www.neuropsychopharmacology.org

# Estimating Endogenous Dopamine Levels at $D_2$ and $D_3$ Receptors in Humans using the Agonist Radiotracer [<sup>11</sup>C]-(+)-PHNO

Fernando Caravaggio<sup>1,2</sup>, Shinichiro Nakajima<sup>1,3</sup>, Carol Borlido<sup>1</sup>, Gary Remington<sup>1,2,3</sup>, Philip Gerretsen<sup>1,2,3</sup>, Alan Wilson<sup>1,2,3</sup>, Sylvain Houle<sup>1,3</sup>, Mahesh Menon<sup>1,3</sup>, David Mamo<sup>1,3</sup> and Ariel Graff-Guerrero<sup>\*,1,2,3</sup>

<sup>1</sup>Research Imaging Centre, Centre for Addiction and Mental Health, Toronto, ON, Canada; <sup>2</sup>Institute of Medical Science, University of Toronto, Toronto, ON, Canada; <sup>3</sup>Department of Psychiatry, University of Toronto, Toronto, ON, Canada

Using positron emission tomography (PET) and an acute dopamine depletion challenge it is possible to estimate endogenous dopamine levels occupying dopamine  $D_{2/3}$  receptors ( $D_{2/3}R$ ) in humans *in vivo*. Our group has developed [<sup>11</sup>C]-(+)-PHNO, the first agonist radiotracer with preferential *in vivo* affinity for  $D_3R$ . Thus, the use of [<sup>11</sup>C]-(+)-PHNO offers the novel possibility of (i) estimating *in vivo* endogenous dopamine levels at  $D_{2/3}R$  using an agonist radiotracer, and (ii) estimating endogenous dopamine levels at  $D_3R$  in extrastriatal regions such as the substantia nigra, hypothalamus, and ventral pallidum. Ten healthy participants underwent a [<sup>11</sup>C]-(+)-PHNO PET scan under baseline conditions and another under acute endogenous dopamine depletion achieved via oral administration of alphamethyl-para-tyrosine (64 mg/kg). [<sup>11</sup>C]-(+)-PHNO binding was sensitive to acute dopamine depletion, allowing *in vivo* estimates of endogenous dopamine in  $D_2R$ -rich regions (caudate and putamen), mixed  $D_{2/3}R$ -rich regions (ventral striatum and globus pallidus), and extrastriatal  $D_3R$ -rich regions (hypothalamus and ventral pallidum). Dopamine depletion decreased self-reported vigor, which was correlated with the reduction in dopamine levels in the globus pallidus. [<sup>11</sup>C]-(+)-PHNO is a suitable radiotracer for use in estimating endogenous dopamine levels at  $D_2R$  and  $D_3R$  in neuropsychiatric populations.

Neuropsychopharmacology (2014) 39, 2769-2776; doi:10.1038/npp.2014.125; published online 25 June 2014

#### INTRODUCTION

The dopamine system has been a key molecular target in understanding the etiology and treatment of numerous neuropsychiatric disorders. Not surprisingly, it has been the most heavily investigated neurotransmitter system in the living human brain using the molecular imaging technique, positron emission tomography (PET) (Banerjee and Prante, 2012). Using radiolabelled dopamine  $D_{2/3}$  receptor  $(D_{2/3}R)$  antagonists, such as [<sup>11</sup>C]-raclopride, [<sup>18</sup>F]-fallypride, and [<sup>11</sup>C]-FLB 457, it has been possible to quantify the availability of  $D_{2/3}R$  in vivo in the brains of healthy persons and persons with neuropsychiatric disease (Gjedde et al, 2005; Newberg et al, 2011; Tatsch and Poepperl, 2012). Studies in vitro demonstrate that  $D_{2/3}R$  exists in multiple affinity states for its endogenous ligand dopamine (Cumming, 2011; Seeman, 2013; van Wieringen et al, 2013). These states seem to affect the binding of agonist, but not antagonist, radioligands in vitro (Cumming, 2011; Seeman, 2013; van Wieringen *et al*, 2013). Therefore, the use of agonist radiotracers for  $D_{2/3}R$  may reveal a more physiologically relevant quantification of  $D_{2/3}R$  availability in the human brain, being sensitive to changes in receptor affinity.

The PET radiotracer  $[^{11}C]$ -(+)-PHNO is the first agonist radiotracer for D<sub>2/3</sub>R, which has preferential affinity for the D<sub>3</sub> receptors (Graff-Guerrero et al, 2010; Graff-Guerrero et al, 2008; Narendran et al, 2006; Rabiner et al, 2009; Wilson *et al*, 2005). This unique property of  $[^{11}C]$ -(+)-PHNO, ~20-40-fold selectivity of  $D_3R$  over  $D_2R$  (Freedman et al, 1994; Gallezot et al, 2012; Rabiner et al, 2009; Searle et al, 2010; Seeman et al, 1993), results in a differential contribution of  $D_2R$  and  $D_3R$  to the [<sup>11</sup>C]-(+)-PHNO signal across different regions of interest (ROIs). The estimated percent of the  $[^{11}C]$ -(+)-PHNO signal *in vivo* in humans attributed to D<sub>3</sub>R across ROIs are: the substantia nigra (SN)  $(\sim 100\%)$ , hypothalamus  $(\sim 100\%)$ , ventral pallidum (VP)  $(\sim 75\%)$ , globus pallidus (GP)  $(\sim 65\%)$ , ventral striatum  $(\sim 26\%)$ , and dorsal caudate-putamen (negligible) (Graff-Guerrero et al, 2010; Searle et al, 2013; Tziortzi et al, 2011).

