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## Immune Response to Tissue Restricted Self-Antigens Induces Airway Inflammation and Fibrosis Following Murine Lung Transplantation

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### Abstract

Immune responses against lung-associated self-antigens (self-Ags) are hypothesized to play a role in the development of chronic lung graft rejection. We determined whether immune responses to lung self-Ags, K-alpha-1-tubulin (K $\alpha$ 1T) and Collagen V (Col-V) in the absence of alloimmunity, could promote airway inflammation and fibrosis. Following syngeneic murine orthotopic lung transplantation (LTx) we administered antibodies (Abs) to either K $\alpha$ 1T or Col-V or in combination to both of these self-Ags. As compared to recipients of isotype control Abs K $\alpha$ 1T Abs and/or Col-V Abs-treated recipients had marked lung graft cellular infiltration and bronchiolar fibrosis. This inflammation was also associated the accumulation of K $\alpha$ 1T and Col-V specific IFN- $\gamma$ + and IL-17+ T cells. Notably, the administration of Abs to K $\alpha$ 1T led to cellular and humoral immune responses to Col-V prior to development of fibrosis, and vice versa, indicating that epitope spreading can occur rapidly in an alloantigen independent manner. Collectively, these data support a model of chronic lung transplant rejection where the progressive loss of self-tolerance through epitope spreading promotes airway fibrosis. Strategies that target autoreactive Abs may be useful to inhibit chronic rejection of lung grafts.

### INTRODUCTION

Long term outcomes following lung transplantation (LTx) remain poor due to development of chronic rejection (1, 2), clinically diagnosed as bronchiolitis obliterans syndrome (BOS). BOS is a fibro-proliferative process characterized by progressive decline in lung function. Several immunological and non-immunological factors have been attributed to BOS (3–7). The link between alloimmunity and chronic rejection is well recognized. This relationship is best exemplified by the finding that acute rejection is a major risk factor for chronic rejection (8). We demonstrated that antibodies (Abs) against self-antigens (self-Ags) such as K-alpha-1tubulin (K $\alpha$ 1T) and Collagen V (Col-V) often precede development of rejection

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#### Disclosure

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(9). We also reported that preemptive Ab depletion in patients with detectable donor specific antibodies (DSA) post-LTx in having normal lung function lowers the risk for chronic rejection (6). However, some patients still developed BOS, despite clearance of DSA. These patients had persistence of Abs to self-Ags. On the other hand, in patients where both DSA and Abs to self-Ags were cleared, there was freedom from BOS suggesting self-Ags may play a pivotal role in the development of chronic rejection.

A link between alloimmunity and immune responses to self-Ags and chronic rejection has been proposed (9, 10). Earlier studies demonstrated that Abs to K $\alpha$ 1T can bind to epithelial cells, activate pro-inflammatory and pro-fibrotic growth factor signaling (11). Oral tolerance to Col-V has been shown to prevent rejection in rat lung allografts (12). Hence, we postulated that immune responses to self-Ags alone may play a pathogenic role for development of chronic lung rejection.

To define the effects of immune responses to self-Ags in the absence of alloimmune responses, we performed syngeneic mouse LTx (13). Syngeneic grafts have no evidence of inflammation for greater than 45 days whereas allografts were rejected by day 7 (13). Our results indicate that administration of Abs to lung associated self-Ags can lead to both cellular and humoral immune responses to other self-Ags expressed in lungs leading to inflammation and fibrosis in the transplanted lung.

## MATERIALS AND METHODS

### Animals and LTx

Six to eight week old male C57Bl/6 (H-2k<sup>b</sup>) were obtained (Jackson Laboratories, Bar Harbor, ME). Orthotopic left LTx was performed using cuff technique (13). For sham experiments, mice were ventilated for 1 hour (duration of mouse LTx). All animal studies performed with sterile precautions and approved by the Animal Studies Committee at Washington University School of Medicine.

### Antibodies to K $\alpha$ 1T and Col-V

Rabbit polyclonal IgG Abs to K $\alpha$ 1T and Col-V were produced against K $\alpha$ 1T and Col-V proteins. Analysis of the specificity of the Abs were done by ELISA with plates coated with purified proteins (Col-V, Col-I and Col-II) (optical densities for Col-V: 0.863, Col-I: 0.124 and Col-II: 0.109). Purified Abs were endotoxin free by limulus amoebocyte lysate assay. Abs to K $\alpha$ 1T or Col-V or both (n=5 per group) were administered intraperitoneally following LTx and to sham surgery mice (200 $\mu$ g/dose) on days 0 and weekly thereafter. Rabbit IgG was used as control.

