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The thromboxane synthase and receptor signaling pathway in cancer: an emerging paradigm in cancer progression and metastasis

Prasanna Ekambara,

Department of Oncology, School of Medicine, Wayne State University, Detroit, MI 48202, USA

Bioactive Lipids Research Program (BLRP), Department of Pathology, School of Medicine, Wayne State University, Detroit, MI 48202, USA

Wanyu Lambiv,

Bioactive Lipids Research Program (BLRP), Department of Pathology, School of Medicine, Wayne State University, Detroit, MI 48202, USA

Rosanna Cazzolli,

Division of Perinatal Research, Kolling Institute for Medical Research, University of Sydney, Sydney, NSW, Australia

Anthony W. Ashton, and

Division of Perinatal Research, Kolling Institute for Medical Research, University of Sydney, Sydney, NSW, Australia

Kenneth V. Honn

Department of Oncology, School of Medicine, Wayne State University, Detroit, MI 48202, USA

Bioactive Lipids Research Program (BLRP), Department of Pathology, School of Medicine, Wayne State University, Detroit, MI 48202, USA

Kenneth V. Honn: k.v.honn@wayne.edu

Abstract

Thromboxane A₂ (TXA₂) is a biologically active metabolite of arachidonic acid formed by the action of the terminal synthase, thromboxane A₂ synthase (TXA₂S), on prostaglandin endoperoxide (PGH₂). TXA₂ is responsible for multiple biological processes through its cell surface receptor, the T-prostanoid (TP) receptor. Thromboxane A₂ synthase and TP are the two necessary components for the functioning of this potent bioactive lipid. Thromboxane A₂ is widely implicated in a range of cardiovascular diseases, owing to its acute and chronic effects in promoting platelet aggregation, vasoconstriction, and proliferation. In recent years, additional functional roles for both TXA₂S and TP in cancer progression have been indicated. Increased cyclooxygenase (COX)-2 expression has been described in a variety of human cancers, which has focused attention on TXA₂ as a downstream metabolite of the COX-2-derived PGH₂. Several studies suggest potential involvement of TXA₂S and TP in tumor progression, especially tumor

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Correspondence to: Kenneth V. Honn, k.v.honn@wayne.edu.

cell proliferation, migration, and invasion that are key steps in cancer progression. In addition, the regulation of neovascularization by TP has been identified as a potent source of control during oncogenesis. There have been several recent reviews of TXA_2S and TP but thus far none have discussed its role in cancer progression and metastasis in depth. This review will focus on some of the more recent findings and advances with a significant emphasis on understanding the functional role of TXA_2S and TP in cancer progression and metastasis.

Keywords

Thromboxane synthase; Thromboxane receptor; Cyclooxygenase; Cancer progression; Metastasis; Angiogenesis; Cell migration; Apoptosis

1 Introduction

Thromboxane A2 (TXA2) was one of the first prostaglandins to be identified from washed platelets (in 1975) and has been widely implicated in a range of cardiovascular diseases, owing to its acute and chronic effects in promoting platelet aggregation, vasoconstriction, and proliferation [1–3]. TXA₂ is a biologically active metabolite of arachidonic acid (AA) formed by the action of TXA₂ synthase (TXA₂S) on prostaglandin endoperoxide (PGH₂) [1, 4, 5]. TXA₂ is highly unstable in aqueous solution, where it spontaneously hydrolyzes to the biologically inactive hemiacetal thromboxane B_2 (TXB₂) with a half-life of 30 s [4]. Due to its short half-life, TXA₂ primarily functions as an autocrine or paracrine mediator in the tissues surrounding its site of production. TXA2 is responsible for multiple biological processes through the cell surface TXA₂ receptor, or T-prostanoid (TP) receptor [6, 7]. TXA2 biosynthesis and TP expression are elevated in numerous cardiovascular and inflammatory diseases [3, 8–10]. It is felt that TXA₂, through its receptors, plays a very important role in the pathogenesis of acute coronary artery syndrome, vessel remodeling, thrombosis, renal, pulmonary, and atherosclerotic cardiovascular diseases primarily through its action as a potent vasoconstrictor and an inducer of platelet aggregation and activation [8, 9, 11–15]. As such TXA₂S inhibition and TP antagonism have become central to the therapy of many diseases including infarction, hypertension, stroke, and renal dysfunction [16–18]. Recently a role for TXA₂ signaling in cancer has become apparent. This review will focus on some of the more recent findings and advances with a major emphasis on understanding the functional role of TXA₂S and TP in cancer progression and metastasis.

1.1 Thromboxane A₂ synthase

The cyclooxygenase enzymes, cyclooxygenase (COX)-1 and COX-2, are responsible for the conversion of AA to PGH₂, the first step in the generation of TXA₂. TXA₂S is an endoplasmic reticulum membrane protein that belongs to the P450 epoxygenase family that catalyzes the conversion of the COX product PGH₂ to TXA₂ [19] (Fig. 1). TXA₂S was first found as a microsomal enzyme in platelets (60 kDa) and is highly expressed in lung, platelets, kidney, stomach, duodenum, colon, and spleen [20–22]. TXA₂S expression is reported to be closely associated with cardiovascular, renal, and inflammatory diseases [21, 23].

1.2 Thromboxane A₂ receptor

 TXA_2 is responsible for multiple biological processes through its cell surface receptor TP. Ligation of TP by TXA2 activates multiple downstream pathways, including phospholipase C, and raises intracellular Ca²⁺ levels leading to vasoconstriction and platelet aggregation [6, 21]. The TXA₂ receptor (TP) is a typical G-protein-coupled receptor expressed as two different isoforms in humans— TP-alpha (TPa) and TP-beta (TPβ) [24, 25]. These TP isoforms arise via alternate splicing of a single gene and share the first 328 amino acids. TP α and TP β differ in the length of the C-terminal cytoplasmic tail with TP α shorter than the TPβ isoform (15 versus 79 residues) [24–26] (Fig. 2). Ligand binding sites for the TP receptor are in its extracellular region and are identical in both splice variants. TP isoforms share both common and distinct signaling pathways depending on the G protein subunits bound to their C-terminal tail. Therefore, differences in function between TP α and TP β may manifest due to the complement of G-proteins associated with the cytoplasmic tail. TP receptors are found in a large number of tissues and cell types. TP mRNAs are expressed widely in platelets and monocytes and in the lung, liver, kidney, cardiovascular system (myocytes, vascular smooth muscle cells, and endothelium), uterus, brain, spleen, thymus, and placenta [6, 26-29].

2 Role of thromboxane A₂ and its receptors in cancer progression

In recent years, several studies have indicated functional roles for both TXA₂S and TP in cancer progression. Increased COX-2 expression has been described in a variety of human cancers and downstream metabolites of COX-2, such as TXA₂, have therefore also become of interest for their potential role in cancer progression [30–33]. Multiple studies have documented roles for TXA₂ signaling in the essential processes of neoplastic transformation, including enhanced tumor cell motility that are key steps in cancer progression. These effects are observed in multiple cancers indicating the extensive nature of these effects and the widespread clinical applicability of targeting these pathways as adjunct therapy in cancer [30–32, 34].

2.1 Role of thromboxane A₂ signaling in prostate cancer

Prostate cancer (PCa) is one of the most common cancers among men in the USA and is the second leading cause of death in this population [35]. Strong correlations exist between diets high in fat (especially from red meat) and PCa [36–38]. As red meat is rich in AA, the presence of enzymes associated with AA metabolism might result in increased synthesis of downstream lipid mediators. Several metabolites of AA, such as 5-HETE and 12-HETE, have already been studied widely for their roles in PCa [39–42]. Numerous studies demonstrate increased COX-2 mRNA and protein expression in PCa correlate with poor prognosis [43, 44]. TXA₂ is derived from the COX product PGH₂ and, as a consequence, both TXA₂S and TP have long been hypothesized to have a possible role in prostate cancer progression.

Prostate tumor progression involves several key steps including cell survival, cell migration, cell invasion, and metastasis. Human PCa cells express functionally active TXA_2S and biosynthesize TXA_2 [45]. As the expression of COX-2 and COX-1 in prostate cancer has

been reported previously, Nie and colleagues investigated their role in synthesis of TXA₂ [45]. Treatment of PC-3 cells with both COX-1 and COX-2 inhibitors reduced TXA₂ synthesis by 95%, similar to the level achieved by direct TXA₂S antagonism. These data suggest that human PCa cells express TXA₂S and that both COX-1 and COX-2 are essential for providing the substrate for TXA₂S mediated TXA₂ biosynthesis in PCa cells [45]. Increased TXA₂S expression and activity in PCa cells augmented cell migration but had minimal effect on cell cycle progression or survival indicating that TXA₂S activity might contribute to PCa progression through modulating cell motility [45].

