

# NIH Public Access

**Author Manuscript**

*AIDS*. Author manuscript; available in PMC 2015 September 24.

Published in final edited form as:

*AIDS*. 2014 September 24; 28(15): 2319–2322. doi:10.1097/QAD.0000000000000419.

# **Novel tetra-peptide insertion in Gag-p6 ALIX-binding motif in HIV-1 subtype C associated with protease inhibitor failure**

**Ujjwal Neogi**a,#, **Shwetha D RAO**b, **Irene BONTELL**<sup>c</sup> , **Jens VERHEYEN**d, **Vasudev R RAO**e, **Sagar C GORE**<sup>f</sup> , **Neelesh SONI**<sup>f</sup> , **Anita SHET**b, **Eugen SCHÜLTER**g, **Maria L. EKSTRAND**h, **Amogne WONDWOSSEN**<sup>i</sup> , **Rolf KAISER**g, **Mallur S. MADHUSUDHAN**f,j,k , **Vinayaka R PRASAD**e, and **Anders SONNERBORG**a,c

<sup>a</sup> Division of Clinical Microbiology, Department of Laboratory Medicine, Karolinska Institutet, Stockholm, Sweden

**b Division of Public Health and Infection, St. John's Research Institute, Bangalore India** 

<sup>c</sup> Unit of Infectious Diseases, Department of Medicine Huddinge, Karolinska Institutet, Stockholm, Sweden

- <sup>d</sup> Institute of Virology, University-Hospital, University Duisburg-Essen, Essen, Germany
- <sup>e</sup> Department of Microbiology and Immunology, Albert Einstein College of Medicine, Bronx, New York.
- f Indian Institute of Science Education and Research, Pune, India
- <sup>g</sup> Institute of Virology, University of Cologne, Cologne, Germany
- h Department of Medicine, University of California, San Francisco, CA, USA
- <sup>i</sup> Department of Medicine, Faculty of Medicine, Addis Ababa University, Addis Ababa, Ethiopia
- <sup>j</sup> Bioinformatics Institute, Singapore
- k Department of Biological Sciences, National University of Singapore, Singapore

# **Abstract**

A novel tetra-peptide insertion was identified in Gag-p6 ALIX-binding region which is appears in protease inhibitor (PI) failure Indian HIV-1C sequences (Odds Ratio 17.1, p<0.001) but naturally present in half of untreated Ethiopian sequences. The insertion will probably restore the ALIX mediated virus release pathway, which is lacking in HIV-1C. The clinical importance of such insertion need to be evaluated in HIV-1C dominating regions were PI-drugs are being scaled up as second line treatment options.

<sup>#</sup>Address correspondence and reprints to Ujjwal Neogi, M.Sc., PhD, Division of Clinical Microbiology, Department of Laboratory Medicine, Karolinska Institutet, Stockholm, Sweden, ujjwal.neogi@ki.se, Ph. +46-858587935, Fax- +46-858587933. Declaration of interests

The author(s) declare that they have no competing interests

#### **Keywords**

Gag-p6; ALIX; HIV-1 subtype C; Protease Inhibitor failure

Subtype specific differences has been observed in Gag-p6 the motifs PTAPP and LYPxnLxxL. Among the subtypes, HIV-1 subtype C (HIV-1C) has a higher frequency of duplications in the PTAPP-motif after ART-failure [1, 2]. Also, a natural deletion of L483Y484 residues has been observed in the LYPxnLxxL motif in >95% of the sequences which abrogates the ALIX-mediated particle release in absence of PTAP/TSG101 pathway[3]. Considerable evidence also suggests that ART-induced changes in the Gag-p6 region may modulate the therapy response and the viral fitness [4]. Study have shown that selective drug pressure leads to accumulations of substitutions and insertions in Gag-p6, at sites distal from the mutations that render the virus highly resistant to PIs [5]. One of the key examples is PTAPP-duplication in TSG101binding motif in the Gag-p6 which affect the virological response to PI-drugs like amprenavir [6].

