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Influence of Single Nucleotide Polymorphisms in COMT, MAO-A and BDNF Genes on Dyskinesias and Levodopa Use in Parkinson's Disease

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Abstract

Background—Clinical heterogeneity in the development of levodopa-induced dyskinesias (LID) suggests endogenous factors play a significant role in determining their overall prevalence.

Objective—We hypothesised that single nucleotide polymorphisms (SNPs) in specific genes may result in a clinical phenotype conducive to an increased risk of LID.

Methods—We examined the influence of SNPs in the catechol-O-methyltransferase (*COMT*), monoamine oxidase A (MAO-A) and brain-derived neurotrophic factor (*BDNF*) genes on LID in a cohort of 285 pathologically confirmed Parkinson's disease patients, using data from their complete disease course.

Results—Dyskinetic patients demonstrated younger age at disease onset (60.3 vs. 66.4 years, p < 0.0001), a longer disease duration (17.0 vs. 12.0 years, $p < 0.0001$) and a higher maximum daily levodopa equivalent dose (LED; 926.7 vs. 617.1 mg/day, p < 0.0001) than patients without dyskinesias. No individual SNP was found to influence prevalence or time to onset of dyskinesias, including after adjustment for known risk factors. We observed that patients carrying alleles conferring both high COMT activity and increased MAO-A mRNA expression received significantly higher maximum and mean daily LEDs than those with low enzyme activity/mRNA expression (max LED: 835 ± 445 vs. 508 ± 316 mg; p = 0.0056, mean LED: 601 ± 335 vs. 398 ± 100 260 mg; $p = 0.025$).

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Conclusions—Individual SNPs in *BDNF, COMT* and *MAO-A* genes did not influence prevalence or time to onset of dyskinesias in this cohort. The possibility that combined *COMT* and *MAO-A* genotype is a significant factor in determining an individual's lifetime levodopa exposure warrants further investigation.

Keywords

Parkinson's disease; Levodopa-induced dyskinesias; Catechol-O-methyltransferase; Monoamine oxidase A; Brain-derived neurotrophic factor

Introduction

Levodopa-induced dyskinesias (LID) are a substantial barrier to effective symptomatic management of Parkinson's disease (PD) and occur as a consequence of chronic levodopa (L-DOPA) treatment. Despite their high prevalence, therapeutic options are currently limited [1]. Epidemiological evidence suggests that the onset and severity of LID varies considerably, with some patients never developing them despite equivalent treatment regimens. This observation suggests endogenous factors play a role in individual susceptibility to LID, and despite established risk factors for developing LID including younger age at PD onset, higher L-DOPA dose, greater disease severity and longer disease duration [2], there has been little research to date regarding genetic susceptibility to LID.

In this study, we used a candidate gene approach to examine functional single nucleotide polymorphisms (SNPs) in which the cellular consequences of the polymorphism have been identified and are directly relevant to dyskinesias. Specifically, we focused on functional SNPs in genes encoding the catechol-O-methyltransferase (COMT) and monoamine oxidase A (MAO-A) enzymes, both of which catabolise dopamine and are central to the therapeutic response of L-DOPA. A valine to methionine substitution at codon 158 of the *COMT* gene produces a Met variant that catabolises dopamine up to four times slower than its Val counterpart [3]. Given the overlapping role of MAO with COMT, we anticipated that a similar finding might be observed for the synonymous substitution of T to G in exon 8 of the *MAO-A* gene, which promotes MAO-A mRNA expression [4].

Finally, a valine to methionine substitution at codon 66 of the brain-derived neurotrophic factor (BDNF) gene has been identified as having a putative role in influencing time to onset of dyskinesia in PD [5]. We hypothesised that these polymorphisms, individually or combined, may contribute to the risk of developing dyskinesias in PD.

Patients and Methods

Case Selection

We identified 285 pathologically confirmed PD cases from the Australian Brain Bank Network (ABBN), Australia and the Queen Square Brain Bank for Neurological Disorders (QSBB), UK with a history of L-DOPA usage and a disease duration of at least 5 years. Patients were excluded if they had confirmed monogenic PD, early symptom onset (40) years of age) or late symptom onset (80 years of age). Approval for the collection of brain tissue as well as retention of and access to clinical records was granted by the Human

Research Ethics Committee of the University of Melbourne (ABBN) and the London Multi-Centre Research Ethics Committee (QSBB).

Genotyping

DNA was extracted using standard methods (QIAamp DNA Kit, Qiagen) and genotyping was performed via the SEQUENOM™ genotyping platform at the Australian Genome Research Facility for all Australian cases. Cases from the UK were genotyped for the Val158Met *COMT* polymorphism (dbSNP rs4680) and the T941G *MAO-A* polymorphism (rs6323) as per Spencer et al. [6]. The *BDNF* Val66Met polymorphism (rs6265) was genotyped as per Foltynie et al. [7].