Like the antagonist radiotracer [<sup>11</sup>C]-raclopride, endogenous dopamine competes with [<sup>11</sup>C]-(+)-PHNO for binding to  $D_{2/3}R$  at baseline (Ginovart *et al*, 2006; Shotbolt *et al*, 2012; Willeit *et al*, 2008). The amount of endogenous dopamine occupying  $D_{2/3}R$  at baseline can be estimated with such radioligands by comparing the percent

<sup>\*</sup>Correspondence: Dr A Graff-Guerrero, Research Imaging Centre, Centre for Addiction and Mental Health, 250 College Street, Toronto, ON, Canada M5T IR8. Tel: +416 535 8501×4834, Fax: +1 416 979 3855, E-mail: ariel.graff@camh.ca

Received 14 January 2014; revised 15 April 2014; accepted 5 May 2014; accepted article preview online 30 May 2014

change in binding potential  $(BP_{ND})$  between a baseline scan and a scan under acute dopamine depletion (Cumming et al, 2002; Laruelle et al, 1997; Verhoeff et al, 2001). Acute dopamine depletion is achieved in humans using alphamethyl-para-tyrosine (AMPT), a competitive inhibitor of tyrosine hydroxylase, which is the rate-limiting enzyme of catecholamine synthesis. Using this paradigm, altered levels of striatal endogenous dopamine occupying D<sub>2/3</sub>R at baseline in neuropsychiatric disorders has been demonstrated (Abi-Dargham et al, 2000; Abi-Dargham et al, 2009; Bloemen et al, 2013; Kegeles et al, 2010; Martinez et al, 2009).

To date, endogenous dopamine levels have not been estimated in humans using an agonist radiotracer for  $D_{2/3}R$ , as opposed to an antagonist. The use of an agonist radiotracer, which should more closely mimic the binding of the endogenous ligand, may offer a more sensitive and functionally significant estimate of endogenous dopamine in humans. Moreover, endogenous dopamine levels have not been estimated in D<sub>3</sub>R-rich regions in humans such as the SN, GP, VP, and hypothalamus. The current investigation sought to validate the use of  $[^{11}C]-(+)$ -PHNO to estimate endogenous dopamine levels at D<sub>2</sub>R and D<sub>3</sub>R in healthy humans.

## MATERIALS AND METHODS

## Subjects

Healthy participants (24) were recruited for the study. Six participants failed screening. Two participants withdrew before the first baseline PET scan. One participant withdrew during the baseline PET scan owing to nausea induced by the  $[{}^{1Y}C]$ -(+)-PHNO injection. One participant was unable to complete the study owing to  $[{}^{11}C]$ -(+)-PHNO tracer problems before the post-AMPT PET scan. One participant withdrew before the post-AMPT PET scan owing to akathisia, and three participants withdrew during the post-AMPT scan owing to feelings of claustrophobia/ anxiety. Ten participants (4 female; mean age =  $29.1 \pm 8.4$ ) in total completed both PET scans, under the baseline and dopamine-depleted condition. All participants were free of any major medical or psychiatric disorder as determined by clinical interview, the Mini-International Neuropsychiatric Interview, basic laboratory tests, and electrocardiogram. Participants were required to have a negative urine screen for drugs of abuse and/or pregnancy at inclusion and before each PET scan. All participants were non-smokers. The study was approved by the Research Ethics Board of the Centre for Addiction and Mental Health (CAMH), Toronto, and no objection letter was issued by Health Canada. All the participants provided written and informed consent.

# Metyrosine/AMPT Administration

Dopamine depletion was induced by oral administration of 64 mg of metyrosineper kilogram of body weight over 25 h. Independent of weight, no participant was dosed above 4500 mg. Metyrosine was administered in six equal doses at the following times: 0900, 1230 hours (post 3.5 h), 1700 hours (post 8 h), and 2100 hours (post 12 h) on day 1, and 0600 hours (post 21 h) and 1000 hours (post 25 h) on day 2. The post-AMPT PET scan was scheduled at 1200 hours, 28 h after the initial metyrosine dose. For two participants, the 1200-hour scan was unavailable, therefore the times for the doses were modified to reflect the 28-h post-AMPT PET scan. The subjects were under direct observation during AMPT administration and slept overnight on an inpatient unit at CAMH to ensure the AMPT dosing schedule and monitor for potential side effects. In addition, subjects were instructed to drink at least 41 of fluids during the 2-day admission to prevent the formation of AMPT crystals in urine. Fluid intake was carefully monitored during the study to ensure compliance. Urine samples were collected at 1400 hours on day 1 and at 0900 hours on day 2 to monitor AMPT crystals in urine. In addition, in order to alkalinize the urine, which increases AMPT solubility, sodium bicarbonate (1.25 g) was given orally at 2200 hours on the evening before day 1 and at 0700 hours on day 1 of administration.

## **Plasma Samples**

Plasma levels of prolactin, homovanillic acid (HVA), and 3-methoxy-4-hydroxyphenethyleneglycol (MHPG) were collected at 0900 hours (day 1, before AMPT administration), 1400 hours (day 1) and 1200 hours (day 2). Plasma levels of AMPT were also collected at 1400 hours (day 1) and 1200 hours (day 2). Plasma prolactin, HVA, MHPG, and AMPT were quantified as previously described (Verhoeff et al, 2002). The times were modified for the two participants with the alternative post-PET scan times.

## **Rating Scales**

Subjects were evaluated by the research psychiatrists (SN, GR, PG, and AG-G) for potential side effects at the following times: 0900 hours (baseline), 1400 hours (post 5h dose), 1200 hours (post 27 h dose), and at 1500 hours (post 30 h dose) The times were modified for the two participants with the alternative post-PET scan times. The presence of adverse effects such as parkinsonian symptoms, acute dystonias, and involuntary movements was monitored using the Simpson Angus Scale (SAS), Barnes Akathisia Scale (BAS), and the Abnormal Involuntary Movement Rating Scale (AIMS). To evaluate changes in energy, mood, and subjective well-being induced by dopamine depletion, the Profile of Mood States (POMS) and the Subjective Well-Being Under Neuroleptic Treatment (SWN) scales were administered.

