Bangladesh	Bangladesh	Pakistan	India	Gambia	Mali	Kenya	Mozambique	All 7 GEMS Sites
Total isolates	1130	519	611	129	91	116	41	105	37	219
<i>Shigella dysenteriae</i>	56 (5.0%) ^a	33 (6.4%) ^b	23 (3.8%) ^c	6 (4.7%) ^c	2 (2.2%) ^c	5 (4.3%) ^c	1 (2.4%) ^c	19 (18.1%) ^c	0	10 (4.6%)
<i>S. boydii</i>	61 (5.4%) ^a	37 (7.1%) ^b	24 (3.9%)	10 (7.8%)	10 (11.0%)	7 (6.0%)	2 (4.9%)	6 (5.7%)	2 (5.4%) ^c	24 (11.0%)
<i>S. sonnei</i> ^d	268 (23.7%) ^a	119 (22.9%) ^b	149 (24.4%)	29 (22.5%)	32 (35.2%)	24 (20.7%)	12 (29.3%)	17 (16.2%)	5 (13.5%)	70 (32.0%)
<i>S. flexneri</i>	745 (65.9%) ^a	330 (63.6%) ^b	415 (67.9%)	84 (65.1%)	47 (51.7%)	80 (69.0%)	26 (63.4%)	63 (60.0%)	30 (81.1%)	115 (52.5%)
<i>S. flexneri</i> ^d serotypes/subserotypes										
1a	3 (0.3%) ^a	1 (0.2%) ^b	2 (0.3%) ^c	1 (0.8%) ^c	0	0	0	0	0	0
1b	85 (7.5%) ^a	55 (10.6%)	30 (4.9%)	12 (9.3%)	1 (1.1%) ^c	15 (12.9%) ^c	8 (19.5%) ^c	15 (14.3%) ^c	4 (10.8%) ^c	19 (8.7%)
2a	228 (20.2%) ^a	101 (19.5%)	127 (20.8%)	21 (16.3%)	24 (26.4%)	35 (30.2%)	5 (12.2%)	2 (1.9%)	14 (37.8%)	21 (9.6%)
2b	123 (10.9%) ^a	12 (2.3%)	111 (18.2%)	0	0	4 (3.5%)	4 (9.8%)	4 (3.8%)	0	9 (4.1%)
3a	106 (9.4%) ^a	47 (9.0%)	59 (9.7%)	12 (9.3%)	11 (12.1%)	5 (4.3%)	2 (4.9%)	14 (13.3%)	3 (8.1%)	17 (7.8%)
3b	1 (0.1%) ^a	0	1 (0.2%)	0	0	0	0	0	0	0
4a	33 (2.9%) ^a	19 (3.7%)	14 (2.3%)	9 (7.0%)	4 (4.4%)	0	1 (2.4%)	5 (4.8%)	0	6 (2.7%)
4b	0	0	0	0	0	0	0	0	0	0
5a	0	0	0	0	0	0	0	0	0	0
5b	3 (0.3%) ^a	0	3 (0.5%)	0	0	0	0	0	0	1 (0.5%)
6	124 (11.0%) ^a	70 (13.5%)	54 (8.9%)	23 (17.8%)	5 (5.5%)	12 (10.3%)	4 (9.8%)	19 (18.1%)	7 (18.9%)	35 (16.0%)
7a ^e	23 (2.0%) ^a	13 (2.5%)	10 (1.6%)	6 (4.7%)	2 (2.2%)	1 (0.9%)	0	4 (3.8%)	0	6 (2.7%)
7b ^e	0	0	0	0	0	0	0	0	0	0
X	11 (1.0%) ^a	11 (2.1%)	0	0	0	7 (6.0%)	2 (4.9%)	0	2 (5.4%)	1 (0.5%)
Y	5 (0.4%) ^a	1 (0.2%)	4 (0.7%)	0	0	1 (0.9%)	0	0	0	0

The distribution of species among the Bangladesh isolates was significantly different ($P = .015$) from the composite of the other 6 GEMS sites. The percentage of the various *S. flexneri* serotypes and subserotypes isolated in Bangladesh was significantly different from the percentage at the other 6 sites for the greater percentage of *S. flexneri* 2b in Bangladesh ($P < .0001$) and the lower percentages of *S. flexneri* 1b ($P = .0003$), *S. flexneri* 6 ($P = .015$), and *S. flexneri* X ($P = .0002$).

