

Effects of transforming growth interacting factor on biological behaviors of gastric carcinoma cells

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Received: 2004-02-27 **Accepted:** 2004-04-05

Abstract

AIM: Transforming growth interacting factor (TGIF) is an inhibitor of both transforming growth factor β (TGF- β) and retinoid signaling pathways. Moreover, the activation of MAPK pathway can prolong its half-life. However, its role in carcinogenesis is still unknown. Thus we attempted to investigate the effect of TGIF on biologic behaviors of gastric carcinoma cells.

METHODS: Gastric carcinoma cell line, SGC-7901, was stably transfected with plasmid PcDNA3.1-TGIF. Western blotting and cell immunohistochemistry screening for the highly expressing clone of TGIF were employed. The growth of transfected cells was investigated by MTT and colony-formation assays, and apoptosis was measured by flow cytometry (FCM) and transmission electron microscopy. Tumorigenicity of the transfectant cells was also analyzed.

RESULTS: TGIF had no effect on the proliferation, cell cycle and apoptosis of SGC-7901 cells, but cellular organelles of cells transfected with TGIF were richer than those of vector control or parental cells. Its clones were smaller than the control ones in plate efficiency, and its tumor tissues also had no obvious necrosis compared with the vector control or parental cells. Moreover, TGIF could resist TGF- β mediated growth inhibition.

CONCLUSION: TGIF may induce differentiation of stomach neoplastic cells. In addition, TGIF can counteract the growth inhibition induced by TGF- β .

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Key words: Gastric carcinoma; TG interacting factor; Cell differentiation; TGF-beta

Hu ZL, Wen JF, Xiao DS, Zhen H, Fu CY. Effects of transforming growth interacting factor on biological behaviors of gastric carcinoma cells. *World J Gastroenterol* 2005; 11(1): 84-88
<http://www.wjgnet.com/1007-9327/11/84.asp>

INTRODUCTION

Gastric carcinoma is one of the most frequent tumors that seriously threatens people's health in China. However, its exact mechanism is still unclear. Many investigations have shown that most neoplasms are associated with TGF- β . Moreover, a

variety of neoplasms are able to resist the growth inhibition mediated by TGF- β ^[1]. The role of TGF- β signaling pathway in the development of stomach neoplasms is worthy of our attention.

TGIF is a nucleoprotein that belongs to the homeobox domain TALE family, which has a three-amino acid insertion between helices 1 and 2 of the homeodomain^[2]. TGIF locates on 18p11.3 and encodes a protein consisting of 272 amino acids. It has been discovered to be involved in many biological processes, such as human and *Drosophila* development. Recent studies have shown that *Drosophila* TGIF is essential for developmentally regulated transcription in its spermatogenesis^[3,4]. Although TGIF homozygous mutant flies are viable and appear morphologically normal, the males are completely sterile. TGIF has also been identified as a small group of genes implicated in the human development disorder holoprosencephaly (HPE), a common structural defect of the developing forebrain in human being^[5]. It has also been suggested to act as a competitive inhibitor of the TALE-class homeodomain protein Meis2 in neuronal cell lines^[6].

In addition, TGIF also participates in a number of distinct pathways. Bertolino *et al*^[2] discovered that TGIF could compete with retinoid for binding sites in promoter, and inhibit the retinoid signaling pathway, while retinoid could inhibit cell proliferation and induce cell differentiation and apoptosis. Recent studies reveal that TGIF is also a transcriptional co-repressor, which inhibits TGF- β signaling pathway^[7-13].

TGIF inhibits TGF- β signaling pathway mainly by histone deacetylase (HDAC) dependent^[7] and HDAC independent mechanisms^[8]. In addition, TGIF also directly inhibits target gene expression via binding to DNA at its HD region consisting of 35-97 amino acid residues^[10]. Recently Lo *et al*^[11] revealed that TGF- β signaling pathway had a cross-talk with EGF/Ras/MAPK signaling pathway that could phosphorylate TGIF, prolong its half-life, and raise its protein level.

TGIF inhibits not only TGF- β signaling pathway but also retinoid signaling pathway. Moreover, EGF/Ras/MAPK signaling pathway prolong half-life of TGIF. But it is still unknown whether TGIF plays a role in carcinogenesis. To determine the role of TGIF in the carcinogenesis, SGC-7901, a moderately differentiated gastric carcinoma cell line, was transfected with TGIF to investigate the effect of TGIF on the biological behaviors of SGC-7901 cells.