### Histology

Mice were sacrificed on day 45 following LTx. Sections were stained with hematoxylin-eosin and trichrome and analyzed blindly. Images were obtained on a Nikon Eclipse microscope (Nikon), and morphometric analysis performed using Nikon Elements software (Nikon).

### **Enzyme Linked Immunosorbent Assay (ELISA) for auto-Abs**

Development of Abs to self-Ags K $\alpha$ 1T, Col-V, Col-I and Col-II was determined by ELISA using 30 and 45 days post-transplant sera (14). Concentration of Abs was calculated from a standard curve of known concentration of Abs and expressed as concentration in  $\mu\text{g/mL}$ .

### **Enzyme-linked immunospot (ELISpot) for cellular immune responses to K $\alpha$ 1T and Col-V**

Cellular immune responses to self-Ags K $\alpha$ 1T, Col-I and Col-V and control Col-II were enumerated using ELISpot (14). All antigens were endotoxin free. Cells cultured in medium or with phytohemagglutinin were used as negative and positive controls respectively. Results were represented as mean spots per million cells (spm)  $\pm$ SD.

### **Col-V polyclonal antisera preadsorption to Col-I**

1mg of human Col-I (Cell Sciences #CRC194) was added to 1ml of Affigel 15 (Biorad # 153-6051) and incubated 4 hours at 4°C. The beads were washed thrice with PBS (1500rpm for 5 min). Four mg of anti-Col-V was added to the beads loaded with Col-I protein and incubated overnight at 4°C, centrifuged (1500rpm for 5 min) at 4°C and the supernatant containing Abs specific to Col-V was collected. 500ng of recombinant Col-V (Sigma #C3657) and Col-I were loaded into 4–20% acrylamide gel and immuno-blotted with pre-absorbed and post-absorbed anti-Col-V. Absorption with Col-I resulted in complete depletion of Col-I Abs.

### **Statistical analysis**

Morphometric analysis of tissue sections was performed using NIS-Elements Software (Nikon, Melville, NY). Analysis was performed on 5 different areas of the lungs and at least 5 fields were analyzed for each and the data presented as the average. Data was analyzed in SPSS v.19 (IBM Inc., Chicago, IL) and GraphPad Prism 5.0 (LaJolla, CA) and represented at mean $\pm$ SD with at least 5 mice in each cohort. ChiSquare tests or Student T-tests were used as appropriate. Significance was set at  $p < 0.05$ .

## **RESULTS**

### **Administration of Abs to lung associated self-Ags (K $\alpha$ 1T and Col-V) induces airway inflammation and fibrosis following syngeneic murine LTx**

To determine whether administration of Abs to lung associated self-Ags in the absence of alloimmunity can lead to the airway fibrosis following murine LTx, Abs to either Col-V or K $\alpha$ 1T or both or control IgG were administered following syngeneic LTx. Syngeneic mouse LTx have demonstrated long-term graft survival (>45 days) with no evidence of rejection (13, 15). In contrast, histopathology of the transplanted lungs on day 45 demonstrated that 4 out of 5 mice that received Abs to both K $\alpha$ 1T and Col-V developed cellular infiltration around bronchioles and fibrosis (Table 1, Figures 1 & 2). Isotype control mice did not demonstrate any lesions (Figure 1) and had >45 days survival. Sham control also did not develop lesions (Figure 1). Lesions were limited to the transplanted left lungs and right lungs were histologically normal suggesting that LTx induced changes are necessary to expose self-Ags so that Abs can bind (16).

### **Pathology is limited to transplanted lungs and does not affect other organs**

To confirm that the administration of Abs to self-Ags following LTx affects the transplanted lung alone, and does not have a systemic response, we analyzed heart, liver and kidney tissues from all mice. None of the other organs, including native right lungs, demonstrated any signs of pathology (data not shown). This demonstrates that Abs to self-Ags induce specific pro-inflammatory and pro-fibrotic response in the transplanted organ alone.