In this study we investigated the consequences of the sequence variations of Gag-p6, using HIV-1C sequences of clinical isolates from India (HIV-1C<sub>IN</sub>; n=158), Ethiopia (HIV-1C<sub>ET</sub>;  $n=73$ ) and Germany (HIV-1 $C<sub>DE</sub>$ ; n=125) and its potential clinical impact. The patients are from several clinical cohorts from India [7, 8], Germany [9, 10] and Ethiopia [11]. Among the patients 61% (215/356) were therapy-naïve. Data were pooled with Gag-p6 sequences (n=8589) of major non-C subtypes and recombinants from the Los Alamos HIV Database.

Gag-p6, protease and partial reverse transcriptase were amplified and sequenced from the plasma viral RNA as described previously [12-15]. Primary and acquired drug resistance mutations (DRMs) were evaluated using World Health Organisation mutations list 2009 [16] and International AIDS Society list 2013[17] respectively. HIV-1 subtyping was performed as described recently using three automated bioinformatics tools [18]. Multiple-template homology models of the p6-ALIX complex were built in MODELLER 9v12 [19] using crystal structure of human ALIX/AIP1 in complex with a peptide fragment of the SIVmac239 and HIV-1 Gag-p6 proteins (PDB codes 2XS1 and 2R02) [20, 21]. The models were analysed for accuracy using the DOPE statistical potential score [22]. The efficacy of the binding was analysed by computing the electrostatics using the Adapted Poisson-Boltzmann Solver software [23] and visualized using Chimera[24]. Statistical analysis was carried out using SPSSv22.0 (IBM Corp, US). The study was approved by respective ethical review committee in India, Germany, Sweden, and Ethiopia. Written informed consent was obtained from the participants.

Cohort characteristics were presented in supplementary digital content (SDC) 1. The multiple sequence alignments of the Gag-p6 of the representative strains were shown in the SDC 2. Duplications of three to thirteen amino acids in the TSG101 binding site were observed more frequently in the HIV-1 $C_{DE}$  (29%) and HIV-1 $C_{ET}$  (25%) sequences than in the HIV-1C<sub>IN</sub> (12%) sequences from therapy-naïve individuals (SDC 2 and Figure 1A). When therapy-naïve and therapy-failure patients were compared, the duplications occurred more frequently in the therapy-failure Indians (12% vs  $25\%$ ; p<0.05) but not in the German cohort (29% vs 33%; p=0.69) (Figure 1A). Interestingly, there are subtype specific

*AIDS*. Author manuscript; available in PMC 2015 September 24.

Neogi et al. Page 3

differences in the accumulation of the PTAPP-duplication after therapy-failure. The duplication occurred in greater frequency in HIV-1C (54%) compared to HIV-1B (9.3%) and HIV-1F1 (17.6%)[2]. Previous controversial findings were shown that duplications of PTAPP were associated with ART in one group of populations but not in others [2, 6, 25-28]. In our study we observed intra-HIV-1C specific preferential duplication in therapyfailure patients compared to therapy-naïve individuals.

A novel tetra-peptide insertion [PYxE; where x represents either arginine (R), lysine (K) or glutamine (Q)] was observed in the C-terminal position of the Gag-p6 in the defective HIV-1C ALIX-binding domain. This PYxE insertion was observed in 52% of the untreated Ethiopian sequences, but significantly less often in the untreated German sequences (16%;  $p<0.001$ ), and even more seldom in the untreated Indian sequences (3%;  $p<0.001$ ) (Figure 1B). When analysing sequences of therapy-naïve individuals obtained from the Los Alamos HIV Database, the frequency of the PYxE insertion was much less common in therapy-naïve patients infected with non-C subtypes (0.1%; n=4263), but also less common in HIV-1C sequences from southern Africa (1%, n=2295) and eastern African (3%, n=61).

The PYxE insertion restores the key Y484 residue (SDC 2 and Figure 1C). The 3Dmolecular models of the Gag-p6-ALIX complex showed a specific interaction involving the inserted Y484 residue of Gag-p6 and the ALIX (Figure 1D). Thus the insertion variant would restore the binding of Gag-p6 to ALIX that was lost due to the L483Y484 deletion in HIV-1C and probably restores the ALIX mediated virus release pathway.