In a study conducted on tissue samples from 46 patients, with well-documented histological and molecular data, increased TXA2S and COX-2 expression were observed in tumors. In the same study, TP expression was higher in malignant tissues (high-grade prostatic intraepithelial neoplasia and cancer glands) [46]. Similarly, TXA₂S mRNA and protein expression were higher in prostate carcinomas compared to matched normal tissues. In contrast, epithelial cells in non-tumoral glands displayed almost non-existent TXA2S and TP expression [46]. The degree of TXA₂S expression correlated with the severity of prostate carcinoma lesions, with advanced stages and poorly differentiated forms having the highest expression levels [45, 46]. Moreover, a significant association between the expression of COX-2/TXA₂S/TP and higher Gleason score/pathologic stage of the tumors was observed [46]. Within the cancer tissue, expression of TXA₂S and TP were localized to areas of perineural invasion, a known mechanism by which PCa cells penetrate the prostatic capsule and spread to other tissues. Strong expression of TXA2S along perineural tracts might be an indicator for its potential involvement in tumor cell invasion and metastasis [45–47]. The enzyme was found to be involved in motility, but not proliferation or survival, of PCa cells [45].

Tumor cell migration is an important step in the metastatic cascade. At this time, tumor cells leave the primary organ, enter the circulation, and colonize distant tissues [48]. Rho GTPases are critical for the dynamic changes in cell shape and adhesion that drive cell migration. Nie and colleagues demonstrated that the TXA₂-TP signaling axis regulated cell migration and cytoskeleton reorganization through promoting Rho-A activation [49]. Recent studies have indicated that the G α 12 family of heterotrimeric G proteins (G α 12 and G α 13) is up-regulated in PCa and that activation of G α 12 signaling promotes PCa cell invasion, through a Rho-dependent pathway. In addition, G α 12 signaling via Rho is required for TXA₂ stimulated invasion of PCa cells [50]. As TP is known to couple to the G α 12 family of heterotrimeric G proteins, the up-regulation of both TP and G α 12 in PCa suggests a possible mechanism by which TP may drive the cell migration and invasion observed in high-grade PCa. This hypothesis needs to be investigated further.

A recent report provided evidence of a novel constitutive interaction between TPa/TP β and protein kinase C-related kinases (PRK1) [51]. PRK1 is a Rho-A effector that has been widely implicated in androgen-associated PCas and ovarian serous carcinomas [51]. It was established that PRK1 directly interacts with endogenously expressed TPa and TP β in both PC-3 and LNCaP cells, and disruption of PRK1 by siRNA substantially impairs cell migration in response to TXA₂ agonist U46619 in these cells [51]. These findings all point towards a possible functional role of TP in cell migration in prostate cancer.

2.2 Role of thromboxane A₂ signaling in breast cancer

Breast cancer is one of the most common cancers among American women and is the second leading cause of death among them in USA [35]. In a cancer-profiling array, TXA₂S mRNA levels were increased in seven of nine breast tumors when compared to their matched normal tissues [45]. A larger study (120 patients) in normal breast and tumor tissues with well-documented histological and molecular data found transcripts of TP and TXA₂S were differentially expressed by quantitative polymerase chain reaction (qPCR) [52]. TXA₂S levels were similar in tumor and normal breast tissues; however, TXA₂S expression was significantly lower in high-grade tumors compared to low-grade tumors. By comparison, higher total TP mRNA expression was observed in the tumor (grade 3 and above) compared with normal mammary tissues [52] and may indicate higher mortality and worse prognosis. TP expression correlated with estrogen receptor status in this study (p=0.0128) but was independent of nodal involvement or primary tumor type (ductal versus lobular) [52]. As in prostate cancer, recent studies suggest the increased expression of Ga12and TP in breast tissue may activate Rho-A to promote cell motility to further exacerbate the progression of breast cancer [53]. These data strongly suggest a role for TXA_2 signaling in the development and progression of breast cancer. Moreover, the association of high TP expression in aggressive tumors with poor prognosis indicates TP may have significant prognostic value in clinical breast cancer.

Abraham et al. analyzed seven prostaglandin pathway genes for single nucleotide polymorphisms that may predispose to breast cancer and found that only PTGIS and TXA₂S polymorphisms showed modest associations [54]. Collectively, these reports suggest that TP may play a role in breast cancer; however, the significance of this role remains uncertain due to the conflicting nature of the reports. The pathogenic role is made less clear by the realization that it is not necessary for both TXA₂S and TP to be expressed in the same tissue to have a possible role in cancer progression. TXA₂ can act as paracrine mediator due to the abundant expression of TXA₂S in platelets and other cell types in the tumor microenvironment. Thus, further pre-clinical investigation into the role of TXA₂ signaling is warranted to define the true role of this bioactive lipid in breast cancer.

2.3 Role of thromboxane A2 signaling in lung cancer

Lung cancer is the second leading cause of cancer in the USA every year and more people die from lung cancer than breast, colon, and prostate cancers combined [35]. The role of COX-2 and prostaglandins in lung cancer is now attracting considerable attention from cancer biologists and the public. A study conducted on 48 samples of non-small cell lung cancer (NSCLC) and matched normal lung tissues observed specific cellular expression patterns of the COX-isoenzymes and terminal synthases of prostanoid synthesis. Increased COX-2 and simultaneous down-regulation of COX-1 expression in NSCLC were identified by immunohistochemistry [55]. Moreover, high levels of TXB₂, the stable metabolite of TXA₂, have been detected in human lung cancer tissues indicating increased TXA₂S activity in lung cancer [56]. An extension of the aforementioned work, in a larger sample size, compared the prostaglandin biosynthesis pathways in small cell lung carcinoma and NSCLC and further correlated their observations with angiogenesis and metastasis [57]. Expression

of TXA₂S, COX-1, COX-2, and microsomal prostaglandin-E synthase were significantly higher in the metastatic cases of NSCLC as compared to non-metastatic cases [57].

TPa expression has been documented in five out of six NSCLC cell lines [58]. Moreover, A549 cells with ectopic TPa expression exhibited greater tumor growth and increased vascularization than the control A549 cells when implanted into nude mice [58]. Over-expression of COX-2 in lung tumors has been widely reported and factors that may cause over-expression of COX-2 in lung tumors are not completely understood. Activation of TPa with the TXA₂ mimetic IBOP induces the expression of COX-2 through activation of four signaling pathways (extracellular signal-regulated kinase (ERK), p38 MAPK, JAK, and β -catenin). In addition, transcription factors such as NFkB, cAMP response element-binding (CREB), C/EBP, and Stat3 are downstream signaling molecules that interact with the COX-2 promoter and play important roles in TPa-mediated expression of COX-2 [59].

Strong evidence suggests that Nurr1 is critical for the proliferation of lung carcinoma cells (H157) through regulation of cyclin D1 expression [60]. Studies have indicated that TP signaling induces Nurr1 expression through a mechanism that bypasses the epidermal growth factor receptor (EGFR) pathway. Moreover, TP agonist-induced proliferation in lung cancer cells promoted cyclin D1 expression through induction of Nurr1 [60]. Nie and colleagues showed that pulmonary metastasis of intravenously injected Lewis lung carcinoma (LLC) is attenuated by TXA₂S inhibitors. Moreover, oral administration of TXA₂S inhibitors CI or furegrelate reduced the number and size of metastatic colonies in the lung after injection of LLC cells [61]. These data confirm that TXA₂S activity is required for development of lung tumor metastasis and that TXA₂S inhibitors are potential antimetastatic agents. These findings shed new light on the contribution of TPα signaling to lung tumorigenesis.

Evading apoptosis (or programmed cell death) is a hallmark of most types of cancer [62] and is a key step in cancer progression. TXA_2S expression inhibits apoptosis in lung cancer cells and TXA_2S inhibition induces apoptosis in NSCLC cells. Moreover, two specific TXA_2 antagonists enhance the effects of cisplatin (a chemotherapeutic agent) [63]. The mechanism by which TXA_2 signaling enhances cell survival and proliferation in lung cancer cells involves reduction of nuclear p27 levels which mitigates apoptosis [64]. While the mechanism inducing apoptosis is not fully defined treating NSCLC cells with the TXA_2S inhibitor 1-BI induces ROS generation and decreases NF- κ B activity and nuclear translocation by decreasing I κ B α phosphorylation [65].

