



Abnormal Calcium Handling and Exaggerated Cardiac Dysfunction in Mice with Defective Vitamin D Signaling

Sangita Choudhury¹, Soochan Bae¹, Qingen Ke¹, Ji Yoo Lee¹, Sylvia S. Singh¹, René St-Arnaud², Federica del Monte¹, Peter M. Kang^{1*}

1 Cardiovascular Institute, Beth Israel Deaconess Medical Center and Harvard Medical School, Boston, Massachusetts, United States of America, **2** Shriners Hospital and Departments of Surgery and Human Genetics, McGill University, Montreal, Canada

Abstract

Aim: Altered vitamin D signaling is associated with cardiac dysfunction, but the pathogenic mechanism is not clearly understood. We examine the mechanism and the role of vitamin D signaling in the development of cardiac dysfunction.

Methods and Results: We analyzed 1α -hydroxylase (1α -OHase) knockout (1α -OHase^{-/-}) mice, which lack 1α -OH enzymes that convert the inactive form to hormonally active form of vitamin D. 1α -OHase^{-/-} mice showed modest cardiac hypertrophy at baseline. Induction of pressure overload by transverse aortic constriction (TAC) demonstrated exaggerated cardiac dysfunction in 1α -OHase^{-/-} mice compared to their WT littermates with a significant increase in fibrosis and expression of inflammatory cytokines. Analysis of calcium (Ca²⁺) transient demonstrated profound Ca²⁺ handling abnormalities in 1α -OHase^{-/-} mouse cardiomyocytes (CMs), and treatment with paricalcitol (PC), an activated vitamin D₃ analog, significantly attenuated defective Ca²⁺ handling in 1α -OHase^{-/-} CMs. We further delineated the effect of vitamin D deficiency condition to TAC by first correcting the vitamin D deficiency in 1α -OHase^{-/-} mice, followed then by either a daily maintenance dose of vitamin D or vehicle (to achieve vitamin D deficiency) at the time of sham or TAC. In mice treated with vitamin D, there was a significant attenuation of TAC-induced cardiac hypertrophy, interstitial fibrosis, inflammatory markers, Ca²⁺ handling abnormalities and cardiac function compared to the vehicle treated animals.

Conclusions: Our results provide insight into the mechanism of cardiac dysfunction, which is associated with severely defective Ca²⁺ handling and defective vitamin D signaling in 1α -OHase^{-/-} mice.

Citation: Choudhury S, Bae S, Ke Q, Lee JY, Singh SS, et al. (2014) Abnormal Calcium Handling and Exaggerated Cardiac Dysfunction in Mice with Defective Vitamin D Signaling. PLoS ONE 9(9): e108382. doi:10.1371/journal.pone.0108382

Editor: Sudhiranjan Gupta, Texas A & M University Health Science Center, United States of America

Received: May 28, 2014; **Accepted:** August 20, 2014; **Published:** September 30, 2014

Copyright: © 2014 Choudhury et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability: The authors confirm that all data underlying the findings are fully available without restriction. All relevant data are within the paper and its Supporting Information files.

Funding: This work was supported in part by NIH Grants R01 HL65742 (P.M.K.) and the World Class University program (R31-20029) from the Ministry of Education, Science and Technology, South Korea (P.M.K.). Support for generating the 1α -OHase^{-/-} mice was provided by a grant from Shriners Hospitals for Children, Canada (R.St-A.). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* Email: pkang@bidmc.harvard.edu

Introduction

Cardiovascular disease (CVD) is the most common cause of mortality and morbidity in the United States and other developed nations. Clinical and epidemiological studies suggest an association between vitamin D deficiency and various cardiovascular disorders [1,2], and vitamin D therapy has been shown to improve cardiovascular function [3]. Particularly, in patients with chronic renal failure (CRF), vitamin D deficiency is uniformly present because the critical conversion of nutritional vitamin D₃ (25(OH)D₃) to the hormonally active form of vitamin D₃ (1,25(OH)₂D₃) occurs primarily in the kidney by 1α -hydroxylase (1α -OHase)[4]. CRF has been shown to be an independent risk factor for CVD, with 10–20 times greater incidence of cardiovascular disease in CRF patients [5]. Clinical studies have also demonstrated that there is an association between improved survival and decreased cardiovascular mortality in hemodialysis patients treated with an activated vitamin D analog, paricalcitol (PC) [6,7]. In fact, similar to CRF patients, 1α -OHase knockout

(1α -OHase^{-/-}) mice show baseline hypertension, cardiac hypertrophy, and impaired cardiac function [8]. Despite these associations between vitamin D deficiency and cardiac dysfunction, the pathogenic mechanism of cardiac dysfunction associated with vitamin D deficiency is not fully elucidated.

Cardiomyocyte (CM) contractility is regulated by Ca²⁺ handling contractile proteins via regulation of intracellular levels of Ca²⁺ [9]. Thus, abnormalities in Ca²⁺ homeostasis, as seen in vitamin D deficiency, may lead to a chronic defect in E–C coupling, which may in turn lead to cardiac dysfunction. Delays in Ca²⁺ transients, for example, have been observed in myocardial tissue obtained from failing hearts [10]. Although it might not be surprising to find abnormal Ca²⁺ handling in a vitamin D deficiency state, the exact mechanism of vitamin D deficiency affecting the Ca²⁺ handling in CMs, however, is poorly understood. In this study, we examined whether abnormal Ca²⁺ homeostasis and Ca²⁺ handling may mediate structural and functional cardiac abnormalities and examined the relationship between vitamin D deficiency and the

development of cardiac hypertrophy in $1\alpha\text{-OHase}^{-/-}$ mice. We hypothesize that restoration of vitamin D signaling improves cardiac functions by restoring Ca^{2+} homeostasis.

Methods

Animal care

Experiments were conducted using 8–10-week old male $1\alpha\text{-OH}^{-/-}$ mice and their littermates in a C57BL/6 background. $1\alpha\text{-OHase}^{+/-}$ and $1\alpha\text{-OHase}^{-/-}$ mice were produced from $1\alpha\text{-OHase}^{+/-}$ [11]. All mice, breeders and offspring were housed at the Animal Research Facility at Beth Israel Deaconess Medical Center (BIDMC) under pathogen-free conditions with a reverse daily 12:12 h light: dark cycle. Euthanasia was performed by CO_2 via a gas cylinder. All experimental procedures were approved by the Institutional Animal Care and Use Committee of BIDMC.

Animal surgery and hemodynamic measurement

TAC was performed in 8–10 week old male C57BL/6 mice and in $1\alpha\text{-OHase}^{-/-}$ [12,13] mice. Cardiac function was analyzed using echocardiography (for baseline cardiac function) and left ventricular (LV) pressure-volume loop measurement (after TAC) as described previously [14,15]. Detailed method is provided in Text S1.

Morphometric analysis of isolated CMs

CMs were enzymatically dissociated from 12 weeks old male mouse hearts according to previously described protocol [16,17]. Detailed method is provided in Text S1.

Serum Ca^{2+} and vitamin D metabolite analysis

Serum $1, 25(\text{OH})_2 \text{D}_3$ ($n=8-10$ replicates per group) was analyzed via enzyme immunoassays using commercial kits (Immunodiagnostic Systems). Serum Ca^{2+} ($n=8-10$) was analyzed via a quantitative Calcium Colorimetric Detection Kit (Bio Vision).

Serum PTH analysis

Serum PTH ($n=8-10$ replicates per group) was analyzed via enzyme immunoassays using commercial kits (Immunoptics Inc.)

Ca^{2+} transients and cell shortening measurements of isolated CMs

Measurement of Ca^{2+} transient and cell shortening in isolated CMs [18]. Briefly, CMs were isolated and contraction parameters and Ca^{2+} transient [Ca^{2+}] were measured in response to electrical stimulation. Caffeine induced [Ca^{2+}] release was measured after a 10-second pause following steady-state stimulation at increasing rates. Detailed methods are described in Text S1.