Abbreviation: GEMS, Global Enteric Multicenter Study.

^a Percent of all 1130 case isolates from the composite of all 7 sites.

^b Percent of the total 519 case isolates from the 6 GEMS sites other than Bangladesh.

^c Percent of the total isolates from the individual GEMS site.

^d Bolded *S. flexneri* serotypes/subserotypes are those proposed for inclusion, along with *S. sonnei*, in a quadrivalent broad-spectrum *Shigella* vaccine.

^e *S. flexneri* 7 strains were previously referred to as *Shigella* "provisional 88-893," "provisional Y394," or "*S. flexneri* 1c."

Table 2. Prevalence of *Shigella sonnei* and *S. flexneri* Serogroups and Proposed Vaccine Component Serotypes of *S. flexneri* Among *Shigella* Isolates From Global Enteric Multicenter Study Cases by Year of the Study

Serogroup, Serotype, or Subserotype	All 7 GEMS Sites			6 GEMS Sites Other Than Bangladesh			Bangladesh		
	Year 1	Year 2	Year 3	Year 1	Year 2	Year 3	Year 1	Year 2	Year 3
Total isolates	457	345	328	214	142	163	243	203	165
<i>S. sonnei</i>	94 (20.6%) ^a	76 (22.0%) ^a	98 (29.9%) ^a	53 (24.8%) ^b	27 (19.0%) ^b	39 (23.9%) ^b	41 (16.9%) ^c	49 (24.1%) ^c	59 (35.8%) ^c
<i>S. flexneri</i>	317 (69.4%) ^a	231 (67.0%) ^a	197 (60.1%) ^a	134 (62.6%) ^b	94 (66.2%) ^b	102 (62.6%) ^b	183 (75.3%) ^c	137 (67.5%) ^c	95 (57.6%) ^c
<i>S. flexneri</i> + <i>S. sonnei</i>	411 (89.9%) ^a	307 (89.0%) ^a	295 (89.9%) ^a	187 (87.4%) ^b	121 (85.2%) ^b	141 (86.5%) ^b	224 (92.2%) ^c	186 (91.6%) ^c	154 (93.3%) ^c
<i>S. flexneri</i> 2a, 3a and 6	183	144	131	97	54	67	86	90	64
As % of all <i>S. flexneri</i>	57.7% ^d	62.3%	66.5%	72.4% ^e	57.5%	65.7%	47.0% ^f	65.7%	67.4%
As % of all isolates	40.0% ^a	41.7%	39.9%	45.3% ^b	38.0%	41.1%	35.4% ^c	44.3%	38.8%
<i>S. sonnei</i> + <i>S. flexneri</i> 2a, 3a and 6	277 (60.6%) ^a	220 (63.8%)	229 (69.8%)	150 (70.1%) ^b	81 (57.0%)	106 (65.0%)	127 (52.3%) ^c	139 (68.5%)	123 (74.6%)
<i>S. sonnei</i> + all <i>S. flexneri</i> serotypes other than <i>S. flexneri</i> 7a	403 (88.2%) ^a	296 (85.8%)	291 (88.7%)	182 (85.1%) ^b	116 (81.7%)	138 (84.7%)	221 (91.0%) ^c	180 (88.7%)	153 (92.7%)

Abbreviation: GEMS, Global Enteric Multicenter Study.

^a Percent of all case isolates for particular study year for all 7 GEMS sites.

^b Percent of all case isolates for particular study year for 6 GEMS sites other than Bangladesh.

^c Percent of all case isolates for particular study year for Bangladesh.

^d Percent of all *S. flexneri* isolates for particular study year for all 7 GEMS sites.

^e Percent of all *S. flexneri* isolates for particular study year for 6 GEMS sites other than Bangladesh.

^f Percent of all *S. flexneri* isolates for particular study year for Bangladesh.

One obstacle to developing serotype-based *Shigella* vaccines is choosing the minimal number from among 50 serotypes. The more serotypes included, the more complex and expensive the vaccine, leading some investigators to pursue common protein vaccines [34]. However, the extensive GEMS serotype data from multiple locations over several years constitutes a hallmark resource to guide vaccine development and provides optimism for serotype-based vaccines.