# **PET Imaging**

Participants underwent two  $[^{11}C]$ -(+)-PHNO PET scans, one under baseline conditions and another at 25 h of starting AMPT-induced dopamine depletion. The radiosynthesis of  $[^{11}C]$ -(+)-PHNO and the acquisition of PET images has been described in detail elsewhere (Graff-Guerrero et al, 2010; Wilson et al, 2000; Wilson et al, 2005). Briefly, images were acquired using a high-resolution head-dedicated PET camera system (CPS-HRRT; Siemens Molecular Imaging, USA), which measures radioactivity in 207 brain slices with a thickness of 1.2 mm each. The inplane resolution was  $\sim$  2.8 mm full-width at half-maximum. Transmission scans were acquired using a  $^{137}$ Cs (T<sub>1/2</sub> = 30.2 years, E = 662 KeV) single-photon point source to provide



**Figure I**  $[^{11}C]$ -(+)-PHNO BP<sub>ND</sub> in each ROI before and after AMPT-induced dopamine depletion. (a) ROIs for which  $[^{11}C]$ -(+)-PHNO BP<sub>ND</sub> before and after dopamine depletion was reliably estimated for all subjects (n=10). (b) ROIs for which  $[^{11}C]$ -(+)-PHNO BP<sub>ND</sub> before and after dopamine depletion was not reliably estimated for all subjects.

attenuation correction, and the emission data were acquired in list mode. The raw data were reconstructed by filteredback projection. For the baseline  $[^{11}C]$ -(+)-PHNO scans, the mean radioactivity dose was  $9.2(\pm 1.2)$  mCi, with a specific activity of  $1113.7(\pm 328.2)$  mCi/µmol, and an injected mass of  $2.1(\pm 0.4)$  µg. For the dopamine-depleted scans, the mean radioactivity dose was  $8.7(\pm 1.5)$  mCi, with a specific activity of  $1024.1(\pm 299.5)$  mCi/µmol, and an injected mass of  $2.1(\pm 0.3) \mu g$ . There was no difference in mean radioactivity dose (t(9) = 0.92, p = 0.38), specific activity (t(9) = 0.96, p = 0.37), or mass injected (t(9)= -0.32, p = 0.75) between the baseline and dopamine depletion scans. [<sup>11</sup>C]-(+)-PHNO scanning data was acquired for 90 min post injection. Once scanning was complete, the data was re-defined into 30 frames (1-15 of 1 min duration and 16–30 of 5 min duration).

#### **Image Analysis**

The ROI-based analysis for  $[^{11}C]-(+)$ -PHNO has been described in detail elsewhere (Graff-Guerrero et al, 2008; Tziortzi et al, 2011). Briefly, time activity curves (TACs) from ROIs were obtained from the dynamic PET images in native space with reference to each subjects co-registered MRI image. The co-registration of each subject's MRI to PET space was done using the normalized mutual information algorithm (Studholme et al, 1997) as implemented in SPM2 (SPM2, Wellcome Department of Cognitive Neurology, London; http://www.fil.ion.ucl.ac.uk/spm). The TACs were analyzed using the Simplified Reference Tissue Method (SRTM) (Lammertsma and Hume, 1996), using the cerebellum as the reference region, to derive a quantitative estimate of binding: the binding potential relative to the non-displaceable compartment (BP<sub>ND</sub>) as defined by the consensus nomenclature for in vivo imaging of reversibly binding radioligands (Innis et al, 2007). The basis function implementation of the SRTM (Gunn et al, 1997) was applied to the dynamic PET images to generate parametric voxelwise BP<sub>ND</sub> maps using PMOD (v2.7, PMOD Technologies, Zurich, Switzerland). These images were spatially normalized into MNI brain space by nearest neighbor interpolation with a voxel size fixed in  $2 \times 2 \times 2 \text{ mm}^3$  using SPM2. Regional  $BP_{ND}$  estimates were then derived from

ROIs defined in MNI space. The ventral striatum and dorsal striatum (dorsal caudate, hereafter caudate, and dorsal putamen, hereafter putamen) were defined according with Mawlawi *et al* (2001). The GP, VP, and hypothalamus ROIs were defined according to the criteria of Tziortzi *et al* (2011).

### **Estimating Endogenous Dopamine Levels**

Our estimate of endogenous dopamine levels at D<sub>2/3</sub>R is based on the occupancy model, in which endogenous dopamine competes with the binding of radiotracers such as  $[^{11}C]$ -(+)-PHNO for  $D_{2/3}R$  at baseline (Laruelle, 2000; Laruelle et al, 1997; Verhoeff et al, 2001). It is assumed by this model that, (i) baseline  $D_{2/3}R$  BP<sub>ND</sub> is confounded by endogenous dopamine, such that the higher the concentration of dopamine the lower the value of  $D_{2/3}R$  BP<sub>ND</sub> will be obtained, (ii) D<sub>2/3</sub>R BP<sub>ND</sub> under depletion more accurately reflects the true status of  $D_{2/3}R$ , and (iii) the fractional increase in  $D_{2/3}R$  BP<sub>ND</sub> after dopamine depletion (ie, 100 × (depletion  $BP_{ND}$  – baseline  $BP_{ND}$ )/baseline  $BP_{ND} = \%$  $\Delta BP_{ND}$ ) is linearly proportional to the baseline dopamine concentration at  $D_{2/3}R$ , provided the process of dopamine depletion does not change the number and affinity of the  $D_{2/3}R$ . Thus, the % $\Delta BP_{ND}$ , under appropriate assumptions, is considered a semiquantitative index of endogenous dopamine levels at  $D_{2/3}R$  (Verhoeff *et al*, 2001).

#### Statistical Analysis

Statistical analyses were conducted using SPSS (v.12.0; SPSS, Chicago, Illinois) and GraphPad (v.5.0; GraphPad Software, La Jolla, California). Normality of variables was determined using the D'Agostino-Pearson test. The significance level for all tests was set at p < 0.05 (two-tailed).

## RESULTS

#### AMPT-Induced $\Delta BP_{ND}$

AMPT-dopamine depletion significantly increased [<sup>11</sup>C]-(+)-PHNO BP<sub>ND</sub> in the caudate (t(9) = 3.36, p = 0.008), putamen (t(9) = 5.84, p = 0.0002), ventral striatum



**Figure 2** Averaged  $[^{11}C]$ -(+)-PHNO voxelwise BP<sub>ND</sub> map of all subjects (n = 10) before and after AMPT-induced dopamine depletion.

(t(9) = 10.87, p = 0.0001), and GP (t(9) = 3.79, p = 0.004)(see Figure 1a).  $[^{11}C]$ -(+)-PHNO BP<sub>ND</sub> did not change in the SN after dopamine depletion (t(9) = 0.29, p = 0.78). The percent change in BP<sub>ND</sub> (% $\Delta$ BP<sub>ND</sub>) at D<sub>2/3</sub>R was ~36% in the ventral striatum,  $\sim 33\%$  in the putamen,  $\sim 33\%$  in the caudate, and  $\sim 11\%$  in the GP (see Figure 2; Supplementary Table 1). Owing to poor model fitting, [<sup>11</sup>C]-(+)-PHNO BP<sub>ND</sub> could not be reliably estimated in the hypothalamus for one subject and in the VP for four subjects (see Supplementary Table 2). AMPT-dopamine depletion significantly increased  $[^{11}C]$ -(+)-PHNO BP<sub>ND</sub> in the hypothalamus (t(8) = 4.96, p = 0.001), observing a % $\Delta BP_{ND}$  of 68.5%. In the VP there was a trend for an increase in  $[^{11}C]$ -(+)-PHNO BP<sub>ND</sub> after dopamine depletion (t(5) = 2.32, p = 0.06), observing a % $\Delta BP_{ND}$  of 64.8% (see Figure 1b).