Given the well-established link between smoking and lung cancer and the role for TXA₂ signaling in lung cancer, it will be obvious to examine if TXA₂ signaling mediates the smoking–lung cancer connection. Indeed, it was shown that significantly higher TXB₂ production in lung cancer tissues from smokers compared to cancer tissues from non-smokers [66]. To examine the causal role for TXA₂ signaling in lung cancer, Huang and colleagues recently correlated TXA₂S, TXB₂, and carcinogen 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone (NNK) in smokers and non-smokers. As expected, significantly higher TXA₂S expression was observed in tumor tissues compared to non-tumor tissues from the same patient. A more interesting link to NNK could be inferred from the observation that

significantly fewer tumors from non-smokers showed TXA₂S expression as compared to tumors from smokers. Further, 100% of tissues from smokers (tumors and non-tumoral tissue) were positive for TXA₂S expression unlike non-smokers where 50% of non-tumoral and 25% of tumoral tissues were negative for TXA₂S expression. NNK increased TXA₂S, TXA₂ synthesis, and TP activation *in vivo* and in lung cancer cells *in vitro*. The increased TXA₂ may subsequently activate CREB through the PI3K/Akt and ERK pathways, thereby contributing to the promotion of survival and growth of lung cancer cells by NNK [67]. Moreover, specific TXA₂S inhibitors and TP antagonists abolished growth and induced apoptosis in lung cancer cells [67] indicating the importance of TXA₂- to NNK-induced tumor growth in lung cancer.

These data were sufficiently provocative that Cathcart and colleagues examined the expression of TXA₂S in NSCLC in 204 patients, to identify if TXA₂S was a prognostic and/or survival factor [68]. They also examined the role of TXA₂S in mediating the migration and growth of NSCLC cells *in vitro*. Though they observed no prognostic role for TXA₂S in NSCLC, they confirmed that both the plasma and protein level of TXA₂S and TXB₂ were significantly higher in tumor tissues than the matched "normal" tissues [68]. Equally their model system confirmed that enhanced TXA₂S expression promotes proliferation, motility, invasiveness, and survival of cancer cells. Inhibition of TXA₂S with selective inhibitors induced apoptosis confirming that it is indeed a potential therapeutic target in NSCLC [68].

2.4 Role of thromboxane A₂ signaling in colon cancer

Colorectal cancer is one of the leading causes of cancer-related deaths in the United States [35]; however, early diagnosis often leads to a complete cure. As in the case of prostate cancer, studies have shown strong correlation between high-fat diets, associated with consumption of red meat, and colon cancer [69]. Consumption of foods rich in AA likely increases synthesis of prostaglandins, such as TXA2, in the presence of enzymes associated with AA metabolism. TXA2S and TXA2 biosynthesis are increased in colon cancer and cause detrimental effects by promoting TP signaling [70]. The recognition that COX signaling was a strong pro-cancer signal for colon tumors is not recent and the correlated expression of COX-2 with colon carcinoma [71] has reinforced the causal relationship between the two. Epidemiologic data showed that up to 40% reduction of mortality in colorectal cancer patients with regular nonsteroidal anti-inflammatory drugs (NSAIDs) use as compared to non-NSAID consumers [71]. Gene transfer of TXA₂S increased colon cancer cell growth in vivo and enhanced angiogenesis [72] (see below). Strong evidence of a role for TXA₂S in colorectal carcinoma was provided when a marked over-expression was observed in different grades of colorectal tumors compared to the paired-normal tissues. The same study found increased expression of TXA2S in colon cancer cell lines and that abrogating TXA₂ signaling with TXA₂S inhibitors, TXA₂S anti-sense as well as TP antagonists reduced proliferation of the colon cancer cell lines [73, 74]. qPCR was used to quantify TP transcript expression from 62 tumors and adjacent normal colon tissues. Results indicated that TP was expressed at significantly higher levels in tumors compared to normal tissues but displayed lower levels of TP expression in cultured colorectal cancer cell lines (HT-29 and HCA-7) [73]. The aforementioned studies have started to carve a role for TXA2

signaling in colon cancer; however, the data presented are all from studies with small sample size and need to be validated in a larger cohort of patients with colon cancer.

2.5 Role of thromboxane A₂ signaling in brain cancer

Gliomas are brain malignancies of glial cell origin, such as oligodendroglioma, that are notorious for recurrence and the rate at which they progress to high-grade malignancies [75]. The capacity of glial tumor cells to migrate and diffusely infiltrate normal brain makes the disease notoriously difficult to cure by surgical eradication. Thus, identification of genes associated with invasion may offer novel strategies for anti-invasive therapies. This prompted McDonough and colleagues to assess glioma cell lines with a migration advantage on a glioma-derived extracellular matrix [76]. The outcome was a genetically stable strain that showed a migration advantage, a slightly arrested growth rate, anchorage-independent growth and was comparable to the parental cells in their tumorigenicity in athymic nude mice. This migration-advantaged strain showed increased TXA₂S expression [76]. When treated with TXA₂S antagonists (such as dazmegrel and furegrelate) the migration of the strain with enhanced mobility was normalized to the rate of the parental cells, suggesting direct regulation of cellular motility by TXA₂S [76]. Giese and colleagues showed TXA₂S expression to be present in a large panel of glioma cell lines but not in normal human astrocytes [77]. Moreover, TXA₂S expression was observed in the parenchyma of glial tumors and in reactive astrocytes but not in quiescent astrocytes and oligodendroglia of normal brain. A wide range of TXB₂ formation in glioma cell lines was observed, the relative expression of which correlated with migration rates of these cells. Not surprisingly, TXA₂S-specific inhibitors effectively blocked the migration advantage and decreased intercellular adhesion in glioma cells [77]. This was the first indication that TXA₂ signaling may play a crucial role in glial tumors and represent a novel strategy for anti-invasive therapies.

Giese and colleagues also revisited phospholipid biosynthesis in glioma by assessing the metabolic activity of the upstream enzymes COX-1 and COX-2. In comparison to inhibiting TXA₂S, the inhibition of COX isoforms produced no significant shift in the phenotype of glioma, despite the robust expression of COX in all grades of glioma [77]. This may highlight a functionally antagonistic role for other downstream metabolites of COX in cancer. In addition, TXA₂S inhibition with furegrelate induced caspase activation, DNA fragmentation, and eventual apoptosis only in glioma-derived cells but not normal astrocytes or fibroblast [77]. These same effects of TXA₂S therapy might sensitize the invasive glioma cells, and angiogenic endothelium to apoptosis [77].

Diagnosis of invasive glioma is often too late for anti-invasive therapy to be effective. Thus, the identification of TXA₂S inhibitors as regulators of glioma cell motility is of little consequence unless the invasive cells are also rendered susceptible to cyto-reductive treatments. In this regard TXA₂ antagonism shows great promise as an adjunct to standard therapy. Pre-treatment of glioma cells with furegrelate, a specific TXA₂S inhibitor, increased radiation sensitivity of cultured glioma cells [78]. The present standard of care in glioblastoma (the most malignant grade 4 glioma) is surgical resection followed by

combined chemo-radiation therapy [79]. Schmidt and colleagues presented compelling data indicating that a single administration of furegrelate to *in vivo* orthotopic glioblastoma in mice significantly reduced tumor size, tumor cell proliferation, and angiogenesis and increased apoptotic cell death [80]. Combining TXA₂S inhibition with an alkylating agent (BCNU) also showed a significant survival effect on an ectopic mouse model of glioma [80]. The ultimate proof of principle for the inclusion of TXA₂S inhibition in glioma therapy will be to demonstrate significant improvements to clinically valid treatment regimens (such as radiation therapy followed by temozolomide treatment) in orthotopic mouse models. If successful, TXA₂S inhibition is likely to be quickly implemented as an adjunct therapy in clinical trials, which raises hopes of finding an effective treatment for glioblastoma patients especially for those with unmethylated MGMT promoter [81].

2.6 Role of thromboxane A₂ signaling in bladder cancer

Over the last decade, there has been an increasing awareness of the role of prostanoids in the development and progression of invasive bladder cancer. Susceptibility to invasive bladder cancer correlates with the –765G>C mutation in the COX-2 promoter [82]. This polymorphism results in an under active promoter and lower COX-2 expression. This change likely causes liberated AA to shunt into a COX-1-based pathway and potentially results in greater synthesis of prostaglandins like TXA₂. TXA₂ release is not observed from unstimulated or challenged "normal" urothelial cells [83] and TXA₂S release and TXA₂S expression are very low in normal bladder tissue [84, 85]. Conversely, TXA₂S expression is an average of 11.6-fold higher (range 5–18-fold) in invasive bladder cancer and transformed bladder epithelial cell lines exhibit greater expression than non-transformed immortalized cells [85]. Focal TXA₂S expression is first observed in carcinoma *in situ* and becomes widespread in invasive and high-grade carcinoma [84]. In both bladder cancer tissue and cell lines, TXA₂S expression is transcriptionally regulated. TXA₂S expression correlates significantly with tumor grade, stage, and overall survival and is an independent marker of invasiveness in the tumor [85].