MATERIALS AND METHODS

Cell line

The human gastric carcinoma cell line, SGC-7901, was cultured in RPMI 1640 medium containing 100 mL/L fetal bovine serum (FBS) supplemented with penicillin and streptomycin. Cultures were incubated in an incubator containing 5 mL/L CO₂ in air at 37 °C.

Plasmid

Plasmid pcDNA3.1-TGIF was a gift from Professor Mouradian, Genetic Pharmacology Unit, Experimental Therapeutics Branch, NINDS, National Institutes of Health, Bethesda, Maryland 20892, USA.

Transfection and selection

Transfection and selection of positive clones were carried out in a 6-well plate. When the cells reached 70% confluence, the transfection process began. Briefly, solution A was prepared by diluting 2 µg of pcDNA3.1-TGIF into 200 µL serum-free medium, and solution B was prepared by diluting 5 µL lipofectamine (Life Technologies) into 200 µL serum-free medium. The two solutions were mixed for 20 min at room temperature, and then 0.6 mL serum-free medium was added to a tube containing the complex, and subsequently added to the rinsed cells. The medium was replaced with fresh and complete medium 18 h after transfection. Seventy-two hours after transfection, it was replaced again with the selective medium containing 800 g/L G418 (Alexis Biochemicals). Once stable transfection was obtained, the cells were maintained in 200 g/L of G418. Meanwhile, SGC-7901 cells were transfected with the empty pcDNA3.1 vector as control.

Western blot analysis

Total proteins were measured using the BCA kit (Pierce) according to the manufacturer's protocol. Forty µg of total proteins was separated by 12% SDS-PAGE under reducing conditions, and transferred to nitrocellulose membrane. The nitrocellulose membrane was then incubated with blocking buffer (PBS containing 5% non-fat milk) for 2 h at room temperature and with goat polyclonal antibody against TGIF overnight at 37 °C with gentle shaking. The membrane was washed with PBS twice for 5 min, and then incubated with rabbit anti-goat IgG conjugated horseradish peroxidase diluted at 1:3 000 (Zhongshan Co, Beijing) for 2 h at room temperature. After washing, TGIF was detected using DAB reagents. Ponceau S was used as a loading control.

Cell immunohistochemical staining

Cells were seeded on slides at a density of 1.8×10^4 and grown for two days. Slides were washed once with PBS and fixed in acetone for 20 min at 4 °C. Fixed cells were washed 3 times with-PBS and nonspecific proteins were blocked using non-immune serum for 30 min at room temperature. Cells were incubated for 1 h with the goat polyclonal antibody against TGIF, then washed twice for 3 min with PBS. Cells were then incubated for 1 h with rabbit anti-goat IgG conjugated horseradish peroxidase diluted at 1:3 000 (Zhongshan Co, Beijing), followed by two washes of 3 min in PBS. Cells were stained with DAB reagent.

MTT assay

Cells were cultured in 96-well plates at a density of 1×10^4 cells per well. Cell survival was measured by MTT assay 24, 48, 72, 96, 120 and 144 h after seeding. MTT assay was used to determine mitochondrial activity, which correlated with the number of viable cells in culture. Briefly, 20 µL of 5 g/L MTT (3-(4,5-dimethyl-thiazolyl-2)-2,5-diphenyltetrazolium bromide) in PBS was added to each well. Cells were incubated with MTT compound for 4 h at 37 °C in a 5 mL/L CO₂ atmosphere, and subsequently 150 µL of DMSO was added to each well. The plates were incubated until MTT was completely resolved and A_{595} was measured.

Flow cytometry analysis

Approximately 5×10^6 centrifugal sedimentation cells were immediately fixed in 700 mL/L ethanol and stored at 4 °C in PBS for fluorescence-activated cell sorting. Flow cytometry analysis was performed on a FACStar flow cytometer (Becton Dickinson). Histograms of cell number logarithmic fluorescence intensity were recorded for 10 000 cells per sample.

Plating efficiency

To determine plating efficiency, cells were seeded in 6-well plates, 1 000 cells per well. After 14 d, the colonies were fixed with 4% methanol and stained with 5% Giemsa solution (Sigma). The number of colonies with a diameter larger than 1 mm was counted. The plating efficiency (PE) was calculated as follows: PE = (colonies formed/cells seeded) × 100%.

Transmission electron microscopy (TEM)

Pellet of the transfected cells was fixed in 2.5 g/L glutaraldehyde, postfixed with 10 g/L osmium tetroxide, treated with 20 g/L uranyl acetate, dehydrated in ethanol, infiltrated with propylene oxide, and embedded in Epon mixture. Ultrathin sections were observed under Opton EM 10C (Germany).