Histology and Western Blots

Hearts were fixed in 10% formalin and paraffin-embedded. Sections were stained with hematoxylin and eosin and Masson Trichrome (MT) at the Histology Core facility at BIDMC [14,15]. Quantification of fibrosis was performed as described previously [16]. To compare the levels of the major proteins involved in Ca^{2+} handling, Western blot analysis was performed on total protein from WT and $1\alpha\text{-OHase}^{-/-}$ mice as described previously [14,15].

Statistical Analysis

Data are reported as mean SEM. Statistical significance was determined by one-way or two-way ANOVA and Student-

Newman-Keuls post hoc test. Values of $p<0.05$ were considered significant.

Results

Exaggerated hypertrophic responses after transverse aortic constriction (TAC) in $1\alpha\text{-OHase}^{-/-}$ mice

At baseline, $1\alpha\text{-OHase}^{-/-}$ mice demonstrated a modest 5% increase in heart weight compared to their WT littermates at 12 weeks of age (**Fig. 1A**). Corresponding isolated adult CMs also demonstrated modest increase in surface areas of $1\alpha\text{-OHase}^{-/-}$ mice compared to the age and gender matched WT mice (**Fig. 1B and 1C**). Cross sectional area of the CMs further confirmed these findings (**Fig. 1D and 1E**). We also observed that baseline systolic cardiac function was modestly reduced in $1\alpha\text{-OHase}^{-/-}$ mice compared with the WT mice (**Fig. 1F**). In addition, the mean arterial pressure (MAP) of $1\alpha\text{-OHase}^{-/-}$ was significantly higher than that of WT littermate mice (**Fig. 1G**). These findings suggested that there is baseline mild cardiac hypertrophy and cardiac dysfunction in $1\alpha\text{-OHase}^{-/-}$ mice compared to the WT mice.

To elucidate the effect of defective vitamin D signaling under pathological conditions, we imposed pressure overload on the heart using TAC. Morphometric analysis demonstrated significant increases in heart weight/body weight (HW/BW) ratio in both $1\alpha\text{-OHase}^{-/-}$ and WT mice compared with the corresponding sham-operated groups after 4 weeks of TAC (**Fig. 2A and 2B**). However, there were exaggerated hypertrophic responses to TAC in $1\alpha\text{-OHase}^{-/-}$ mice heart compared to the WT mice. Cardiac functional analysis using pressure-volume (PV) loop measurement revealed that $1\alpha\text{-OHase}^{-/-}$ mice demonstrated significantly greater cardiac dysfunction after TAC, as observed by reduction in stroke volume and cardiac output, compared to their WT littermates (**Fig. 2C and 2D**).

We further analyzed these hearts for the presence of interstitial fibrosis using Masson-Trichrome staining. We found that both WT and $1\alpha\text{-OHase}^{-/-}$ mice showed presence of fibrosis after TAC, (**Fig. 2E**) which was significantly increased in $1\alpha\text{-OHase}^{-/-}$ mice compared with those in WT mice (WT = $11.9\pm 1.5\%$ vs. $1\alpha\text{-OHase}^{-/-}$ = $21.5\pm 1.2\%$; $p<0.05$). These findings demonstrated that there were exaggerated pathological responses to TAC in $1\alpha\text{-OHase}^{-/-}$ mice that are associated with greater decreased cardiac function and more aggressive development of cardiac fibrosis.

Increased fetal gene activation and inflammatory responses after TAC in $1\alpha\text{-OHase}^{-/-}$ mice

We determined the biochemical markers for cardiac hypertrophy and heart failure by measuring the level of “fetal” genes, such as atrial natriuretic factor (ANF) and brain natriuretic peptide (BNP). Ventricular ANF was significantly expressed in all TAC groups. However, there was a 8-fold greater increase in ANF mRNA levels after TAC in $1\alpha\text{-OHase}^{-/-}$ mice compared to the WT littermates (**Fig. 3A**). Similarly, there was also a greater increase in BNP expression in $1\alpha\text{-OHase}^{-/-}$ mice compared to the WT mice hearts after TAC (**Fig. 3B**). Another marker for cardiac hypertrophy, the ratio of $\beta\text{-MHC}$ to total MHC expression, was also increased after TAC in both groups. Yet, there were again dramatic and exaggerated increases observed in $1\alpha\text{-OHase}^{-/-}$ mice compared to the WT mice after TAC (**Fig. 3C**).

It's known that vitamin D, a steroid hormone, affects immune regulation by preventing excessive expression of inflammatory cytokines and increasing the ‘oxidative burst’ potential of macrophages [19]. Interestingly, we observed a reduction in

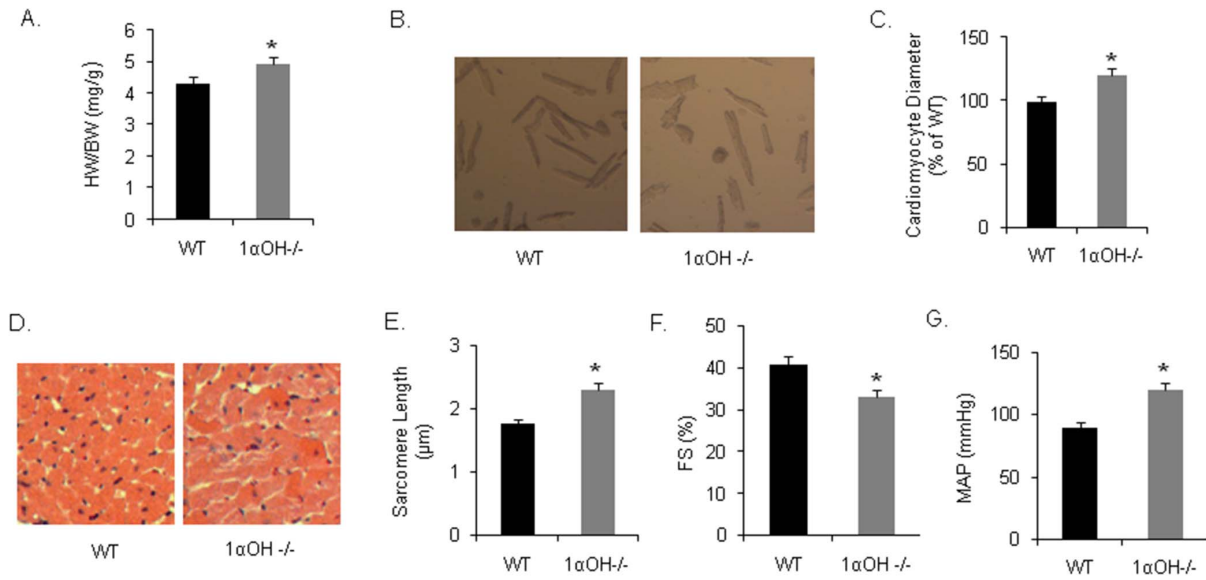


Figure 1. Physiological, echocardiographic, and hemodynamic parameters in $1\alpha\text{OHase}^{-/-}$ mice at baseline: (A) Baseline HW/BW of $1\alpha\text{OHase}^{-/-}$ mice. *, $p < 0.05$ compared to WT. N = 8–10. (B and C) Isolated adult CMs from $1\alpha\text{OHase}^{-/-}$ mice showing increased surface area compared to WT CMs. *, $p < 0.05$ compared to WT. N = 8–10. (D) Cross sectional area of mice heart showing CM surface area. (E) Resting sarcomere length of $1\alpha\text{OHase}^{-/-}$ and WT CMs. *, $p < 0.05$ compared to WT. N = 8–10. (F) Baseline fractional shortening (FS) of WT and KO mice. *, $p < 0.05$ compared to WT. N = 8–10. (G) MAP of WT and $1\alpha\text{OHase}^{-/-}$ mice at baseline. *, $p < 0.05$ compared to WT. N = 8–10. doi:10.1371/journal.pone.0108382.g001