## **Plasma Results**

The average plasma concentration of AMPT after 27 h of oral administration was  $24(\pm 11)\mu$ g/ml. On the basis of this average plasma concentration of AMPT, the average tyrosine hydroxylase inhibition in our sample can be estimated to be ~80% (Engelman *et al*, 1968; Laruelle *et al*, 1997; Udenfriend *et al*, 1965). AMPT-dopamine

Table I	Effect of Alpha-Methyl-Para-Tyrosine (AMPT) on '	Various
Plasma Le	evels $(n = 10)$	

Plasma levels	Baseline	AMPT 5 h	<b>AMPT</b> 27 h	%∆ Over 27 h	P-value				
Homovanillic Acid (HVA)									
nmol/l	81.8 (42.8)	37.0 (15.0)	32.4 (27.2)	- 60	0.009				
3-Methoxy-4-hydroxphenyglycol (MHPG)									
nmol/l	188.3 (99.2)	160.5 (89.2)	109.5 (60.3)	- 42	0.0003				
Alpha-methyl-prara-tyrosine (AMPT)									
µmol/l		63.1 (38.2)	123.0 (53.0)	+ 95	0.009				

Data are given as mean and SD in parenthesis.

depletion significantly decreased plasma levels of HVA (t(9) = 3.32, p = 0.009) and MHPG (t(9) = 5.72, p = 0.0003) compared with baseline (see Table 1). AMPT-dopamine depletion significantly increased plasma levels of prolactin (t(9) = 5.83, p = 0.0003). Baseline levels of HVA and MHPG did not correlate with baseline [ $^{11}C$ ]-(+)-PHNO BP<sub>ND</sub> in any ROI. Nor did changes in HVA and MHPG levels induced by AMPT correlate with changes in [ $^{11}C$ ]-(+)-PHNO BP<sub>ND</sub>. However, baseline levels of prolactin correlated with baseline [ $^{11}C$ ]-(+)-PHNO BP<sub>ND</sub> in the caudate (r = 0.71, p = 0.02). Changes in prolactin levels induced by AMPT correlated with changes in [ $^{11}C$ ]-(+)-PHNO BP<sub>ND</sub> in the caudate (r = 0.76, p = 0.01) and putamen (r = 0.82, p = 0.004).

#### SUV before and after AMPT

The averaged standard uptake values (SUVs) of  $[^{11}C]$ -(+)-PHNO in each ROI before and after AMPT-dopamine depletion are presented in Supplementary Figure 1.

#### Self-Reported Measures

*POMS.* AMPT-dopamine depletion significantly increased self-reported fatigue (t(9) = 3.50, p = 0.0068), decreased vigor (t(9) = 3.75, p = 0.0046), and increased tension (t(9) = 2.31, p = 0.046). However, changes in self-reported tension did not survive false discovery rate (FDR) correction (see Table 2). We investigated whether changes in self-reported fatigue and vigor were related to changes in [<sup>11</sup>C]-(+)-PHNO BP<sub>ND</sub> in our ROIs. Change in vigor scores were negatively correlated with change in [<sup>11</sup>C]-(+)-PHNO BP<sub>ND</sub> in the GP (r = -0.77, p = 0.009) (see Figure 3).

SWN. AMPT-dopamine depletion significantly decreased self-reported physical functioning (t(9) = 3.28, p = 0.009), negative sum scores (t(9) = 3.22, p = 0.01), and emotion regulation (t(9) = 2.26, p = 0.049). However, only changes in negative sum scores survived FDR correction (see Table 2). Change in negative sum scores were not correlated with change in [ $^{11}$ C]-(+)-PHNO BP<sub>ND</sub> in any ROI.

**Table 2** Effect of Alpha-Methyl-Para-Tyrosine (AMPT) on Self-Reported Measures (n = 10)

Scales	Baseline scores	AMPT 5 h scores	AMPT 27 h scores	%∆ Scores over 27 h	P-value	FDR P Threshold
Profile of mood states (POMS	S)					
Fatigue	9.6 (3.6)	10.4 (3.7)	5.4 (6.  )	+ 66.4	0.0068 <sup>a</sup>	0.008
Vigor	25.7 (9.2)	23.4 (5.9)	19.4 (7.1)	-21.8	0.0046ª	0.017
Tension	.  (2.3)	11.9 (3.3)	13 (4.5)	+ 15.5	0.0463	0.025
Depression	16.3 (3.1)	16.0 (3.1)	18.9 (7.9)	+ 13.4	0.1817	0.033
Confusion	11.9 (2.1)	11.7 (2.3)	3.3 (4.7)	+ 11.6	0.2606	0.042
Anger	12.5 (1.1)	3 (2.0)	13.8 (4.8)	+ 9.3	0.3464	0.050
Subjective well-being under ne	euroleptic treatment (SWI	N)				
Physical functioning	22.2 (1.5)	22 (1.6)	19.9 (2.8)	- 10.5	0.0094	0.007
Negative sum	57.7 (2.3)	56.9 (3.0)	54.2 (4.4)	- 6.10	0.0105 <sup>a</sup>	0.014
Emotion egulation	22.4 (1.3)	21.8 (1.8)	21 (2.5)	- 6.36	0.0498	0.021
Mental functioning	22 (1.6)	21.7 (2.1)	20.3 (3.2)	- 7.91	0.0523	0.029
Positive sum	52.6 (5.5)	51.5 (6.1)	48.3 (10.8)	- 8.88	0.0867	0.036
Self-control	22.3 (1.4)	22.2 (1.4)	20.9 (3.8)	- 6.58	0.1914	0.043
Social integration	21.4 (2)	20.7 (2.4)	20.4 (3.2)	- 4.71	0.2289	0.050

Abbreviation: FDR, false discovery rate.