Tumor development in athymic nude mice

Nine female nude mice (BALB/c-nude, 4-6 wk old, weighing 16-18 g) were divided into 3 groups, 3 mice each group, and inoculated subcutaneously at the left flank with TGIF transfectant, vector control and parental cells (7×10^6 cells suspended in 0.2 mL of phosphate-buffered saline) and monitored for tumor development. Tumor size and animal weight were measured weekly. The nude mice were sacrificed and tumors were removed 35 d after inoculation.

Statistical analysis

F test was used. *P* value less than 0.05 was considered statistically significant.

RESULTS

Construction of cell clones stably expressing TGIF protein

After transfection of SGC-7901 cells with a vector encoding TGIF, we identified two cell clones that constitutively overexpressed this protein by cell immunohistochemistry (Figure 1) and Western blot analyses (Figure 2). We selected one of them for further experiment.

Effect of TGIF on growth of SGC-7901 cells

To determine the impact of TGIF on cell growth *in vitro*, we examined the rate of cell growth with MTT assay. As shown in Figure 3A, the growth rate of cells overexpressing TGIF had no distinct changes compared with blank and negative controls. After incubating with 10 ng/mL TGF-β 1 for the indicated time, both parental and vector control cells were found to have lower growth rates. However, TGIF transfected cells had no distinct changes (Figure 3B) and the difference was significant ($P < 0.05$).

Table 1 Effects of TGIF on cell cycle and apoptosis by flow cytometry in blank, vector control and TGIF expressing cells

	TGF-β 1 (-) (%)				TGF-β 1 (+) (%)			
	G1	G2	S	Apoptosis	G1	G2	S	Apoptosis
SGC-7901	55.2	15.8	29	1.35	63.2	14.2	22.6	2.7
PcDNA3.1	50.1	15.8	34.2	0.36	60.3	13.9	25.8	0.54
TGIF	50.9	18.4	30.6	0.41	50.1	21.2	28.7	0.67



Figure 1 Expression of TGIF protein by cell immunohistochemistry. A: cells stably transfected with pcDNA3.1-TGIF; B: cells stably transfected with PcDNA3.1; C: parental cells.

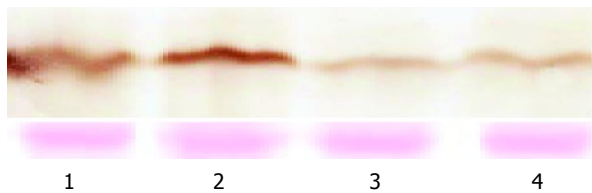


Figure 2 Expression of TGIF protein by Western blotting. Lanes 1 and 2: TGIF expressing cells; lane 3: vector control cells, lane 4: parental cells. The lower panel was stained with Ponceau S as a loading control.

Effect of TGIF on cell cycle and apoptosis rate of SGC-7901 cells

To confirm the effect of TGIF on proliferation of SGC-7901 cells, the cell cycle distribution and apoptosis were determined by flow cytometry. As shown in Table 1, TGIF had no effect on cell cycle and apoptosis rate of SGC-7901 cells. In parental and vector control cells, cell content of G1 phase obviously increased after treatment with TGF-β 1. However, TGIF expressing cells

had no distinct change in cell content of G1 phase. All groups had a slight increase in apoptosis rate after incubation with 10 μg/L TGF-β 1 for 72 h.

Plating efficiency

Plating efficiency in parent, vector control and TGIF transfected cells was 15.1%, 12.4% and 16.9% respectively (Figure 4), and the difference had no statistical significance ($P>0.05$). However, clones of TGIF transfected cells were smaller than those of the parental and vector control cells (Figure 4, upper panel).

Effect of TGIF on ultrastructure of SGC-7901 cells

Apoptotic body was not found in parental, vector control or TGIF transfected cells under TEM, but there were more cell organellae in TGIF transfected cells compared with the blank and negative control cells (Figure 5).