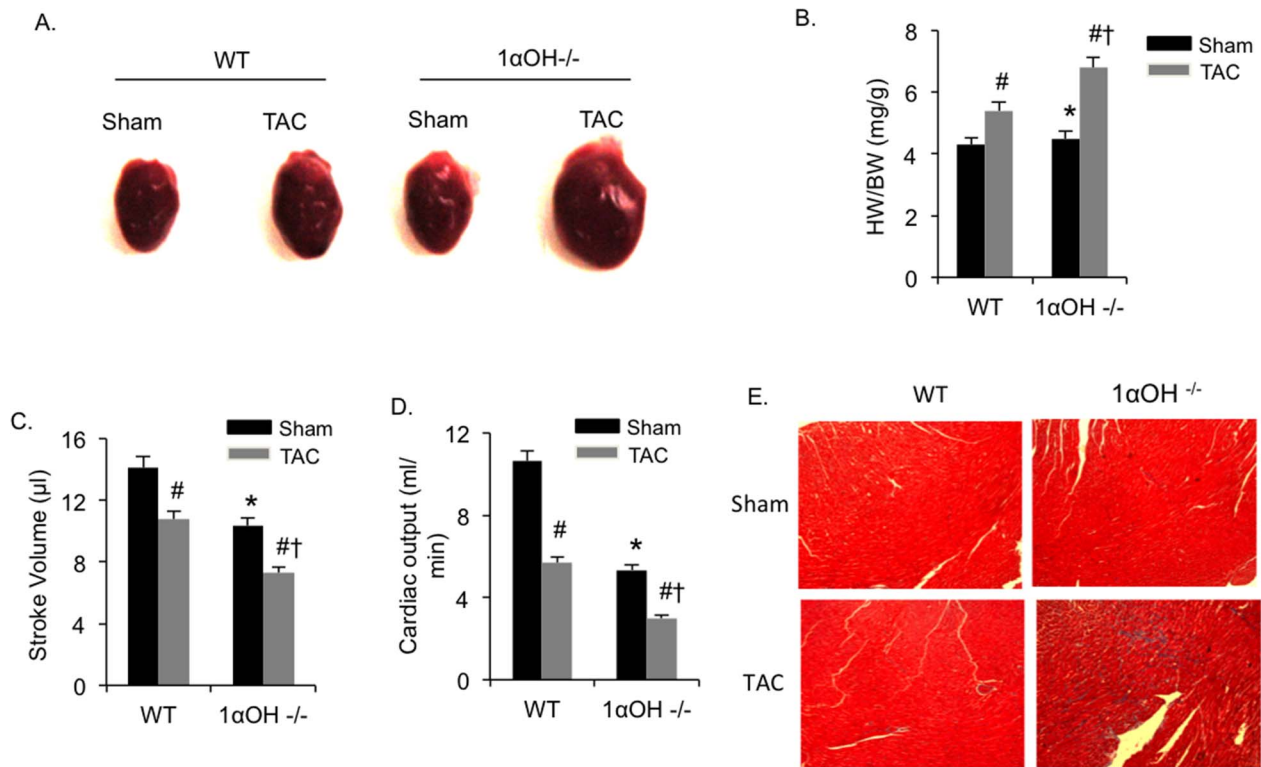


Figure 2. In response to pressure overload mutant mice showing exaggerated hypertrophy. (A) Representative hearts after 4 weeks of sham or TAC in WT and $1\alpha\text{OHase}^{-/-}$ mice. (B) HW/BW after 4 weeks of sham or TAC in WT and $1\alpha\text{OHase}^{-/-}$ mice. *, $p < 0.05$ compared to WT. #, $p < 0.05$ compared to sham. †, $p < 0.05$ compared to WT TAC. N = 8–10. (C and D) Stroke volume (C) and cardiac output (D) in WT and $1\alpha\text{OHase}^{-/-}$ mice after 4 weeks of sham or TAC. *, $p < 0.05$ compared to WT. #, $p < 0.05$ compared to sham. †, $p < 0.05$ compared to WT TAC. N = 8–10. (E) Representative Masson-Trichrome heart staining in $1\alpha\text{OHase}^{-/-}$ mice after 4 weeks of sham or TAC. doi:10.1371/journal.pone.0108382.g002

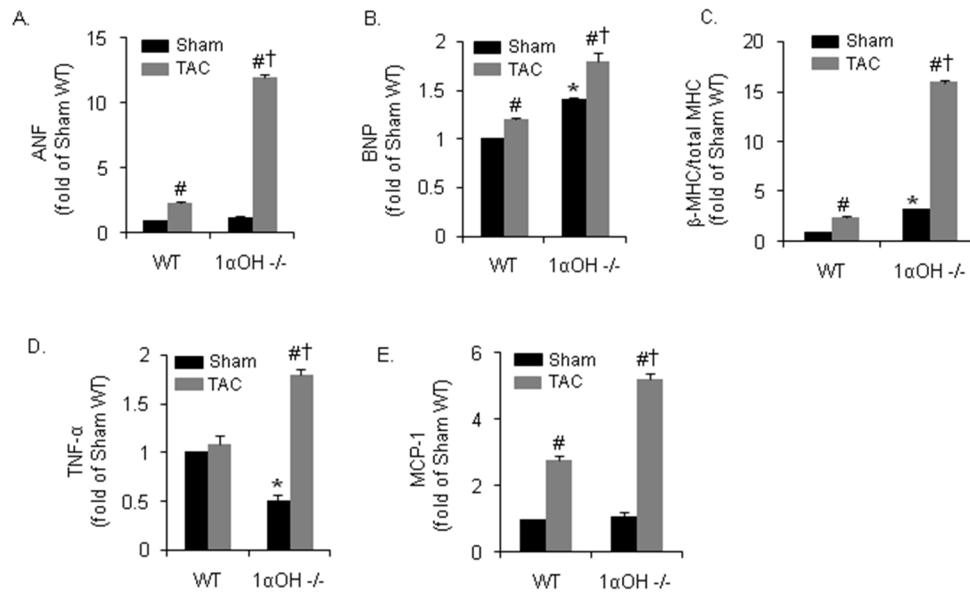


Figure 3. Biochemical findings after 4 weeks of Pressure overload in $1\alpha\text{OHase}^{-/-}$ mice. (A–C) Real-time PCR mRNA expression of ANF (A), BNP (B) and of $\beta\text{-MHC}$ to total MHC ratio (C) in WT and $1\alpha\text{OHase}^{-/-}$ mice after 4 weeks of sham or TAC. *, $p < 0.05$ compared to WT. #, $p < 0.05$ compared to sham. †, $p < 0.05$ compared to WT TAC. N = 8–10. (D and E) Real-time PCR mRNA expression of inflammatory cytokines TNF- α (D) and MCP-1 (E) in WT and $1\alpha\text{OHase}^{-/-}$ mice after 4 weeks of sham or TAC. *, $p < 0.05$ compared to WT. #, $p < 0.05$ compared to sham. †, $p < 0.05$ compared to WT TAC. N = 8–10. doi:10.1371/journal.pone.0108382.g003

tumor necrosis factor- α (TNF- α) expression at baseline in $1\alpha\text{OHase}^{-/-}$ mice compared to WT mice. Although no significant differences were observed in TNF- α expression after TAC in WT mice, $1\alpha\text{OHase}^{-/-}$ mice hearts showed enhanced activation of TNF- α after TAC (Fig. 3D). Another inflammatory marker, monocyte chemoattractant protein-1 (MCP-1), was activated by TAC in both mice, but with significantly greater activation was observed in $1\alpha\text{OHase}^{-/-}$ mice after TAC (Fig. 3E). These findings demonstrated that there is an exaggerated biochemical evidence of cardiac dysfunction and uncontrolled immune response associated with TAC in $1\alpha\text{OHase}^{-/-}$ mice.