### **Increase in self-reactive CD4+ T cells and Abs in LTx treated with Abs to self-Ags**

To determine the development of cellular responses against self-Ags, splenocytes from syngeneic LTx mice or sham control following Ab administration were isolated on day 45 and stimulated with K $\alpha$ 1T, Col-V, Col-I or Col-II. IFN- $\gamma$ , IL-4, IL-17 secreting cells were enumerated using ELISpot. As shown in Figure 3, administration of Abs to self-Ags K $\alpha$ 1T and Col-V, either alone or in combination, resulted in a significant increase in IL-17 secreting cells to K $\alpha$ 1T and Col-V compared to control and sham mice (in spm $\pm$ SD for Col-V+K $\alpha$ 1T, isotype control and sham K $\alpha$ 1T and Col-V: Col-V - 53 $\pm$ 9 vs. 2 $\pm$ 1 vs. 1 $\pm$ 2, p=0.012; K $\alpha$ 1T - 67 $\pm$ 10 vs. 8 $\pm$ 4 vs. 3 $\pm$ 2, p=0.029). There was also a significant increase in self-Ag specific IFN- $\gamma$  secreting cells (p=0.011) and IL-4 secreting cells (p=0.002) (Figure 3) in Abs administered LTx.

Another important finding is that administration of Abs to one of the self-Ags lead to immune responses to other tissue restricted self-Ags. As shown in Figure 3. Abs to self-Ags induced development of cellular responses (IL-17, IFN- $\gamma$  and IL-4 secreting cells) not only to the self-Ags to which the Ab was specific but also to other self-Ag Col-I. In addition to cellular immune responses, the mice also developed humoral responses to other self-Ags (Figure 4).

To eliminate the possibility the Col-I Ab reactivity within lung grafts was due to cross-reactivity to Col-I in our Col-V antiserum we preabsorbed these polyclonal Abs to Col-I proteins (Figure S1). Col-V antiserum preabsorbed to Col-I was then administered to three syngenic LTx and 30 days later lung grafts were analyzed for lesion development. Similar to recipients that received unabsorbed antiserum (Figure 4 versus Figure S2) recipients or preabsorbed serum also generated lesions. In addition, Abs (Figure S3) as well as IL-17 and IFN- $\gamma$ -mediated T cell responses (data not shown) could be detected against Col-V, K $\alpha$ 1T and Col-I. However, there was no difference in Col-II specific IL-17, IFN- $\gamma$  and IL-4 secreting cells. This demonstrates that Abs to self-Ags following LTx can induce both cellular and humoral immune responses to those self-Ags and also to other lung associated self-Ags in a tissue restricted manner.

One concern is that administration of rabbit Abs could result in the development of Abs against the rabbit proteins that could influence the outcome. However, ELISA performed using the serum from the LTx mice against plates coated with rabbit immunoglobulin (Sigma, St Louis, MO) demonstrated no detectable Abs against the rabbit immunoglobulins (OD 0.084).

## Cellular immune responses to self-Ags precede the development of humoral immune responses

In order to determine the kinetics of cellular and humoral immune responses to self-Ags in the syngeneic murine LTx following administration of Abs to self-Ags we analyzed the cellular immune responses against K $\alpha$ 1T and Col-V on day 7 and 15 post-transplantation by ELISpot (IFN- $\gamma$  and IL-4). Ab responses to K $\alpha$ 1T and Col-V were analyzed on day 7, 15 and 21 by ELISA. As shown in Figure 5A there was significant increases in the frequency of CD4 T cells secreting IFN- $\gamma$  and IL-4 reactive to K $\alpha$ 1T (IFN- $\gamma$ : 74 $\pm$ 46, IL-4: 24 $\pm$ 9) and Col-V (IFN- $\gamma$ : 384 $\pm$ 303, IL-4: 26 $\pm$ 20) at day 15. However, Abs to K $\alpha$ 1T and Col-V in the sera of the mice were detectable only on day 21 post-LTx (Figure 5B). These results demonstrate that cellular immune responses to self-Ags precede humoral immunity.

## Determination of reactivity to individual chains of Collagen V

In order to determine the reactivity of the sera to the individual Col-V chains, 1 $\mu$ g of the purified  $\alpha$ 1 and  $\alpha$ 2 chain of the Col-V (kindly provided by Dr. William Burlingham, University of Wisconsin) was electrophoresed on 4–12% Bis-Tris SDS-PAGE and transferred to a nitrocellulose membrane. The membranes were blocked with 5% skim milk powder and probed with 1:250 diluted serum collected at 0, 7, 14, 21 and 28 days post-LTx following administration of Abs against K $\alpha$ 1T and Col-V and developed with HRP conjugated anti-mouse Ab. Western blot (Figure S4) demonstrated that mice administered with polyclonal Abs against self-Ags developed Abs against the  $\alpha$ 1 chain of the Col-V. There was no reactivity against the  $\alpha$ 2 chain of Col-V. These results demonstrate that Abs developed against the self-Ags in the mouse model of LTx is against the  $\alpha$ 1 chain of Col-V which is similar to that has been seen following human LTx.

