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## PROGRESS IN PIG-TO-NONHUMAN PRIMATE TRANSPLANTATION MODELS (1998–2013): A COMPREHENSIVE REVIEW OF THE LITERATURE

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### Abstract

The pig-to-nonhuman primate model is the standard choice for *in vivo* studies of organ and cell xenotransplantation. In 1998 Lambripts and his colleagues surveyed the entire world literature and reported all experimental studies in this model. With the increasing number of genetically-engineered pigs that have become available during the past few years, this model is being utilized ever more frequently. We have now reviewed the literature again and have compiled the data we have been able to find for the period January 1<sup>st</sup> 1998 to December 31<sup>st</sup> 2013, a period of 16 years. The data are presented for transplants of the heart (heterotopic and orthotopic), kidney, liver, lung, islets, neuronal cells, hepatocytes, corneas, artery patches, and skin. Heart, kidney, and, particularly, islet xenograft survival have increased significantly since 1998, and the reasons for this are briefly discussed. A comment on the limitations of the model has been made, particularly with regard to these will affect progression of xenotransplantation towards the clinic.

### Keywords

Baboons; islets; monkeys; nonhuman primates; pigs; xenotransplantation; Cornea; Heart; Kidney; Liver; Lung; Nonhuman primate; Pancreatic islets

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#### Statement re authors' contributions

DKCC conceived the study. All authors contributed to the literature search and collection of data. DKCC and VS wrote the original draft, and all authors contributed to the final manuscript.

#### Statement regarding conflict of interest

The authors report no conflicts of interest.

## Introduction

Because of the immunologic similarities of Old World nonhuman primates (NHP), e.g., baboons, rhesus monkeys, and cynomolgus monkeys, to humans, the NHP represents the preferred surrogate for humans in exploring the response to pig organ or cell transplantation. The pig-to-NHP model was introduced into xenotransplantation research in the mid-1980s (1, 2), and has become the standard model for testing the primate immune response to organs and cells from pigs with genetic manipulations and/or the effect of novel immunosuppressive regimens.

Early experience in this model was comprehensively reviewed by Lambrigts et al., in 1998 (3), but has not been reviewed fully since then. With the aim of assessing progress in the 16 years that have elapsed since 1998, we have therefore attempted to search the literature for experience of pig organ (heart, kidney, liver, lung), islet, neuronal cell, hepatocyte, cornea, and artery patch transplantation. Others have relatively recently reviewed cornea (4–6) and islet (7) xenotransplantation, and their data have been included here. Brief mention has also been made of skin transplantation in the pig-to-NHP model. The early studies on the transfusion of pig red blood cells into nonhuman primates have been reviewed elsewhere (8), but have not been included here.

In 1998, the only genetically-engineered pigs available were those expressing a human complement-regulatory protein, e.g., CD55 (hDAF) (9). Research in the NHP model was greatly facilitated by the generosity of David White and his colleagues at Imutran and Novartis in making these pigs available to the research community.

The major innovations that have been introduced into the field since 1998 include (i) many new genetic modifications in pigs (reviewed by Ekser et al (10), including  $\alpha 1,3$ -galactosyltransferase gene-knockout (GTKO) pigs (11–15) and (ii) T cell costimulatory blockade agents, first introduced into NHP xenotransplantation models by Buhler et al., in 2000 (16). In 2013, the first pigs that did not express the important N-glycolylneuraminic acid epitopes (NeuGc-KO pigs) became available (17) but, as this oligosaccharide is expressed in all mammals except humans, its relevance cannot be explored in the pig-to-NHP model (discussed in (18)).

The literature has been reviewed from January 1<sup>st</sup> 1998 to December 31<sup>st</sup> 2013. On occasion, the same series of experiments has been reported in more than one paper, e.g., one reporting the overall results, one concentrating attention on the histopathology, etc. At times, it has been difficult to determine whether the experiments included in a report are the same as, or overlap with, those reported previously, and so there may be some duplication. If the report is of relevance to the pig-to-NHP model, we have attempted to be comprehensive, but we cannot guarantee we have included all publications.

We have not included reports of studies in NHPs that did not undergo organ or cell transplantation, e.g., immunoadsorption of anti-pig antibodies alone, or reports of *ex vivo* blood perfusion of pig organs, which has been a relatively common form of experimentation with regard to assessment of pig lungs and livers. Nor have we included bone marrow or hematopoietic cell xenotransplantation between pig and NHP unless it was associated with

an organ graft. We have not included papers published in languages other than English, nor abstracts of congresses, and have not always reviewed publications that did not present new data, or presented *in vitro* data from *in vivo* studies if the actual results of the transplants were not reported. If a short publication in *Transplantation Proceedings* was followed by a full publication in another journal, we have not always included reference to the preliminary publication.

### Heart xenotransplantation (Table 1)

More progress has been made in pig heart transplantation than in the transplantation of other vital organs. The introduction of GTKO pigs was important (reviewed in (19)); hyperacute rejection, already minimized by the transplantation of hearts from pigs transgenic for a human complement-regulatory protein, was virtually eliminated, particularly when GTKO pigs expressed a human complement-regulatory protein. With adequate exogenous immunosuppressive therapy, the incidence of delayed xenograft rejection (acute humoral xenograft rejection, acute vascular rejection) was also greatly reduced. However, a new phenomenon, thrombotic microangiopathy, was reported (20), stimulating the development of pigs transgenic for one or more human coagulation-regulatory proteins, e.g., thrombomodulin, CD39, endothelial cell protein C receptor, which are only now being explored in the pig-to-NHP model.

In 1998, the longest survival of a heterotopically-placed (non-life-supporting) heart was reported to be 31 days (3), whereas by the end of 2013 this has been extended to >12 months (19,21–24) (Table 1A). Survival after orthotopic (life-supporting) pig heart transplantation has been extended from a maximum of 19 days (3) to 57 days (Table 1B).

### Kidney xenotransplantation (Table 2)

Progress in the pig kidney-to-NHP model has been slower than in the pig heart-to-NHP model, though this conclusion may be misleading since the kidney is transplanted as a life-supporting organ whereas in the majority of cases the heart is transplanted as a heterotopic, non-life-supporting organ. However, the complications of consumptive coagulopathy appear to develop more rapidly when the kidney is transplanted (25). For reasons not fully understood, this model may therefore be a more difficult one than when the heart is transplanted. The longest life-supporting kidney graft survival in 1998 was reported to be 23 days (3), but this has been extended to 90 days (26).

With regard to co-transplantation of pig kidney and thymic tissue, which has resulted in a maximum kidney graft survival of 83 days (27), there have been studies of pig thymic grafts in NHPs in the absence of kidney transplants (28–31). Six baboons underwent a regimen aimed towards inducing tolerance, three of which received fetal or neonatal pig thymic tissue transplants (31). There was some *in vitro* evidence that the thymic tissue induced xenogeneic hyporesponsiveness.

### **Liver xenotransplantation (Table 3)**

Special problems relating to pig liver transplantation have proved a major barrier to progress, largely because the genetic manipulations of the organ-source pigs have to date largely been directed towards extending survival of heart and kidney grafts. The rapid development of thrombocytopenia in the recipient NHP following pig liver transplantation remains unresolved. Nevertheless, graft survival has been extended from <3 days in 1998 (3) to 10 days today (32).

### **Lung xenotransplantation (Table 4)**

The lung continues to provide major barriers, and considerable attention is currently being directed towards overcoming them. Most studies have been of *ex vivo* pig lung perfusion with human blood, which are not reviewed here. Despite major efforts, to date, pig lung graft survival after transplantation into NHPs has been extended only from 9 hours in 1998 (3) to 5 days today.

### **Pancreatic islet xenotransplantation (Table 5)**

More progress has been made in the transplantation of pig cells than pig organs. Although the instant blood-mediated inflammatory reaction remains a major barrier after islet transplantation into the portal vein, pig islet transplantation in NHPs has been successful for >1 year.

These reports and those for neuronal cell transplantation (see below) suggest that, with the possible exception of corneal xenotransplantation (see below), these cellular transplants are likely to be the first introduced into the clinic. Indeed, clinical trials of encapsulated wild-type pig islets have already been undertaken (33–35) but detailed reports have been scarce.