TP expression is also increased in invasive bladder cancer compared to "normal" adjacent and remote bladder tissue. TP expression is increased at the protein, but not them RNA, level prompting speculation that post-translational mechanisms stabilize the protein [85]. However, the increased TP expression in bladder cancer was documented using antibodies/ probes that bind the common domain. More recently, the significance of individual TP isoforms in bladder cancer has been investigated. Increased expression of the humanspecific isoform TP β , but not TP α , is enhanced in epithelial and stromal compartments in invasive bladder cancer and correlates with cancer grade. Like expression of TXA₂S, TP expression, and in particular TP β expression, correlates well with increased growth rate, invasiveness, and shorter survival time [86]. TP β expression was only observed in bladder cancer cell lines and inhibition of TP β produces apoptosis associated with suppression of ERK and focal adhesion kinase (FAK) signaling. Conversely, ectopic expression of TP β in "normal" bladder epithelium stimulates growth, migration, and invasive potential [86]. Moreover, it enables "normal" bladder cancer to form xenografts in nude mice that result in highly dedifferentiated tumors [86]. TP signaling through the $G\alpha 12$ family was implicated in the enhanced motility of bladder cancer but no further information was provided on the

pathways responsible for proliferation. This is one of the few cancers for which roles of the two TP isoforms have been identified and provides evidence for distinct roles for TP α and TP β in the pathogenesis of cancer.

While the changes in TXA₂/TP expression are pronounced in bladder cancer, the real question is: Does targeting this pathway arrest/perturb tumor growth? Cells overexpressing TXA₂S or TP grow at an accelerated rate and are highly invasive [85, 86]. Pharmacological antagonism of TXA₂S or TP *in vitro* slows proliferation of bladder cancer cell lines two- to three fold and ablates migration and invasion [85, 86]. Further, depriving bladder cancer cells from TP signaling (either pharmacological inhibition or small interfering RNA (siRNA) knockdown approaches) promotes caspase processing and the onset of apoptosis [84] and sensitizes the cells to the effects of other chemotherapeutic agents including paclitaxel and cisplatin [84]. These data suggest that enhanced TXA₂ levels due to TXA₂S overexpression were activating TP-dependent pathways of cell proliferation.

2.7 Modulation of endothelial cell migration/angiogenesis by thromboxane A₂

Neovascularization, or the formation of new blood vessels, is central to the pathogenesis of most solid tumors and their sequelae, including metastasis. It occurs through a number of mechanisms including angiogenesis, vasculogenesis, and intussusception (reviewed in [87]). The level of new blood vessel growth is dependent upon the balance between proand antiangiogenic factors present in the tissue. TXA₂ has both pro- and anti-angiogenic effects in multiple experimental systems. We have found that TP stimulation reduces spontaneous endothelial cell (EC) migration by 58% and in vitro capillary formation by 85% [88] as well as abrogating the pro-angiogenic effects of vascular endothelial growth factor (VEGF)-A [89] and fibroblast growth factor-2 (FGF-2) [90] in vitro and in vivo. Moreover, TP stimulation results in the destruction of established EC networks through increased apoptosis [91]. The regulatory pathways for each of the effects are highly stimulus specific and share few features in common. Antagonism of nitric oxide and FAK [89], stabilization of p53/ antagonism of integrin $\alpha\nu\beta3$ [90], and inhibition of intercellular communication [88] are just some of the mechanisms employed by TXA₂ to prevent angiogenesis. A number of reports from other groups support the concept that TXA₂ is an anti-angiogenic stimulus. TXA₂ suppresses angiogenesis and promotes vascular degeneration in the retina through induction of calpain-dependent neuro-retinovascular EC death [92]. TP stimulation mediates EC apoptosis associated with diabetes [93, 94] and acts in an additive/synergistic manner with platelet releasate [95] and neurokinin B [96] to promote EC injury and inhibit angiogenesis. In addition, the TP ligands isoprostane 8-iso-PGF_{2a}, 8-iso-PGE₂, and 8-iso-PGA₂ all inhibit spontaneous and VEGF-induced migration and differentiation of human coronary ECs in vitro and sprouting angiogenesis from cardiac explants ex vivo without influencing apoptosis [97]. These effects are sensitive to inhibition of Rho kinase suggesting deranged actin metabolism was involved, later proven by alterations to stress fiber formation. Collectively, these data support the hypothesis that TXA₂ may discourage re-vascularization of infarcted/ hypoxic tissue and may promote the regression of vessels through the induction of apoptosis in exposed vascular beds. Thus, blocking TP seems an appropriate course of action in diseases such as myocardial infarction, where re-vascularization is to be encouraged.

Conversely, a number of groups have shown TXA₂ is pro-angiogenic. Robust TXA₂S expression is observed in the tumor-associated endothelial cells of lung cancer but not normal lung tissue [57]. These findings support the view that TXA_2 may promote angiogenesis, thereby accelerating cancer progression. Some of the most convincing data have correlated the overexpression of TXA2 synthase in tumor cells with enhanced angiogenesis, shortened survival time, and increased tumor growth rate [72]. However, an important limitation of these findings was that the authors did not distinguish whether the effects of TXA2 on angiogenesis were due to direct actions on endothelial cells or an autocrine effect on the tumor cells. Indeed, Tai and colleagues [58] reported that TXA2 stimulation of human lung cancer cells enhances production of the pro-angiogenic protein VEGF, a key stimulus for tumor angiogenesis. Stimulation of TPa increased VEGF expression at both the mRNA and protein levels via a mechanism involving activation of ERK, PKA, EGFR, and Src kinases. Xenografts of A549-TPa cells induced greater tumor growth and increased vascularization in nude mice than control A549 cells [58]. However, evidence of increased neovascularization was only macroscopic and could have easily resulted from hemorrhage into the growing tumors. These data indicate that the autocrine effects of TP on the tumor itself are not just a regulator of tumor growth but also a significant regulator of neovascularization.

TXA₂ also has direct pro-angiogenic effects on the endothelium. Pro-angiogenic factors (FGF-2 and VEGF) promote TXA₂ release from EC up to five fold [61]. TXA₂ stimulation promotes sprouting in corneal neovascularization and rat aortic ring assays, induces vascularization of the rodent ovary, and stimulates the differentiation and migration of endothelial cells *in vitro* [46, 61, 72, 98, 99]. Furthermore, antagonizing TP/TXA₂ synthesis attenuates endothelial chemotaxis to FGF-2 and VEGF, differentiation of endothelium on matrigel *in vitro*, and angiogenesis in the corneal neovascularization and rat aortic ring assays [61, 100] [98]. These activities are not overcome by incubation with VEGF [100] indicating the strength of the inhibition of angiogenesis by these agents and/or essential contribution of TXA₂ to the pro-angiogenic effects of other factors. These antagonists do not affect EC viability or adhesion indicating their effects on quiescent ECs would be minimal [100]. If these data hold true, then the inhibition of TXA₂ synthesis or TP signaling during diseases, such as cancer, would prevent vessel formation, slowing tumor growth, and prolonging survival.

TP α and TP β have almost identical coupling to heterotrimeric G-proteins resulting in misplaced complacency regarding their distinct roles in disease. We believe the dichotomy in the reported effects of TP stimulation on angiogenesis result from the use of models that lack TP β expression. The relative importance of both TP isoforms to EC biology, especially TP β , is largely unexplored. Our data also show that expressing TP β in the endothelium from TP null mice inhibits migration and differentiation *in vitro*, and transgenic mice overexpressing TP β in endothelial cells display reduced angiogenesis in matrigel plug models[89, 90]; however, only humans have TP β with both rat and mouse TP similar to the human TP α isoform (78% homology). Thus, the small animal models upon which the proangiogenic properties of TXA₂ are based are flawed as they do not account for TP β making comparisons difficult. More importantly, our data indicate that the anti-angiogenic

properties of TP β dominate in EC expressing both isoforms [89, 90]. The primary deficit in the original report of the TP β transgenic mouse was decreased placental size with microscopic evidence of ischemia [101]. Our seminal observations extend these findings and suggest that vessel formation in the developing placenta was most likely suppressed by TP β . Further, the amplified signaling of TP α -TP β heterodimers in response to isoprostanes [102] may be the reason for the highly potent nature of these TP agonists as anti-angiogenic compounds [97]. Thus, the existence of two TP isoforms in humans may have implications for the role of TXA₂ in vascular remodeling in angiogenic diseases such as cancer.