## Critical role of T cells in the induction of immune response to self-Ags and development of fibroproliferation

In order to determine the role of T cells in the development of immune responses to self-Ags and development of fibroproliferation, we performed syngeneic LTx in B6.129P2.Tcrb<sup>tm1Mom</sup>Tcrd<sup>tm1Mom</sup>/J mice (Jackson Laboratory, Bar Harbor, MA) and administered Abs to K $\alpha$ 1T and Col-V. Histopathology of the lungs on day 45 demonstrated no evidence of rejection including lack of cellular infiltration and lack of fibroproliferation (Figure 6). Analysis of the serum by ELISA was also negative against the self-Ags (K $\alpha$ 1T and Col-V). Further, T cell responses were undetectable by ELISpot against K $\alpha$ 1T and Col-V. These results demonstrate that T cell responses play a crucial role in the induction of immune response against self-Ags and development of fibroproliferation.

## DISCUSSION

Lung transplantation is lifesaving in patients with end stage pulmonary diseases. However, long term outcomes continue to be affected by chronic rejection (BOS) (1). Both immune and non-immune risk factors have been linked to development of chronic rejection following human LTx. Chronic rejection is characterized by immune mediated tissue inflammatory responses that lead to cellular infiltration, damage to parenchyma leading to fibroproliferative changes around the terminal vessels and bronchioles. There is growing

evidence that besides alloimmune responses (5, 17, 18) immune responses developed against lung associated self-Ags – K $\alpha$ 1T and Col-V (6, 9, 10, 19) may play an important role in the immunopathogenesis of BOS following human LTx. In this study, we demonstrate that immune responses to lung tissue restricted self-Ags alone in the absence of alloimmune responses can induce inflammation and fibroproliferative changes. Administration of Abs to K $\alpha$ 1T, Col-V alone or in combination resulted in cellular infiltration, inflammation and fibrosis around the terminal airways. Further, there was induction of self-Ag specific IL-17 and IFN- $\gamma$  (Figure 3) responses following syngeneic LTx. In our earlier studies we observed that administration of anti-keratin Abs into the native lung does not induce OAD lesions (14).

An important finding is that administration of Abs specific to one of the lung associated self-Ag lead to “spreading” of immune responses to other self-Ags, i.e. mice administered Abs against K $\alpha$ 1T developed both cellular and humoral immune response not only to K $\alpha$ 1T but also to Col-V and Col-I. This was, however, a tissue-restricted response as seen by an absence of immune responses to Col-II, which is not expressed much in the lung tissues as well as the lack of inflammation in other organs. In order to rule out that this is not due to cross reactivity to Col-I in the polyclonal Abs to Col-V administered, we performed absorption of the sera with Col-I. The resulting polyclonal containing only Col-V specificity when administered to murine LTx demonstrated OAD with fibrosis on day 30 and further developed Abs to K $\alpha$ 1T as well as Col-I. This strongly suggests that the induction of immune responses to Col-I is not due to cross reactivity but due to spreading of the immune responses. Further, the development of Ab responses to Col-V was restricted to  $\alpha$ 1 chain of Col-V as seen in human LTx.

In a previous study on LTx patients – presence of pre-transplant Abs to K $\alpha$ 1T or, Col-V increased the risk of PGD and BOS. Thus, when patients develop *de novo* Abs against one of these Ags, the inflammatory response may facilitate the induction of IL-17 and IFN- $\gamma$  response to other self-Ags that may further perpetuate each other leading to chronic rejection. Studies have also demonstrated that immune responses to cardiac myosin play a crucial role in chronic rejection of heart allografts (20, 21) that is mediated through the production of auto-Abs (22). Further, immunization with cardiac myosin can lead to induction of chronic rejection in a syngeneic heart transplant model (23) and modulation of immune responses against cardiac myosin can prolong the heart graft survival (22). Although, in this study, we determined immune responses against only K $\alpha$ 1T, Col-V and Col-I, it is possible that there may be development of autoimmune response against other unknown lung self-Ags.