Defective Ca^{2+} handling and modulation of Ca^{2+} regulatory protein expressions in $1\alpha\text{OHase}^{-/-}$ mice CMs

We found that the circulating 1, 25(OH) vitamin D₃ level was undetectable in $1\alpha\text{OHase}^{-/-}$ mice and significantly reduced in heterozygous $1\alpha\text{OHase}^{+/-}$ littermates (Fig. 4A), which coincide with previous findings [20,21]. Without vitamin D supplementation, $1\alpha\text{OHase}^{-/-}$ mice at 12 weeks showed severe hypocalcemia with plasma Ca^{2+} concentrations as low as 1.2 mmol/L in contrast to the WT littermates, which exhibited normal plasma Ca^{2+} concentrations (WT = 2.5 mmol/L vs. $1\alpha\text{OHase}^{-/-}$ = 1.2 mmol/L; $P < 0.05$). Supplementation of vitamin D using injection of PC (200 ng/kg), an activated vitamin D₃ analog, starting at 4 weeks after birth normalized the plasma Ca^{2+} concentration in $1\alpha\text{OHase}^{-/-}$ mice by 12 weeks (Fig. 4B).

To examine the effect of the vitamin D deficiency in CMs, contraction parameters of individual isolated CM from $1\alpha\text{OHase}^{-/-}$ mice and their WT littermates were studied by measuring sarcomere shortening (contraction) and relengthening (relaxation) in response to electrical stimulation (see Text S1 for detail). Baseline $1\alpha\text{OHase}^{-/-}$ CM sarcomeres were significantly larger compared to WT (Fig. 4C, Table S1). Additionally, $1\alpha\text{OHase}^{-/-}$ CMs showed a significant decrease in peak shortening, as well as the rate of cell shortening and rate of cell relengthening

compared to WT CMs (Fig. 4D and 4E). Peak shortening is calculated from the percent shortening of length data (Fig. 4D). The highest and lowest value reached by the transient.

We then investigated Ca^{2+} handling properties of individual CM from $1\alpha\text{OHase}^{-/-}$ mice (see Text S1 for detail). Analysis of Ca^{2+} transients indicated decreased peak systolic Ca^{2+} , measured as Fura-2 ratios in CM from $1\alpha\text{OHase}^{-/-}$ compared to WT group CM (Fig. 4F). The rate of Ca^{2+} transient decay (τ , exponential decay of time constant, the speed of relaxation/calcium uptake) was also significantly reduced in $1\alpha\text{OHase}^{-/-}$ CMs (Fig. 4G) which was calculated by the ionoptix program from the transient, with a reduced time to peak (TTP) and a decrease in relaxation rate (Ca^{2+} efflux) (Fig. 4H and 4I). Caffeine causes Ca^{2+} release from the sarcoplasmic reticulum (SR) of mammalian muscle. Thus, to evaluate the SR Ca^{2+} load, we tested caffeine-induced Ca^{2+} release amount from SR in WT and $1\alpha\text{OHase}^{-/-}$ CMs. We found that CMs from $1\alpha\text{OHase}^{-/-}$ mice presented a decreased caffeine-induced Ca^{2+} transient compared to WT CMs (Fig. 4J). These findings demonstrated profound Ca^{2+} handling abnormalities in $1\alpha\text{OHase}^{-/-}$ mice CMs, which may contribute to the development of fulminant heart failure after TAC.

The expression level and the activity of sarcoplasmic reticulum Ca^{2+} ATPase (SERCA) is significantly decreased in pressure overload-induced hypertrophy and during heart failure in animals and human, which correlate with decreased myocardial dysfunction [22,23]. To further elucidate the mechanism underlying the defective Ca^{2+} handling, we examined the SERCA2a protein expression level in these mice. We found that $1\alpha\text{OHase}^{-/-}$ mice showed significant decreased SERCA2a expression after TAC compared to WT mice (Fig. 5A and 5B). Phospholamban (PLB) interacts with SERCA and inhibits its Ca^{2+} transport rate, and the relative abundance of SERCA and PLB maintains Ca^{2+} homeostasis and CM contractility. Therefore, SERCA/PLB ratio reflects Ca^{2+} transport capacity of the SR [24]. At baseline, $1\alpha\text{OHase}^{-/-}$

