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# **Cross-Relaxation Imaging of Human Patellar Cartilage In-Vivo at 3.0T**

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# **Abstract**

**Objective—**To compare quantitative magnetization transfer (qMT) parameters of patellar cartilage measured using cross relaxation imaging (CRI) in asymptomatic volunteers and patients with osteoarthritis.

**Design—**The study was performed with Institutional Review Board approval and with all subjects signing informed consent. CRI of the knee joint was performed at 3.0T on 20 asymptomatic volunteers and 11 patients with osteoarthritis. The fraction of macromolecular bound protons (f), the exchange rate constant between macromolecular bound protons and free water protons (k), and the  $T_2$  relaxation time of macromolecular bound protons  $(T_2^B)$  of patellar cartilage were measured. Mann-Whitney-Wilcoxon rank-sum tests were used to compare qMT parameters between asymptomatic volunteers and patients with osteoarthritis.

**Results—**Average f, k, and  $T_2^B$  of patellar cartilage was 12.46%, 7.22 s<sup>-1</sup>, and 6.49 µs respectively for asymptomatic volunteers and 12.80%, 6.13 s<sup>-1</sup>, and 6.80 µs respectively for patients with osteoarthritis. There were statistically significant differences between groups of subjects for k (p<0.01) and  $T_2^B$  (p<0.0001) but not f (p=0.38) of patellar cartilage.

Competing Interests

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None of the authors have any conflicts of interests to declare.

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**Conclusion—**Patients with osteoarthritis had significantly lower k and significantly higher T<sub>2</sub><sup>B</sup> of patellar cartilage than asymptomatic volunteers which suggests that qMT parameters can detect changes in the macromolecular matrix of degenerative cartilage. Key Words: Cartilage; MRI; Osteoarthritis; Magnetization Transfer

# **Introduction**

Osteoarthritis is one of the most prevalent chronic diseases in the United States and worldwide [1]. Characteristic changes in the macromolecular matrix of articular cartilage occur during osteoarthritis including a decrease in proteoglycan content and disruption of the highly organized collagen fiber network [2–4]. Techniques to non-invasively assess the cartilage macromolecular matrix would be beneficial in osteoarthritis research studies to monitor disease-related and treatment-related changes in the composition and ultra-structure of cartilage [5]. Sensitive methods to detect early cartilage degeneration would also be useful in clinical practice to identify the cause of joint pain in symptomatic patients [6] and to allow early initiation of interventions such as weight loss, aerobic activity, and range of motion and strengthening exercises which may alleviate symptoms and potentially slow the rate of joint degeneration [7].

Various quantitative magnetic resonance (MR) imaging techniques have been used to evaluate articular cartilage. Multiple techniques including gadolinium enhanced spin-lattice relaxation time  $(T_1)$  imaging [8, 9], sodium imaging [10, 11], spin-lattice relaxation time in the rotating frame  $(T_{1rbo})$  imaging [12, 13], and chemical exchange-dependent saturation transfer (CEST) imaging [14, 15] have been shown to be sensitive for detecting changes in the proteoglycan content of cartilage. However, only a few MR techniques including spinspin relaxation time  $(T_2)$  imaging [16–18] and diffusion tensor imaging [19–22] have been used to identify alterations in cartilage ultra-structure, and all currently used methods have limitations. T2 relaxation time is a nonspecific parameter which is influenced by multiple factors including organization of the collagen fiber network [16–18], water and macromolecular content [23–26], and orientation of cartilage relative to the main magnetic field [27]. While diffusion tensor imaging may provide sensitive and specific information regarding cartilage ultra-structure, it is technically challenging and typically requires the use of high field strength scanners and custom made coils which has limited its use for evaluating human articular cartilage in-vivo [19–22].