The mechanisms by which Abs against self-Ags are induced, or how they act once formed are not well understood. Absence of any immunological or pathological changes in sham animals (ventilation alone) suggests that early post-LTx inflammatory response may be necessary. Ischemia reperfusion injury during the transplant may lead to exposure of cryptic self-Ags to the immune system. There is evidence that following rat LTx Col-V is released into bronchoalveolar lavage fluid (16, 24). It is important to note that such a phenomenon is not only limited to allografts but also in syngeneic grafts (16). Hence, we hypothesize that Abs administered to syngeneic LTx have the capacity to bind the exposed self-Ags resulting

in pro-inflammatory and pro-fibrotic responses. The critical role for autoimmune responses in the peri-transplant period influencing such changes is supported by studies in human LTx where pre-existing autoimmunity prior to transplant increased the risk of PGD, alloimmunity and BOS (10).

Th17 cells specific to self-Ags play an important role in development of self-Ags specific immune responses. Although we specifically didn't block Th17, previous studies from our laboratory, and others, have shown that blocking of Th17 can prevent OAD (14, 25). In addition, the balancing role of IFN- $\gamma$  and IL-17 responses in development of self-Ag induced airway fibroproliferative responses is an area of future investigation.

It is still not clear how the immune response spread to other tissue-restricted antigens. We propose that once Abs are bound to one of the self-Ags, it leads to opsonization of the membrane by Ag presenting cells leading to further activation, phenotypic switch leading to immune response to all the antigens present on the phagocytized membrane leading to spreading of immune responses to other self-Ags, and likely to alloantigen's in allografts. These mechanistic studies are currently being investigated.

In conclusion, this study demonstrates that immune responses to lung associated self-Ags in the absence of alloimmunity can lead to airway inflammation and fibrosis in murine syngeneic transplants. This supports the hypothesis that *de novo* development of Abs to lung associated self-Ags following LTx can induce cellular immune responses which predisposes to development of chronic rejection. Strategies to detect and deplete such Abs during post-transplant may represent a novel approach to prevent BOS following human LTx.

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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## Abbreviations

<b>Abs</b>	antibodies
<b>BOS</b>	bronchiolitis obliterans syndrome
<b>Col-V</b>	Collagen V
<b>DSA</b>	donor specific antibodies
<b>K<math>\alpha</math>1T</b>	K-alpha-1-Tubulin
<b>LTx</b>	lung transplant
<b>self-Ags</b>	self-antigens