During GEMS, the fearsome *S. dysenteriae* 1 serotype, which caused protracted pandemics of severe disease from the 1960s to the 1990s in Central America [35], Asia [36], and Africa [37, 38], was not isolated nor were there *S. dysenteriae* 1 outbreaks reported during 2006–2013. This suggests one may exclude *S. dysenteriae* 1 from a multivalent vaccine to prevent endemic shigellosis in developing countries. Nevertheless, should public health authorities deem it critical that a *S. dysenteriae* 1 vaccine be available for future resurgences of pandemic Shiga dysentery, a monovalent vaccine could be stockpiled [39]. Since absence of Shiga disease precludes a controlled field trial, prelicensure efficacy must be demonstrated in alternative ways (eg, volunteer challenges with a nontoxigenic strain) [40].

Although *S. dysenteriae* has 15 and *S. boydii* has 19 distinct serotypes, they accounted for only 5.0% and 5.4%, respectively, of all *Shigella* case isolates. Assuming distributions do not change dramatically over longer time periods, excluding these serotypes from a vaccine would have little impact on breadth of coverage. In contrast, *S. flexneri* serotypes/subserotypes comprised 65.9% of all *Shigella* case isolates, making it imperative that coverage be provided against the most important *S. flexneri* subserotypes. Interestingly, a mere 5 of the 15 currently recognized *S. flexneri* serotypes/subserotypes accounted for 89.4% of *S. flexneri* isolates, including (in rank order) *S. flexneri* 2a, *S. flexneri* 6, *S. flexneri* 2b, *S. flexneri* 3a, and *S. flexneri* 1b. Nevertheless, the relative distribution of *S. flexneri* subserotypes may change over time in various geographic locales. Thus, it will be prudent for *Shigella* vaccines to provide coverage against all 15 *S. flexneri* serotypes/subserotypes.

CVD investigators devised a strategy to achieve broad-spectrum coverage against all *S. flexneri* serotypes except uncommon *S. flexneri* 7a by presenting to the immune system a mix of the following 3 serotypes: *S. flexneri* 2a, *S. flexneri* 3a, and *S. flexneri* 6 [6, 41]. *Shigella flexneri* 6 is common, and its O antigen is distinct from other *S. flexneri*. Indeed, genomic evidence indicates *S. flexneri* 6 might more appropriately be classified as a *S. boydii* serotype; however, for historical and practical reasons, it retains designation as a *S. flexneri* serotype.

Shigella flexneri serotypes/subserotypes other than *S. flexneri* 6 have O antigens that share a common backbone structure that consists of tetrasaccharide repeats of 3 rhamnose residues linked to 1 N-acetylglucosamine [6]. Genes encoding the enzymes that synthesize the tetrasaccharide backbone reside in the *S. flexneri*

Table 3. Twelve Serotypes and Subserotypes of *Shigella flexneri* Not in the Quadrivalent Vaccine and O Group Antigens That They Share With the Vaccine Serotypes

Serotypes in the Quadrivalent <i>Shigella</i> Vaccine (and their O antigens)	12 Serotypes and Subserotypes of <i>S. flexneri</i> Not in the Quadrivalent Vaccine											
	1a	1b	2b	3b	4a	4b	5a	5b	7a	7b	X	Y
<i>S. flexneri</i> 2a (type antigen II and group antigen "3,4")	Group antigen "3,4" ^a	Group antigen "6" ^a	Type antigen II ^a	Group antigen "3,4" ^a	Group antigen "3,4" ^a	Group antigen "6" ^a	Group antigen "3,4" ^a	Group antigen "7,8" ^a	...	Group antigen "6" ^a	Group antigen "7,8" ^a	Group antigen "3,4" ^a
<i>S. flexneri</i> 3a (type antigen III and group antigens "6" and "7,8")	Group antigen "6" ^a	Group antigen "6" ^a	Group antigen "7,8" ^a	Type antigen III ^a	Group antigen "6" ^a	Group antigen "6" ^a	Group antigen "7,8" ^a	Group antigen "7,8" ^a	...	Group antigen "6" ^a	Group antigen "7,8" ^a	Group antigen "7,8" ^a
<i>S. flexneri</i> 6 (type antigen VI)	Group antigen "6" ^a	Group antigen "6" ^a	Group antigen "6" ^a	Group antigen "6" ^a	Group antigen "6" ^a	Group antigen "6" ^a	Group antigen "6" ^a	Group antigen "6" ^a	...	Group antigen "6" ^a	Group antigen "6" ^a	Group antigen "6" ^a

^a *S. flexneri* type and group antigens that are shared with the *S. flexneri* serotypes/subserotypes that are in the vaccine and that constitute the immunologic basis for cross protection.