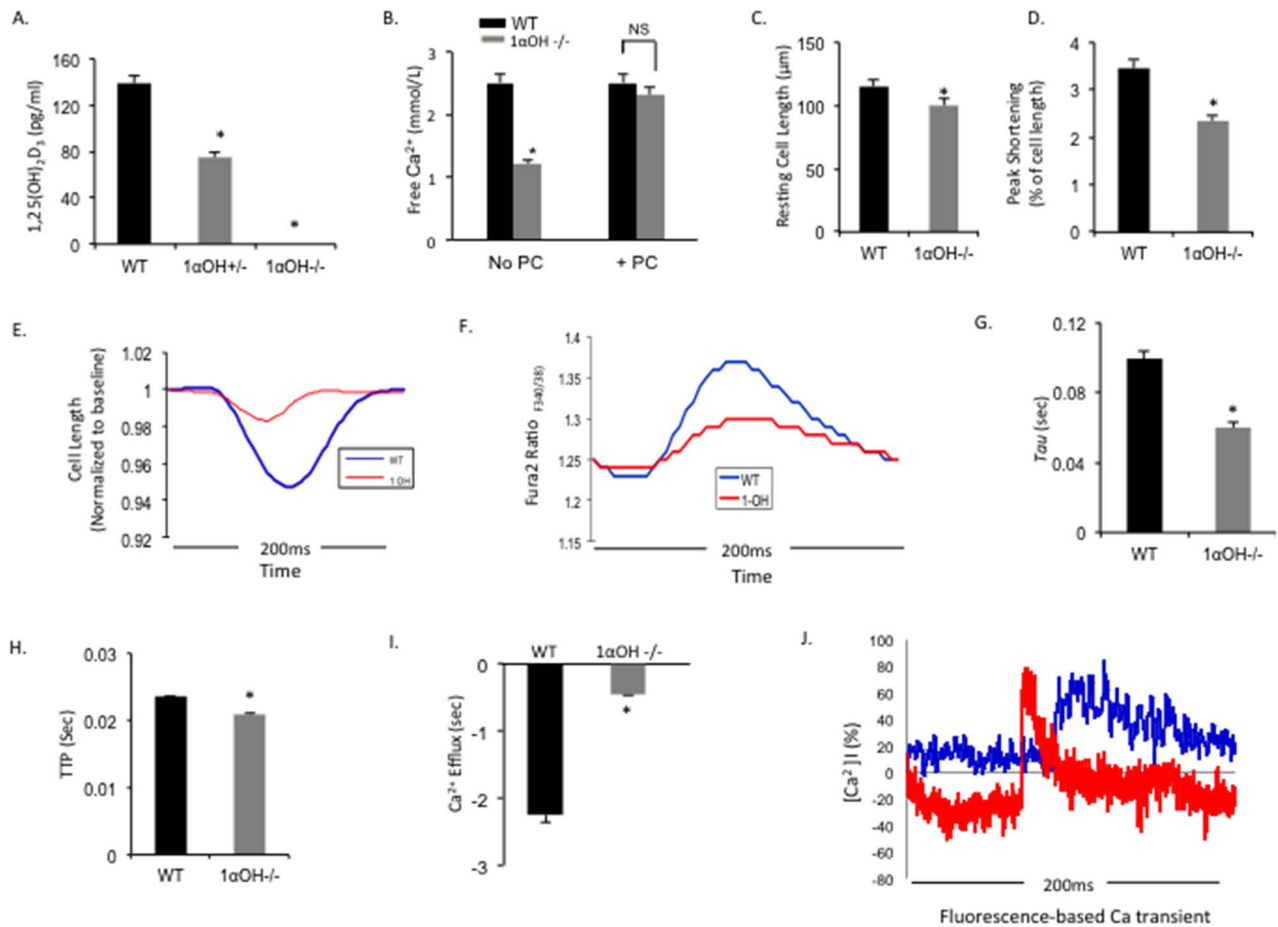


Figure 4. Contractile function and Ca²⁺ transient parameters in isolated CMs from 1 α OHase^{-/-} mice. (A and B) Baseline 1, 25(OH)₂D₃ (A) and free Ca²⁺ (B) level in WT and 1 α OHase^{-/-} mice serums. *, $p < 0.05$ compared to WT, N=6–8 mice. (C) Resting cell length (D) and peak shortening (% of cell length) from isolated CMs obtained from WT and 1 α OHase^{-/-} mice. *, $p < 0.05$ compared to WT. Ten twitches per CMs were collected for each mouse heart. N=6–8 mice/group. (E) Continuous measurement of cell contractility from isolated CMs obtained from WT (blue line) and 1 α OHase^{-/-} (red line) mice. *, $p < 0.05$ compared to WT. Ten twitches per CMs were collected for each mouse heart. N=6–8 mice/group. (F) Representative Fura 2 ratio (F340/380) from isolated CMs obtained from WT (blue line) and 1 α OHase^{-/-} (red line) mice. Data shows that the twitch peak amplitudes are significantly different between WT and 1 α OHase^{-/-} CMs, which indicate severely defective Ca²⁺ handling in 1 α OHase^{-/-} CMs. (G–I) The rate of Ca²⁺ transient decay (*Tau*) (G), time to peak contraction (TTP) (H), and return velocity to baseline or Ca²⁺ efflux in WT and 1 α OHase^{-/-} CMs. *, $p < 0.05$ compared to WT. N=10 CMs/mouse heart. N=6–8 mice/group. (J) Representative traces of Ca²⁺ transients evoked by 10 mM caffeine recorded in WT (blue line) and 1 α OHase^{-/-} (red line) mice CMs. doi:10.1371/journal.pone.0108382.g004

mouse hearts showed significant increase in PLB, which decreased after TAC (Fig. 5A and 5B). SERCA/PLB ratio was also significantly decreased in 1 α -OHase^{-/-} mice compared to WT mice (Fig. 5C). This may explain the difference in SR Ca²⁺ load and contractile function between WT and 1 α -OHase^{-/-} mice CMs, and the depressed levels of SERCA2a may contribute to the severity of heart failure after TAC in 1 α -OHase^{-/-} mice.

The Na⁺-Ca²⁺-exchanger (NCX) is the dominant myocardial Ca²⁺ efflux mechanism. Ca²⁺ removal from the cytosol occurs by activity of the SR Ca²⁺ pump and by exchange of Ca²⁺ for sodium by the sarcolemmal NCX [25]. The decrease in PLB might lead to a higher activation of the remaining SERCA pumps and increase the activity of NCX [26]. At baseline, there were no significant difference in expression of NCX between the WT and 1 α -OHase^{-/-} mice (Fig. 5A and 5B). However, we found significant up-regulation of NCX expression in banded WT and 1 α -OHase^{-/-} mice with significant 4-folds increase seen in 1 α -OHase^{-/-} mice. A relative increase in NCX is expected when SR function is impaired [27]. The function of ryanodine receptor 2

(RyR2) (the cardiac isoform) is to allow Ca²⁺-induced Ca²⁺ release that brings about contraction, while myocyte relaxation results in RyR2 closure accompanied by the Ca²⁺ re-uptake into SR through the SERCA [28]. After TAC, there was a moderate decrease in RyR2 expression in both WT and 1 α -OHase^{-/-} mice (Fig. 5A and 5B).

Treatment with PC attenuates cardiac dysfunction after TAC in 1 α -OHase^{-/-} mice

The responses of 1 α -OHase^{-/-} mice to TAC in our previous experiments may be compounded by their baseline cardiac hypertrophy. Thus, we further determined the specific role of defective vitamin D to pathological conditions by first using “rescue” vitamin D protocol as described previously in 1 α -OHase^{-/-} mice, where vitamin D deficiency and its non-cardiac phenotype were both corrected [29]. From 28–30 days of life, 1 α -OHase^{-/-} mice were treated with an initial “rescue” dose of 500 ng/kg of PC, followed by a daily “maintenance” dose of 200 ng/kg from 5 to 8 weeks of age. At 8 weeks, the animals were

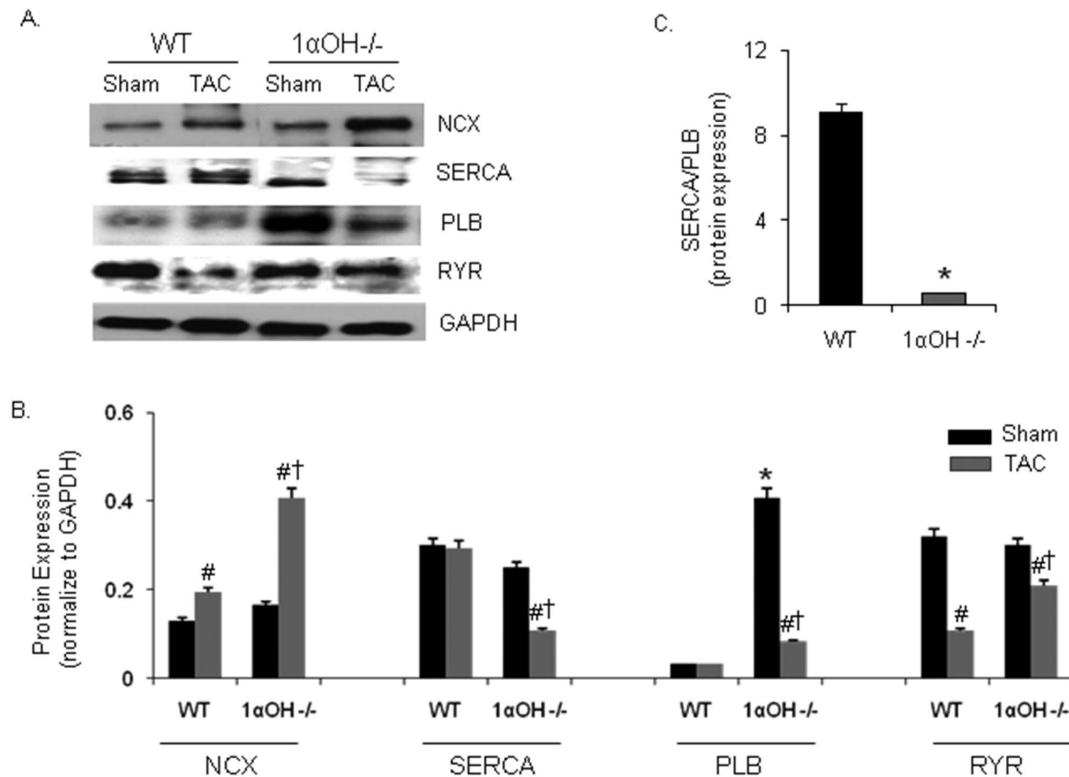


Figure 5. Expressions of Ca²⁺ regulatory protein in mice with altered metabolism of Vitamin D. (A) Representative western blots of various Ca²⁺ handling proteins. (B) Quantitative analysis of various Ca²⁺ handling proteins. NCX = Na⁺/Ca²⁺ exchanger, SERCA2a = sarcoplasmic reticulum Ca²⁺ ATPase, PLB = phospholamban, and RYR = ryanodine receptor. GAPDH = glyceraldehyde 3-phosphate dehydrogenase. GAPDH was used as internal loading control. *, $p < 0.05$ compared to WT. #, $p < 0.05$ compared to sham. †, $p < 0.05$ compared to WT TAC. N = 8–10. (C) Quantitative analysis of SERCA/PLB protein expression ratio. *, $p < 0.05$ compared to WT. #, $p < 0.05$ compared to sham, N = 8–10. doi:10.1371/journal.pone.0108382.g005

divided into the following groups: 1) Vehicle + Sham, 2) Vehicle + TAC, 3) PC + Sham, and 4) PC+TAC. For the vehicle conditions, maintenance doses of PC were changed to vehicle injections, while the PC dose was maintained for PC groups. TAC resulted in a significant increase in HW/BW and LW/BW ratios in mice receiving vehicle for 4 weeks compared to the sham operated mice (Fig. 6A and 6B). In mice treated with PC, there was a significant 18% attenuation of TAC-induced HW/BW ratio increase compared to the vehicle treated animals. Histological examination demonstrated that interstitial fibrosis induced by TAC in vehicle-administered 1α-OHase^{-/-} mice were significantly inhibited in PC treated group after TAC (Fig. 6C). TAC-induced increases in ANF and β-MHC gene expression, which were both attenuated by PC treatment (Fig. 6D and 6E).