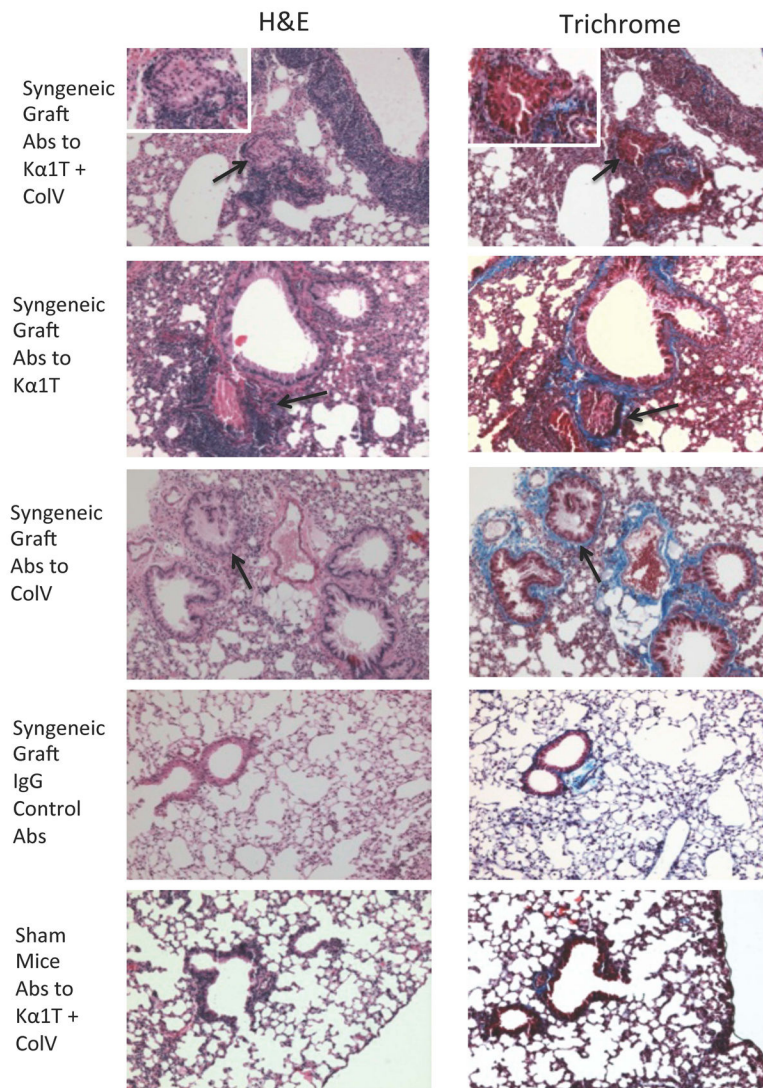


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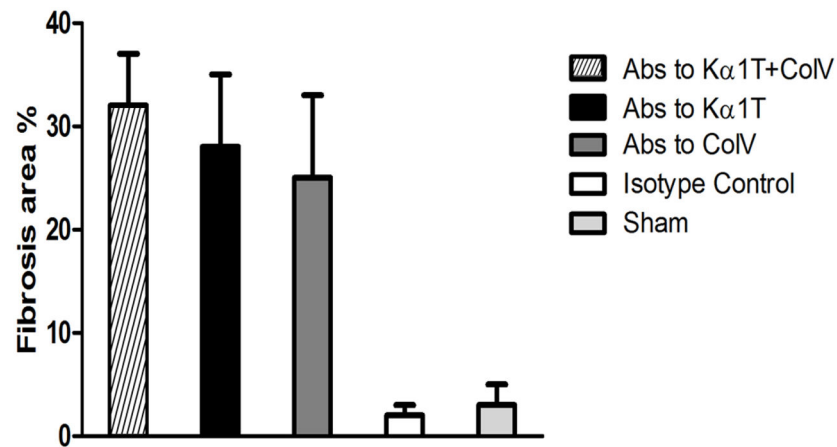
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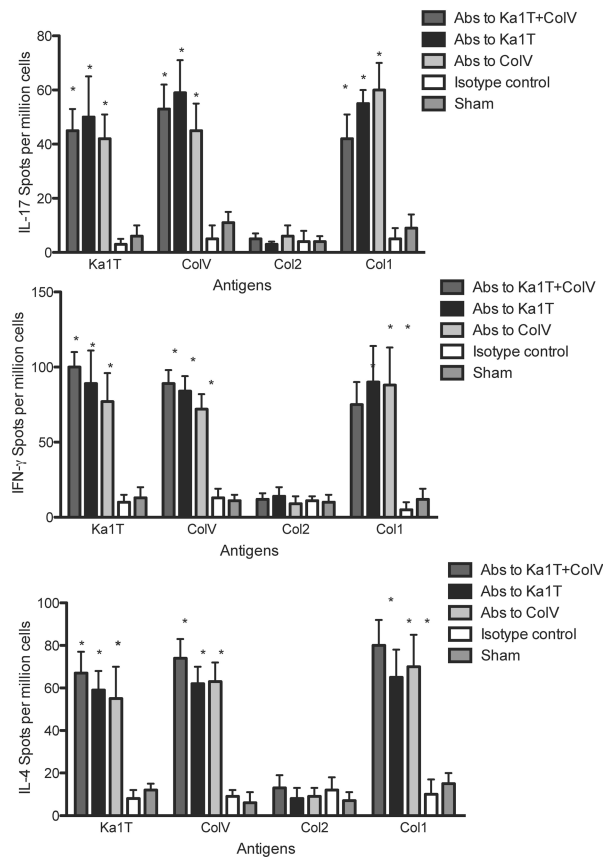
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**Figure 1.** Representative histology on day 45 from mice that underwent syngeneic LTx (B1/6 to B1/6) and were administered Abs to K $\alpha$ 1T and Col-V, or K $\alpha$ 1T alone, or Col-V alone or isotype control Abs, or sham surgery (ventilation alone) mice that were administered Abs to K $\alpha$ 1T and Col-V. Figures on left represent H&E stains and right represent Trichrome stains. Images at 10x magnification and insets at 40x. Five mice in each group.