chromosomal *rfb* locus. *Shigella flexneri* Y's O antigen consists of tetrasaccharide repeats without further modifications. However, lysogenic bacteriophages that encode enzymes able to decorate the tetrasaccharide backbone at specific sites with O-acetyl or D-glucose moieties create new epitopes or ablate others and result in modified saccharide structures that represent the other *S. flexneri* serotypes [6]. The epitopes created by these O-acetyl and D-glucose groups also constitute group antigens shared among different *S. flexneri* serotypes. If the shared group and type-specific antigens induce cross-protective immunity, the number of subserotypes required for a broadly effective *Shigella* vaccine can be minimized. Thus, a multivalent vaccine that includes *S. flexneri* 2a and *S. flexneri* 3a (Table 3), in addition to cross protecting against *S. flexneri* 2b (via type 2 antigen) and *S. flexneri* 3b (via type 3 antigen), would provide shared group antigens that could elicit cross protection against *S. flexneri* 1a, 3b, 4a, 5a, and Y (via group antigen 3,4); against *S. flexneri* 1b, 3b, 4b, and 7b (via group antigen 6); and against *S. flexneri* 2b, 5b, and X (via group 7,8). No cross protection could accrue against *S. flexneri* 6, as its O tetrasaccharide structure (rhamnose-rhamnose-D-galactose-N-acetylgalactosamine) is distinct and lacks the group antigens shared by other *S. flexneri* serotypes.

Noriega et al [41] used the guinea pig Sereny keratoconjunctivitis model to measure cross-reacting serological responses and cross protection when animals immunized mucosally with a live bivalent vaccine containing *S. flexneri* 2a and *S. flexneri* 3a were challenged with other *S. flexneri* subserotypes. Significant cross protection was demonstrated against challenge with heterologous serotypes/subserotypes including *S. flexneri* Y, 1b, 2b, and 5b; protection against *S. flexneri* 1a was borderline ($P = .065$). As expected, no cross protection was observed in immunized animals challenged with *S. flexneri* 6 [41].

Following *S. flexneri* 2a illness or vaccination with live oral vaccine expressing *S. flexneri* 2a O antigen, the human immune system mounts cross-reacting antibody responses against other *S. flexneri* serotypes that share type or group antigens [42]. If the cross protection observed in animals can be extrapolated to humans, a multivalent vaccine that includes O antigens of *S. sonnei*, *S. flexneri* 2a, *S. flexneri* 3a, and *S. flexneri* 6 would provide direct coverage against approximately 64% of the GEMS *Shigella* strains, and cross protection could provide up to 88% overall coverage. Indeed, only *S. flexneri* 7a (merely 2.0% of GEMS isolates) lacks any of the mentioned shared group antigens; *S. flexneri* 7b expresses group antigen 6. Excluding *S. flexneri* 7a from a multivalent vaccine would have little impact on global breadth of coverage.

Table 2 displays GEMS serotypes in relation to the proposed quadrivalent vaccine composition (*S. flexneri* 2a, *S. flexneri* 3a, *S. flexneri* 6 plus *S. sonnei*) to estimate breadth of coverage and possible variation over time from the perspective of all 7 GEMS

sites, the site with the most *Shigella* cases (Bangladesh), and the other 6 sites. Only minimal changes in serotypes are seen from year to year; matching of serotypes between circulating strains and vaccine composition would provide 52%–75% direct protection and, with shared group antigens, 82%–93% coverage can be achieved via cross protection.

One other multicenter study used systematic surveillance to detect *Shigella* cases, quantify the burden of shigellosis, and identify serotypes [2]. Von Seidlein et al [2] used a different definition of diarrheal illness as the eligibility criterion for enrollment, obtained strains from older subjects as well as children aged <5 years, maintained surveillance for different time periods (1–3 years, depending on the site), and worked only in Asia (China, Thailand, Vietnam, Indonesia, Bangladesh, and Pakistan) but used microbiological methods similar to GEMS and similarly included sites in Pakistan and Bangladesh. Among the total 2927 *Shigella* isolates reported by Von Seidlein et al [2], 90% were either *S. flexneri* (68%) or *S. sonnei* (22%), similar to GEMS; 51% of their *S. flexneri* isolates were *S. flexneri* 2a, *S. flexneri* 3a, or *S. flexneri* 6. Importantly, like GEMS, they found no *S. dysenteriae* 1 among their 110 *S. dysenteriae* isolates [2]. Via direct or via shared group antigens, the quadrivalent vaccine would cover at least 84.7% of the 2819 fully serotyped *Shigella* strains isolated by Von Seidlein et al [2].