Data are given as mean raw scores with SD in parenthesis.

<sup>a</sup>Denotes significance after FDR correction.



Figure 3 Correlation between percent change in self-reported vigor and  $[^{11}C]\text{-}(+)\text{-}PHNO\ BP_{ND}$  in the globus pallidus.

## **Presence of Adverse Events**

There was a trend for significant change in total scores on the SAS (t(9) = 2.25, p = 0.05), but no change in the sum of global clinical scores on the BAS (t(9) = 1.00, p = 0.34), as assessed from baseline to 30 h post AMPT-dopamine depletion. There was also no change in a total score of zero on the AIMS. However, like previous dopamine-depletion studies, it was clinically observed that subjects experienced strong AMPT side effects. This is indicated by our four participant withdrawals before the post-AMPT PET scan (one owing to severe akathisia and three owing to feelings of claustrophobia/anxiety). Notably, subject #4 and #5 presented with clinically notable akathisia and all subjects reported fatigue (for a list of all subjects and their  $BP_{ND}$  data, see Supplementary Table 2). Perhaps these observations were not captured by our scales due to the timing of when these scales were administered throughout the study. Urine testing revealed that subject #8 developed a significant level of crystalluria given AMPT-dopamine depletion, and was treated after the post-PET scan accordingly.

## DISCUSSION

The present investigation is the first to estimate endogenous dopamine levels at  $D_{2/3}R$  using an agonist radiotracer: [<sup>11</sup>C]-(+)-PHNO. Hypothetically, an agonist radiotracer for  $D_{2/3}R$ should be more sensitive to changes in endogenous dopamine levels than an antagonist radiotracer. Notably, the change in striatal  $BP_{ND}$  of  $[^{11}C]$ -(+)-PHNO after dopamine depletion reported here ( $\sim 30\%$ ) is much greater than has been previously reported in healthy controls using the antagonist radiotracer [<sup>11</sup>C]-raclopride ( $\sim 5.7-18.3\%$ ) (Kegeles *et al*, 2010; Martinez et al, 2009; Verhoeff et al, 2003; Verhoeff et al, 2002; Verhoeff *et al*, 2001). This may be due to  $[^{11}C]-(+)-$ PHNO being more sensitive than [<sup>11</sup>C]-raclopride to changes in endogenous dopamine levels. This is consistent with the finding that an amphetamine challenge in healthy controls displaced the BP<sub>ND</sub> of  $[^{11}C]$ -(+)-PHNO greater than  $[^{11}C]$ -raclopride (Shotbolt *et al*, 2012; Willeit *et al*, 2008). However, it is worth noting that although this increased sensitivity to endogenous dopamine has been demonstrated in vivo in humans, rodents (Kiss et al, 2011), and cats (Ginovart et al, 2006), this finding is not ubiquitous across studies (Galineau et al, 2006; McCormick et al, 2011). Furthermore, there are



1 Cai avaggio

differences in the amount of AMPT administered across studies. However, in our sample we achieved a similar average plasma concentration of AMPT, and likely tyrosine hydroxylase inhibition, as previous studies (Kegeles *et al*, 2010; Martinez *et al*, 2009; Verhoeff *et al*, 2003; Verhoeff *et al*, 2002; Verhoeff *et al*, 2001). Finally, we cannot rule out that differences in reported dopamine estimation may be explained by differences in PET camera resolution. Directly comparing the change in  $[^{11}C]$ -(+)-PHNO and  $[^{11}C]$ -raclopride BP<sub>ND</sub> after dopamine depletion in the same persons using a high-resolution PET camera is warranted.

The preferential affinity of  $[^{11}C]$ -(+)-PHNO for D<sub>3</sub>R over D<sub>2</sub>R affords the current investigation of the ability to estimate, for the first time *in vivo* in humans, endogenous dopamine levels at D<sub>3</sub>R in select extrastriatal regions. The SN, GP, hypothalamus, and VP constitute those ROIs for which the majority of the  $[^{11}C]$ -(+)-PHNO BP<sub>ND</sub> signal is due to D<sub>3</sub>R binding. In the SN, we did not observe a significant change in  $[^{11}C]$ -(+)-PHNO BP<sub>ND</sub> after dopamine depletion. Thus, our findings suggest that acute dopamine depletion with AMPT does not alter dopamine levels in the SN (see Supplementary Text for discussion).

The magnitude of  $\%\Delta BP_{ND}$  varied across ROIs. Differences in the concentrations of dopamine in these regions may explain some of the observed difference in the  $\%\Delta BP_{ND}$ . Investigations in rodent brains and post-mortem human brains generally support that the regional concentrations of dopamine are: VS > putamen > or = caudate > GP >hypothalamus > SN (Adolfsson et al, 1979; Versteeg et al, 1976). In our current investigation, the magnitude of  $\%\Delta BP_{ND}$ generally followed this difference in regional dopamine concentration: VS > putamen = caudate > GP > SN. It has been suggested that the dopamine concentration in the human GP is one-third of that in the striatum (Adolfsson *et al*, 1979). Notably, the magnitude of  $\&\Delta BP_{ND}$  between the GP and striatal ROIs differed by one-third. Finally, our observation of a  $\sim$  33–36%  $\Delta$ BP<sub>ND</sub> in the striatum ROIs is in accordance with the 34% change in specific binding seen with  $[^{11}C]$ -(+)-PHNO in rodent striata after dopamine depletion with AMPT and reserpine ex vivo (Wilson et al, 2005).

However, differences in regional concentrations in dopamine cannot explain the large  $\%\Delta BP_{ND}$  observed in the hypothalamus. We do not think this observation can be easily explained by the greater affinity of [<sup>11</sup>C]-(+)-PHNO for D<sub>3</sub>R over D<sub>2</sub>R, as the  $\%\Delta BP_{ND}$  varied across all the D<sub>3</sub>Rrich regions. Likewise, any potential non-tracer conditions at D<sub>3</sub>R cannot readily explain the large magnitude of  $\%\Delta BP_{ND}$  observed in the hypothalamus and VP, especially as such conditions should reduce changes in BP<sub>ND</sub> due to dopamine (Laruelle, 2000). Future studies are required to replicate these observations with larger sample sizes.