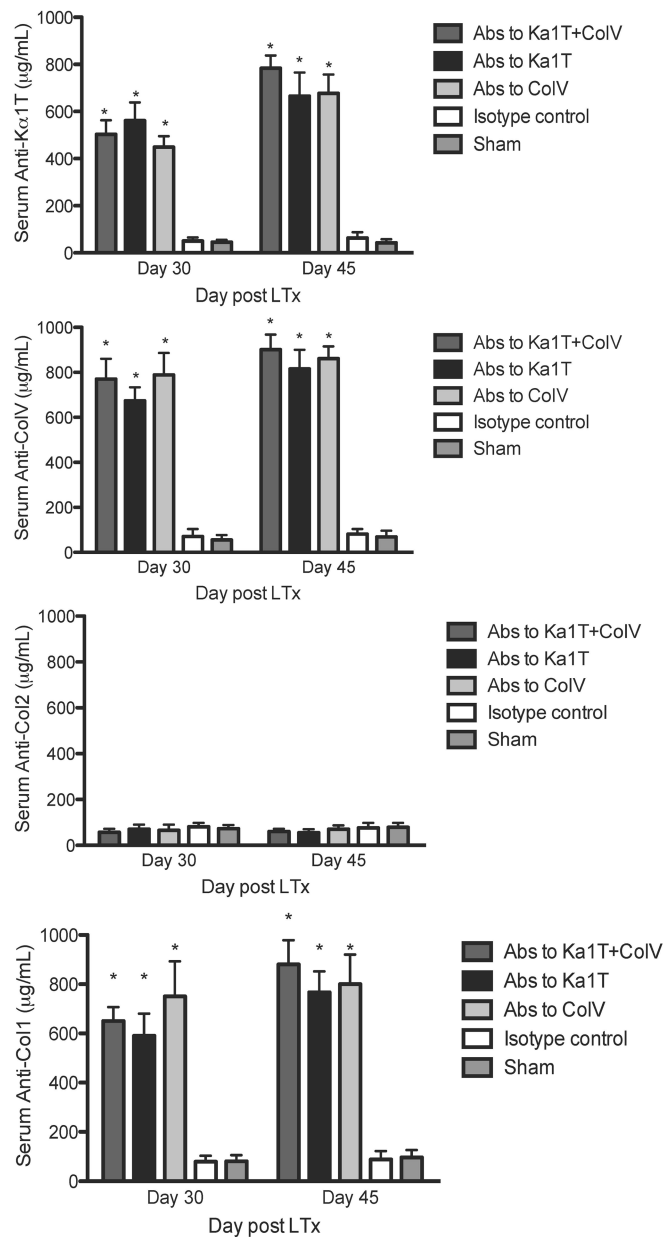


**Figure 2.** Morphometric analysis of lung tissue slides with trichrome stains to quantify extent of fibrosis. Histological images were captured on a Nikon Eclipse microscope and analyzed by Nikon Elements software. Images from transplanted lung tissue in at least 3 different areas for each mouse in each group (total of 5 mice per group) were analyzed to quantify peribronchial fibrotic (blue areas around bronchioles in trichrome stains) per high-power field and represented as a percentage area of total.



**Figure 3.**

Cellular immune responses against self-Ags. IL-17, IFN- $\gamma$ , IL-10 and IL-4 ELISpot responses against Col-V, Ka1T, Col-II or Col-I in mice that underwent syngeneic LTx (B1/6 to B1/6) and were administered Abs to Ka1T and Col-V, or Ka1T alone, or Col-V alone or isotype control Abs, or sham surgery (ventilation alone) mice that were administered Abs to Ka1T and Col-V. Represented as mean spots per million cells of an average of three wells per mice with at least 5 mice in each group. \*represents  $p < 0.05$  compared to isotype control and sham surgery mice.



**Figure 4.** Humoral immune responses against self-Ags. ELISA to determine serum Abs (on day 30 and day 45) against Col-V, Kα1T, Col-II or Col-I in mice that underwent syngeneic LTx (B1/6 to B1/6) and were administered Abs to Kα1T and Col-V, or Kα1T alone, or Col-V alone or isotype control Abs, or sham surgery (ventilation alone) mice that were administered Abs to Kα1T and Col-V. Represented in µg/mL with at least 5 mice in each group. \*represents p<0.05 compared to isotype control and sham surgery mice.