Quantitative magnetization transfer (qMT) imaging is MR technique which utilizes a twopool model of magnetization exchange to acquire information regarding the cartilage macromolecular matrix [28–30]. qMT imaging techniques typically require multiple MTcontrast images with different magnetization preparatory pulses resulting in long scan times which have limited cartilage assessment to ex-vivo specimens [31–33]. Cross-relaxation imaging (CRI) is a qMT method which can create three-dimensional parametric maps of articular cartilage measuring the fraction of macromolecular bound protons (f), the exchange rate constant between macromolecular bound protons and free water protons  $(k)$ , and the  $T<sub>2</sub>$ relaxation time of macromolecular bound protons ( ${\rm T_2^B}$ ) with high resolution and relatively short scan time based upon a limited number of MT-contrast images [34–36]. The parameter f provides an indirect measure of macromolecular content, while the parameters k, and  $T_2^B$ 

reflect the efficiency of magnetization exchange between macromolecular bound protons and free water protons and the spin diffusion between proton sites in macromolecules respectively which may be influenced by macromolecular organization and ultra-structure [30, 37, 38]. We have developed a CRI protocol for evaluating human patellar cartilage invivo at 3.0T which can provide robust measurements of f, k, and  $T_2^B$  in a 19 minute scan time. This study was performed to compare qMT parameters of patellar cartilage measured using CRI in asymptomatic volunteers and patients with osteoarthritis to determine whether the MR technique can detect changes in the macromolecular matrix of degenerative cartilage.

# **Materials and Methods**

#### **Study Group**

The study was performed in compliance with HIPAA regulations and with approval from our Institutional Review Board. All subjects signed informed consent prior to their participation in the study. The study group consisted of 20 asymptomatic volunteers (15 males and five females between 23 and 45 years of age with an average age of 32.3 years) and 11 patients with osteoarthritis of the knee joint (seven males and four females between 45 and 62 years of age with an average age of 52.6 years). All patients with osteoarthritis of the knee joint complained of chronic knee pain and stiffness for a minimum of six months and showed definitive grade 2 osteophytes with no associated joint space loss on standing anterior-posterior radiographs of the knee [46, 47] . All patients had mild osteoarthritis within the patellofemoral compartment with 6 patients showing small grade 1 osteophytes and 5 patients showing definitive grade 2 osteophytes on the patella and femoral trochlea and no patients showing joint space narrowing on axial radiographs of the knee [46].

#### **MR Examination**

An MR examination of the knee joint was performed on all subjects in the study group using a 3.0T scanner (Discovery MR750, GE Healthcare, Waukesha, WI) and an 8-channel phased-array extremity coil (In Vivo, Orlando, FL). Foam padding was used to firmly secure the knee within the coil to minimize subject motion during the MR examination. All MR examinations consisted of the following sequences performed in the axial plane through the patellofemoral compartment of the knee joint: 1) the CRI protocol, 2) a frequency-selective fat-suppressed T2-weighted fast spin-echo sequence acquired using a 4050 ms repetition time, 85 ms echo time, 90° excitation flip angle, 31 kHz bandwidth, 14 cm field of view, 256 x 256 matrix, 4 mm slice thickness, and four signal averages, and 3) an SPGR sequence with iterative decomposition of water and fat with echo asymmetry and least-squares estimation (IDEAL) fat-water separation [48] acquired using a 12.4 ms repetition time, 3.4 ms, 4.2 ms, and 5.0 ms echo times, 14° excitation flip angle, 31 kHz bandwidth, 14 cm field of view, 256 x 256 matrix, 4 mm slice thickness, and one signal average. A frequencyselective fat-suppressed three-dimensional intermediate-weighted fast spin-echo sequence was also performed in the sagittal plane through the knee joint using a 2217 ms repetition time, 23.6 ms echo time, 90° excitation flip angle, 31 kHz bandwidth , 15 cm field of view, 256 x 256 matrix, 1 mm slice thickness, and one signal average. Coronal and axial reformat images were created from the volumetric fast spin-echo source data.