The mechanisms by which the two TP isoforms regulate angiogenesis are still somewhat unclear. TP α does not share the same angioregulatory activity as TP β indicating that the divergent tail residues are the source of these properties. The tail of TP β contains multiple sites that could regulate receptor signaling but few are well characterized. Residues TP β 355–TP β 366 and TP β 337–TP β 344 are important for ligand-induced [103] and tonic [104] internalization and mediate interactions with Nm23-H2 [105] and Rab11 [106]. These data indicate the protein interactions of the two tails are different and have binding sites for signaling molecules that may regulate angiogenesis. Further, Kinsella and colleagues recently reported PRK1 and angio-associated migratory cell protein as the first non-G-protein signaling proteins that couple to TP isoforms that have direct effects on cell motility [51, 107]. These data highlight the role of non-G-protein-mediated mechanisms in the regulation of angiogenesis by TP isoforms and further strengthen the notion of divergent pathological roles for TP α and TP β in disease.

3 Summary and future directions

In summary, these findings suggest TXA₂S and TP not only play an important role in cardiovascular disorders but also in cancer progression and metastasis. Over the past decade, several advances have been made in the field of TXA₂ research that have significantly increased our knowledge and understanding of the underlying mechanisms of TXA₂S and TP signaling. COX-2 expression has been found to be upregulated in a variety of cancers and TXA₂S and TXA₂ being a downstream metabolite of COX-2 is also over expressed in various cancers. Cancer progression involves several key steps such as angiogenesis, cell survival, cell migration, cell invasion, and metastasis; studies have shown that both TXA₂S and TP play important roles in one or more of these key process. It is of great importance to further determine whether and how prostanoids, such as TXA_2 , mediate the effects of COX-2 in cancer, potentially leading to a more targeted approach for cancer prevention and treatment. The failure of existing TXA₂S inhibitors and TXA₂ antagonists to show robust clinical benefit in treatment of cardiovascular disorders has largely been due to co-incident use of NSAIDs, such as aspirin, as part of the standard of care. Conversely, current therapeutic regimens for cancer incorporate few agents in this class making TXA₂S/TP antagonism an attractive target with potentially significant therapeutic benefit. Moreover, in contrast to NSAIDs, TXA2S/TP inhibitors do not prevent other COX-derived anti-tumor products, such as PGI₂, with beneficial effects from being synthesized [108, 109]. Hence, future work should focus on the advantages of directly targeting TXA₂S and/or antagonizing TP for its functional role in cancer progression.

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References

- Hamberg M, Svensson J, Samuelsson B. Thromboxanes: A new group of biologically active compounds derived from prostaglandin endoperoxides. Proceedings of the National Academy of Sciences of the United States of America. 1975; 72(8):2994–2998. [PubMed: 1059088]
- Moncada S, Needleman P, Bunting S, Vane JR. Prostaglandin endoperoxide and thromboxane generating systems and their selective inhibition. Prostaglandins. 1976; 12(3):323–335. [PubMed: 968048]
- Smyth EM. Thromboxane and the thromboxane receptor in cardiovascular disease. Clinical Lipidology. 2010; 5(2):209–219. [PubMed: 20543887]
- Needleman P, Minkes M, Raz A. Thromboxanes: Selective biosynthesis and distinct biological properties. Science. 1976; 193(4248):163–165. [PubMed: 945611]
- Needleman P, Moncada S, Bunting S, Vane JR, Hamberg M, Samuelsson B. Identification of an enzyme in platelet microsomes which generates thromboxane A2 from prostaglandin endoperoxides. Nature. 1976; 261(5561):558–560. [PubMed: 934294]
- Halushka PV, Allan CJ, Davis-Bruno KL. Thromboxane A2 receptors. Journal of Lipid Mediators and Cell Signalling. 1995; 12(2–3):361–378. [PubMed: 8777579]
- Jones RL, Wilson NH, Armstrong RA. Characterization of thromboxane receptors in human platelets. Advances in Experimental Medicine and Biology. 1985; 192:67–81. [PubMed: 3010674]
- 8. Fitzgerald DJ, Roy L, Catella F, FitzGerald GA. Platelet activation in unstable coronary disease. The New England Journal of Medicine. 1986; 315(16):983–989. [PubMed: 3531859]
- Katugampola SD, Davenport AP. Thromboxane receptor density is increased in human cardiovascular disease with evidence for inhibition at therapeutic concentrations by the AT(1) receptor antagonist losartan. British Journal of Pharmacology. 2001; 134(7):1385–1392. [PubMed: 11724743]
- Neri Serneri GG, Gensini GF, Abbate R, Mugnaini C, Favilla S, Brunelli C, et al. Increased fibrinopeptide A formation and thromboxane A2 production in patients with ischemic heart disease: Relationships to coronary pathoanatomy, risk factors, and clinical manifestations. American Heart Journal. 1981; 101(2):185–194. [PubMed: 6451162]
- Fuse S, Kamiya T. Plasma thromboxane B2 concentration in pulmonary hypertension associated with congenital heart disease. Circulation. 1994; 90(6):2952–2955. [PubMed: 7994842]
- Gresele P, Deckmyn H, Nenci GG, Vermylen J. Thromboxane synthase inhibitors, thromboxane receptor antagonists and dual blockers in thrombotic disorders. Trends in Pharmacological Sciences. 1991; 12(4):158–163. [PubMed: 1829559]
- Kobayashi T, Tahara Y, Matsumoto M, Iguchi M, Sano H, Murayama T, et al. Roles of thromboxane A(2) and prostacyclin in the development of atherosclerosis in apoE-deficient mice. The Journal of Clinical Investigation. 2004; 114(6):784–794. [PubMed: 15372102]
- Mehta JL, Lawson D, Mehta P, Saldeen T. Increased prostacyclin and thromboxane A2 biosynthesis in atherosclerosis. Proceedings of the National Academy of Sciences of the United States of America. 1988; 85(12):4511–4515. [PubMed: 3132716]
- Willerson JT, Yao SK, Ferguson JJ, Anderson HV, Golino P, Buja LM. Unstable angina pectoris and the progression to acute myocardial infarction. Role of platelets and platelet-derived mediators. Texas Heart Institute Journal. 1991; 18(4):243–247. [PubMed: 15227406]
- Lariviere R, Moreau C, Rodrigue ME, Lebel M. Thromboxane blockade reduces blood pressure and progression of renal failure independent of endothelin-1 in uremic rats. Prostaglandins, Leukotrienes, and Essential Fatty Acids. 2004; 71(2):103–109.