Preclinical and clinical evidence indicates that ShET1 contributes to the watery diarrhea observed early in *S. flexneri* 2a clinical illness and to diarrheal adverse reactions associated with certain live oral vaccines [26, 27, 43]; deleting *set* and *sen* diminishes vaccine reactogenicity [43, 44]. We confirmed that ShET1 genes are common in *S. flexneri* 2a and 2b isolates (117/119, 98.3%) but rare among other *S. flexneri* subserotypes (4/137, 2.9%) or other species (2/132, 1.5%).

Serotyping of the GEMS *Shigella* isolates offers optimism that a quadrivalent vaccine containing *S. sonnei* and 3 serotype/subserotypes of *S. flexneri* (*S. flexneri* 2a, *S. flexneri* 3a, and *S. flexneri* 6) can provide broad coverage against *Shigella*, which causes the majority of endemic pediatric shigellosis in the developing world, and also can provide broad coverage for travelers [45, 46].

Notes

Acknowledgments. The authors acknowledge the exceptional serotyping prowess of Ms Evangeline Sowers at the Centers for Disease Control and Prevention (CDC).

Disclaimer. The findings and conclusions in this report are those of the author(s) and do not necessarily represent the views of the CDC.

Financial support. This work was supported by grant 38774 from the Bill & Melinda Gates Foundation to M. M. L.

Potential conflicts of interest. All authors: No potential conflicts of interest.

All authors have submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest. Conflicts that the editors consider relevant to the content of the manuscript have been disclosed.