### **Neuronal cell xenotransplantation**

A field in which there were no reports in NHP models in 1998 is pig neuronal cell transplantation. Research has been largely limited to one European consortium, but graft function in monkeys with a Parkinson-like disorder has been documented for periods >1 year (36, 37).

### **Hepatocyte xenotransplantation**

Although the transplantation of pig hepatocytes may have some advantages over liver xenotransplantation, few studies have been undertaken in NHP models to date. However, hepatocyte function has been documented for >80 days after transplantation and for 253 days when a second transplant was performed (38).

### **Corneal xenotransplantation (Table 6)**

To our knowledge, no studies of pig corneal transplantation in NHPs had been reported before 1998, whereas a number of studies have been published since then, with encouraging

results. Anterior lamellar keratoplasty using decellularized corneas from wild-type pigs has resulted in graft transparency for >1 year.

### Artery patch xenotransplantation (Table 7)

Because of the development of thrombotic microangiopathy and/or consumptive coagulopathy, the assessment of immunosuppressive regimens or immunomodulatory approaches directed to the T cell response have been difficult to assess following pig-to-NHP organ transplantation. At our own center, Ezzelarab et al. have introduced a simple pig artery patch model which allows the adaptive immune response and, in particular, the production of T cell-dependent elicited anti-pig antibodies to be followed in the absence of the complicating factors of coagulation dysfunction (39). No such model had been reported in 1998, but today artery patch xenotransplantation is proving of value in assessing various immunosuppressive regimens.

### Skin xenotransplantation

In very complex experiments involving transduction of baboon bone marrow cells with SLA class II genes, followed by bone marrow transplantation and either pig kidney or skin transplantation, Ierino et al reported pig skin graft survival for 17 (control) or 21 days (40). Wiener et al reported prolonged GTKO pig skin graft survival in baboons (41). The GTKO skin grafts survived for up to 14 days whereas wild-type pig skin grafts were rejected by day 4.

### Discussion

In summary, we were unable to identify any reports on experimental studies of pig thymic tissue, neuronal cell, hepatocyte, corneal, skin, or artery patch transplantation in NHPs before 1998. Pig heart, kidney, and, particularly, islet graft survival have increased significantly since 1998. This has been associated largely with the increasing availability of pigs with genetic manipulations aimed at protecting the pig tissues from the primate immune response. Both GTKO and the introduction of human complement- and coagulation-regulatory transgenes have played a role, particularly with regard to heart transplantation. Although encouraging results have been achieved after the transplantation of wild-type pig islets into NHPs, the transplantation of islets from genetically-engineered pigs may allow a reduction in the intensity of the immunosuppressive therapy required to prevent graft loss.

Many successful immunosuppressive regimens in the pig-to-NHP model have been based on costimulation blockade with an anti-CD154 monoclonal antibody (mAb), which is unlikely to be available for clinical use because of its thrombogenic effects. Attention is now being directed towards replacing this agent with others, e.g., an anti-CD40 mAb (+/- an agent that blocks the CD28/B7 costimulation pathway). This problem will have to be resolved if approval for clinical trials is to be obtained by most national regulatory administrations, e.g., the Food and Drug Administration (FDA) in the USA. Recent studies by Mohiuddin and his colleagues (21–24) suggest that an anti-CD40 mAb-based regimen is likely to be successful, but this agent is not yet approved for clinical use. Blockade of the CD28/B7 pathway alone would appear to be inadequate (Iwase H et al, submitted).

A second topic that needs to be addressed before clinical trials are likely to be fully successful is that of expression of NeuGc in pigs (and in all NHPs), but not in humans (18). However, we would suggest that a successful GTKO pig transplant in a NHP provides an indicator of the likelihood of success of a GTKO/NeuGc-KO pig transplant in a human. We therefore continue to believe that the pig-to-NHP model provides very important data that cannot be obtained from *in vitro*, *ex vivo* perfusion, or other *in vivo* models. This particularly applies to the efficacy and safety testing of immunosuppressive protocols.

There are, of course, other differences in the biological (e.g., physiologic, immunologic) responses of NHPs and humans to transplanted pig organs, tissues, or cells, and resolutions to some of these may not be possible until clinical trials are undertaken. Nevertheless, we believe it is important to accumulate as many data from the pig-to-NHP model before proceeding to clinical trials, as this will provide the greatest likelihood of success. Whatever barriers identified in the pig-to-NHP model should be addressed before proceeding to the possibly more complex pig-to-human studies. Even though the management of a patient with a pig graft will undoubtedly be easier in many respects, failure from graft rejection or from the complications of excessive immunosuppressive therapy in the pig-to-NHP model is unlikely to be fully reversed in the pig-to-human model.

It is of interest to note the relative publication rates of the papers surveyed. For example, more papers relating to kidney xenotransplantation were published during the period covered by this survey than those relating to other organs. However, there was a marked reduction in the number of papers published after 2005 (Figure 1B). This may be associated with the increased problems related to pig kidney than to heart transplantation, where consumptive coagulopathy appears to be less problematic. However, after a peak period of publications in 2005, the rate of publication of papers reporting studies of pig heart transplantation also fell (Figure 1A).

The decline is possibly more likely a consequence of lack of sufficient funding for xenotransplantation research during the past few years, and this may also be a major factor in the reduction in publications relating to kidney transplantation after 2005 (Figure 1B). In turn, this decline in funding was at least in part associated with reluctance on the part of certain commercial/industrial sponsors to continue to participate in the development of xenotransplantation in view of the scares being propagated at that time relating to the transfer of porcine endogenous retroviruses (PERV) with the graft to the recipient. Since the first report of the potential of pig-to-human infection by PERV in 1996, much subsequent research has demonstrated that the risk of cross-species transmission is unlikely and, in any event, manageable (42). Of note, regulatory authorities are largely concentrating their attention on proper patient monitoring and archiving of samples rather than on preventing the introduction of xenotransplantation.

In contrast, pig islet xenotransplantation, which has provided very encouraging results, was reported fairly consistently throughout the period with between 1 and 4 publications per year (Figure 1C).

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## Abbreviations

<b>Gal</b>	galactose- $\alpha$ 1,3-galactose
<b>GTKO</b>	$\alpha$ 1,3-galactosyltransferase gene-knockout
<b>mAb</b>	monoclonal antibody
<b>NeuGc</b>	N-glycolylneuraminic acid
<b>NHP</b>	nonhuman primate
<b>PERV</b>	porcine endogenous retroviruses

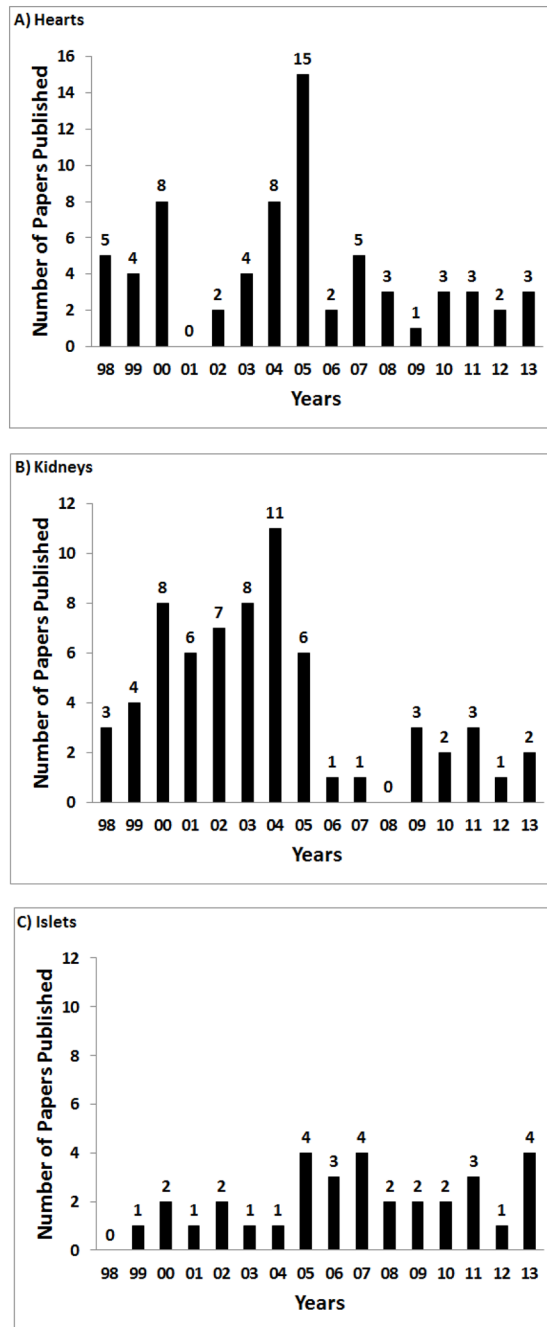
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**Figure 1.** Numbers of publications relating to pig-to-nonhuman primate heart (**A, top**), kidney (**B, middle**), and islet (**C, bottom**) transplantation during the period 1998–2013.

