



Genome Assembly of Methicillin-Resistant Quality Control Strain Staphylococcus aureus CDC73-57501 (ATCC 29247)

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Staphylococcus aureus is a major cause of bacterial infections in the United States, with high percentages of serious infections resistant to a variety of β -lactam antibiotics. Here, we present the scaffolded genome assembly into 16 contigs of *S. aureus* CDC73-57501 (ATCC 29247), a methicillin-resistant quality control strain.

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B oth a commensal and a pathogen, *Staphyloccus aureus* is a well-known Gram-positive bacterium that often inhabits human skin and mucosa (1, 2). This continual contact with human skin is often blamed for the antibiotic resistance developed by so many staphylococci (3, 4). Strains resistant to all β -lactams, often referred to simply as community-acquired methicillin-resistant *S. aureus* (CA-MRSA), have become common among U.S. urban populations since the 1980s (5). A recent report by the U.S. Centers for Disease Control and Prevention listed MRSA as causing nearly 80,500 severe infections and 11,300 deaths in 2011 alone (6). Here, we present a scaffolded genome assembly of *S. aureus* CDC73-57501 (ATCC 29247), a methicillin-resistant strain used in diagnostic and quality control measures.

High-quality genomic DNA was extracted from purified isolates of each strain using a Qiagen Genomic-tip 500 at the USAMRIID Diagnostic Systems Division (DSD). Specifically, a 100-mL bacterial culture was grown to stationary phase and nucleic acid was extracted per the manufacturer's recommendations. Sequence data generated included a combination of Illumina and 454 technologies (7, 8). For each genome, we constructed and sequenced an Illumina library of 100-bp reads at 669-fold genome coverage and a separate long-insert paired-end library (6,988 \pm 1,747-bp insert and 14-fold genome coverage) (Roche 454 Titanium platform). The two libraries were assembled together in Newbler (version 2.6; Roche), and the consensus sequences were computationally shredded into 2-kbp overlapping fake reads (shreds). The raw reads were also assembled in Velvet (version 1.2.08), and those consensus sequences were computationally shredded into 1.5-kbp overlapping shreds (9). Draft data from all platforms were then assembled together with Allpaths (version 39750), and the consensus sequences were computationally shredded into 10-kbp overlapping shreds (10). We then integrated the Newbler consensus shreds, Velvet consensus shreds, Allpaths

consensus shreds, and a subset of the long-insert read pairs using parallel Phrap (version SPS-4.24; High Performance Software, LLC). Possible misassemblies were corrected and some gap closure accomplished with manual editing in Consed (11–13).

Automatic annotation of the *S. aureus* 73-57501 genome utilized an Ergatis-based workflow at LANL with minor manual curation. The final assembly is 2,927,712 bp long (32.9% G+C content) with 2,778 coding, 7 rRNA, and 58 tRNA sequences. Preliminary review revealed *mecI* (methicillin resistance), *tetM* (tetracycline resistance), and three penicillin binding proteins, as well as the regulator *agrA* often associated with virulence (14).

Nucleotide sequence accession number. The annotated genome assembly of *S. aureus* 73-57501 is available in GenBank under the accession number JOVN00000000.

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REFERENCES

- Archer GL. 1998. Staphylococcus aureus: a well-armed pathogen. Clin. Infect. Dis. 26:1179–1181. http://dx.doi.org/10.1086/520289.
- Lowy FD. 1998. Staphylococcus aureus infections. N. Engl. J. Med. 339: 520–532. http://dx.doi.org/10.1056/NEJM199808203390806.
- Bae T, Banger AK, Wallace A, Glass EM, Åslund F, Schneewind O, Missiakas DM. 2004. *Staphylococcus aureus* virulence genes identified by bursa aurealis mutagenesis and nematode killing. Proc. Natl. Acad. Sci. U. S. A. 101:12312–12317. http://dx.doi.org/10.1073/pnas.0404728101.
- 4. Neu HC. 1992. The crisis in antibiotic resistance. Science 257:1064–1073. http://dx.doi.org/10.1126/science.257.5073.1064.

- Dukic VM, Lauderdale DS, Wilder J, Daum RS, David MZ. 2013. Epidemics of community-associated methicillin-resistant *Staphylococcus aureus* in the United States: a meta-analysis. PLoS One 8:e52722. http:// dx.doi.org/10.1371/journal.pone.0052722.
- U.S. Centers for Disease Control and Prevention. 2013. Antibiotic resistance threats in the United States. U.S. Centers for Disease Control and Prevention, Atlanta, GA. http://www.cdc.gov/drugresistance/threat -report-2013/pdf/ar-threats-2013-508.pdf.
- Bennett S. 2004. Solexa Ltd. Pharmacogenomics 5:433–438. http:// dx.doi.org/10.1517/14622416.5.4.433.
- 8. Margulies M, Egholm M, Altman WE, Attiya S, Bader JS, Bemben LA, Berka J, Braverman MS, Chen Y-J, Chen Z, Dewell SB, Du L, Fierro JM, Gomes XV, Godwin BC, He W, Helgesen S, Ho CH, Irzyk GP, Jando SC, Alenquer MLI, Jarvie TP, Jirage KB, Kim J-B, Knight JR, Lanza JR, Leamon JH, Lefkowitz SM, Lei M, Li J, Lohman KL, Lu H, Makhijani VB, McDade KE, McKenna MP, Myers EW, Nickerson E, Nobile JR, Plant R, Puc BP, Ronan MT, Roth GT, Sarkis GJ, Simons JF, Simpson JW, Srinivasan M, Tartaro KR, Tomasz A, Vogt KA, Volkmer GA, Wang SH, Wang Y, Weiner MP, Yu P, Begley RF, Rothberg JM. 2005. Genome sequencing in microfabricated high-density picolitre reactors. Nature 437:376–380. http://dx.doi.org/10.1038/nature03959.
- Zerbino DR, Birney E. 2008. Velvet: algorithms for *de novo* short read assembly using de Bruijn graphs. Genome Res. 18:821–829. http:// dx.doi.org/10.1101/gr.074492.107.
- Butler J, MacCallum I, Kleber M, Shlyakhter IA, Belmonte MK, Lander ES, Nusbaum C, Jaffe DB. 2008. ALLPATHS: *de novo* assembly of wholegenome shotgun microreads. Genome Res. 18:810–820. http:// dx.doi.org/10.1101/gr.7337908.
- Ewing B, Hillier L, Wendl MC, Green P. 1998. Base-calling of automated sequencer traces using phred. I. Accuracy assessment. Genome Res. 8:175–185. http://dx.doi.org/10.1101/gr.8.3.175.
- Ewing B, Green P. 1998. Base-calling of automated sequencer traces using phred. II. Error probabilities. Genome Res. 8:186–194.
- Gordon D, Abajian C, Green P. 1998. Consed: a graphical tool for sequence finishing. Genome Res. 8:195–202. http://dx.doi.org/10.1101/ gr.8.3.195.
- Jarraud S, Mougel C, Thioulouse J, Lina G, Meugnier H, Forey F, Nesme X, Etienne J, Vandenesch F. 2002. Relationships between *Staphylococcus aureus* genetic background, virulence factors, *agr* groups (alleles), and human disease. Infect. Immun. 70:631–641. http://dx.doi.org/ 10.1128/IAI.70.2.631-641.2002.