We found that decreased fractional shortening after TAC in vehicle treated mice was significantly improved with vitamin D replacement in 1α-OHase^{-/-} mice (Fig. 6G). In addition, we observed reduced stroke volume and cardiac output in vehicle treated group after TAC. These parameters also improved after vitamin D replacement (Fig. 6G and 6H). Thus, vitamin D replacement resulted in significant attenuation of cardiac dysfunction after TAC in 1α-OHase^{-/-} mice.

Treatment with PC attenuates defective Ca²⁺ handling associated with defective vitamin D signaling

To further assess the effect of vitamin D replacement on CMs contractility in 1α-OHase^{-/-} mice contraction parameters, we performed Ca²⁺ handling analysis of isolated CMs in 1α-OHase^{-/-}

mice with or without PC treatment. We found that vitamin D replacement attenuated hypocontractility and increased peak shortening in PC treated 1α-OHase^{-/-} mice (Fig. 6I and 6J). Rate of relaxation was significantly increased with PC treatment and the rate of Ca²⁺ transient decay (*tau*) was significantly improved (Fig. 6K and 6L). Treatment with PC also normalized the PTH level in 1α-OHase^{-/-} mice (Fig. 7). These findings suggest that the pathological response in 1α-OHase^{-/-} mice is mainly due to defective vitamin D signaling and associated Ca²⁺ handling, and correcting these abnormalities could rescue these pathological responses.

Discussion

In this study, we demonstrated that vitamin D deficiency is associated with cardiac hypertrophy at baseline results in an exaggerated progression to heart failure after pressure overload in 1α-OHase^{-/-} mice. We also found that there is defective Ca²⁺ handling in 1α-OHase^{-/-} mice, with structural and functional cardiac abnormalities in these mice. These cardiac Ca²⁺ handling abnormalities were completely corrected with vitamin D replacement in 1α-OHase^{-/-} mice. Thus, our studies strongly support the notion that vitamin D deficiency is an under-recognized, non-classic risk factor for developing heart failure that is readily correctable.

Vitamin D is known for its primary role in Ca²⁺ and bone homeostasis [30]. Biological activities of vitamin D are mediated by a hormonally active 1, 25-dihydroxyvitamin D₃ that is converted from 25-hydroxyvitamin D₃ by 1α-OHase. It binds to

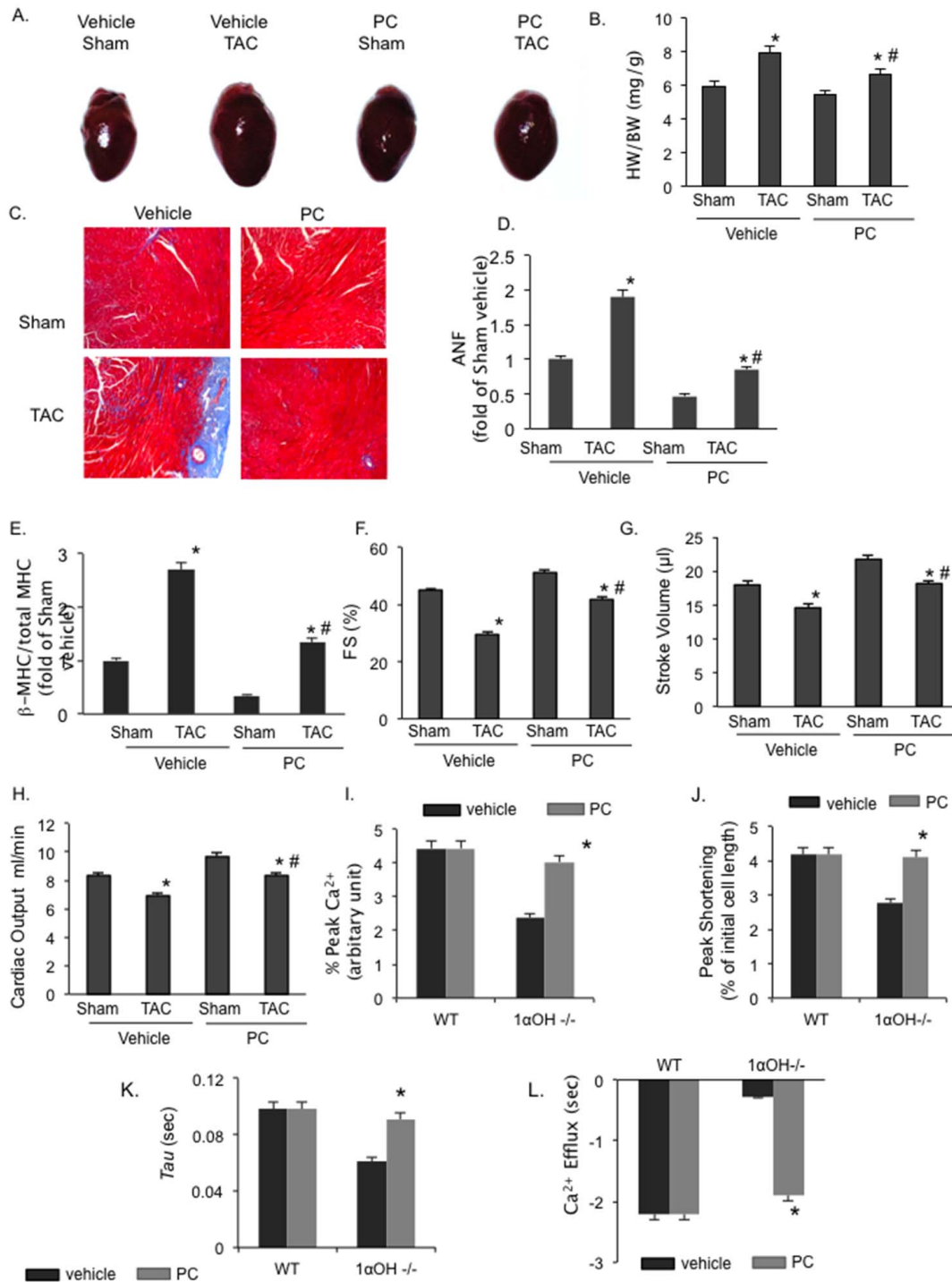


Figure 6. Effect of paricalcitol on cardiac dysfunction parameters. (A) Representative hearts in $1\alpha\text{-OHase}^{-/-}$ mice with or without PC treatment after 4 weeks of sham or TAC. (B) HW/BW ratio in $1\alpha\text{-OHase}^{-/-}$ mice with or without PC treatment after 4 weeks of sham or TAC. *, $p < 0.05$ compared to Sham. #, $p < 0.05$ compared to vehicle TAC. N = 8–10. (C) Representative Masson-Trichrome heart staining in $1\alpha\text{-OHase}^{-/-}$ mice with or without PC treatment after 4 weeks of sham or TAC. (D and E) Real-time PCR mRNA expression of ANF (D) and of $\beta\text{-MHC}$ to total MHC ratio (E) after 4 weeks of sham or TAC in $1\alpha\text{-OHase}^{-/-}$ mice with or without PC treatment. *, $p < 0.05$ compared to Sham. #, $p < 0.05$ compared to vehicle TAC. N = 8–10. (F–H) Fractional shortening (F), stroke volume (G) and cardiac output (H) in $1\alpha\text{-OHase}^{-/-}$ mice with or without PC treatment after 4 weeks of sham or TAC. *, $p < 0.05$ compared to Sham. #, $p < 0.05$ compared to vehicle TAC. N = 8–10. Effect of paricalcitol on defective calcium handling in $1\alpha\text{-OHase}^{-/-}$ mice. (I–L). The % peak Ca^{2+} (I), % peak shortening (J), the rate of Ca^{2+} transient decay (Tau) (K), and the return velocity to baseline or Ca^{2+} efflux (L) in $1\alpha\text{-OHase}^{-/-}$ mice with or without PC treatment. *, $p < 0.05$ compared to vehicle treated mice group, N = 20 CMTs/mouse from 6–8 mice/group. doi:10.1371/journal.pone.0108382.g006