**Table 1A**

Heterotopic transplantation of pig hearts in NHPs (1998–2013)

FIRST AUTHOR (Year)	DONOR (pig)	RECIPIENT (n)	IMMUNOSUPPRESSIVE THERAPY	SURVIVAL-RANGE (MEDIAN) (Days, unless otherwise stated)
Simon (1998)	WT	Baboon (n=2)	Intravenous infusion of synthetic Gal oligosaccharides	<1 (4–6 hours)
Waterworth (1998)	CD55	Baboon (n=3)	CyP, CsA, CS	>2, 13, >21
Lin (1998)	CD55/CD59	Baboon (n=5) Baboon (n=6)	CyP, CsA, CS CyP, CsA, CS, Ig-depleted (immunoabsorption column)	<1–5 1–29
Bhatti (1999)	CD55	Baboon (n=14)	CyP, CsA, CS, MMF	10–99 (26)
Crespo (1999)	CD55	Baboon (n=13)	Not stated	<3 (n=2) 3–7 (n=11)
Kozłowski (1999)	WT (MSw)	Baboon (n=2)	TBI, TI, pig BMTx, splenectomy, immunoabsorption, ATG, CsA, MMF, CS, 15-deoxyspergualin	8, 15
Romano (1999)	WT	Baboon (n=1)	Intravenous infusion of synthetic Gal oligosaccharide	<1 (<18 hours)
Buhler (2000) Alwayn (2000)	WT (MSw)	Baboon (n=2)	TBI, TI, splenectomy, immunoabsorption, ATG, CVF, CsA or anti-CD154mAb, MMF or 15-deoxyspergualin (not clearly stated) +/- pig hematopoietic stem cells (n=1)	Not applicable (study of hemostasis)
Manez (2000)	WT CD55	Baboon (n=10) Baboon (n=10)	None (n=5) Immunoabsorption (n=5) None (n=5) Immunoabsorption (n=5)	<96 hours 87.6+/-35 hours 89.6+/-42 hours 101.6+/-23 hours
Lin (2000)	CD55/CD59	Baboon (n=5) Baboon (n=4)	CyP, CsA, CS Immunoabsorption, CyP, CsA CS	<1–10 (3) 9–39
Brenner (2000)	WT	Cynomolgus (n=1) Rhesus (n=2) Rhesus (n=4)	Immunoabsorption Immunoabsorption No immunoabsorption	} } } } 140+/-35 minutes 78+/-28 minutes
Lam (2002)	CD55	Cynomolgus (n=7)	CyP, CsA, CS, MMF	6–36 (23)
Schuurman (2002) (based on previous publications)	WT (n=7) CD55 (n=55) WT (n=5) CD55 (n=28)	Cynomolgus (n=62) Baboon (n=33)	CyP, CsA, CS, splenectomy +/- rapa +/- MMF +/- sCR1 CyP, CsA, CS, splenectomy +/- rapa +/- MMF +/- sCR1	WT: HAR 57% CD55: HAR 7% WT: HAR 20% CD55: HAR 11%.
Ashton-Chess (2003)	CD55	Baboon (n=2) Baboon (n=2) Baboon (n=9)	None Immunoabsorption Cyp, CsA, MMF, CS	4, 5 4, 6 6–29 (14)
Domenech (2003a,b)	CD55	Baboons (n=8)	CyP (high dose), CsA, CS, GAS914 (n=6) CyP (low dose), CsA, CS, GAS914 (n=2)	6–60 (27) 5, 7
Lam (2003)	CD55	Rhesus (n=2)	ATG, sCR1, tacrolimus, MMF, CS, GAS914	<1
Schirmer (2004)	CD46	Baboon (n=9) Baboon (n=9)	Anti-CD20mAb, tacrolimus, rapa, CS, TPC, clopidogrel, aspirin	15–30 (22) 4–53 (15)

FIRST AUTHOR (Year)	DONOR (pig)	RECIPIENT (n)	IMMUNOSUPPRESSIVE THERAPY	SURVIVAL-RANGE (MEDIAN) (Days, unless otherwise stated)
			Anti-CD20mAb, tacrolimus, rapa, CS, TPC	
Manez (2004a)	CD 55 CD55/CD46	Baboon (n=5) Baboon (n=5)	CsA, GAS914 CsA	5-8 (6) 4-9 (7)
Manez (2004b)	WT CD55 WT CD55	Baboon (n=5) Baboon (n=6) Baboon (n=5) Baboon (n=7)	None None Immunoabsorption Immunoabsorption	HAR in 3 of 5 3, 4 No HAR <1-<5 No HAR <2-5 No HAR <4-<6
McGregor (2004)	CD46	Baboon (n=10)	ATG, splenectomy, anti-CD20mAb, tacrolimus, rapa, CS, TPC	56-113 (76)
Houser (2004) Kuwaki (2004)	CD55	Baboon (n=10)	ATG, anti-CD2mAb, TI, CVF, anti-CD154mAb, MMF, CS	4-139 (27)
Lam (2004a) Lam (2004b)	CD55	Cynomolgus (n=15)	ATG or CyP, CsA or tacrolimus, MMF, CS, immunoabsorption, +/- GAS914	No GAS914: HAR in 4 of 6 4, 78 GAS914: HAR in 0 of 9 0-36 (20)
Chan (2005)	CD55	Cynomolgus (n=4)	CyP, CsA, MMF, CS, GAS914 +/- sCR1	20, 22, 35, 36
Stalder (2005)	CD55	Cynomolgus (n=6)	CyP or ATG, CsA or tacrolimus, MMF, CS, GAS914 +/- sCR1	2-36 (mean 30.5)
Teotia (2005)	CD46	Baboon (n=16)	Anti-CD20mAb, tacrolimus, rapa, CS, TPC	6-113 (mean 71)
Dor (2005)*	GTKO (MSw)	Baboon (n=8)	ATG, anti-CD154mAb, MMF, CS ATG, anti-CD154mAb, MMF, CS, recombinant human antithrombin III	16-179
Kuwaki (2005) Tseng (2005) Hisashi (2008) Shimizu (2008)	GTKO (MSw)	Baboon (n=8)	ATG, Anti-CD2mAb, TI, CVF, anti-CD154mAb, MMF, CS	>16-179 (63)
Moscoco (2005)	CD55 CD55/CD46	Baboon (n=9) Baboon (n=5)	CyP, CsA, MMF, CS, GAS914 (n=5) CsA, GAS914 (n=4) CsA, GAS914	50+/-19 6+/-1 6+/-2
Weaver (2005)	CD46	Baboons (n=8)	Anti-CD20mAb, tacrolimus, rapa, CS, TPC	0-92 (64)
Wu (2005)	CD55 CD46	Baboon (n=13) Baboon (n=5)	CyP, CsA, MMF, CS +/- anti-CD20mAb +/- ATG +/- GAS914 or TPC (n=10) ATG, anti-CD154mAb +/- anti-CD20mAb +/- CTLA4-Fc +/- GAS914 or TPC (n=8)	2-36 (12) 0-11 (6)
McGregor (2005)	CD46	Baboon (n=7)	Splenectomy, ATG, anti-CD20mAb, tacrolimus, rapa, CS, TPC	15-137 (96)
Byrne (2005)	CD46	Baboon (n=9) Baboon (n=13) Baboon (n=9)	Splenectomy, anti-CD20mAb, tacrolimus, rapa, TPC, warfarin + ATG or CyP for rejection episodes Splenectomy, anti-CD20mAb, tacrolimus, rapa, TPC, low molecular weight heparin+ ATG or CyP for rejection episodes Splenectomy, anti-CD20mAb, tacrolimus, rapa, TPC	3-62 (20) 5-109 (18) 4-53 (15)
Brenner (2005)	CD55	Baboon (n=4)	Immunoabsorption, CyP, CsA, CS, MMF	2-8 (10)
Davila (2006)	CD46	Baboon (n=1) Baboon (n=2) Baboon (n=1)	Splenectomy TPC, splenectomy TPC, anti-CD20mAb, splenectomy	5 6, 7 7