Clinical Data

A systematic review of patients' medical records was performed by movement disorder specialists (K.B., H.L. and S.O'S.) and data including age at PD onset, disease duration, prevalence and time of onset of dyskinesia, and dopaminergic medication history were recorded. Levodopa equivalent dose (LED), which accounts for other antiparkinsonian drugs, was calculated as per Tomlinson et al. [8]. An approximation of the cumulative lifetime L-DOPA dose was estimated using previously published methods [9]. Mean daily LED was obtained using the calculated lifetime estimate and adjusting for the number of years of L-DOPA treatment.

Statistical Analysis

We constructed Kaplan-Meier survival curves, with the time-dependent variable set as the first recorded incidence of dyskinesias in dyskinetic patients, or disease duration for patients without dyskinesia. Time zero was set as age of PD onset, as this data was more reliably recorded than age at first L-DOPA administration. We used Cox proportional hazards regression and generalised linear modelling to examine the relationship between genotype and time to onset of LID, adjusting for established risk factors. As the distribution of LED was skewed, we used logarithmic transformation to normalise the distribution prior to linear modelling. Statistical analysis was performed using GraphPad Prism (version 5.0, GraphPad Software, San Diego, Calif., USA) or SPSS (version 20, IBM SPSS, New York, N.Y., USA).

Results

Patients in our combined cohort demonstrated typical PD demographics, with a mean age of onset of 63.0 ± 9.2 years, a mean disease duration of 14.8 ± 6.4 years and a mean maximum daily LED of 794.2 \pm 431.6 mg/day. Dyskinesias were reported in 61.3% of patients. Dyskinetic patients demonstrated established risk factors for dyskinesias, including younger age of PD onset (60.3 vs. 66.4 years, p < 0.0001), a longer disease duration (17.0 vs. 12.0 years, $p < 0.0001$) and a higher maximum daily LED (926.7 vs. 617.1 mg/day, $p < 0.0001$) than patients without dyskinesias.

When patients were stratified according to COMT, MAO-A or BDNF genotype, no individual genotype was found to independently influence the prevalence or time to onset of

dyskinesias (fig. 1). Individual genotypes were compared for each gene, and genotypes were pooled to examine the effect of homozygosity for either allele. No difference was observed for any clinical feature, including age at onset, disease duration and maximum or mean daily LED (table 1).

We considered that individual functional SNPs may be unlikely to exert a significantly large effect on dyskinesia in isolation, and that given the overlapping function of COMT and MAO-A, double homozygosity for both SNPs may exert a greater effect than homozygosity for either SNP individually. However, pooling genotypes demonstrated that patients with alleles conferring high COMT activity (Val/Val) and high MAO-A mRNA expression (T males; TT females) did not have a statistically significantly increased prevalence of dyskinesias (58.0 vs. 43.8%, $\chi^2 = 0.62$, d.f. = 1, p = 0.43) compared to those with low enzyme activity/mRNA expression. Interestingly, we observed that patients homozygous for high COMT activity and high MAO-A mRNA expression alleles received significantly higher maximum daily LED (835 \pm 445 mg) than those homozygous for low activity/ expression alleles (508 \pm 316 mg; p = 0.0056), as well as a higher mean daily LED (601 \pm 335 vs. 398 ± 260 mg; p = 0.0248). Generalised linear modelling confirmed the association between pooled COMT and MAO-A genotype and LED, with low activity/expression allele carriers receiving a mean maximum daily LED 43% lower ($p = 0.003$) than other genotype groups, after controlling for disease duration.

Discussion

In this study of pathologically confirmed PD, polymorphisms in COMT, MAO-A and BDNF genes, either individually or combined, had no influence on the prevalence or time to onset of LID (fig. 1). While COMT and BDNF have previously been implicated in the pathophysiology of dyskinesias, MAO-A is a novel candidate. COMT is primarily responsible for the breakdown of L-DOPA when DOPA decarboxylase inhibitors are coadministered with L-DOPA, blocking DOPA decarboxylase activity. The COMT rs4680 SNP has been associated with an increased risk of dyskinesias in a recent prospective study [10], although earlier cross-sectional studies failed to find a statistically significant effect [11].