Among the HIV-1 $C_{IN}$  sequences from therapy failed patients, the PYxE insertion was found significantly more often in PI-failure patients (6/10) as compared to those failing a non-PI containing regimen  $(5/62)$  (Odds ratio; 95% CI: 17.1; 3.6 – 81.4; p<0.001). Among the other clinical and demographic parameters, the median CD4+ T-cell count was significantly lower among the individuals with the PYxE insertion in the ALIX-motif compared to those without the insertion (73 vs 160 cells/mm<sup>3</sup>; p<0.001). That Indian patients with the PYxE insertion had significantly lower CD4<sup>+</sup> T-cell counts than those without, could possibly indicate that the virus with the insertion is more pathogenic. This suggestion was further supported by the very low CD4+ T-cell counts in the three therapy-naïve Indian individuals with the insertion before initiating ART (22, 90 and 56 cells/mm<sup>3</sup>). We further followed those three patients for two years after initiation of therapy. None of the patients gain optimal CD4+ T-cell count at two years  $(<$ 350 cells/mm<sup>3</sup>) (data not shown).

In conclusion, for the first time we have identified a PYxE tetra-peptide insertion in the ALIX-binding motif, which appeared in PI-therapy-failure cases in  $HIV-1C<sub>IN</sub>$  sequences, but occurred naturally in more than half of the therapy-naïve  $HIV-1C<sub>ET</sub>$  sequences examined. We therefore hypothesised that the genetic background might have influenced the preferential selection of these insertions. The insertion probably restores the ALIX-mediated virus release pathway, which is lacking in HIV-1C and the virus with this insertion might be more pathogenic. To better elucidate the clinical importance of such insertion, in countries with a high prevalence of HIV-1C, further investigations are needed when the PI-drugs are used in the ART regimen.

# **Supplementary Material**

Refer to Web version on PubMed Central for supplementary material.

## **Acknowledgments**

The study was partially funded by European Union FP7, Swedish International Developing Agency, CHAIN FP7 EU, the Swedish Civil Contingencies Agency [SWE-2009-151], and the Swedish Research Council [521-2012-3476 and 2007-7092]. MLE acknowledge the funds received from R01MH067513 from the National Institute of Mental Health (NIMH) (Bethesda, MD, USA) for the study Examining ART Adherence Issues in Bangalore India. V.R.P. and V.R.R. wish to acknowledge support from the National Institute of Health, USA (NIH) grant 1R01MH083579 (to V.R.P.).