- Willerson JT, Buja LM. Potential of combined thromboxane A2 and serotonin antagonists to prevent the development of unstable angina and acute myocardial infarction. Texas Heart Institute Journal. 1990; 17(3):157–164. [PubMed: 15227166]
- Willerson JT, Golino P, Eidt J, Yao SK, Buja LM. Potential usefulness of combined thromboxane A2 and serotonin receptor blockade for preventing the conversion from chronic to acute coronary artery disease syndromes. The American Journal of Cardiology. 1990; 66(16):48G–53G.
- Haurand M, Ullrich V. Isolation and characterization of thromboxane synthase from human platelets as a cytochrome P-450 enzyme. Journal of Biological Chemistry. 1985; 260(28):15059– 15067. [PubMed: 2999104]
- Ohashi K, Ruan KH, Kulmacz RJ, Wu KK, Wang LH. Primary structure of human thromboxane synthase determined from the cDNA sequence. Journal of Biological Chemistry. 1992; 267(2): 789–793. [PubMed: 1730669]
- Shen RF, Tai HH. Thromboxanes: Synthase and receptors. Journal of Biomedical Science. 1998; 5(3):153–172. [PubMed: 9678486]
- Yokoyama C, Miyata A, Ihara H, Ullrich V, Tanabe T. Molecular cloning of human platelet thromboxane A synthase. Biochemical and Biophysical Research Communications. 1991; 178(3): 1479–1484. [PubMed: 1714723]
- Tanabe T, Ullrich V. Prostacyclin and thromboxane synthases. Journal of Lipid Mediators and Cell Signalling. 1995; 12(2–3):243–255. [PubMed: 8777569]
- 24. Hirata M, Hayashi Y, Ushikubi F, Yokota Y, Kageyama R, Nakanishi S, et al. Cloning and expression of cDNA for a human thromboxane A2 receptor. Nature. 1991; 349(6310):617–620. [PubMed: 1825698]
- Raychowdhury MK, Yukawa M, Collins LJ, McGrail SH, Kent KC, Ware JA. Alternative splicing produces a divergent cytoplasmic tail in the human endothelial thromboxane A2 receptor. Journal of Biological Chemistry. 1994; 269(30):19256–19261. [Comparative Study]. [PubMed: 8034687]
- Kinsella BT. Thromboxane A2 signalling in humans: A 'Tail' of two receptors. Biochemical Society Transactions. 2001; 29(Pt 6):641–654. [PubMed: 11709048]
- Miggin SM, Kinsella BT. Expression and tissue distribution of the mRNAs encoding the human thromboxane A2 receptor (TP) alpha and beta isoforms. Biochimica et Biophysica Acta. 1998; 1425(3):543–559. [Research Support, Non-U.S. Gov't]. [PubMed: 9838218]
- Nakahata N. Thromboxane A2: Physiology/pathophysiology, cellular signal transduction and pharmacology. Pharmacology and Therapeutics. 2008; 118(1):18–35. [PubMed: 18374420]
- 29. Namba T, Narumiya S. Thromboxane A2 receptor; structure, function and tissue distribution. Nihon Rinsho. 1993; 51(1):233–240. [PubMed: 8433523]
- 30. Honn KV, Bockman RS, Marnett LJ. Prostaglandins and cancer: A review of tumor initiation through tumor metastasis. Prostaglandins. 1981; 21(5):833–864. [PubMed: 6280245]
- 31. Honn KV, Busse WD, Sloane BF. Prostacyclin and thromboxanes. Implications for their role in tumor cell metastasis. Biochemical Pharmacology. 1983; 32(1):1–11. [PubMed: 6338880]
- 32. Menter, DG.; Neagos, J.; Dunn, R.; Pallazo, TT.; Chen, T.; Taylor, JD., et al. Tumor Cell induced Platelet aggregration: Inhibition by prostacylin, thromboxane A2 and phosphodiesterase inhibitors. In: Powles, TJ.; Bockman, RS.; Honn, KV.; Ramwell, PW., editors. Prostaglandins and cancer. New York: Alan Liss, Inc.; 1982. p. 369-374.
- Nie D, Honn KV. Cyclooxygenase, lipoxygenase and tumor angiogenesis. Cellular and Molecular Life Sciences. 2002; 59(5):799–807. [PubMed: 12088280]
- Honn KV, Meyer J. Thromboxanes and prostacyclin: Positive and negative modulators of tumor growth. Biochemical and Biophysical Research Communications. 1981; 102(4):1122–1129. [PubMed: 6274330]
- 35. U.S.C.S.W.Group. United States Cancer Statistics: 1999–2007 Incidence and Mortality Web-based Report. Atlanta: U.S. Department of Health and Human Services, Centers for Disease Control and Prevention and National Cancer Institute; 2010.
- Bairati I, Meyer F, Fradet Y, Moore L. Dietary fat and advanced prostate cancer. The Journal of Urology. 1998; 159(4):1271–1275. [Research Support, Non-U.S. Gov't]. [PubMed: 9507851]
- Rose DP, Connolly JM. Dietary fat, fatty acids and prostate cancer. Lipids. 1992; 27(10):798–803. [Review]. [PubMed: 1435098]

- West DW, Slattery ML, Robison LM, French TK, Mahoney AW. Adult dietary intake and prostate cancer risk in Utah: A case-control study with special emphasis on aggressive tumors. Cancer Causes & Control. 1991; 2(2):85–94. [Research Support, U.S. Gov't, P.H.S.]. [PubMed: 1873441]
- 39. Gupta S, Srivastava M, Ahmad N, Sakamoto K, Bostwick DG, Mukhtar H. Lipoxygenase-5 is over expressed in prostate adenocarcinoma. Cancer. 2001; 91(4):737–743. [PubMed: 11241241]
- 40. Nie D, Che M, Grignon D, Tang K, Honn KV. Role of eicosanoids in prostate cancer progression. Cancer Metastasis Reviews. 2001; 20(3–4):195–206. [PubMed: 12085962]
- Nie D, Hillman GG, Geddes T, Tang K, Pierson C, Grignon DJ, et al. Platelet-type 12lipoxygenase regulates angiogenesis in human prostate carcinoma. Advances in Experimental Medicine and Biology. 1999; 469:623–630. [PubMed: 10667391]
- Nie D, Nemeth J, Qiao Y, Zacharek A, Li L, Hanna K, et al. Increased metastatic potential in human prostate carcinoma cells by overexpression of arachidonate 12-lipoxygenase. Clinical & Experimental Metastasis. 2003; 20(7):657–663. [PubMed: 14669797]
- Gupta S, Srivastava M, Ahmad N, Bostwick DG, Mukhtar H. Over-expression of cyclooxygenase-2 in human prostate adenocarcinoma. The Prostate. 2000; 42(1):73–78. [PubMed: 10579801]
- Lee LM, Pan CC, Cheng CJ, Chi CW, Liu TY. Expression of cyclooxygenase-2 in prostate adenocarcinoma and benign prostatic hyperplasia. Anticancer Research. 2001; 21:1291–1294. [PubMed: 11396201]
- Nie D, Che M, Zacharek A, Qiao Y, Li L, Li X, et al. Differential expression of thromboxane synthase in prostate carcinoma: Role in tumor cell motility. American Journal of Pathology. 2004; 164(2):429–439. [PubMed: 14742249]
- 46. Dassesse T, de Leval X, de Leval L, Pirotte B, Castronovo V, Waltregny D. Activation of the thromboxane A2 pathway in human prostate cancer correlates with tumor Gleason score and pathologic stage. Eur Urol. 2006; 50(5):1021–1031. discussion 1031. [PubMed: 16522350]
- Villers A, McNeal JE, Redwine EA, Freiha FS, Stamey TA. The role of perineural space invasion in the local spread of prostatic adenocarcinoma. Journal of Urology. 1989; 142(3):763–768. [PubMed: 2769857]
- Pantel K, Brakenhoff RH. Dissecting the metastatic cascade. Nature reviews. Cancer. 2004; 4(6): 448–456. [Review].
- Nie D, Guo Y, Yang D, Tang Y, Chen Y, Wang MT, et al. Thromboxane A2 receptors in prostate carcinoma: Expression and its role in regulating cell motility via small GTPase Rho. Cancer Research. 2008; 68(1):115–121. [PubMed: 18172303]
- Kelly P, Stemmle LN, Madden JF, Fields TA, Daaka Y, Casey PJ. A role for the G12 family of heterotrimeric G proteins in prostate cancer invasion. Journal of Biological Chemistry. 2006; 281(36):26483–26490. [PubMed: 16787920]
- 51. Turner EC, Kavanagh DJ, Mulvaney EP, McLean C, Wikstrom K, Reid HM, et al. Identification of an interaction between the TPalpha and TPbeta isoforms of the human thromboxane A2 receptor with protein kinase C-related kinase (PRK) 1: Implications for prostate cancer. Journal of Biological Chemistry. 2011; 286(17):15440–15457. [PubMed: 21357687]
- 52. Watkins G, Douglas-Jones A, Mansel RE, Jiang WG. Expression of thromboxane synthase, TBXAS1 and the thromboxane A2 receptor, TBXA2R, in human breast cancer. International Seminars in Surgical Oncology. 2005; 2:23. [PubMed: 16250911]
- 53. Kelly P, Moeller BJ, Juneja J, Booden MA, Der CJ, Daaka Y, et al. The G12 family of heterotrimeric G proteins promotes breast cancer invasion and metastasis. Proceedings of the National Academy of Sciences of the United States of America. 2006; 103(21):8173–8178. [PubMed: 16705036]
- 54. Abraham JE, Harrington P, Driver KE, Tyrer J, Easton DF, Dunning AM, et al. Common polymorphisms in the prostaglandin pathway genes and their association with breast cancer susceptibility and survival. Clinical Cancer Research. 2009; 15(6):2181–2191. [PubMed: 19276290]
- Ermert L, Dierkes C, Ermert M. Immunohistochemical expression of cyclooxygenase isoenzymes and downstream enzymes in human lung tumors. Clinical Cancer Research. 2003; 9(5):1604– 1610. [PubMed: 12738712]