MAO-A has a similar role to COMT in catabolising monoamines, and the T allele of the T941G MAO-A SNP, which confers increased mRNA expression, has been investigated as a risk factor for developing PD [12]. Despite this overlap in function, we found no evidence to suggest that this SNP individually influences dyskinesias. Similarly, met allele carriage of the BDNF Val66Met polymorphism has been associated with an earlier time to onset of dyskinesias [5]; however, our study found no association between BDNF genotype and prevalence or time to onset of dyskinesias, which may be a reflection of different study designs.

Our data suggest a putative relationship between combined COMT and MAO-A genotype and total lifetime L-DOPA exposure. Patients with alleles conferring high COMT activity and high MAO-A mRNA expression received significantly higher L-DOPA exposure than low activity/expression carriers. Although these data should be interpreted with caution

given the small sample size of this subgroup of double homozygotes, we note that if we reduce our p value to $p < 0.01$ to correct for multiple comparisons, our data remain statistically significant.

In a clinical setting, patients with increased COMT enzyme activity and increased MAO-A expression will metabolise L-DOPA faster, which may result in increased wearing off, necessitating higher or more frequent L-DOPA doses over the day, resulting in the higher lifetime exposure to L-DOPA observed in our study. This finding warrants replication in a larger cohort, although it is difficult to increase the sample size of a post-mortem cohort without adding cases from additional tissue banks, introducing genetic diversity that can obscure results.

The present study has some important limitations. The data were collected retrospectively using clinical case notes with the implicit assumption that a reasonably consistent threshold would apply for specialists to report dyskinesias. While our patient demographics appear to represent a typical PD population on the basis of age of onset and disease duration, there is likely to be a selection bias in cases submitted for brain banking.

In summary, we find no evidence to suggest the SNPs in COMT, MAO-A or BDNF examined in this study influence the prevalence or time to onset of LID. Our data suggest that combined COMT and MAO-A genotype may influence L-DOPA use, but further studies in different populations are required to clarify this effect.

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Fig. 1. Survival curves of time to onset of dyskinesia according to *COMT* **(a),** *MAO***-***A* **(b) or** *BDNF* **(c) genotype.**

Kaplan-Meier curves demonstrating duration from PD onset to first reported incidence of LID in patients stratified according to genotype. For the *BDNF* Val66Met polymorphism, given the low prevalence of the variant Met allele in European populations, Met/Met homozygous patients were pooled with Val/Met heterozygotes for statistical analysis. As the *MAO-A* gene is X-linked, males and females were analysed both separately and with male hemizygotes pooled with female homozygotes to directly compare the effects of T or G allele carriage.

Time to onset of dyskinesia, years 9.7±4.9 8.4±4.3 9.2±5.6 8.4±4.3 9.2±5.6 8.0±4.6 9.2±5.6 8.7±4.9 9.3±5.6 9.54
U 1.7±4.9 8.1±4.2 9.1±4.2 9.2±5.6 8.4±4.3 9.2±5.6 7.9±4.3 9.2±5.6 8.0±4.5 9.2±5.6 8.1±4.2 8.1±4.2 Dyskinesia reported, % 58.5 59.6 63.9 0.78*** 55.4 75.0 56.2 0.12*** 62.0 62.0 >0.90*** Mean dialy LED, mg/day 583±304 581±2035 521±312 520±275 530±275 530±275 590±275 590±275 590±2035 590±202 590±335 0.06
Mean dialy LED, mg/day 500±30±31 501±202 501±312 531±235 590±202 590±3035 590±302

 $8.0 + 4.3$

 0.67

 $8.4 + 4.2$

 $7.9 + 4.9$

 $8.7 + 4.9$

Time to onset of dyskinesia, years

 $^{\ast}0.90^{\ast}$ 0.62

 62.0

 $7.7 + 3.9$

 $8.1 + 4.2$ 62.0

 $8.0 + 4.6$

 $9.2 + 5.6$

0.06 0.08

 $619 + 335$ 845±465

497±202 $721 + 374$

0.83 0.33

590±299

 $551 + 235$

 $551 + 312$

 $520 + 275$

593±299

 $588 + 304$

Mean daily LED, mg/day Dyskinesia reported, %

 0.12 ^{*} 0.58

56.2

75.0

55.4

 $0.78*$ 0.30

63.9

59.6

58.5

Table 1

Maximum daily LED, mg/day 667±474 8687 101±454 845±424 845±454 845±454 845±464 845±464 845±465 0.08
Maximum daily LED, mg/day 814±430 $687 + 316$ Values are mean ± SD. Statistical analysis was non-parametric Kruskal-Wallis or Mann-Whitney test. Values are mean ± SD. Statistical analysis was non-parametric Kruskal-Wallis or Mann-Whitney test. 710±356 0.11 $783 + 424$ 846±464 667±294 Maximum daily LED, mg/day

p value from a χ^2 (Fisher's exact) test.