FIRST AUTHOR (Year)	DONOR (pig)	RECIPIENT (n)	IMMUNOSUPPRESSIVE THERAPY	SURVIVAL-RANGE (MEDIAN) (Days, unless otherwise stated)
Byrne (2006)	CD46	Baboon (n=63)	Splenectomy, TPC, anti-CD20mAb, tacrolimus, rapa, CS +/- aspirin/ clopidogrel or Lovenox or warfarin	0-139 (96)
Ricci (2007) (based on previous publications)	CD46	Baboon (n=64)	Groups 1-4 (n=40): (Low maintenance IS) Splenectomy, anti-CD20mAb, tacrolimus, rapa, CS, TPC +/- Lovenox or aspirin + clopidogrel or coumadin Group 5 (n=15): (High maintenance IS) Splenectomy, anti-CD20mAb, tacrolimus, rapa, CS, TPC, Lovenox +/- ATG as induction of treatment of rejection	Groups 1-4: 0-137 (30) Group 5: 0-139 (96)
Wu (2007)	CD55 CD46	Baboon (n=20) Baboon (n=3)	CyP, CsA, MMF (n=18); ATG, anti-CD154mAb (n=10); GAS914 (n=8) or TPC (n=3)+/- immunoadsorption; complement inhibitors (n= 12)	Technical failure (n=1) HAR (n=10) Early graft failure 1-3 (n=4) AHXR 6-36 (15) (n=8)
Zahorsky-Reeves (2007)	CD55	Cynomolgus (n=3)	ATG or CyP, CsA, MMF, CS +/- GAS914	36, 39, 78
Byrne (2008)	CD46 GTKO	Baboon (n=4) Baboon (n=8)	Splenectomy +/- TPC +/- anti-CD20mAb Splenectomy, ATG, anti-CD20mAb, tacrolimus rapa	5-7 0-128 (25)
Ezzelarab (2009)	GTKO	Baboon (n=9)	ATG, CVF, anti-CD154mAb, MMF, CS	2-56
Bauer (2010) (Intrathoracic)	WT GTKO/CD46	Baboon (n=2) Baboon (n=2)	None ATG, anti-CD20mAb, tacrolimus, rapa, MMF, CS, bortezomib, immunoadsorption (n=1)	Euthanized after weaning from cardiopulmonary bypass <1, 50
Tazelaar (2011) (partially based on previous publications)	CD46 CD46 GTKO +/- CD55	Baboon (n=11) Baboon (n=8) Baboon (n=5)	ATG, anti-CD20mAb, tacrolimus, rapa, TPC Immunoadsorption, CyP, CsA ATG, anti-CD20mAb, tacrolimus, rapa	15-109 (41) 8-42 (13) 18-71 (26)
Mohiuddin (2012) Corcoran (2010) Horvath (2010)	GTKO/CD46	Baboon (n=2) Baboon (n=2) Baboon (n=9)	No IS ATG, CVF, anti-CD154mAb, MMF, CS ATG, anti-CD20mAb, CVF, anti-CD154mAb, MMF, CS	< 1 8, 8 36-236 (71)
McGregor (2012)	GTKO GTKO/CD55	Baboon (n=6) Baboon (n=5)	ATG, anti-CD20mAb, tacrolimus, rapa, CS ATG, anti-CD20mAb, tacrolimus, rapa, CS	<1-128 (21) 15-52 (28)
Kim (2013)	GTKO	Cynomolgus (n=4)	ATG, anti-CD20mAb, CVF, anti-CD154mAb, tacrolimus, CS	11-24 (14)
Mohiuddin (2013)	GTKO/CD46	Baboon (n=9)	ATG, anti-CD20mAb, CVF, anti-CD40mAb (either 3A8 [n=3] or 2C10R4 [n=6]), MMF, CS	3A8 = 21, 21, 28 (21) 2C10R4 = >30, >40, 60, 107, 146, 149 (84)
Mohiuddin (2013/4)	GTKO/CD46/TBM	Baboon (n=5)	ATG, anti-CD20mAb, CVF, anti-CD40mAb, MMF, CS	0->380 (4 ongoing at 77-380 days)

## Abbreviations:

ATG = anti-thymocyte globulin; BMTx = bone marrow transplant; CD46 = membrane cofactor protein; CD55 = decay-accelerating factor; CD59 = protectin, membrane inhibitor of reactive lysis; CS = corticosteroids; CsA = cyclosporine; CTLA4-Fc = CTLA4 covalently linked to a human immunoglobulin Fc molecule; CVF = cobra venom factor; CyP = cyclophosphamide; EGF = early graft failure; GAS914 = a soluble

glycoconjugate comprising Gal on a poly-L-lysine backbone; GTKO =  $\alpha$ 1,3-galactosyltransferase gene-knockout; HAR = hyperacute rejection; IS = immunosuppressive therapy; LoCD2b = rat anti-primate CD2b monoclonal antibody; MMF = mycophenolate mofetil (or analog, e.g., mycophenolate sodium); MSw = miniature swine (MGH herd); Rapa = rapamycin (or derivative, e.g., RAD); sCR1 = soluble complement receptor type 1 (in some papers described as TP10); TBI = total body irradiation; TI = thymic irradiation, TPC= an  $\alpha$ Gal-polyethylene glycol polymer conjugate.

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**Table 1B**

Orthotopic transplantation of pig hearts in NHPs (1998–2013)

FIRST AUTHOR (Year)	DONOR (pig)	RECIPIENT (n)	IMMUNOSUPPRESSIVE THERAPY	SURVIVAL-RANGE (MEDIAN) (Days unless otherwise stated)
Schmoeckel (1998)	CD55	Baboon (n=10)	CyP, CsA, CS	<1 (x5) 4–9 (x5)
Waterworth (1998)	CD55	Baboon (n=5)	CyP, CsA, CS	>1, >1, 5, 5, 9
Xu (1998)	WT	Baboon (n=2)	Immunoabsorption (through another donor organ), TBI, CsA, methotrexate,	18, 19
Brenner (2000a)	WT	Cynomolgus (n=2)	Immunoabsorption	130+/-21 <u>minutes</u>
Brenner (2000b,c)	WT	Baboon (n=4)	None (n=1) Immunoabsorption (n=3)	29 <u>minutes</u> <2, 11, 21 <u>hours</u>
Vial (2000)	CD55	Baboon (n=1)	CyP, CsA, MMF, CS	39
Schuurman (2002) (based on previous publications)	CD55	Baboon (n=16)	CyP, CsA, CS, splenectomy +/- rapa +/- MMF +/- sCR1	HAR 1 (6%)
Brandl (2005)	CD55	Baboon (n=4)	ATG, tacrolimus, rapa, CS, GAS914 +/- CyP	1–25 (9)
Brenner (2005)	CD55	Baboon (n=4)	CyP, CsA, MMF, CS	<1, 11, 13, 20
Bauer (2005)	CD55	Baboon (n=9)	ATG, tacrolimus or CsA, rapa, CS, GAS914,	Not stated
Brandl (2007)	CD46 or CD55	Baboon (n=2) Baboon (n=2) Baboon (n=4) Baboon (n=5)	ATG, tacrolimus, rapa, CS, GAS914 ATG, anti-CD20mAb, tacrolimus, rapa, CS, GAS914 ATG, low-dose CyP, tacrolimus, rapa, CS, GAS914 ATG, CyP, tacrolimus, rapa, CS, anti-HLA-DR antibody, +/- GAS914+TPC	1, 9 <2 (1 technical failure) <1, 14, 25 (1 technical failure) <1–4
Bauer (2007)	CD55 CD46	Baboon (n=6) Baboon (n=6)	Not stated	Not stated
Bauer (2011)	CD46	Baboon (n=6)	ATG +/- CyP, tacrolimus, rapa, CS, GAS914 or TPC, anti-HLA antibody	Not stated
Byrne (2011)	CD46 or CD55 or CTKO/CD55		ATG or CyP, tacrolimus, rapa +/- anti-CD20mAb +/- GAS914 or TPC	0–57 (6)