The CRI protocol consisted of six MT-prepared SPGR scans and four non-MT-prepared SPGR scans. The MT-prepared SPGR scans were performed with different combinations of MT offset frequencies and flip angles (2.5 kHz/1550°, 5 kHz/1550°, 10 kHz/1550°, 20 kHz/ 1550°, 2.5 kHz/890°, and 5 kHz/890°) with an 18-ms Fermi MT pulse, 42 ms repetition time, 3.2 ms echo time, and 13° excitation flip angle. The non-MT-prepared SPGR scans were performed over a range of excitation flip angles (4°, 10°, 20°, and 30°) with a 24 ms repetition time and 3.2 ms echo time. Actual flip angle imaging (AFI) was also performed for flip angle mapping using an SPGR scan consisting of two identical radiofrequency pulses followed by repetition times of 30 ms and 150 ms acquired with a 2.2 ms echo time and 55° excitation flip angle [45]. All SPGR scans were performed using a 31 kHz bandwidth, 14 cm field of view, 256 x 256 matrix for the MT-prepared and non-MTprepared scans and 128 x 128 matrix for the AFI scans, 4 mm slice thickness, 10 slices, and one signal average. Total scan time for the CRI protocol was 19 minutes. In order to measure MTR, one additional MT-prepared SPGR scan was performed using the same imaging parameters except for a 250 kHz offset frequency and 1550° flip angle to create negligible MT effect.

#### **Cartilage qMT Parameter Map Reconstruction**

Quantitative MR parameter maps of patellar cartilage were reconstructed using in-house software developed in MATLAB (MATLAB 2011b, MathWorks Inc, Natick, MA). Image registration software (FLIRT, Functional Magnetic Resonance Imaging of the Brain Analysis Group, Oxford University, United Kingdom) was used to correct for any subject motion which may have occurred between the multiple scans. MT-prepared SPGR scans, non-MT-prepared SPGR scans, and AFI scans were co-registered using the IDEAL-SPGR scan as the reference. The MT-prepared and non-MT-prepared SPGR datasets were simultaneously fitted on a pixel-by-pixel basis using a non-linear least squares two-pool model to create cartilage f, k and  $T_2^B$  maps [35]. Both excitation flip angle and MT saturation power were corrected for each pixel using the flip angle maps acquired from the AFI scans. In addition, cartilage MTR maps were created using a pixel-by-pixel measurement of the difference in the signal of the SPGR scan with negligible MT effect (250kHz/1550°) and the SPGR scan with strongest MT effect (2.5kHz/1550°) divided by the signal of the SPGR scan with negligible MT effect [43, 49].

#### **Comparison of Morphologic and Quantitative MR Parameters Between Groups of Subjects**

Morphologic joint analysis was performed by a fellowship-trained musculoskeletal radiologist with 12 years of clinical experience who was blinded to whether a subject was an asymptomatic volunteer or patient with osteoarthritis. The radiologist used the axial fatsuppressed T2-weighted fast spin-echo, axial IDEAL-SPGR, and multi-planar fatsuppressed three-dimensional intermediate-weighted fast spin-echo images to grade the severity of degeneration within the knee joint using the Boston-Leeds Osteoarthritis Knee (BLOK) scoring system [50]. A patellar BLOK score and total knee BLOK score was calculated for each subject using the semi-quantitative scoring system.

Quantitative cartilage analysis was performed by a research assistant with four years of cartilage segmentation experience under the supervision of the fellowship-trained

musculoskeletal radiologist using software developed in MATLAB (MATLAB 2011b, MathWorks Inc, Natick, MA). Regions of interest were placed around the patellar cartilage on each slice of the IDEAL-SPGR images of each subject to create a three-dimensional contour of patellar cartilage. The three-dimensional contour was then superimposed over the cartilage MTR, f, k, and  $T_2{}^{\rm B}$  maps to measure the average quantitative MR parameters of patellar cartilage. Non-parametric two-tailed Mann-Whitney-Wilcoxon rank-sum tests were used to compare MTR, f, k, and  $T_2^B$  of patellar cartilage between asymptomatic volunteers and patients with osteoarthritis. Two-tailed Mann-Whitney-Wilcoxon rank-sum tests were also used to compare MTR, f, k, and  $T_2^B$  of patellar cartilage in patients with osteoarthritis who had small grade 1 osteophytes and definitive grade 2 osteophytes on the patella and femoral trochlea. Spearman's rank correlation coefficients were used to determine the association between MTR, f, k, and  $T_2^B$  of patellar cartilage and the patellar and total knee BLOK scores in patients with osteoarthritis.