Despite following the ROI segmentation guidelines of Tziortzi *et al* (2011), we were unable to reliably estimate  $[^{11}C]$ -(+)-PHNO BP<sub>ND</sub> in the VP and hypothalamus in all our subjects. Despite this, we observed an increase in  $[^{11}C]$ -(+)-PHNO BP<sub>ND</sub> in both of these D<sub>3</sub>R-rich regions. Thus, although estimates of endogenous dopamine levels in D<sub>3</sub>R-rich regions may be achieved with  $[^{11}C]$ -(+)-PHNO, these estimates could not be achieved in the hypothalamus and VP for all subjects due to poor SRTM model fitting associated with noise in the TAC and no washout of the signal. With our experience with  $[^{11}C]$ -(+)-PHNO, our

group has noted the reliability of fitting in these regions is less than for the other ROIs. Thus, this may have contributed to the low statistical significance of the AMPT effect despite a high average change in BP<sub>ND</sub>. However, it is reassuring that our reported BP<sub>ND</sub> values for these regions are in accordance with the reports of other studies (Tziortzi *et al*, 2011). Moreover, we may have been underpowered to detect significant changes in these regions, given their reliability of fitting. Notably, no study has ever published test-retest reliability data for the hypothalamus and VP ROIs with [<sup>11</sup>C]-(+)-PHNO. Although this poses a limitation to our current investigation, it does not detract from the fact that after AMPT, large average increases in BP<sub>ND</sub> were observed in these ROIs. Future studies would benefit from reporting test-retest reliability of fitting for these ROIs.

Consistent with previous reports (Verhoeff et al, 2003), AMPT significantly decreased self-reported vigor and increased self-reported fatigue in healthy participants. Interestingly, the change in vigor scores under dopamine depletion was related to the change in  $[^{11}C]$ -(+)-PHNO  $BP_{ND}$  in the GP. Several case studies have reported that unilateral or bilateral lesions to the GP, in particular the internal segment, can result in profound apathy and lack of motivation (Adam et al, 2013; Singh et al, 2011; Vijayaraghavan et al, 2008). It has also been demonstrated that D<sub>3</sub>R are more abundant in the internal segment of the GP than D<sub>2</sub>R (Gurevich and Joyce, 1999). Thus, we speculate that the significant decrease in vigor and motivation seen in participants after dopamine depletion may be mediated by reduced dopaminergic signaling at D<sub>3</sub>R in the GP, as captured by the change in  $[^{11}C]$ -(+)-PHNO BP<sub>ND</sub>.

There are several limitations with the current investigation worth addressing. First, our study was only single-blind and not placebo controlled, a limitation shared by all previous dopamine PET studies using AMPT. Second, we have not collected arterial plasma data, quantifying BP<sub>ND</sub> using the SRTM rather than using full-kinetic modeling. The SRTM approach to estimate  $BP_{ND}$  with the cerebellum as reference has a good correlation with the BP as estimated with a full-kinetic analysis and it was validated in controls for  $[^{11}C]$ -(+)-PHNO (Ginovart *et al*, 2007). The full-kinetic analysis would allow direct estimation of the BP in the ROIs without the potential bias induced by specific binding in the reference region, if any. Simply, we cannot rule out (quantitatively) that AMPT did not exert an effect in the reference tissue, and poses as a current limitation to our study. Finally, it has been noted that the injected mass of  $[^{11}C]$ -(+)-PHNO was not within ideal radiotracer conditions (ie, <1.5 ng/kg) and consequently could lead to an underestimation of the occupancy by endogenous dopamine (see appendix in (Shotbolt et al, 2012)). Unfortunately the specific activity required to obtain tracer conditions is not possible with the available radiosynthesis method. We do not believe this unavoidable technical limitation substantially changes the conclusion of our study. As the radiotracer mass injected was similar in both PET scan sessions, we are underestimating the apparent occupancy of the competitor dopamine by a similar factor in both scans. Furthermore, the PET scans were performed >24h apart avoiding any carry-over effect.

In conclusion, the current investigation is the first to estimate endogenous dopamine levels at  $D_2R$  and  $D_3R$  in

healthy humans using the agonist radiotracer  $[^{11}C]$ -(+)-PHNO.  $[^{11}C]$ -(+)-PHNO is sensitive to acute dopamine depletion in humans, and future studies employing this radiotracer to estimate endogenous dopamine levels at D<sub>2</sub>R and D<sub>3</sub>R in neuropsychiatric populations are warranted.

### FUNDING AND DISCLOSURE

This study was funded by Canadian Institutes of Health Research (MOP-114989) and US National Institute of Health (RO1MH084886-01A2). Dr Nakajima reports having received grants from the Japan Society for the Promotion of Science and Inokashira Hospital Research Fund and speaker's honoraria from GlaxoSmith Kline, Janssen Pharmaceutical, Pfizer, and Yoshitomiyakuhin within the past 3 years. Dr Graff-Guerrerro currently receives research support from the following external funding agencies: Canadian Institutes of Health Research, the US National Institute of Health, and the Mexico Instituto de Ciencia y Tecnologia para la Capital del Conocimiento en el Distrito Federal (ICyTDF). He has also received professional services compensation from Abbott Laboratories, Gedeon-Richter Plc, and Lundbeck; grant support from Janssen; and speaker compensation from Eli Lilly. Dr Remington has received research support from the Canadian Diabetes Association, the Canadian Institutes of Health Research, Medicure, Neurocrine Biosciences, Novartis, Research Hospital Fund-Canada Foundation for Innovation, and the Schizophrenia Society of Ontario and has served as a speaker consultant for Novartis, or Laboratorios FarmacéuticosRovi, Synchroneuron, and Roche. Mr Caravaggio, Ms Carol Borlido, Dr Gerretsen, Dr Sylvain Houle, and Dr Wilson report no biomedical financial interests or potential conflicts of interest relevant to the current study.

#### ACKNOWLEDGEMENTS

We gratefully acknowledge Dr Isabelle Boileau for her insights and disclosing data. The authors thank the PET Centre staff at the Centre for Addiction and Mental Health, including Alvina Ng and Laura Nguyen, for technical assistance in data collection. We also thank Ms Wanna Mar, Ms Kathryn Kalahani-Bargis, and Ms Zhe Feng for their assistance in participant recruitment.