- 56. Kreutzer M, Fauti T, Kaddatz K, Seifart C, Neubauer A, Schweer H, et al. Specific components of prostanoid-signaling pathways are present in non-small cell lung cancer cells. Oncology Reports. 2007; 18(2):497–501. [PubMed: 17611676]
- 57. Yoshimoto A, Kasahara K, Kawashima A, Fujimura M, Nakao S. Characterization of the prostaglandin biosynthetic pathway in non-small cell lung cancer: A comparison with small cell lung cancer and correlation with angiogenesis, angiogenic factors and metastases. Oncology Reports. 2005; 13(6):1049–1057. [PubMed: 15870920]
- Wei J, Yan W, Li X, Ding Y, Tai HH. Thromboxane receptor alpha mediates tumor growth and angiogenesis via induction of vascular endothelial growth factor expression in human lung cancer cells. Lung Cancer. 2010; 69(1):26–32. [PubMed: 19853959]
- Wei J, Yan W, Li X, Chang WC, Tai HH. Activation of thromboxane receptor alpha induces expression of cyclooxygenase-2 through multiple signaling pathways in A549 human lung adenocarcinoma cells. Biochemical Pharmacology. 2007; 74(5):787–800. [PubMed: 17632087]
- Li X, Tai HH. Activation of thromboxane A(2) receptors induces orphan nuclear receptor Nurr1 expression and stimulates cell proliferation in human lung cancer cells. Carcinogenesis. 2009; 30(9):1606–1613. [PubMed: 19570744]
- Nie D, Lamberti M, Zacharek A, Li L, Szekeres K, Tang K, et al. Thromboxane A(2) regulation of endothelial cell migration, angiogenesis, and tumor metastasis. Biochemical and Biophysical Research Communications. 2000; 267(1):245–251. [PubMed: 10623605]
- 62. Hanahan D, Weinberg RA. The hallmarks of cancer. Cell. 2000; 100(1):57–70. [Review]. [PubMed: 10647931]
- Fujimura M, Kasahara K, Shirasaki H, Heki U, Iwasa K, Ueda A, et al. Up-regulation of ICH-1 L protein by thromboxane A2 antagonists enhances cisplatin-induced apoptosis in non-small-cell lung-cancer cell lines. Journal of Cancer Research and Clinical Oncology. 1999; 125(7):389–394. [PubMed: 10394958]
- Leung KC, Hsin MK, Chan JS, Yip JH, Li M, Leung BC, et al. Inhibition of thromboxane synthase induces lung cancer cell death via increasing the nuclear p27. Experimental cell research. 2009; 315(17):2974–2981. [Research Support, Non-U.S. Gov't]. [PubMed: 19576211]
- Leung KC, Li MY, Leung BC, Hsin MK, Mok TS, Underwood MJ, et al. Thromboxane synthase suppression induces lung cancer cell apoptosis via inhibiting NF-kappaB. Experimental Cell Research. 2010; 316(20):3468–3477. [PubMed: 20647010]
- McLemore TL, Hubbard WC, Litterst CL, Liu MC, Miller S, McMahon NA, et al. Profiles of prostaglandin biosynthesis in normal lung and tumor tissue from lung cancer patients. Cancer Research. 1988; 48(11):3140–3147. [PubMed: 3130187]
- Huang RY, Li MY, Hsin MK, Underwood MJ, Ma LT, Mok TS, et al. 4-Methylnitrosamino-1-3pyridyl-1-butanone (NNK) promotes lung cancer cell survival by stimulating thromboxane A2 and its receptor. Oncogene. 2011; 30(1):106–116. [Research Support, Non-U.S. Gov't]. [PubMed: 20818420]
- Cathcart MC, Gately K, Cummins R, Kay E, O'Byrne KJ, Pidgeon GP. Examination of thromboxane synthase as a prognostic factor and therapeutic target in non-small cell lung cancer. Molecular Cancer. 2011; 10:25. [PubMed: 21388528]
- McAfee AJ, McSorley EM, Cuskelly GJ, Moss BW, Wallace JM, Bonham MP, et al. Red meat consumption: An overview of the risks and benefits. Meat science. 2010; 84(1):1–13. [PubMed: 20374748]
- Bing RJ, Miyataka M, Rich KA, Hanson N, Wang X, Slosser HD, et al. Nitric oxide, prostanoids, cyclooxygenase, and angiogenesis in colon and breast cancer. Clinical Cancer Research. 2001; 7(11):3385–3392. [PubMed: 11705852]
- Tsujii M, Kawano S, DuBois RN. Cyclooxygenase-2 expression in human colon cancer cells increases metastatic potential. Proceedings of the National Academy of Sciences of the United States of America. 1997; 94(7):3336–3340. [PubMed: 9096394]
- 72. Pradono P, Tazawa R, Maemondo M, Tanaka M, Usui K, Saijo Y, et al. Gene transfer of thromboxane A(2) synthase and prostaglandin I(2) synthase antithetically altered tumor angiogenesis and tumor growth. Cancer Research. 2002; 62(1):63–66. [PubMed: 11782360]

- Gustafsson A, Hansson E, Kressner U, Nordgren S, Andersson M, Lonnroth C, et al. Prostanoid receptor expression in colorectal cancer related to tumor stage, differentiation and progression. Acta Oncologica. 2007; 46(8):1107–1112. [PubMed: 17943472]
- 74. Sakai H, Suzuki T, Takahashi Y, Ukai M, Tauchi K, Fujii T, et al. Upregulation of thromboxane synthase in human colorectal carcinoma and the cancer cell proliferation by thromboxane A2. FEBS Letters. 2006; 580(14):3368–3374. [PubMed: 16709411]
- 75. Sathornsumetee S, Rich JN. New treatment strategies for malignant gliomas. Expert Review of Anticancer Therapy. 2006; 6(7):1087–1104. [Review]. [PubMed: 16831080]
- McDonough W, Tran N, Giese A, Norman SA, Berens ME. Altered gene expression in human astrocytoma cells selected for migration: I. Thromboxane synthase. Journal of Neuropathology and Experimental Neurology. 1998; 57(5):449–455. [PubMed: 9596415]
- 77. Giese A, Hagel C, Kim EL, Zapf S, Djawaheri J, Berens ME, et al. Thromboxane synthase regulates the migratory phenotype of human glioma cells. Neuro-Oncology. 1999; 1(1):3–13. [PubMed: 11550298]
- 78. Schauff AK, Kim EL, Leppert J, Nadrowitz R, Wuestenberg R, Brockmann MA, et al. Inhibition of invasion-associated thromboxane synthase sensitizes experimental gliomas to gamma-radiation. Journal of Neuro-Oncology. 2009; 91(3):241–249. [PubMed: 18825315]
- Stupp R, Mason WP, van den Bent MJ, Weller M, Fisher B, Taphoorn MJ, et al. Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. The New England Journal of Medicine. 2005; 352(10):987–996. [PubMed: 15758009]
- Schmidt NO, Ziu M, Cargioli T, Westphal M, Giese A, Black PM, et al. Inhibition of thromboxane synthase activity improves glioblastoma response to alkylation chemotherapy. Translational Oncology. 2010; 3(1):43–49. [PubMed: 20165694]
- Hegi ME, Diserens AC, Gorlia T, Hamou MF, de Tribolet N, Weller M, et al. MGMT gene silencing and benefit from temozolomide in glioblastoma. The New England Journal of Medicine. 2005; 352(10):997–1003. [PubMed: 15758010]
- Gangwar R, Mandhani A, Mittal RD. Functional polymorphisms of cyclooxygenase-2 (COX-2) gene and risk for urinary bladder cancer in North India. Surgery. 2011; 149(1):126–134. [Comparative Study]. [PubMed: 20627335]
- Banon A, Zenser TV, Thomasson DL, Davis BB. Eicosanoid synthesis by cultured human urothelial cells: Potential role in bladder cancer. Cancer Research. 1986; 46(11):5676–5681. [PubMed: 3093068]
- Moussa O, Riker JM, Klein J, Fraig M, Halushka PV, Watson DK. Inhibition of thromboxane synthase activity modulates bladder cancer cell responses to chemotherapeutic agents. Oncogene. 2008; 27(1):55–62. [PubMed: 17603559]
- Moussa O, Yordy JS, Abol-Enein H, Sinha D, Bissada NK, Halushka PV, et al. Prognostic and functional significance of thromboxane synthase gene overexpression in invasive bladder cancer. Cancer Research. 2005; 65(24):11581–11587. [PubMed: 16357168]
- Moussa O, Ashton AW, Fraig M, Garrett-Mayer E, Ghoneim MA, Halushka PV, et al. Novel role of thromboxane receptors beta isoformin bladder cancer pathogenesis. Cancer Research. 2008; 68(11):4097–4104. [PubMed: 18519668]
- Patan S. Vasculogenesis and angiogenesis. Cancer Treatment and Research. 2004; 117:3–32. [PubMed: 15015550]
- Ashton AW, Yokota R, John G, Zhao S, Suadicani SO, Spray DC, et al. Inhibition of endothelial cell migration, intercellular communication, and vascular tube formation by thromboxane A(2). Journal of Biological Chemistry. 1999; 274(50):35562–35570. [PubMed: 10585431]
- Ashton AW, Ware JA. Thromboxane A₂ receptor signaling inhibits vascular endothelial growth factor–induced endothelial cell differentiation and migration. Circulation Research. 2004; 95:372– 379. [PubMed: 15242977]
- 90. Ashton AW, Cheng Y, Helisch A, Ware JA. Thromboxane A₂ receptor agonists antagonize the proangiogenic effects of fibroblast growth factor-2: role of receptor internalization, thrombospondin-1, and ανβ3. Circulation Research. 2004; 94:735–742. [PubMed: 14963009]