Non-parametric statistical analysis was chosen due to the presence of outliers and nonconstant variance of qMT parameters in the subject populations which violated the assumptions of normality and homoscedasticity. The Bonferonni method was used to account for comparison of four quantitative MR parameters between groups of subjects with statistical significance defined as a p-value less than 0.01 due to

#### **Assessment of Repeatability of Cartilage Quantitative MR Measurements**

The CRI protocol was performed twice on both knee joints of five asymptomatic volunteers (five males between 28 and 32 years of age with an average age of 29.2 years) with the subjects taken out of the scanner and allowed to rest in a sitting position for 10 minutes between the scans for a total of four scans per subject. Average MTR, f, k, and  $T_2^{\ B}$  of patellar cartilage were measured using the previously described methods. Repeatability of cartilage qMT measurements was assessed using conventional [51–53] and standardized [54] coefficients of variance. Confidence intervals for coefficient of variance were calculated using the approximate pivotal method [55]. Bland-Altman analysis was also performed to assess the estimated bias or mean of differences and 95% limits of agreement for the repeat cartilage qMT measurements obtained using the two CRI scans [56]. Student t-tests were used to determine the significance of the estimated bias of the repeat measurements by testing whether the mean of differences was equal to 0. Significance of the estimated bias was also determined by observing whether the 95% confidence intervals of the mean of differences on the Bland-Altman plots included 0.

#### **Assessment of Signal-to-Noise Ratio (SNR) of the CRI Protocol**

Monte Carlo simulations were used to compare the signal-to-noise ratio (SNR) to the variations due to error in MTR, f, k, and  $T_2^B$  [57]. One hundred digital phantoms, each containing 2000 pixels, were created using the average MTR, f, k, and  $T_2^{\ B}$  of all subjects in the study. Different levels of normally distributed noise were added to the digital signals to simulate SNR levels from 0 to 160 in increments of 20. For each SNR level, the digital phantom data was used to obtain estimates of MTR, f, k, and  $T_2^B$  [35]. The average MTR, f, k, and  $T_2^{\; {\rm B}}$  were calculated over each digital phantom at each SNR level. Standard deviations of the averages were taken at each SNR level. The standard deviations were then

normalized as a percentage of the average MTR, f, k, and  $T_2{}^{\rm B}$  values. The results were compared to the observed differences in MTR, f, k, and  $T_2^B$  between asymptomatic volunteers and patients with osteoarthritis at the reference SNR for the CRI protocol. The reference SNR for the CRI protocol was determined using the addition/subtraction method in which two identical SPGR scans with negligible MT effect were performed on an asymptomatic volunteer one immediately following the other. Signal was defined as the signal of patellar cartilage on the addition images divided by 2, while noise was defined as the standard deviation of patellar cartilage on the subtraction images divided by the square root of 2 [58].

# **Results**

All asymptomatic volunteers had a patellar BLOK score and total knee BLOK score of 0 indicating no degeneration within the knee joint. Patients with osteoarthritis had patellar BLOK scores ranging between 6 and 13 with an average value of 8.0 and total knee BLOK scores ranging between 27 and 58 with an average value of 36.1. All patients with osteoarthritis had osteophytes and focal areas of partial-thickness cartilage loss on the patella with two patients also having subchondral bone marrow edema.