#### REFERENCES

- Abi-Dargham A, Rodenhiser J, Printz D, Zea-Ponce Y, Gil R, Kegeles LS *et al* (2000). Increased baseline occupancy of D2 receptors by dopamine in schizophrenia. *Proc Natl Acad Sci USA* 97: 8104–8109.
- Abi-Dargham A, van de Giessen E, Slifstein M, Kegeles LS, Laruelle M (2009). Baseline and amphetamine-stimulated dopamine activity are related in drug-naive schizophrenic subjects. *Biol Psychiatry* 65: 1091–1093.
- Adam R, Leff A, Sinha N, Turner C, Bays P, Draganski B et al (2013). Dopamine reverses reward insensitivity in apathy following globus pallidus lesions. *Cortex* **49**: 1292–1303.
- Adolfsson R, Gottfries CG, Roos BE, Winblad B (1979). Postmortem distribution of dopamine and homovanillic acid in

human brain, variations related to age, and a review of the literature. *J Neural Transm* **45**: 81–105.

- Banerjee A, Prante O (2012). Subtype-selective dopamine receptor radioligands for PET imaging: current status and recent developments. *Curr Med Chem* **19**: 3957–3966.
- Bloemen OJ, de Koning MB, Gleich T, Meijer J, de Haan L, Linszen DH *et al* (2013). Striatal dopamine D2/3 receptor binding following dopamine depletion in subjects at ultra high risk for psychosis. *Eur Neuropsychopharmacol* 23: 126–132.
- Cumming P (2011). Absolute abundances and affinity states of dopamine receptors in mammalian brain: a review. *Synapse* **65**: 892–909.
- Cumming P, Wong DF, Dannals RF, Gillings N, Hilton J, Scheffel U *et al* (2002). The competition between endogenous dopamine and radioligands for specific binding to dopamine receptors. *Ann N Y Acad Sci* **965**: 440–450.
- Engelman K, Jequier E, Udenfriend S, Sjoerdsma A (1968). Metabolism of alpha-methyltyrosine in man: relationship to its potency as an inhibitor of catecholamine biosynthesis. *J Clin Invest* **47**: 568–576.
- Freedman SB, Patel S, Marwood R, Emms F, Seabrook GR, Knowles MR *et al* (1994). Expression and pharmacological characterization of the human D3 dopamine receptor. *J Pharmacol Exp Ther* **268**: 417–426.
- Galineau L, Wilson AA, Garcia A, Houle S, Kapur S, Ginovart N (2006). In vivo characterization of the pharmacokinetics and pharmacological properties of [11C]-(+)-PHNO in rats using an intracerebral beta-sensitive system. *Synapse* **60**: 172–183.
- Gallezot JD, Beaver JD, Gunn RN, Nabulsi N, Weinzimmer D, Singhal T *et al* (2012). Affinity and selectivity of [(1)(1)C]-(+)-PHNO for the D3 and D2 receptors in the rhesus monkey brain *in vivo. Synapse* **66**: 489–500.
- Ginovart N, Galineau L, Willeit M, Mizrahi R, Bloomfield PM, Seeman P et al (2006). Binding characteristics and sensitivity to endogenous dopamine of [11C]-(+)-PHNO, a new agonist radiotracer for imaging the high-affinity state of D2 receptors in vivo using positron emission tomography. J Neurochem 97: 1089-1103.
- Ginovart N, Willeit M, Rusjan P, Graff A, Bloomfield PM, Houle S *et al* (2007). Positron emission tomography quantification of [11C]-(+)-PHNO binding in the human brain. *J Cereb Blood Flow Metab* 27: 857–871.
- Gjedde A, Wong DF, Rosa-Neto P, Cumming P (2005). Mapping neuroreceptors at work: on the definition and interpretation of binding potentials after 20 years of progress. *Int Rev Neurobiol* **63**: 1–20.
- Graff-Guerrero A, Redden L, Abi-Saab W, Katz DA, Houle S, Barsoum P et al (2010). Blockade of [11C](+)-PHNO binding in human subjects by the dopamine D3 receptor antagonist ABT-925. Int J Neuropsychopharmacol 13: 273–287.
- Graff-Guerrero A, Willeit M, Ginovart N, Mamo D, Mizrahi R, Rusjan P *et al* (2008). Brain region binding of the D2/3 agonist [11C]-(+)-PHNO and the D2/3 antagonist [11C]raclopride in healthy humans. *Hum Brain Mapp* **29**: 400-410.
- Gunn RN, Lammertsma AA, Hume SP, Cunningham VJ (1997). Parametric imaging of ligand-receptor binding in PET using a simplified reference region model. *Neuroimage* 6: 279–287.
- Gurevich EV, Joyce JN (1999). Distribution of dopamine D3 receptor expressing neurons in the human forebrain: comparison with D2 receptor expressing neurons. *Neuropsychopharmacology* **20**: 60–80.
- Innis RB, Cunningham VJ, Delforge J, Fujita M, Gjedde A, Gunn RN *et al* (2007). Consensus nomenclature for in vivo imaging of reversibly binding radioligands. *J Cereb Blood Flow Metab* 27: 1533–1539.
- Kegeles LS, Abi-Dargham A, Frankle WG, Gil R, Cooper TB, Slifstein M *et al* (2010). Increased synaptic dopamine function in

associative regions of the striatum in schizophrenia. Arch Gen Psychiatry 67: 231–239.