References

1. Kotloff KL, Nataro JP, Blackwelder WC, et al. Burden and aetiology of diarrhoeal disease in infants and young children in developing countries (the Global Enteric Multicenter Study, GEMS): a prospective, case-control study. *Lancet* **2013**;382:209–22.
2. von Seidlein L, Kim DR, Ali M, et al. A multicentre study of *Shigella* diarrhoea in six Asian countries: disease burden, clinical manifestations, and microbiology. *PLoS Med* **2006**; 3:e353.
3. Lindsay B, Ochieng JB, Ikumapayi UN, et al. Quantitative PCR for detection of *Shigella* improves ascertainment of *Shigella* burden in children with moderate-to-severe diarrhea in low-income countries. *J Clin Microbiol* **2013**; 51:1740–6.
4. Barry EM, Pasetti MF, Szein MB, Fasano A, Kotloff KL, Levine MM. Progress and pitfalls in *Shigella* vaccine research. *Nat Rev Gastroenterol Hepatol* **2013**; 10:245.
5. Levine MM. Enteric infections and the vaccines to counter them: future directions. *Vaccine* **2006**; 24:3865–73.
6. Levine MM, Kotloff KL, Barry EM, Pasetti MF, Szein MB. Clinical trials of *Shigella* vaccines: two steps forward and one step back on a long, hard road. *Nat Rev Microbiol* **2007**; 5:540–53.
7. Foster RA, Carlin NI, Majcher M, Tabor H, Ng LK, Widmalm G. Structural elucidation of the O-antigen of the *Shigella flexneri* provisional serotype 88–893: structural and serological similarities with *S. flexneri* provisional serotype Y394 (1c). *Carbohydr Res* **2011**; 346:872–6.
8. Formal SB, Oaks EV, Olsen RE, Wingfield Eggleston M, Snoy PJ, Cogan JP. Effect of prior infection with virulent *Shigella flexneri* 2a on the resistance of monkeys to subsequent infection with *Shigella sonnei*. *J Infect Dis* **1991**; 164:533–7.
9. DuPont HL, Hornick RB, Snyder MJ, Libonati JP, Formal SB, Gangarosa EJ. Immunity in shigellosis. II. Protection induced by oral live vaccine or primary infection. *J Infect Dis* **1972**; 125:12–6.
10. Herrington DA, Van de Verg L, Formal SB, et al. Studies in volunteers to evaluate candidate *Shigella* vaccines: further experience with a bivalent *Salmonella typhi-Shigella sonnei* vaccine and protection conferred by previous *Shigella sonnei* disease. *Vaccine* **1990**; 8:353–7.
11. Kotloff KL, Nataro JP, Losonsky GA, et al. A modified *Shigella* volunteer challenge model in which the inoculum is administered with bicarbonate buffer: clinical experience and implications for *Shigella* infectivity. *Vaccine* **1995**; 13:1488–94.
12. Ferreccio C, Prado V, Ojeda A, et al. Epidemiologic patterns of acute diarrhea and endemic *Shigella* infections in a poor periurban setting in Santiago, Chile. *Am J Epidemiol* **1991**; 134:614–27.
13. Cohen D, Green MS, Block C, Slepion R, Ofek I. Prospective study of the association between serum antibodies to lipopolysaccharide O antigen and the attack rate of shigellosis. *J Clin Microbiol* **1991**; 29:386–9.
14. Cohen D, Green MS, Block C, Rouach T, Ofek I. Serum antibodies to lipopolysaccharide and natural immunity to shigellosis in an Israeli military population. *J Infect Dis* **1988**; 157:1068–71.
15. Mel DM, Terzin AL, Vuksic L. Studies on vaccination against bacillary dysentery. 3. Effective oral immunization against *Shigella flexneri* 2a in a field trial. *Bull WHO* **1965**; 32:647–55.
16. Mel DM, Arsic BL, Nikolic BD, Radovanovic ML. Studies on vaccination against bacillary dysentery. 4. Oral immunization with live monotypic and combined vaccines. *Bull WHO* **1968**; 39:375–80.
17. Mel DM, Gangarosa EJ, Radovanovic ML, Arsic BL, Litvinjenko S. Studies on vaccination against bacillary dysentery. 6. Protection of children by oral immunization with streptomycin-dependent *Shigella* strains. *Bull WHO* **1971**; 45:457–64.
18. Mel DM, Arsic BL, Radovanovic ML, Litvinjenko S. Live oral *Shigella* vaccine: vaccination schedule and the effect of booster dose. *Acta Microbiol Acad Sci Hung* **1974**; 21:109–14.
19. Cohen D, Ashkenazi S, Green MS, et al. Double-blind vaccine-controlled randomised efficacy trial of an investigational *Shigella sonnei* conjugate vaccine in young adults. *Lancet* **1997**; 349:155–9.
20. Passwell JH, Ashkenzi S, Banet-Levi Y, et al. Age-related efficacy of *Shigella* O-specific polysaccharide conjugates in 1–4-year-old Israeli children. *Vaccine* **2010**; 28:2231–5.
21. Levine MM, Kotloff KL, Nataro JP, Muhsen K. The Global Enteric Multicenter Study (GEMS): Impetus, rationale, and genesis. *Clin Infect Dis* **2012**; 55(suppl 4):S215–24.
22. Panchalingam S, Antonio M, Hossain A, et al. Diagnostic microbiologic methods in the GEMS-1 case/control study. *Clin Infect Dis* **2012**; 55(suppl 4):S294–302.
23. Farag TH, Nasrin D, Wu Y, et al. Some epidemiologic, clinical, microbiologic, and organizational assumptions that influenced the design and performance of the Global Enteric Multicenter Study (GEMS). *Clin Infect Dis* **2012**; 55(suppl 4):S225–31.
24. Kotloff KL, Blackwelder WC, Nasrin D, et al. The Global Enteric Multicenter Study (GEMS) of diarrheal disease in infants and young children in developing countries: epidemiologic and clinical methods of the case/control study. *Clin Infect Dis* **2012**; 55(suppl 4):S232–45.
25. Morris GK, Koehler JA, Gangarosa EJ, Sharrar RG. Comparison of media for direct isolation and transport of *Shigellae* from fecal specimens. *Appl Microbiol* **1970**; 19:434–7.
26. Fasano A, Noriega FR, Maneval DR Jr, et al. *Shigella* enterotoxin 1: an enterotoxin of *Shigella flexneri* 2a active in rabbit small intestine in vivo and in vitro. *J Clin Invest* **1995**; 95:2853–61.
27. Fasano A, Noriega FR, Liao FM, Wang W, Levine MM. Effect of *Shigella* enterotoxin 1 (ShET1) on rabbit intestine in vitro and in vivo. *Gut* **1997**; 40:505–11.
28. Noriega FR, Liao FM, Formal SB, Fasano A, Levine MM. Prevalence of *Shigella* enterotoxin 1 among *Shigella* clinical isolates of diverse serotypes. *J Infect Dis* **1995**; 172:1408–10.
29. Roy S, Thanasekaran K, Dutta Roy AR, Sehgal SC. Distribution of *Shigella* enterotoxin genes and secreted autotransporter toxin gene among diverse species and serotypes of *Shigella* isolated from Andaman Islands, India. *Trop Med Int Health* **2006**; 11:1694–8.
30. Farag TH, Faruque AS, Wu Y, et al. Housefly population density correlates with shigellosis among children in Mirzapur, Bangladesh: a time series analysis. *PLoS Negl Trop Dis* **2013**; 7:e2280.
31. Hyma KE, Lacher DW, Nelson AM, et al. Evolutionary genetics of a new pathogenic *Escherichia* species: *Escherichia albertii* and related *Shigella boydii* strains. *J Bacteriol* **2005**; 187:619–28.
32. Al-Hasani K, Rajakumar K, Bulach D, Robins-Browne R, Adler B, Sakkellaris H. Genetic organization of the she pathogenicity island in *Shigella flexneri* 2a. *Microb Pathog* **2001**; 30:1–8.
33. Coster TS, Hoge CW, Van DeVerg LL, et al. Vaccination against shigellosis with attenuated *Shigella flexneri* 2a strain SC602. *Infect Immun* **1999**; 67:3437–43.
34. Martinez-Becerra FJ, Kissmann JM, Diaz-McNair J, et al. Broadly protective *Shigella* vaccine based on type III secretion apparatus proteins. *Infect Immun* **2012**; 80:1222–31.
35. Gangarosa EJ, Perera DR, Mata LJ, Mendizabal-Morris C, Guzman G, Reller LB. Epidemic Shiga bacillus dysentery in Central America. II. Epidemiologic studies in 1969. *J Infect Dis* **1970**; 122:181–90.
36. Rahaman MM, Khan MM, Aziz KMS, Islam MS, Kibriya AK. An outbreak of dysentery caused by *Shigella dysenteriae* type 1 on a Coral Island in the Bay of Bengal. *J Infect Dis* **1975**; 132:15–9.
37. Ebright JR, Moore EC, Sanborn WR, Schaberg D, Kyle J, Ishida K. Epidemic Shiga bacillus dysentery in Central Africa. *Am J Trop Med Hyg* **1984**; 33:1192–7.
38. Birmingham ME, Lee LA, Ntakibirora M, Bizimana F, Deming MS. A household survey of dysentery in Burundi: implications for the current pandemic in sub-Saharan Africa. *Bull World Health Organ* **1997**; 75:45–53.
39. Martin S, Costa A, Perea W. Stockpiling oral cholera vaccine. *Bull World Health Organ* **2012**; 90:714.
40. Samandari T, Kotloff KL, Losonsky GA, et al. Production of IFN-gamma and IL-10 to *Shigella* invasins by mononuclear cells from