Figure 5A

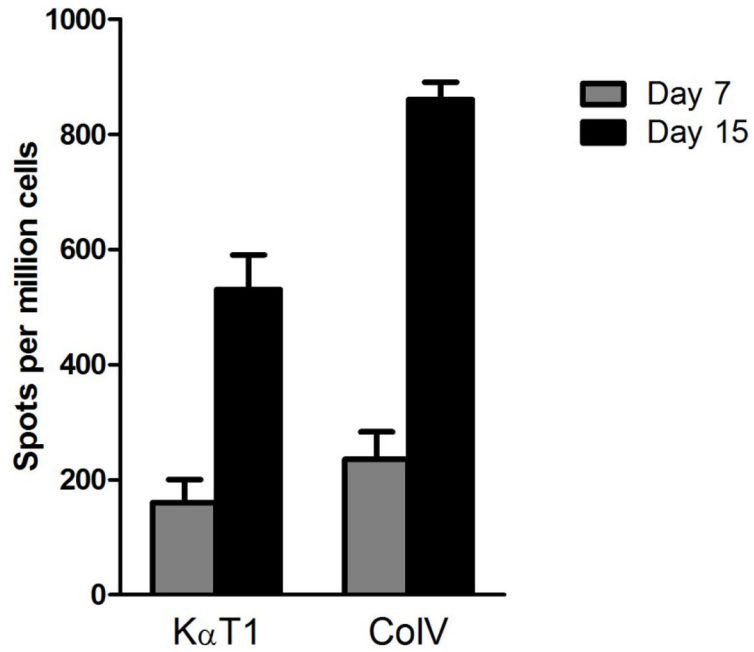
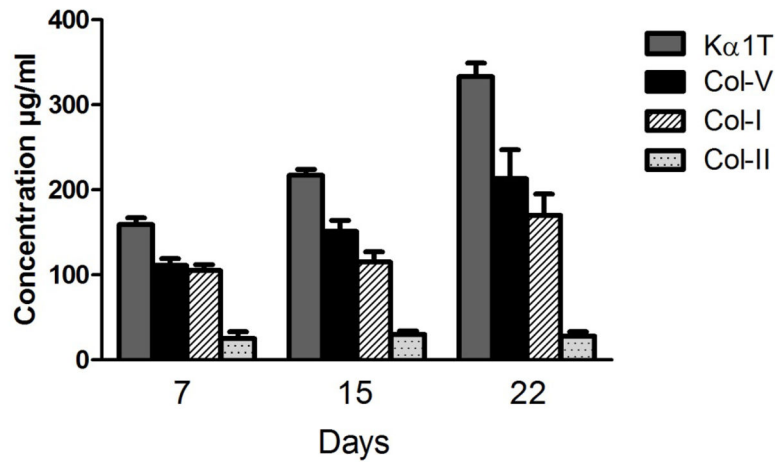


Figure 5B

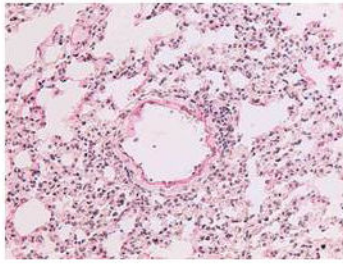
**Figure 5.**

Kinetics of cellular and humoral immune responses against self-Ags. **(A)** IFN- $\gamma$ , and IL-4 ELISpot responses against Col-V, and K $\alpha$ 1T, in mice that underwent syngeneic LTx (B1/6 to B1/6) and were administered Abs to K $\alpha$ 1T and Col-V were analyzed on day 7 and day 15. Represented as mean spots per million cells of an average of three wells per mice with at least 5 mice in each group. **(B)** ELISA was used to determine serum Abs (on day 7,15, 22) against Col-V, K $\alpha$ 1T, Col-II or Col-I in mice that underwent syngeneic LTx (B1/6 to B1/6)

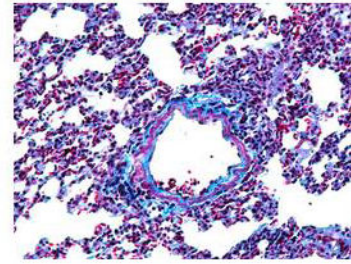


and were administered Abs to K $\alpha$ 1T and Col-V. Represented in  $\mu\text{g/mL}$  with at least 5 mice in each group.

## H&amp;E



## Trichrome

**Figure 6.**

Representative histology on day 45 from mice that underwent syngeneic LTx (B1/6 to B6.192P2.Tcrb<sup>tm1Mom</sup> Tcrd<sup>tm1Mom/J</sup>) and were administered Abs to K $\alpha$ 1T and Col-V. Figures on left represent H&E stains and right represent Trichrome stains. Images at 10x magnification. Five mice in each group.

**Table 1**

Prevalence of airway inflammation and fibrosis in mice at day 45 following administration of either Abs to K $\alpha$ 1T or Col-V or both or isotype control or sham surgery mice

Mouse group	Airway inflammation	Fibrosis
Abs to Col-V+K $\alpha$ 1T	4/5 (80%)	4/5 (80%)
Abs to K $\alpha$ 1T	4/5 (80%)	4/5 (80%)
Abs to Col-V	4/5 (80%)	4/5 (80%)
Isotype control Abs	1/5 (20%)	1/5 (0%)
Sham mice	1/5 (20%)	0/5 (0%)