Abbreviations as for Table 1A

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Table 2

Transplantation of pig kidneys into NHPs (1998–2013)

FIRST AUTHOR (YEAR)	DONOR (PIG)	RECIPIENT (n)	IMMUNOSUPPRESSIVE THERAPY	SURVIVAL-RANGE (MEDIAN) (Days)
Ierino (1998)	WT (MSw)	Baboon (n=3)	TBI, BMTx (autologous, transfected with SLA class II (n=2)), splenectomy, apheresis, immunoadsorption, sCR-1, CsA, MMF, CS, 15-deoxyspergualin	8, 12, 13
Xu (1998)	WT (MSw)	Cynomolgus (n=10)	Immunoadsorption (Gal column n=4; pig liver n=6)	2–12 (mean 7)
Zaidi (1998)	WT	Cynomolgus (n=6)	CyP, CsA, CS	<1–30 (7)
	CD55	Cynomolgus (n=7)		6–35 (13)
Schmoeckel (1999)	CD55	Cynomolgus (n=11)	Splenectomy, CyP, CsA, MMF, CS (n=5) CyP, CsA, MMF, CS (n=6)	Median 43 Median 15 (but 4 deaths with functioning kidney)
Ierino (1999) [Ierino (1998)]	WT (MSw)	Baboons (n=4; life-supporting in only one)	SLA class II gene transduction of baboon CD34 <sup>+</sup> and CD34 <sup>-</sup> bone marrow cells (n=3)	Uncertain
Kozlowski (1999)	WT (MSw)	Baboon (n=2)	TBI, TI, pig BMTx, splenectomy, Immunoadsorption, ATG, CsA, CS, MMF and/or 15-DSG	9, 11
	WT (MSw)	Baboon (n=2)	As above (modified)	3, 6
Meyer (1999)	WT	Baboon (n=5)	Plasmapheresis, immunoadsorption	1–5 (3)
	vWD	Baboon (n=5)		1–5 (4)
Loss (2000)	WT	Cynomolgus (n=7)	(Non-life supporting - study of effect of cold ischemia on HAR)	<1
	WT	Cynomolgus (n=8)	Life-supporting - study of effect of cold ischemia on HAR	<1
Cowan (2000)	WT	Baboon (n=4)	None	<1
	CD55/H-transferase	Baboon (n=2)		2
	CD55/CD59/H-transferase	Baboon (n=2)		2, 3
	WT	Baboon (n=4)		3–5
Cozzi (2000) [Bhatti (1998)]	WT	Cynomolgus (n=5)	CyP, CsA, CS, splenectomy	0–30 (0)
	CD55	Cynomolgus (n=9)	CyP, CsA, CS, splenectomy	5–78 (39)
	WT	Cynomolgus (n=7)	CyP, CsA, CS	6–35 (13)
Dehoux (2000)	WT	Baboon (n=4)	None (splenectomy in 1)	<1
	WT (MSw)	Baboon (n=5)	anti-IgMmAb	4–6 (4)
Buhler (2000)	WT (MSw)	Baboon (n=1)	TBI, TI, ATG, splenectomy, immunoadsorption, CVF, CsA or anti-CD154mAb, MMF or 15-deoxyspergualin (not clearly stated)	Not applicable (study of hemostasis)

FIRST AUTHOR (YEAR)	DONOR (PIG)	RECIPIENT (n)	IMMUNOSUPPRESSIVE THERAPY	SURVIVAL-RANGE (MEDIAN) (Days)
		Baboon (n=3)	As above + pig hematopoietic cells	0, 1, 7, 8
		Cynomolgus (n=4)	Immunoabsorption, splenectomy, CsA, 15-deoxyspergualin	0-15 (9.5)
		Cynomolgus (n=10)	As above + TBI, TI, ATG, BMTx (n=1)	9, 11
		Baboon (n=2)		
Shimizu (2000)	WT (MSw)	Baboon (n=4)	As above + TBI, TI, ATG, MMF or brequinar, BMTx (n=1)	3, 6, 6, 14
		Baboon (n=5)	As above + TI, ATG (n=1), MMF, sCRI or CVF, BMTx	6-13 (8)
		Baboon (n=2)	CyP, CsA or tacrolimus, CS, plasmapheresis	6, 7
		Baboon (n=2)	As above + additional plasmapheresis	<1
		Cynomolgus (n=8)		1-11 (3.5)
Loss (2000)	WT	Cynomolgus (n=9)	CyP, CsA, CS	1-68 (11)
	CD55			
Przemek (2001)	WT	Cynomolgus (n=7)	CyP, CsA, CS	Not stated (study of cardio-circulatory parameters)
	CD55	Cynomolgus (n=6)		
		Baboon (n=3)	Splenectomy, TBI, TI, ATG, immunoabsorption, CVF, anti-CD154mAb, MMF, CS	4, 6, 8
Bühler (2001)	WT (MSw)	Baboon (n=2)	As above + CyP, but no TBI	7, 13
		Baboon (n=3)		28, 29, 29
	CD55		CyP, CsA, CS	3, 4
		Cynomolgus (n=2)		
Vangerow (2001)	CD55	Cynomolgus (n=4)	As above + treatment of rejection with CyP, CS	9, 11, 11, 15
		Cynomolgus (n=4)	As above + treatment of rejection with CyP, CS, C1-INH	18, 21, 28, 68
		Cynomolgus (n=12) (donor kidney <50g)	CsA, CyP, CS:	<1-<15 (3)
Loss (2001)	WT	Cynomolgus (n=3) : (donor kidney >70g)	CsA, CyP, CS	1, 4, 11
		Cynomolgus (n=7)	CyP, CsA, CS (Non life-supporting graft. Study on transmission of porcine endogenous retrovirus and porcine chimerism)	4-287 (28)
Loss (2001)	WT	Cynomolgus (n=5)	CyP, CsA, CS (Study on transmission of porcine endogenous retrovirus and porcine chimerism)	1-11 (3)
		Baboon (n=5)	CyP, CsA, CS	3-7 (mean 5+/-1)
		Baboon (n=6)	Splenectomy, immunoabsorption, CyP, CsA, CS	6-12 (mean 10+/-1)
Dean (2001)	CD55	Baboon (n=3)	Splenectomy, intensive immunoabsorption, CyP, CsA, CS	13-15 (mean 14+/-0.3)