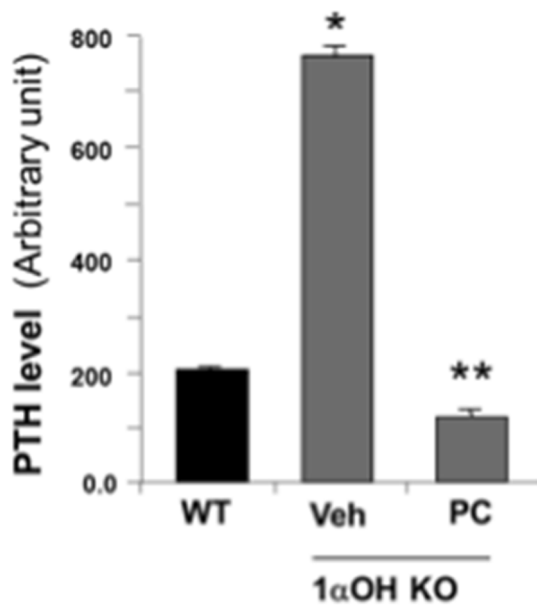


Figure 7. Paricalcitol restore the parathyroid hormone in 1 α -OHase^{-/-} mice this model. PTH level in 1 α -OHase^{-/-} mice with or without PC treatment. *, $p < 0.05$ compared to WT. **, $p < 0.05$ compared to vehicle. N = 8–10.
doi:10.1371/journal.pone.0108382.g007

a specific high-affinity vitamin D receptor (VDR), a member of the superfamily of nuclear receptors for steroid hormones [4]. Vitamin D plays an important physiological role in controlling cardiac functions and vitamin D-dependent signaling systems are present in cardiac myocytes and fibroblasts [31]. Vitamin D deficiency has been associated with abnormal cardiac relaxation, proliferation, and increased cardiac renin gene expression [32,33]. Previously, we showed that vitamin D therapy prevents the progression to cardiac hypertrophy [34], and attenuates the development of heart failure in salt sensitive rat model [35]. In addition, 1 α -OHase^{-/-} mice developed hypertension, cardiac hypertrophy, and impaired cardiac systolic function, possibly due to the activation of the RAS [8]. Clinically, vitamin D deficiency has been associated with the increased prevalence of myocardial dysfunctions and heart failure [36,37]. Since the critical conversion of the storage form to the active form of vitamin D by 1 α -OHase occurs in the kidney, patients with CKD are typically vitamin D deficient. In fact, the prevalence of cardiac hypertrophy and cardiac dysfunction is over 80% in these patients [38,39], and vitamin D therapy has been shown to improve survival and decrease cardiovascular mortality [6,7]. These findings suggest that the role of vitamin D signaling may be significant in the heart, and vitamin D deficiency may offer novel therapeutic target in the treatment of heart failure.

References

- Wang TJ, Pencina MJ, Booth SL, Jacques PF, Ingelsson E, et al. (2008) Vitamin D deficiency and risk of cardiovascular disease. *Circulation* 117: 503–511.
- Zittermann A (2006) Vitamin D and disease prevention with special reference to cardiovascular disease. *Prog Biophys Mol Biol* 92: 39–48.
- Park CW, Oh YS, Shin YS, Kim CM, Kim YS, et al. (1999) Intravenous calcitriol regresses myocardial hypertrophy in hemodialysis patients with secondary hyperparathyroidism. *Am J Kidney Dis* 33: 73–81.
- Demay MB (2006) Mechanism of vitamin D receptor action. *Ann N Y Acad Sci* 1068: 204–213.
- McCullough PA, Jurkovic CT, Pergola PE, McGill JB, Brown WW, et al. (2007) Independent components of chronic kidney disease as a cardiovascular risk state:

CM contraction is regulated by the interplay between Ca²⁺, contractile proteins, and the intracellular handling of Ca²⁺, whereas abnormal Ca²⁺ homeostasis is primarily responsible for depression of CM contractility [22,23]. In several animal and human models of cardiac hypertrophy and heart failure, the whole-cell [Ca²⁺]_i transient is altered [40,41]. Decreases in SERCA pump expression and activity have been observed in a variety of animal and human models of heart failure and a degree of decrease in SERCA level and its activity, closely correlate with a decreased myocardial function [27]. Thus, these data suggest that alterations in Ca²⁺ handling proteins are important contributors to cardiac dysfunction and heart failure. We found profound Ca²⁺ transient abnormalities, contractile abnormalities and altered expression of various Ca²⁺ handling regulatory proteins in vitamin D deficient CMs, which became exaggerated after TAC. Abnormalities in Ca²⁺ homeostasis, as seen in vitamin D deficiency, may lead to a chronic defect in E–C coupling, which may in turn lead to cardiac dysfunction. Particularly, decreased SERCA and SERCA/PLB expression in 1 α -OHase^{-/-} mice supports the hypothesis that alterations in abundance or activity of molecules that regulate systolic and diastolic Ca²⁺ are centrally involved in depressed contractility in hypertrophied and failing hearts.

The key role of cardiac hypertrophy in the pathogenesis of heart disease underscores the need to identify the cellular and molecular mechanisms responsible for both cardiac hypertrophy and its progression to heart failure. A better understanding of disease progression may facilitate the development of novel therapeutic modalities, as well as the development of better guidelines for the prevention of cardiac hypertrophy. Our findings contribute to a better understanding of disease progression that are involved in vitamin D signaling and the development of heart failure. Our finding provides evidences that Ca²⁺ homeostasis and Ca²⁺ handling mediate structural and functional cardiac abnormalities in vitamin D deficient 1 α -OHase^{-/-} mice and development of cardiac hypertrophy.

Supporting Information

Table S1 Comparison of cardiomyocytes resting sarcomere length in 0.5 Hz paced cells in WT and 1 α OH^{-/-}. Sarcomere length of 1 α -OHase^{-/-} and WT CMs, raw value in 0.5 Hz paced.
(DOCX)

Text S1 Supplemental Information.
(DOC)

Author Contributions

Conceived and designed the experiments: SC RSA PMK. Performed the experiments: SC SB QK JYL SSS. Analyzed the data: SC FdM. Contributed reagents/materials/analysis tools: SC SB QK JYL SSS. Wrote the paper: SC PMK.

results from the Kidney Early Evaluation Program (KEEP). *Arch Intern Med* 167: 1122–1129.

- Teng M, Wolf M, Lowrie E, Ofsthun N, Lazarus JM, et al. (2003) Survival of patients undergoing hemodialysis with paricalcitol or calcitriol therapy. *N Engl J Med* 349: 446–456.
- Teng M, Wolf M, Ofsthun MN, Lazarus JM, Hernan MA, et al. (2005) Activated injectable vitamin D and hemodialysis survival: a historical cohort study. *J Am Soc Nephrol* 16: 1115–1125.
- Zhou C, Lu F, Cao K, Xu D, Goltzman D, et al. (2008) Calcium-independent and 1,25(OH)₂D₃-dependent regulation of the renin-angiotensin system in 1 α -hydroxylase knockout mice. *Kidney Int* 74: 170–179.