- 91. Gao Y, Yokota R, Tang S, Ashton AW, Ware JA. Reversal of angiogenesis *in vitro*, induction of apoptosis, and inhibition of AKT phosphorylation in endothelial cells by thromboxane A(2). Circulation Research. 2000; 87(9):739–745. [PubMed: 11055976]
- 92. Beauchamp MH, Martinez-Bermudez AK, Gobeil F Jr, Marrache AM, Hou X, Speranza G, et al. Role of thromboxane in retinal microvascular degeneration in oxygen-induced retinopathy. Journal of Applied Physiology. 2001; 90(6):2279–2288. [PubMed: 11356793]
- 93. De La Cruz JP, Moreno A, Ruiz-Ruiz MI, Sanchez De La Cuesta F. Effect of DT-TX 30, a combined thromboxane synthase inhibitor and thromboxane receptor antagonist, on retinal vascularity in experimental diabetes mellitus. Thrombosis Research. 2000; 97(3):125–131. [PubMed: 10680643]
- 94. Zou MH, Shi C, Cohen RA. High glucose via peroxynitrite causes tyrosine nitration and inactivation of prostacyclin synthase that is associated with thromboxane/prostaglandin H(2) receptor-mediated apoptosis and adhesion molecule expression in cultured human aortic endothelial cells. Diabetes. 2002; 51(1):198–203. [PubMed: 11756341]
- 95. Kishi Y, Numano F. *In vitro* study of vascular endothelial injury by activated platelets and its prevention. Atherosclerosis. 1989; 76(2–3):95–101. [PubMed: 2543423]
- 96. Pal S, Wu J, Murray JK, Gellman SH, Wozniak MA, Keely PJ, et al. An antiangiogenic neurokinin-B/ thromboxane A2 regulatory axis. The Journal of Cell Biology. 2006; 174(7):1047– 1058. [PubMed: 17000881]
- 97. Benndorf RA, Schwedhelm E, Gnann A, Taheri R, Kom G, Didié M, et al. Isoprostanes inhibit vascular endothelial growth factor–induced endothelial cell migration, tube formation, and cardiac vessel sprouting *in vitro*, as well as angiogenesis in vivo via activation of the thromboxane A2 receptor. A potential link between oxidative stress and impaired angiogenesis. Circulation Research. 2008; 103:1037–1046. [PubMed: 18802021]
- Daniel TO, Liu H, Morrow JD, Crews BC, Marnett LJ. Thromboxane A2 is a mediator of cyclooxygenase-2-dependent endothelial migration and angiogenesis. Cancer Research. 1999; 59(18):4574–4577. [PubMed: 10493510]
- Sakurai T, Tamura K, Kogo H. Stimulatory effects of eicosanoids on ovarian angiogenesis in early luteal phase in cyclooxygenase-2 inhibitor-treated rats. European Journal of Pharmacology. 2005; 516(2):158–164. [PubMed: 15921676]
- 100. de Leval X, Dassesse T, Dogne JM, Waltregny D, Bellahcene A, Benoit V, et al. Evaluation of original dual thromboxane A2 modulators as antiangiogenic agents. Journal of Pharmacology and Experimental Therapeutics. 2006; 318(3):1057–1067. [PubMed: 16720756]
- 101. Rocca B, Loeb AL, Strauss JF 3rd, Vezza R, Habib A, Li H, et al. Directed vascular expression of the thromboxane A2 receptor results in intrauterine growth retardation. Nature Medicine. 2000; 6(2):219–221.
- 102. Wilson SJ, McGinley K, Huang AJ, Smyth EM. Heterodimerization of the alpha and beta isoforms of the human thromboxane receptor enhances isoprostane signaling. Biochemical and Biophysical Research Communications. 2007; 352(2):397–403. [PubMed: 17134677]
- 103. Parent JL, Labrecque P, Orsini MJ, Benovic JL. Internalization of the TXA2 receptor alpha and beta isoforms. Role of the differentially spliced cooh terminus in agonist-promoted receptor internalization. Journal of Biological Chemistry. 1999; 274(13):8941–8948. [PubMed: 10085139]
- 104. Parent JL, Labrecque P, Driss Rochdi M, Benovic JL. Role of the differentially spliced carboxyl terminus in thromboxane A2 receptor trafficking: Identification of a distinct motif for tonic internalization. Journal of Biological Chemistry. 2001; 276(10):7079–7085. [PubMed: 11112783]
- 105. Rochdi MD, Laroche G, Dupre E, Giguere P, Lebel A, Watier V, et al. Nm23-H2 interacts with a G protein-coupled receptor to regulate its endocytosis through an Rac1-dependent mechanism. Journal of Biological Chemistry. 2004; 279(18):18981–18989. [PubMed: 14976202]
- 106. Theriault C, Rochdi MD, Parent JL. Role of the Rab11-associated intracellular pool of receptors formed by constitutive endocytosis of the beta isoform of the thromboxane A2 receptor (TP beta). Biochemistry. 2004; 43(19):5600–5607. [PubMed: 15134434]

- 107. Reid HM, Wikstrom K, Kavanagh DJ, Mulvaney EP, Kinsella BT. Interaction of angio-associated migratory cell protein with the TPalpha and TPbeta isoforms of the human thromboxane A receptor. Cellular Signalling. 2011; 23(4):700–717. [PubMed: 21172430]
- 108. Honn KV, Cicone B, Skoff A. Prostacyclin: A potent antimetastatic agent. Science. 1981; 212(4500):1270–1272. [PubMed: 7015512]
- Menter DG, Onoda JM, Taylor JD, Honn KV. Effects of prostacyclin on tumor cell-induced platelet aggregation. Cancer Research. 1984; 44(2):450–456. [PubMed: 6362850]

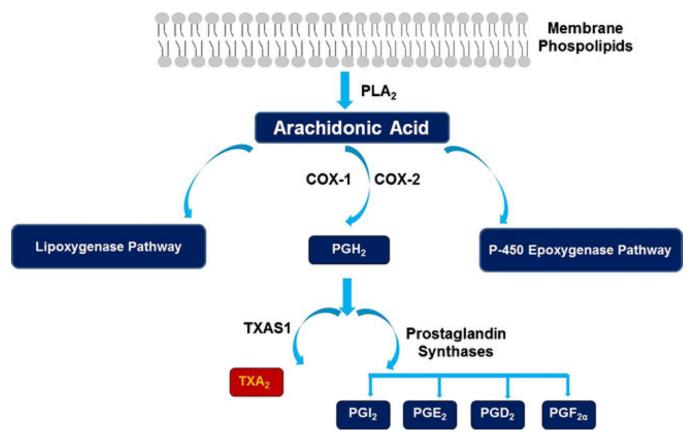


Fig. 1.

Generation of the prostanoids through metabolism of arachidonic acid. Arachidonic acid can be metabolized into different bioactive lipids by one of three distinct signaling pathways; the cyclooxygenase (COX), the lipoxygenase, and the P-450 epoxygenase pathways. The COXisoforms-COX-1 and COX-2 are responsible for the conversion of AA to PGH₂, the first step in the generation of TXA₂. Thromboxane synthase (TXA₂S) then catalyzes the conversion of the COX product, PGH₂ to TXA₂

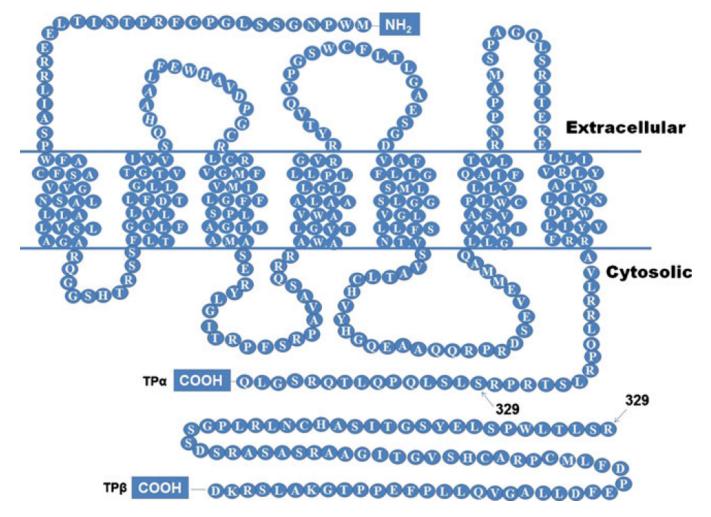


Fig. 2.

Structures of TP α and TP β receptor proteins. The TXA₂ receptor (TP) is a typical G-proteincoupled receptor that is expressed as two different isoforms in humans—*TP* α and *TP* β . TP isoforms share the first 328 amino acids but differ in the length of the C-terminal cytoplasmic tail with TP α shorter than the TP β isoform (15 *versus* 79 residues) [24, 25]