FIRST AUTHOR (YEAR)	DONOR (PIG)	RECIPIENT (n)	IMMUNOSUPPRESSIVE THERAPY	SURVIVAL-RANGE (MEDIAN) (Days)
McInnes (2002)	WT	Cynomolgus (n=11)	CyP, CsA, CS, splenectomy +/- rapa +/-MMF +/-sCRI (Study on post-transplant lymphoproliferative disease)	<1-78 (6.5)
	CD55	Cynomolgus (n=234)		
Hecker (2002)	WT	Cynomolgus (n=3)	CyP, CsA, MMF, CS, CI-INH	5, 13, 15
Richards (2002)	CD55	Cynomolgus (n=20)	CyP, CsA, CS +/-rapa +/-MMF +/-sCRI	4-60 (31)
	WT	Cynomolgus (n=11)	CyP, CsA, CS, splenectomy +/- rapa +/-MMF +/-sCRI	HAR: n=3
Schuurman (2002)	CD55	Cynomolgus (n=234)		HAR: n=0
	Ghanekar (2002)	CD55	Baboon (n=2)	CsA, GAS914, CS, CyP, rapa
Baboon (n=4)			ATG, CsA, GAS914, CS, rapa	20, 22, 23, 26
Baboon (n=3)			ATG, CsA, GAS914, CS	18, 21, 22
Baboon (n=5)			Anti-IgM mAb	4-6(4)
Cowan (2002)	CD55/CD59	Baboon (n=1)	Low molecular weight heparin	<1-6
		Baboon (n=3)	Low molecular weight heparin, recombinant human antithrombin III	
		Baboon (n=1)	Recombinant human antithrombin III	
Zhong (2003)	CD55	Baboon (n=4)	CyP, CsA, CS, rapa	4, 4, 26, 40
		Baboon (n=12)	CyP, CsA, CS, rapa, GAS914	7-37 (14)
Barth (2003)	CD55	Baboon (n=5)	(Thymokidneys), thymectomy/TI, splenectomy, immunoadsorption, anti-CD2mAb, ATG/anti-CD3IT, anti-CD154mAb, CyP, CVF, MMF, CS	24-229 (27)
		WT (MSw)	TI, splenectomy, immunoadsorption, ATG, anti-CD154mAb, CyP, CVF, MMF, CS	7, 13
Gollaekner (2003)	CD55	Baboons (n=2)	As above + BSA-Gal	28, 29, 29
	WT (MSw)	Baboons (n=3)		
Cozzi (2003)	CD55	Baboons (n=4)	Splenectomy, CsA, CyP, Cs, MMF	8, 9, 11, 11
	WT	Cynomolgus (n=10)		
Ashton-Chess (2003)	CD55	Baboon (n=7)	Immunoadsorption, CyP, CsA, MMF, CS	2-51 (22)
	CD55	Baboon (n=4)	As above	4-8 (5)
	CD55	Baboon (n=4)	As above, but no immunoadsorption	5, 5, 5, 9
	CD55	Baboon (n=4)	As above, but immunoadsorption x1	7, 8, 10, 12
	CD55	Cynomolgus (n=5)	Methotrexate (6 doses), CsA, MMF, CS	5, 5, 7, 10
Cozzi (2003)	CD55	Cynomolgus (n=3)	Methotrexate (4 doses), CsA, MPS, CS	6, 9, 9, 10, 39
		Cynomolgus (n=2)	Methotrexate (5 doses), CsA, CS	0, 16, 34
				16, 41

FIRST AUTHOR (YEAR)	DONOR (PIG)	RECIPIENT (n)	IMMUNOSUPPRESSIVE THERAPY	SURVIVAL-RANGE (MEDIAN) (Days)
Gollackner (2003)	WT (MSw)	Baboon (n=3)	Immunoadsorption, splenectomy, Tl, ATG, CVF, anti-CD154mAb, MMF, CS, CyP	6, 7, 13
		Baboon (n=2)	As above + BSA-Gal	8, 11
		Baboon (n=2)	As above + BSA-Gal, but no CyP	9, 12
Kaosalla (2003)	WT (MSw) CD55	Baboon (n=3)	Immunoadsorption, splenectomy, Tl, ATG, CVF, CyP, anti-CD154mAb, MMF, CS	6, 7, 13
		Baboon (n=3)		28, 29, 29
Ashton-Chess (2004)	CD55/CD59	Baboon (n=2)	None	5, 6
		Baboon (n=4)	CsA, MMF, CS, mitoxantrone	6, 7, 8, 10 (8)
Loveland (2004)	WT CD46	Baboon (n=2)	As above, but CyP not mitoxantrone	9, 9
		Baboon (n=7)	(Non-life-supporting) None	Not stated
Key (2004)	CD55	Baboon (n=9)		
		Cynomolgus (n=52)	Various	1-53 (17.5)
Lam (2004)	CD55	Cynomolgus (n=4)	CyP, CsA, MMF, CS	<0, <0, <0, 4
		Cynomolgus (n=4)	As above + GAS914	6, 12, 31, 37
Cozzi (2004)	CD55 (Donor pig pretreated with carbon monoxide)	Cynomolgus (n=5)	CyP, CsA, MMF, CS, GAS914	2-37 (12)
		Cynomolgus (n=7)	CyP, CsA, MMF, CS	1-90 (48)
Baldan (2004)	CD55	Cynomolgus (n=10)	As above + methorexate	0-39 (13)
		Cynomolgus (n=5)	As above + GAS914	7-38 (21)
		Cynomolgus (n=6)	(Donor pig pretreated with carbon monoxide) As above + GAS914	2-37 (8.5)
		Cynomolgus (n=2)	As above + recombinant human antithrombin III	23, 23
Diaz (2004)	CD55	Baboon (n=9)	Not stated	Not stated
Garcia (2004)	WT CD55/CD59	Baboons (n=27)	CyP/ATG, CsA/taecrolimus, rapa/MMF, CS, GAS914	4-75 (mean 21)
		Baboon (n=2)	None	<1
Menoret (2004)	WT (MSw) CD55/CD59	Baboon (n=2)		5, 6
		Baboon (n=2)		
Gollackner (2004)	WT (MSw) CD55	Baboon (n=6)	WBI/CyP, IAD, SpX, Tl, ATG, CVF, anti-CD154, MMF, CS	4-13 (6.5)
		Baboon (n=3)		28, 29, 29
		Baboon (n=4)	Same, in addition BSA-Gal +/- CyP	8, 9, 11, 11
Ghanekar (2004)	CD55	Baboon	Various (based on Ghanekar 2002 and Zhong 2003)	

FIRST AUTHOR (YEAR)	DONOR (PIG)	RECIPIENT (n)	IMMUNOSUPPRESSIVE THERAPY	SURVIVAL-RANGE (MEDIAN) (Days)
Yamada (2005)	GTKO (MSw)	Baboon (n=6)	(Vascularized thymic lobe) +/- WB1, thymectomy, splenectomy, anti-CD2mAb, anti-CD154mAb, MMF, CS, +/- CVF	4-68 (32)
		Baboon (n=5)	(Thymokidney) As above	16-83 (26)
		Baboon (n=3)	(Kidney) As above	20-34 (33)
Chen (2005)	GTKO	Baboon (n=3)	ATG, tacrolimus, CS	8, 10, 11
		Baboon (n=3)	As above + MMF, CVF	9, 13, 16
Moscoso (2005)	CD55	Baboon (n=8)	GAS914, CyP, CsA, MMF, CS	11±9
		Baboon (n=5)	Anti-CD25mAb, FTY, CsA, CS	12±10
Sun (2005)	CD55 (from supplier A)	Baboon (n=4)	CyP, CsA, rapa	4, 4, 26, 40
		Baboon (n=10)	As above + GAS914	9-37 (17)
		Baboon (n=3)	ATG, CsA, rapa, GAS914	20, 23, 26
		Baboon (n=1)	ATG, tacrolimus, MMF, GAS914/TPC	75
		Baboon (n=5)		7-16 (13)
Shimizu (2005)	CD55	Baboon (n=6)	(Thymokidney) Splenectomy, thymectomy/TI, immunoadsorption, ATG/anti-CD3IT, CyP, CVF, MMF, anti-CD154mAb	9-27 (12)
		Baboon (n=10)		2-30 (16)
		Cynomolgus (n=3)	CyP, CsA, MMF, CS, GAS	6, 12, 31
Lam (2005)	CD55	Cynomolgus (n=2)	As above + sCR-1	3, 15
		Cynomolgus (n=3)	As above + GAS914 + sCR-1	10, 20, 32, 37
Chen (2006)	CD55	Baboon (n=5)	GAS914/TPC, ATG, tacrolimus, MMF, CS	7-75 (13)
		Baboon (n=2)	GAS914/TPC, ATG, tacrolimus, CVF, anti-CD20mAb	8, 14
Cavicchioni (2007)	CD55	Cynomolgus (n=7)	CyP, CsA, MMF, CS	1-90 (20)
		Cynomolgus (n=8)	As above, but methotrexate not CyP	0-39 (10)
Yazaki (2009)	CD55/endo-β-galactosidase C	Baboon (n=4)	Splenectomy, CyP, tacrolimus, CS	2, 8, 9, 11
		Baboon (n=6)	Not stated	Not stated
Knosalla (2009)	GTKO (MSw)	Baboon (n=14)	(+/- vascularized thymic lobe or thymokidney) Not stated	Not stated
		Baboon (n=7)	Thymectomy, splenectomy, TBI (n=1), ATG +/- anti-CD2mAb, anti-CD154mAb, tacrolimus, MMF, anti-CD20mAb	18-83 (49)
Lin (2010)	GTKO/CD46	Baboon (n=1)	None	<1
		Baboon (n=1)		2