- volunteers orally inoculated with a shiga toxin-deleted *Shigella dysenteriae* type 1 strain. *J Immunol* **2000**; 164:2221–32.
41. Noriega FR, Liao FM, Maneval DR, Ren S, Formal SB, Levine MM. Strategy for cross-protection among *Shigella flexneri* serotypes. *Infect Immun* **1999**; 67:782–8.
 42. Van de Verg LL, Bendiuk NO, Kotloff K, et al. Cross-reactivity of *Shigella flexneri* serotype 2a O antigen antibodies following immunization or infection. *Vaccine* **1996**; 14:1062–8.
 43. Kotloff KL, Pasetti MF, Barry EM, et al. Deletion in the *Shigella* enterotoxin genes further attenuates *Shigella flexneri* 2a bearing guanine auxotrophy in a phase 1 trial of CVD 1204 and CVD 1208. *J Infect Dis* **2004**; 190:1745–54.
 44. Kotloff KL, Simon JK, Pasetti MF, et al. Safety and immunogenicity of CVD 1208S, a live, oral *AguaBA Äsen Äset Shigella flexneri* 2a vaccine grown on animal-free media. *Human Vaccines* **2007**; 3:268–75.
 45. Hyams KC, Bourgeois AL, Merrell BR, et al. Diarrheal disease during Operation Desert Shield. *N Engl J Med* **1991**; 325:1423–8.
 46. Thornton SA, Sherman SS, Farkas T, Zhong W, Torres P, Jiang X. Gastroenteritis in US marines during Operation Iraqi Freedom. *Clin Infect Dis* **2005**; 40:519–25.