Figure 1 shows box plots illustrating the distribution of MTR, f, k, and  $T_2^B$  values of patellar cartilage for asymptomatic volunteers and patients with osteoarthritis. Table 1 shows average MTR, f, k, and  $T_2^B$  values of patellar cartilage with standard deviations and 95% confidence intervals for asymptomatic volunteers and patients with osteoarthritis. There was significantly lower k (p<0.01) and significantly higher  $T_2^B$  (p<0.0001) of patellar cartilage in patients with osteoarthritis than asymptomatic volunteers. There was no significant difference in MTR ( $p=0.52$ ) and f ( $p=0.38$ ) of patellar cartilage between groups of subjects. There was no significantly difference in MTR (p=0.100), f (p=0.13), k (p=1.00), and  $T_2^B$  $(p=1.00)$  of patellar cartilage between patients with osteoarthritis who had small grade 1 osteophytes and definitive grade 2 osteophytes on the patella and femoral trochlea. There was no significant correlation in patients with osteoarthritis between patellar BLOK scores and total knee BLOK scores and MTR ( $p=0.56$  and  $p=0.77$  respectively), f ( $p=0.45$  and p=0.77 respectively), k (p=0.51 and p=0.53 respectively), and  $T_2^B$  (p=0.83 and p=0.82 respectively) of patellar cartilage. Figures 2, 3, and 4 illustrate examples of differences in the qMT parameters f, k, and  $T_2^B$  of patellar cartilage between asymptomatic volunteers and patients with osteoarthritis.

Conventional coefficients of variance for repeat qMT measurements obtained using the two CRI scans performed on the same knee of the same subject were 0.45% for MTR (95% confidence intervals 0.44% to 0.48%), 2.21% for f (95% confidence intervals 2.15% to 2.35%), 4.56% for k (95% confidence intervals 4.44% to 4.87%), and 0.72% for  $T_2^{\ B}$  (95% confidence intervals 0.70% to 0.77%). Standardized coefficients of variance for repeat qMT measurements were 10.28% for MTR (95% confidence intervals 10.02% to 10.96%), 9.55% for f (95% confidence intervals 9.30% to 10.18%), 6.22% for k (95% confidence intervals 6.06% to 6.63%), and 10.81% for  $T_2^B$  (95% confidence intervals 10.53% to 11.53. Figure 5 shows the Bland-Altman plots with estimated bias or mean of differences with 95% confidence intervals and 95% limits of agreement for repeat qMT measurements. The

estimated biases for repeat measurements on Bland-Altman analysis were 0.01 for MTR, −0.24 for f, −0.05 for k, and 0.02 for  $T_2^B$ . The estimated biases for the repeat measurements were not statistically significant with the p-values for the student t-tests ranging between 0.06 and 0.84 and all 95% confidence intervals for the mean of differences including 0.

The relationships between signal-to-noise ratio (SNR) and variations due to error in MTR, f, k, and  $T_2^B$  are illustrated in Figure 6. The normalized standard deviation for a given parameter may be interpreted as the percent change expected due to noise and data fit instability at a certain SNR level. At the reference SNR of 75 for the CRI protocol, the percent change expected due to noise and data fit instability was approximately 0.1% in MTR, 0.1% in f, 1% in k, and 0.1% in  $T_2^B$ . In comparison, the percent change of the differences in qMT values of patellar cartilage between asymptomatic volunteers and patients with osteoarthritis was 0.1% for MTR, 2.8% in f,  $-15.9\%$  in k, and 4.7% in  $T_2^B$ .

# **Discussion**

Our study has demonstrated the feasibility of using CRI to measure the qMT parameters f, k, and  $T_2^B$  of the articular cartilage of the knee joint in human subjects which has never been previously performed to the best of our knowledge. Our study found significant differences in f and  $T_2^B$  of patellar cartilage between asymptomatic volunteers and patients with osteoarthritis which suggests that qMT parameters can detect changes in the macromolecular matrix of degenerative cartilage. MTR could not distinguish between groups of subjects which is similar to the findings of a previous study which found no significant difference in MTR of patellar cartilage between asymptomatic volunteers and patients with osteoarthritis [59]. These results suggest that a more detailed analysis of the magnetization exchange between macromolecular bound protons and free water protons provided by CRI is needed to detect changes in the cartilage macromolecular matrix.