FIRST AUTHOR (YEAR)	DONOR (PIG)	RECIPIENT (n)	IMMUNOSUPPRESSIVE THERAPY	SURVIVAL-RANGE (MEDIAN) (Days)
	WT	Baboon (n=1)	ATG, anti-CD154, MMF, CVF, CS	6
	GTKO	Baboon (n=1)		7
	GTKO/CD46	Baboon (n=1)		4
Griesemer (2010)	GTKO (MSw)	Baboon (n=4)	As above, but no CVF or CS	9, 10, 10, 16
		Baboon (n=2)	TBI, TI, splenectomy, ATG, anti-CD20mAb, tacrolimus, GTKO BMTx +/- CVF (n=1)	8, 11
Simioni (2011)	CD55	Cynomolgus (n=2)	Extracorporeal kidney perfusion, GAS914, CyP, CsA, MMF, CS	28, 55
		Cynomolgus (n=2)	As above + recombinant human antithrombin III + recombinant human activated protein C	12, 34
		Cynomolgus (n=9)	As above + recombinant human activated protein C	8-37 (20)
Nishimura (2011)	GTKO (MSw)	Baboon (n=2)	(Thymokidney) Thymectomy, splenectomy, anti-CD3IT +/- anti-CD20mAb, ATG, anti-CD20mAb, tacrolimus, MMF, anti-CD154mAb	15, 15
		Baboon (n=2)	(Thymokidney) As above, but no anti-CD3IT	14, 14
Le Bas Bernadet (2011)	GTKO/CD55/CD59/H-transferrase	Baboon (n=2)	None	3, 4
		Baboon (n=4)	Splenectomy (n=2), CyP, tacrolimus, MMF, CS, C1-INH	4, 12, 13, 15
Shimizu (2012)	GTKO (MSw)	Baboon (n=2)	ATG, anti-CD20mAb, anti-CD154, MMF, CS	20-33
		Baboon (n=1)	As above + thymectomy, splenectomy, TBI	34
Pintore (2013)	GTKO/CD55/CD59/CD39/H-transferrase	Baboon (n=4)	(Vascular thymic lobe or thymokidney) Thymectomy, splenectomy, ATG, anti-CD20mAb, anti-CD154mAb, MMF, CS +/- TBI (n=1)	56, 68, 81, 83
		Cynomolgus (n=5)	CyP, CsA, MMF, CS	18±3.2 (16)
Spiezia (2013)	CD55 (n=4) GTKO/CD55 (n=2) GTKO/CD55/CD59/H-transferrase	Cynomolgus (n=3)	Anti-CD20mAb, CsA, MMF, CS	13±2.3 (12)
		Cynomolgus (n=8)	CyP or anti-CD20mAb, CsA, MMF, CS	19, 30, 34, 55
		Cynomolgus (n=2)		12, 13
				8, 8

Abbreviations (if not used previously):

Anti-CD3IT = anti-CD3 immunotoxin; BSA-Gal = bovine serum albumin conjugated to Gal oligosaccharides; C1-INH = complement component 1 inhibitor; FTY = sphingosine 1-phosphate receptor agonist; vWD = pigs homozygous for von Willebrand disease

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**Table 3**

Transplantation of pig livers into NHPs (1998–2013)

FIRST AUTHOR (Year)	DONOR (pig)	RECIPIENT (n)	IMMUNOSUPPRESSIVE THERAPY	SURVIVAL – RANGE (MEDIAN) (Hours)
Luo (1998)	WT	Baboons (n=2) Rhesus (n=6)	CyP, CsA, CS None (n=3) Cyp, CsA, CS, Dashen (traditional Chinese medicine) (n=3)	<2 <6
Ramirez (2000)	WT CD55	Baboon (n=3) Baboon (n=2)	CyP, CsA, CS	<12 96,192
Ramirez (2005)	WT CD46/CD59/FT	Baboon (n=4) Baboon (n=5)	Anti-CD25mAb, CyP, anti-CD20mAb, CsA, MMF, CS	<16 13–24 (20)
Ekser (2010)	WT GTKO GTKO/CD46	Baboon (n=1) Baboon (n=2) Baboon (n=8)	ATG, CyP, CVF, tacrolimus, MMF, CS	<24 <24,144 <24–168 (144)
Kim (2012)	GTKO	Baboon (n=3)	ATG, LoCd2b, CVF, anti-CD154mAb, azathioprine, tacrolimus, CS	72–216

Abbreviations as used previously

Table 3: References

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**Table 4**

## Transplantation of pig lungs into NHPs (1998–2013)

FIRST AUTHOR (Year)	DONOR (pig)	RECIPIENT (n)	IMMUNOSUPPRESSIVE THERAPY	SURVIVAL (Hours)
Daggett (1998)	WT	Baboon (n=7)	Pre-Tx perfusion with pig lungs (n=4), CsA, azathioprine, CS (n=7)	<11
Yeatman (1998)	WT or CD55/CD59	Baboon (n=10)	CsA, azathioprine, CS +/- CVF	<3
Lau (2000)	CD55/CD59	Baboon (n=9)	Immunodepletion (3 different methods), CsA, CS, CyP or azathioprine	<24 (details not stated)
Gaca (2002)	CD46	Baboon (n=7)	Immunodepletion, dexamethasone, CsA, CyP, indomethacin, azathioprine +/- anti-human GPIb mAb	<9
Gonzalez-Stawinski (2002)	CD46	Baboon (n=8; control 4, experimental 4)	Anti-Gal antibody depletion, CsA, CS, CyP or azathioprine, gamma globulin, anti-CD20 mAb, splenectomy	<16
Lau (2003)	vWF-deficient	Baboon (n=5)	CsA, CS, azathioprine	<5
Gaca (2006)	CD46	Baboon (n=5)	Immunodepletion +/- anti-human C5a mAb (n=3)	<12
Cantu (2007)	WT/macrophage and vWF-deficient	Baboon (n=15)	Immunodepletion	19–109
Nguyen (2007)	CD46 or GTKO	Baboon (n=6)	CS (details not stated)	<4
Bush (2011)	GTKO/CD55, PIM-depleted (Clodronate liposomes to donor pig)	Baboon (n=2)	CsA, azathioprine, CS	3–48

Abbreviations (if not used previously)

PIM = pulmonary intravascular macrophages

Table 4: References

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**Table 5**

Transplantation of pig pancreatic islets in NHPs (1998–2013)