- Kiss B, Horti F, Bobok A (2011). In vitro and in vivo comparison of [(3)H](+)-PHNO and [(3)H]raclopride binding to rat striatum and lobes 9 and 10 of the cerebellum: a method to distinguish dopamine D(3) from D(2) receptor sites. *Synapse* **65**: 467–478.
- Lammertsma AA, Hume SP (1996). Simplified reference tissue model for PET receptor studies. *Neuroimage* 4(Pt 1): 153-158.
- Laruelle M (2000). Îmaging synaptic neurotransmission with in vivo binding competition techniques: a critical review. J Cereb Blood Flow Metab 20: 423-451.
- Laruelle M, D'Souza CD, Baldwin RM, Abi-Dargham A, Kanes SJ, Fingado CL *et al* (1997). Imaging D2 receptor occupancy by endogenous dopamine in humans. *Neuropsychopharmacology* **17**: 162–174.
- Martinez D, Greene K, Broft A, Kumar D, Liu F, Narendran R *et al* (2009). Lower level of endogenous dopamine in patients with cocaine dependence: findings from PET imaging of D(2)/D(3) receptors following acute dopamine depletion. *Am J Psychiatry* **166**: 1170–1177.
- Mawlawi O, Martinez D, Slifstein M, Broft A, Chatterjee R, Hwang DR *et al* (2001). Imaging human mesolimbic dopamine transmission with positron emission tomography: I. Accuracy and precision of D(2) receptor parameter measurements in ventral striatum. *J Cereb Blood Flow Metab* **21**: 1034–1057.
- McCormick PN, Ginovart N, Wilson AA (2011). Isoflurane anaesthesia differentially affects the amphetamine sensitivity of agonist and antagonist D2/D3 positron emission tomography radiotracers: implications for in vivo imaging of dopamine release. *Mol Imaging Biol* 13: 737–746.
- Narendran R, Slifstein M, Guillin O, Hwang Y, Hwang DR, Scher E et al (2006). Dopamine (D2/3) receptor agonist positron emission tomography radiotracer [11C]-(+)-PHNO is a D3 receptor preferring agonist *in vivo*. Synapse **60**: 485–495.
- Newberg AB, Moss AS, Monti DA, Alavi A (2011). Positron emission tomography in psychiatric disorders. *Ann N Y Acad Sci* **25**: 1749–6632.
- Rabiner EA, Slifstein M, Nobrega J, Plisson C, Huiban M, Raymond R *et al* (2009). *In vivo* quantification of regional dopamine-D3 receptor binding potential of (+)-PHNO: studies in non-human primates and transgenic mice. *Synapse* **63**: 782–793.
- Searle G, Beaver JD, Comley RA, Bani M, Tziortzi A, Slifstein M *et al* (2010). Imaging dopamine D3 receptors in the human brain with positron emission tomography, [11C]PHNO, and a selective D3 receptor antagonist. *Biol Psychiatry* **68**: 392–399.
- Searle GE, Beaver JD, Tziortzi A, Comley RA, Bani M, Ghibellini G et al (2013). Mathematical modelling of [(1)(1)C]-(+)-PHNO human competition studies. *Neuroimage* 68: 119-132.
- Seeman P (2013). Schizophrenia and dopamine receptors. Eur Neuropsychopharmacol 23: 999-1009.
- Seeman P, Ulpian C, Larsen RD, Anderson PS (1993). Dopamine receptors labelled by PHNO. Synapse 14: 254-262.
- Shotbolt P, Tziortzi AC, Searle GE, Colasanti A, van der Aart J, Abanades S *et al* (2012). Within-subject comparison of [(11)C]-(+)-PHNO and [(11)C]raclopride sensitivity to acute ampheta-

mine challenge in healthy humans. *J Cereb Blood Flow Metab* **32**: 127–136.

- Singh A, Mahgoub N, Klimstra S (2011). Apathy associated with a unilateral globus pallidus lesion: a case report. *Int J Geriatr Psychiatry* **26**: 765–766.
- Studholme C, Hill DL, Hawkes DJ (1997). Automated threedimensional registration of magnetic resonance and positron emission tomography brain images by multiresolution optimization of voxel similarity measures. *Med Phys* 24: 25-35.
- Tatsch K, Poepperl G (2012). Quantitative approaches to dopaminergic brain imaging. *Q J Nucl Med Mol Imaging* 56: 27–38.
- Tziortzi AC, Searle GE, Tzimopoulou S, Salinas C, Beaver JD, Jenkinson M *et al* (2011). Imaging dopamine receptors in humans with [11C]-(+)-PHNO: dissection of D3 signal and anatomy. *Neuroimage* 54: 264–277.
- Udenfriend S, Zaltzman-Nirenberg P, Nagatsu T (1965). Inhibitors of purified beef adrenal tyrosine hydroxylase. *Biochem Pharmacol* 14: 837–845.
- van Wieringen JP, Booij J, Shalgunov V, Elsinga P, Michel MC (2013). Agonist high- and low-affinity states of dopamine D(2) receptors: methods of detection and clinical implications. Naunyn Schmiedebergs Arch Pharmacol 386: 135-154.
- Verhoeff NP, Christensen BK, Hussey D, Lee M, Papatheodorou G, Kopala L *et al* (2003). Effects of catecholamine depletion on D2 receptor binding, mood, and attentiveness in humans: a replication study. *Pharmacol Biochem Behav* 74: 425–432.
- Verhoeff NP, Hussey D, Lee M, Tauscher J, Papatheodorou G, Wilson AA *et al* (2002). Dopamine depletion results in increased neostriatal D(2), but not D(1), receptor binding in humans. *Mol Psychiatry* 7: 322–328.
- Verhoeff NP, Kapur S, Hussey D, Lee M, Christensen B, Psych C et al (2001). A simple method to measure baseline occupancy of neostriatal dopamine D2 receptors by dopamine in vivo in healthy subjects. *Neuropsychopharmacology* **25**: 213–223.
- Versteeg DH, Van Der Gugten J, De Jong W, Palkovits M (1976). Regional concentrations of noradrenaline and dopamine in rat brain. *Brain Res* 113: 563–574.
- Vijayaraghavan L, Vaidya JG, Humphreys CT, Beglinger LJ, Paradiso S (2008). Emotional and motivational changes after bilateral lesions of the globus pallidus. *Neuropsychology* **22**: 412–418.
- Willeit M, Ginovart N, Graff A, Rusjan P, Vitcu I, Houle S et al (2008). First human evidence of d-amphetamine induced displacement of a D2/3 agonist radioligand: A [11C]-(+)-PHNO positron emission tomography study. Neuropsychopharmacology 33: 279–289.
- Wilson AA, Garcia A, Jin L, Houle S (2000). Radiotracer synthesis from [(11)C]-iodomethane: a remarkably simple captive solvent method. *Nucl Med Biol* 27: 529–532.
- Wilson AA, McCormick P, Kapur S, Willeit M, Garcia A, Hussey D et al (2005). Radiosynthesis and evaluation of [11C]-(+)-4-propyl-3,4,4a,5,6,10b-hexahydro-2H-naphtho[1,2-b][1,4]oxazin-9-ol as a potential radiotracer for in vivo imaging of the dopamine D2 high-affinity state with positron emission tomography. J Med Chem 48: 4153-4160.

Supplementary Information accompanies the paper on the Neuropsychopharmacology website (http://www.nature.com/npp)

npg