Our study found significantly lower k and significantly higher  $T_2^B$  of patellar cartilage in patients with osteoarthritis when compared to asymptomatic volunteers. Few previous studies have investigated k and  $T_2^B$  of cartilage, and thus the exact mechanisms of the observed changes in the qMT parameters remain unknown. However, previous ex-vivo studies have shown that k decreases [60, 61] and  $T_2^{\;B}$  increases [60] with proteoglycan loss due to trypsin degradation of bovine cartilage specimens. Furthermore, thermal denaturation of collagen solution has been shown to cause an increase in  $T_2^B$  although the change was accompanied by a decrease in f [60]. Thus, the decrease in k and increase in  $T_2^{\ B}$  with cartilage degeneration in our study may be due to the combined effects of proteoglycan loss and collagen denaturation which both occur during the early stages of osteoarthritis [2–4]. Fragmentation of collagen decreases proton binding sites on the macromolecule and may thereby reduce the exchange rate between macromolecular bound and free water protons. Fragmentation of collagen may also increase the mobility of the macromolecule within cartilage and thereby increase  $T_2^B$ . Decreased organization of the cartilage matrix due to collagen denaturation and proteoglycan loss may increase spin diffusion of macromolecularbound protons, the primary mechanism defining  $T_2$  relaxation time in the semisolid fraction, and thereby also increase  $T_2^B$  [38].

Our study found no significant difference in f of patellar cartilage in patients with osteoarthritis when compared to asymptomatic volunteers. The factors responsible for the measured f values of cartilage are incompletely understood. A previous study using CRI to investigate ex-vivo human cadaveric cartilage specimens found a moderate correlation between f and the proteoglycan content of cartilage [36]. However, the two-pool model used in the study fixed  $T_2^B$  which has been shown in our study to change with cartilage degeneration. Another study using proteoglycan and collagen phantoms and a similar CRI protocol as our study has documented large increases in f with increasing collagen concentration but only negligible increases in f with increasing proteoglycan concentration. The same study found no change in f with proteoglycan loss due to trypsin degradation of ex-vivo bovine cartilage specimens suggesting that proteoglycan content has a minimal effect on f [60]. Proteoglycan has an abundance of macromolecular bound protons, but its concentration within cartilage is lower [2] and its protons are more mobile [62] when compared to collagen which may limit its contribution to f. The absence of changes in f with cartilage degeneration in our study may be due to the fact that f is primarily a measure of the collagen content of cartilage which decreases by only 5% during the late stages of osteoarthritis [63]. However, additional studies are needed to investigate the influence of collagen and proteoglycan on the measured f values of both normal and degenerative cartilage.

The CRI protocol used in our study had adequate SNR to detect differences in qMT parameters between groups of subjects and high repeatability with conventional coefficients of variance which compared favorably with other quantitative cartilage imaging techniques [51–53]. However, k was more sensitive to measurement errors when compared to f and  $T_2^B$ with higher percent change expected due to noise and data fit instability. These results are consistent with the findings of a previous study investigating qMT parameters within neural tissue which showed that small fluctuations in MT signals due to measurement error had a more significant effect on k than f and  $T_2^B$  [64]. One method to improve measurements of k of articular cartilage would be to acquire SPGR scans at higher MT flip angles than those used in our study. However, using higher MT flip angles would increase specific absorption rate (SAR) so SAR would need to be reduced by using lower field strength scanners at the expense of decreased SNR or longer repetition times at the expense of increased scan time.

Our study has several limitations. One limitation was the relatively small number of subjects which prevented detailed analysis of inter-group variability in qMT parameters and identification of thresholds for k and  $T_2^B$  which could be considered diagnostic for cartilage degeneration. Furthermore, the small number of patients with osteoarthritis provided low statistical power for comparison of qMT parameters with radiographic and MR parameters of cartilage degeneration. Another limitation was that not all patients with osteoarthritis in our study had definitive grade 2 osteophytes within the patellofemoral compartment which is considered to be the radiographic hallmark of the disease [47]. However, all patients did have osteoarthritis of the whole knee joint diagnosed using standardized clinical and radiographic criteria [47, 65]. Furthermore, a group of patients with mild osteoarthritis within the patellofemoral compartment was desired to determine whether CRI could detect macromolecular changes associated with early patellar cartilage degeneration. Another limitation of our study was that it could not identify the mechanisms responsible for changes