## **References**

- 1. Marlowe N, Flys T, Hackett J Jr. Schumaker M, Jackson JB, Eshleman SH. Analysis of insertions and deletions in the gag p6 region of diverse HIV type 1 strains. AIDS Res Hum Retroviruses. 2004; 20:1119–1125. [PubMed: 15585104]
- 2. Martins AN, Arruda MB, Pires AF, Tanuri A, Brindeiro RM. Accumulation of P(T/S)AP late domain duplications in HIV type 1 subtypes B, C, and F derived from individuals failing ARV therapy and ARV drug-naive patients. AIDS Res Hum Retroviruses. 2011; 27:687–692. [PubMed: 21083435]
- 3. Patil A, Bhattacharya J. Natural deletion of L35Y36 in p6 gag eliminate LYPXnL/ALIX auxiliary virus release pathway in HIV-1 subtype C. Virus Res. 2012; 170:154–158. [PubMed: 22981647]
- 4. Dam E, Quercia R, Glass B, Descamps D, Launay O, Duval X, et al. Gag mutations strongly contribute to HIV-1 resistance to protease inhibitors in highly drug-experienced patients besides compensating for fitness loss. PLoS Pathog. 2009; 5:e1000345. [PubMed: 19300491]
- 5. Fun A, Wensing AM, Verheyen J, Nijhuis M. Human Immunodeficiency Virus Gag and protease: partners in resistance. Retrovirology. 2012; 9:63. [PubMed: 22867298]
- 6. Lastere S, Dalban C, Collin G, Descamps D, Girard PM, Clavel F, et al. Impact of insertions in the HIV-1 p6 PTAPP region on the virological response to amprenavir. Antivir Ther. 2004; 9:221–227. [PubMed: 15134184]
- 7. Shet A, Antony J, Arumugam K, Kumar Dodderi S, Rodrigues R, DeCosta A. Influence of adverse drug reactions on treatment success: prospective cohort analysis of HIV-infected individuals initiating first-line antiretroviral therapy in India. PLoS One. 2014; 9:e91028. [PubMed: 24614165]
- 8. Ekstrand ML, Chandy S, Heylen E, Steward W, Singh G. Developing useful highly active antiretroviral therapy adherence measures for India: the Prerana study. J Acquir Immune Defic Syndr. 2010; 53:415–416. [PubMed: 20190588]
- 9. Balduin M, Oette M, Daumer MP, Hoffmann D, Pfister HJ, Kaiser R. Prevalence of minor variants of HIV strains at reverse transcriptase position 103 in therapy-naive patients and their impact on the virological failure. J Clin Virol. 2009; 45:34–38. [PubMed: 19375978]
- 10. Oette M, Reuter S, Kaiser R, Lengauer T, Fatkenheuer G, Knechten H, et al. Epidemiology of transmitted drug resistance in chronically HIV-infected patients in Germany: the RESINA study 2001-2009. Intervirology. 2012; 55:154–159. [PubMed: 22286886]
- 11. Abdurahman S, Barqasho B, Nowak P, Cuong do D, Amogne W, Larsson M, et al. Pattern of microbial translocation in patients living with HIV-1 from Vietnam, Ethiopia and Sweden. J Int AIDS Soc. 2014; 17:18841. [PubMed: 24461466]
- 12. Bontell I, Cuong do D, Agneskog E, Diwan V, Larsson M, Sonnerborg A. Transmitted drug resistance and phylogenetic analysis of HIV CRF01\_AE in Northern Vietnam. Infect Genet Evol. 2012; 12:448–452. [PubMed: 21620998]
- 13. Neogi U, Prarthana BS, Gupta S, D'Souza G, De Costa A, Kuttiatt VS, et al. Naturally occurring polymorphisms and primary drug resistance profile among antiretroviral-naive individuals in Bangalore, India. AIDS Res Hum Retroviruses. 2010; 26:1097–1101. [PubMed: 20836706]