FIRST AUTHOR (Year)	DONOR (Pig)	RECIPIENT (n)	IMMUNOSUPPRESSIVE THERAPY	SURVIVAL – RANGE (MEDIAN) (Days)
Söderlund (1999)	WT fetal islet-like cell clusters	Cynomolgus (n=8) Cynomolgus (n=6)	None CsA, 15-deoxyspergualin	<6 <12
Rijkelijkhuisen (2000a)	WT adult	Cynomolgus (n=4)	CyP, CsA, CS	4–11
Rijkelijkhuisen (2000b)	WT adult	Cynomolgus (n=4)	CyP, CsA, CS	4–11
Jonker (2001)	WT adult	Rhesus (n=4)	ATG, CsA, CS, anti-IL2RmAb	21–53
Buhler (2002)	WT adult WT adult islets + PBPC infusion	Baboon (n=3) Baboon (n=2)	Splenectomy, ATG, CsA, azathioprine Splenectomy, multiple immunoadsorptions, TBI, TI, ATG, CVF, anti-CD154mAb, CsA, MMF, CS	<2 12, 28
Cantarovich (2002)	WT adult	Baboon (n=4) Cynomolgus (n=1)	ATG, CsA, MMF, CS ATG, deoxyspergualin, MMF, CS	<2 <2
Rijkelijkhuisen (2003)	WT adult	Cynomolgus (n=4) Rhesus (n=4)	CyP, CsA, CS ATG, CsA, CS, anti-IL2RmAb	4–11 21–53
Kirchhof (2004)	WT adult	Rhesus (n=6)	None	<1–>3
Elliott (2005a)	WT neonatal	Cynomolgus (n=2)	Encapsulation, nicotinamide	56
Isaac (2005)	WT neonatal + Sertoli cells	Cynomolgus (n=7)	None	<56
Elliott (2005b)	WT neonatal	Cynomolgus (n=8)	Encapsulation, nicotinamide	>252
Komoda (2005)	WT adult GnT-III adult	Cynomolgus (n=3) Cynomolgus (n=4)	None None	1–3 1–5
Hering (2006)	WT adult	Cynomolgus (n=3) Cynomolgus (n=4) Cynomolgus (n= 5)	Anti-CD25mAb, FTY720, rapamycin Anti-CD25mAb, FTY720, rapamycin, anti-CD154mAb Anti-CD25mAb, FTY720/tacrolimus, rapamycin, anti-CD154mAb, leflunomide	24–45 47–187 68–>158
Cardona (2006)	WT neonatal	Rhesus (n=9)	Anti-CD25mAb, anti-CD154mAb, CTLA4Ig (belatacept), rapamycin	4–>260 (140)
Dufrane (2006)	WT adult	Cynomolgus (n=2) Cynomolgus (n=12)	None Encapsulation	<7 <7–60 (30)
Gianello (2007)	WT adult	Cynomolgus (n=4) Cynomolgus (n=4)	Encapsulation, (renal subcapsular) Encapsulation (subcutaneous with islet mono-layer device)	Data not available
Rood (2007)	WT and GTKO adult	Cynomolgus (n=2) Cynomolgus (n=4) Cynomolgus (n=4)	ATG, anti-CD20mAb, tacrolimus, rapamycin ATG, CVF, anti-CD154mAb, MMF or rapamycin + tacrolimus ATG, anti-CD154mAb, MMF	<5 >58 (partial function) 5–7
Cardona (2007)	WT adult	Rhesus (n=5)	Anti-CD25mAb, anti-CD154mAb, CTLA4-Ig (belatacept), rapamycin	3–76
Rogers (2007)	WT embryonic pancreatic primordia	Rhesus (n=3)	No IS, multiple transplants	78–409
Casu (2008)	WT adult	Cynomolgus (n=9)	ATG, anti-CD154mAb, MMF	Partial function >60
Garkavenko (2008)	WT neonatal	Cynomolgus (n=12)	No IS	>180
van der Windt (2009)	CD46 adult	Cynomolgus (n=9)	ATG, anti-CD154mAb, MMF	5–396 (46)

FIRST AUTHOR (Year)	DONOR (Pig)	RECIPIENT (n)	IMMUNOSUPPRESSIVE THERAPY	SURVIVAL – RANGE (MEDIAN) (Days)
Hecht (2009)	Fetal pancreatic fragments	Cynomolgus (n=2)	ATG, anti-CD25mAb, anti-CD20mAb, FTY720, rapamycin, CTLA4-Ig	280, 380
Igarashi (2010)	WT adult	Cynomolgus (n=4) Cynomolgus (n=5)	Microencapsulation Macrodevice	>42 >180
Dufrane (2010)	WT adult	Cynomolgus (n=4) Cynomolgus (n=5)	Microencapsulation Macrodevice	14 136–180
Rogers (2011)	WT embryonic pancreatic primordia WT adult	Rhesus (n=3) Rhesus (n=3)	None None	56 (Experiments were electively terminated)
Thompson (2011a)	WT neonatal	Rhesus (n=9)	Anti-CD25mAb, anti-CD40mAb, rapamycin, CTLA4-Ig (belatacept)	47–203 (80)
Thompson (2011b)	GTKO neonatal (n=5) WT neonatal (n=5)	Rhesus	Anti-CD154mAb, anti-LFA1mAb, MMF, CTLA4-Ig (belatacept)	50–249 (137)
Thompson (2012)	WT neonatal	Rhesus (n=3) Rhesus (n=5) Rhesus (n=5)	MMF, CTLA4-Ig (belatacept), anti-LFA-1mAb, basiliximab MMF, CTLA4-Ig (belatacept), anti-LFA-1mAb, basiliximab, tacrolimus MMF, CTLA4-Ig (belatacept), anti-LFA-1mAb, alefacept, tacrolimus	<50 (none engrafted) 46–99 92–114
Kim (2013)	WT adult Msw	Rhesus (n=3)	None	<5
Lee (2013)	WT adult	Rhesus (n=2)	Not stated	>120
Vériter (2013)	WT adult	Cynomolgus (n=4) Cynomolgus (n=6)	Coencapsulation with bone marrow-driven stem cells Coencapsulation with adipose-derived stem cells	1–217 14–224
Graham (2013)	WT adult	Cynomolgus (n=not stated) Cynomolgus (n=1)	Various (review of previous data from Hering [2006] and additional data) Anti-CD25mAb, CTLA4-Ig (abatacept), tacrolimus, rapamycin	Various >180

Abbreviations (if not used previously):

GnT-III= N-acetylglucosaminyltransferase III

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**Table 6**

Transplantation of pig corneas into NHPs (1998–2013)

FIRST AUTHOR (Year)	DONOR (Pig)	RECIPIENT (n)	IMMUNOSUPPRESSIVE THERAPY	SURVIVAL - RANGE (MEDIAN) (Days)
Amano (2003)	WT	Cynomolgus (n=6)	None (Stromal disk into stromal pocket)	60–180 (165)
Pan (2007)	WT	Rhesus (n=6) Rhesus (n=4) Rhesus (n=4)	None (PKP) Local betamethasone (PKP) None (ALK)	12–18 (15) 129–276 (183) >90
Li (2011)	WT	Rhesus (n=5) Rhesus (n=5) Rhesus (n=5)	None (ALK, dehydrated) None (ALK, fresh) Local triamcinolone (ALK, dehydrated)	>180 >180 >180
Choi (2011)	WT	Rhesus (n=5) Rhesus (n=4)	Local prednisone + dexamethasone, systemic CS (ALK, decellularized) Local prednisone + dexamethasone, systemic CS (ALK, fresh)	195–>391 194–>398
Jie (2013)	WT	Rhesus (n=6) Rhesus (n=6)	CYP +bone marrow Tx (PKP) CYP (PKP)	30–42 (36) 12–20 (19)
Choi (2013)	PKP allograft, following previous decellularized WT pig ALK	Rhesus (n=5)	Local prednisone + dexamethasone, systemic CS	35, 49, >324, >379, >421. Previous xeno-sensitization did not influence outcome

Abbreviations (if not used previously):

ALK = anterior lamellar keratoplasty

PKP = penetrating keratoplasty

Table 6: References

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**Table 7**

Transplantation of pig artery patches into NHPs (1998–2013)

FIRST AUTHOR (Year)	DONOR (Pig)	RECIPIENT (n)	IMMUNOSUPPRESSIVE THERAPY	SURVIVAL - RANGE (MEDIAN) (Days)
Ezzelarab (2012)	GTKO	Baboon (n=3) Baboon (n=5) Baboon (n=1) Baboon (n=1) Baboon (n=4)	No IS Anti-CD154mAb Anti-CD154 mAb, ATG, MMF CTLA4-Ig ATG, MMF, CTLA4-Ig	14->28 2->28 >28 >28 14->28

Abbreviations as used previously

Table 7: References

Ezzelarab MB, Eksler B, Echeverri G, et al. Costimulation blockade in pig artery patch xenotransplantation - a simple model to monitor the adaptive immune response in nonhuman primates. *Xenotransplantation*. 2012; 4:221–32.