in k and  $T_2^B$  between asymptomatic volunteers and patients with osteoarthritis. Additional studies correlating qMT parameters in ex-vivo cartilage specimens with proteoglycan and collagen content measured using biochemical assays and tissue ultra-structure measured using polarized light microscopy are needed to investigate the factors responsible for changes in qMT parameters at various stages of cartilage degeneration. A final limitation was that the CRI protocol used in our study had a relatively long scan time which limited qMT assessment to 10 slices through patellar cartilage. We are currently investigating various methods to reduce the scan time of the CRI protocol including use of a smaller number of optimized magnetization preparatory pulses, more rapid methods for flip angle mapping, and compressed sensing with parallel imaging to provide complete anatomic coverage of the knee joint in the sagittal plane.

In conclusion, our study has shown that patients with osteoarthritis have significantly lower k and significantly higher  $T_2^B$  of patellar cartilage than asymptomatic volunteers which suggests that qMT parameters can detect changes in the macromolecular matrix of degenerative cartilage. CRI may provide a new quantitative MR technique to identify patients with early cartilage degeneration and to monitor disease-related and treatmentrelated changes in the macromolecular matrix of articular cartilage in osteoarthritis research studies. However, further studies are needed to better understand the fundamental mechanisms responsible for changes in qMT parameters in patients with osteoarthritis and to identify thresholds of k and  $T_2^B$  which could be considered diagnostic for cartilage degeneration.

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#### **Figure 1.**

Box plots illustrating the distribution of (A) MTR, (B) f, (C) k, and (D)  $T_2^{\ B}$  values of patellar cartilage in asymptomatic volunteers and patients with osteoarthritis. The boxes indicate interquartile ranges, the blue dots within the boxes indicate average values, the red lines within the boxes indicate median values, the whiskers extend to the most extreme data points not considered outliers (1.5 of the interquartile range between the first and third quartiles), and the crosses indicate outliers.



# **Figure 2.**

Comparison of f of patellar cartilage in (A) a 25 year old male asymmetric volunteer and (B) a 52 year old male patient with osteoarthritis. Note that there are no visible differences in f between the asymptomatic volunteer and the patient with osteoarthritis.



# **Figure 3.**

Comparison of k of patellar cartilage in (A) a 27 year old male asymmetric volunteer and (B) a 45 year old male patient with osteoarthritis. Note that the patient with osteoarthritis has lower k of patellar cartilage than the asymptomatic volunteer.



# **Figure 4.**

Comparison of  $T_2^B$  of patellar cartilage in (A) a 27 year old male asymmetric volunteer and (B) a 45 year old male patient with osteoarthritis. Note that the patient with osteoarthritis has higher  $T_2^B$  of patellar cartilage than the asymptomatic volunteer.

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#### **Figure 5.**

Bland-Altman plots for (A) MTR, (B) f, (C) k, and (D)  $T_2^B$ . The two solid blue lines indicate 95% limits of agreement, and the dotted blue line indicates the estimated bias or mean of differences. The two solid red lines indicate 95% confidence intervals (CI) of the mean of differences which can be used to determine the significance of the estimated bias. Note that for all qMT parameters, the 95% CI of the mean of differences includes 0 indicating no statistically significant estimated bias between the repeat measurements.



# **Figure 6.**

Relationship between normalized standard deviation due to error and SNR for MTR, f, k, and  $T_2^{\;B}$  of patellar cartilage. The reference SNR of 75 for the CRI protocol (dotted black vertical line) provides low percent change due to error in all qMT parameters.

# **Table 1**

Average MTR, f, k, and  $T_2^B$  values of patellar cartilage with standard deviations (SD) and 95% confidence intervals (CI) in asymptomatic volunteers and patients with osteoarthritis