9. Wehrens XH, Lehnart SE, Reiken SR, Marks AR (2004) Ca²⁺/calmodulin-dependent protein kinase II phosphorylation regulates the cardiac ryanodine receptor. *Circ Res* 94: e61–70.
10. Chien KR, Ross J, Jr., Hoshijima M (2003) Calcium and heart failure: the cycle game. *Nat Med* 9: 508–509.
11. Dardenne O, Prud'homme J, Arabian A, Glorieux FH, St-Arnaud R (2001) Targeted inactivation of the 25-hydroxyvitamin D(3)-1(alpha)-hydroxylase gene (CYP27B1) creates an animal model of pseudovitamin D-deficiency rickets. *Endocrinology* 142: 3135–3141.
12. McMullen JR, Sherwood MC, Tarnavski O, Zhang L, Dorfman AL, et al. (2004) Inhibition of mTOR signaling with rapamycin regresses established cardiac hypertrophy induced by pressure overload. *Circulation* 109: 3050–3055.
13. McMullen JR, Shioi T, Zhang L, Tarnavski O, Sherwood MC, et al. (2003) Phosphoinositide 3-kinase(p110alpha) plays a critical role for the induction of physiological, but not pathological, cardiac hypertrophy. *Proc Natl Acad Sci U S A* 100: 12355–12360.
14. Bae S, Siu PM, Choudhury S, Ke Q, Choi JH, et al. (2010) Delayed activation of caspase-independent apoptosis during heart failure in transgenic mice overexpressing caspase inhibitor Crma. *Am J Physiol Heart Circ Physiol* 299: H1374–1381.
15. Choudhury S, Bae S, Ke Q, Lee JY, Kim J, et al. (2011) Mitochondria to nucleus translocation of AIF in mice lacking Hsp70 during ischemia/reperfusion. *Basic Res Cardiol* 106: 397–407.
16. Rigor DL, Bodyak N, Bae S, Choi JH, Zhang L, et al. (2009) Phosphoinositide 3-kinase Akt signaling pathway interacts with protein kinase Cbeta2 in the regulation of physiologic developmental hypertrophy and heart function. *Am J Physiol Heart Circ Physiol* 296: H566–572.
17. Shioi T, Kang PM, Douglas PS, Hampe J, Yballe CM, et al. (2000) The conserved phosphoinositide 3-kinase pathway determines heart size in mice. *EMBO J* 19: 2537–2548.
18. Bassani JW, Bassani RA, Bers DM (1995) Calibration of indo-1 and resting intracellular [Ca]_i in intact rabbit cardiac myocytes. *Biophys J* 68: 1453–1460.
19. Di Rosa M, Malaguarnera M, Nicoletti F, Malaguarnera L (2011) Vitamin D3: a helpful immuno-modulator. *Immunology* 134: 123–139.
20. Hoenderop JG, Dardenne O, Van Abel M, Van Der Kemp AW, Van Os CH, et al. (2002) Modulation of renal Ca²⁺ transport protein genes by dietary Ca²⁺ and 1,25-dihydroxyvitamin D₃ in 25-hydroxyvitamin D₃-1alpha-hydroxylase knockout mice. *FASEB J* 16: 1398–1406.
21. Xue Y, Karaplis AC, Hendy GN, Goltzman D, Miao D (2005) Genetic models show that parathyroid hormone and 1,25-dihydroxyvitamin D₃ play distinct and synergistic roles in postnatal mineral ion homeostasis and skeletal development. *Hum Mol Genet* 14: 1515–1528.
22. Del Monte F, Hajjar RJ (2008) Intracellular devastation in heart failure. *Heart Fail Rev* 13: 151–162.
23. Morgan JP (1991) Abnormal intracellular modulation of calcium as a major cause of cardiac contractile dysfunction. *N Engl J Med* 325: 625–632.
24. Pieske B, Kretschmann B, Meyer M, Holubarsch C, Weirich J, et al. (1995) Alterations in intracellular calcium handling associated with the inverse force-frequency relation in human dilated cardiomyopathy. *Circulation* 92: 1169–1178.
25. Bers DM (2000) Calcium fluxes involved in control of cardiac myocyte contraction. *Circ Res* 87: 275–281.
26. Kubo H, Margulies KB, Piacentino V, 3rd, Gaughan JP, Houser SR (2001) Patients with end-stage congestive heart failure treated with beta-adrenergic receptor antagonists have improved ventricular myocyte calcium regulatory protein abundance. *Circulation* 104: 1012–1018.
27. Hasenfuss G (1998) Alterations of calcium-regulatory proteins in heart failure. *Cardiovasc Res* 37: 279–289.
28. Asahi M, Sugita Y, Kurzydowski K, De Leon S, Tada M, et al. (2003) Sarcolipin regulates sarco(endo)plasmic reticulum Ca²⁺-ATPase (SERCA) by binding to transmembrane helices alone or in association with phospholamban. *Proc Natl Acad Sci U S A* 100: 5040–5045.
29. Dardenne O, Prud'homme J, Hacking SA, Glorieux FH, St-Arnaud R (2003) Rescue of the pseudo-vitamin D deficiency rickets phenotype of CYP27B1-deficient mice by treatment with 1,25-dihydroxyvitamin D₃: biochemical, histomorphometric, and biomechanical analyses. *J Bone Miner Res* 18: 637–643.
30. Dusso AS, Brown AJ, Slatopolsky E (2005) Vitamin D. *Am J Physiol Renal Physiol* 289: F8–28.
31. Chen S, Glenn DJ, Ni W, Grigsby CL, Olsen K, et al. (2008) Expression of the vitamin d receptor is increased in the hypertrophic heart. *Hypertension* 52: 1106–1112.
32. Lee W, Kang PM (2010) Vitamin D deficiency and cardiovascular disease: Is there a role for vitamin D therapy in heart failure? *Curr Opin Investig Drugs* 11: 309–314.
33. Xiang W, Kong J, Chen S, Cao L-P, Qiao G, et al. (2005) Cardiac hypertrophy in vitamin D receptor knockout mice: role of the systemic and cardiac renin-angiotensin systems. *American Journal of Physiology - Endocrinology And Metabolism* 288: E125–E132.
34. Bodyak N, Ayus JC, Achinger S, Shivalingappa V, Ke Q, et al. (2007) Activated vitamin D attenuates left ventricular abnormalities induced by dietary sodium in Dahl salt-sensitive animals. *Proc Natl Acad Sci U S A* 104: 16810–16815.
35. Bae S, Yalamarti B, Ke Q, Choudhury S, Yu H, et al. (2011) Preventing progression of cardiac hypertrophy and development of heart failure by paricalcitol therapy in rats. *Cardiovasc Res* 91: 632–639.
36. Gotsman I, Shauer A, Zwas DR, Hellman Y, Keren A, et al. (2012) Vitamin D deficiency is a predictor of reduced survival in patients with heart failure; vitamin D supplementation improves outcome. *Eur J Heart Fail* 14: 357–366.
37. Holick MF (2007) Vitamin D deficiency. *N Engl J Med* 357: 266–281.
38. Pilz S, Tomaschitz A, Marz W, Drechsler C, Ritz E, et al. (2011) Vitamin D, cardiovascular disease and mortality. *Clin Endocrinol (Oxf)* 75: 575–584.
39. Vacek JL, Vanga SR, Good M, Lai SM, Lakkireddy D, et al. (2012) Vitamin D Deficiency and Supplementation and Relation to Cardiovascular Health. *Am J Cardiol* 109: 359–363.
40. Beuckelmann DJ, Nabauer M, Erdmann E (1992) Intracellular calcium handling in isolated ventricular myocytes from patients with terminal heart failure. *Circulation* 85: 1046–1055.
41. Perreault CL, Shannon RP, Komamura K, Vatner SF, Morgan JP (1992) Abnormalities in intracellular calcium regulation and contractile function in myocardium from dogs with pacing-induced heart failure. *J Clin Invest* 89: 932–938.