Neogi et al. Page 5

- 14. Neogi U, Sahoo PN, Kumar R, De Costa A, Shet A. Characterization of HIV type 1 subtype C protease gene: selection of L63P mutation in protease inhibitor-naive Indian patients. AIDS Res Hum Retroviruses. 2011; 27:1249–1253. [PubMed: 21453185]
- 15. Verheyen J, Litau E, Sing T, Daumer M, Balduin M, Oette M, et al. Compensatory mutations at the HIV cleavage sites p7/p1 and p1/p6-gag in therapy-naive and therapy-experienced patients. Antivir Ther. 2006; 11:879–887. [PubMed: 17302250]
- 16. Bennett DE, Camacho RJ, Otelea D, Kuritzkes DR, Fleury H, Kiuchi M, et al. Drug resistance mutations for surveillance of transmitted HIV-1 drug-resistance: 2009 update. PLoS One 2009. 4:e4724.
- 17. Johnson VA, Calvez V, Gunthard HF, Paredes R, Pillay D, Shafer RW, et al. Update of the drug resistance mutations in HIV-1: March 2013. Top Antivir Med 2013. 21:6–14.
- 18. Neogi U, Haggblom A, Santacatterina M, Bratt G, Gisslen M, Albert J, et al. Temporal Trends in the Swedish HIV-1 Epidemic: Increase in Non-B Subtypes and Recombinant Forms over Three Decades. PLoS One. 2014; 9:e99390. [PubMed: 24922326]
- 19. Sali A, Blundell TL. Comparative protein modelling by satisfaction of spatial restraints. J Mol Biol. 1993; 234:779–815. [PubMed: 8254673]
- 20. Zhai Q, Landesman MB, Robinson H, Sundquist WI, Hill CP. Identification and structural characterization of the ALIX-binding late domains of simian immunodeficiency virus SIVmac239 and SIVagmTan-1. J Virol. 2011; 85:632–637. [PubMed: 20962096]
- 21. Zhai Q, Fisher RD, Chung HY, Myszka DG, Sundquist WI, Hill CP. Structural and functional studies of ALIX interactions with YPX(n)L late domains of HIV-1 and EIAV. Nat Struct Mol Biol. 2008; 15:43–49. [PubMed: 18066081]
- 22. Shen MY, Sali A. Statistical potential for assessment and prediction of protein structures. Protein Sci. 2006; 15:2507–2524. [PubMed: 17075131]
- 23. Baker NA, Sept D, Joseph S, Holst MJ, McCammon JA. Electrostatics of nanosystems: application to microtubules and the ribosome. Proc Natl Acad Sci U S A. 2001; 98:10037–10041. [PubMed: 11517324]
- 24. Pettersen EF, Goddard TD, Huang CC, Couch GS, Greenblatt DM, Meng EC, et al. UCSF Chimera--a visualization system for exploratory research and analysis. J Comput Chem. 2004; 25:1605–1612. [PubMed: 15264254]
- 25. Brumme ZL, Chan KJ, Dong WW, Wynhoven B, Mo T, Hogg RS, et al. Prevalence and clinical implications of insertions in the HIV-1 p6Gag N-terminal region in drug-naive individuals initiating antiretroviral therapy. Antivir Ther. 2003; 8:91–96. [PubMed: 12741620]
- 26. Gallego O, de Mendoza C, Corral A, Soriano V. Changes in the human immunodeficiency virus p7-p1-p6 gag gene in drug-naive and pretreated patients. J Clin Microbiol. 2003; 41:1245–1247. [PubMed: 12624058]
- 27. Peters S, Munoz M, Yerly S, Sanchez-Merino V, Lopez-Galindez C, Perrin L, et al. Resistance to nucleoside analog reverse transcriptase inhibitors mediated by human immunodeficiency virus type 1 p6 protein. J Virol. 2001; 75:9644–9653. [PubMed: 11559796]
- 28. Ibe S, Shibata N, Utsumi M, Kaneda T. Selection of human immunodeficiency virus type 1 variants with an insertion mutation in the p6(gag) and p6(pol) genes under highly active antiretroviral therapy. Microbiol Immunol. 2003; 47:71–79. [PubMed: 12636256]

Neogi et al. Page 6



#### **Figure 1.**

Prevalence of **(A)** duplications in the TSG101 binding PTAPP motif and **(B)** insertions in the ALIX-binding LYPxnLxxL motif. Statistically significant differences are marked. **(C)** Different types of ALIX-binding sites in lentiviral Gag-p6 region as describe by Zhai *et al* 2011 [20]. Representative clinical isolates from the cohort are presented which had PYxE insertion. All the strains had the key residues conserved (highlighted). The 3D-molecular models of the Gag-p6-ALIX complex **(D)** The ALIX is shown in surface representation, interacting with the Gag-p6 protein PYKE shown in ribbon and stick (side-chain only) representation. The residues that belong to the ALIX binding motif and the insertions are all labelled in black. The residues in ALIX that is crucial to mediate the interaction with Gagp6 are labelled in magenta. The surface of the ALIX is coloured according to the electrostatic potential with blue indicating positively charged regions and red indicating negatively charged regions. The intensity of the colour reflects charge intensity. The models also establish that the Y484 residue in the insertions play a role similar to the Tyr residue in the wild type HIV-1B/SIV Gag-p6 protein by forming a specific hydrogen bond with the ALIX. Similar results were observed with PYRE and PYQE insertions (data not shown).