Effect of TGIF on SGC-7901 cell growth in vivo

We examined the effect of TGIF expression on SGC-7901 cell growth in athymic mice. Tumors were palpable in the first week

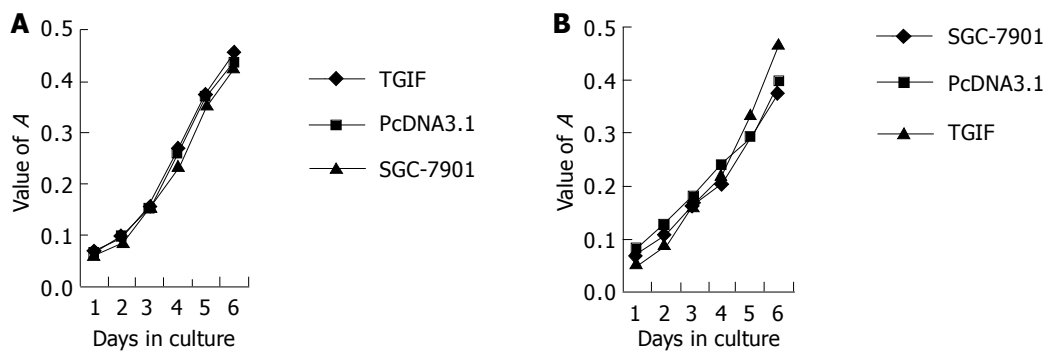


Figure 3 Proliferation rate of TGIF transfected, vector control and parental cells. A: Without 10 μg/L TGF-1; B: with 10 μg/L TGF-β 1.

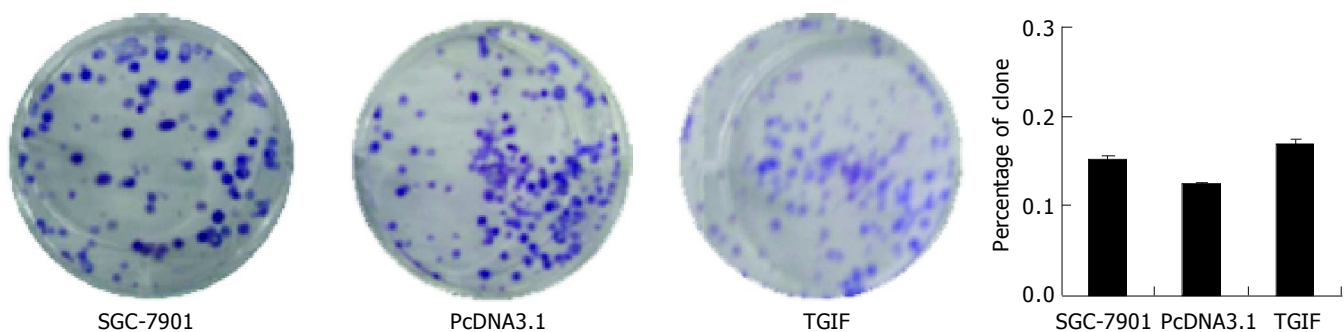


Figure 4 Plating efficiency in parental, vector control and TGIF transfected cells.

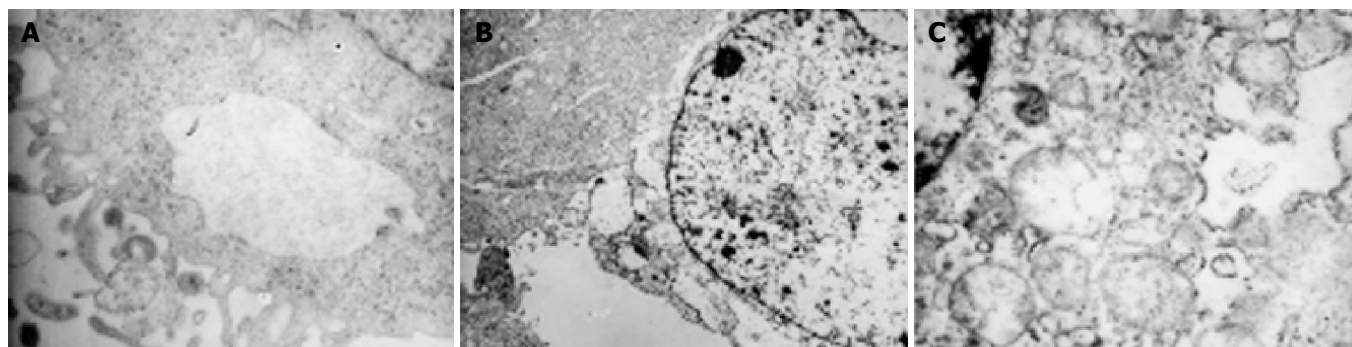


Figure 5 Morphology of blank, negative control and TGIF transfectant cells by TEM $\times 15\ 000$. A: parental cell; B: vector control cell; C: TGIF transfectant cell.



Figure 6 Tumor development in nude mice. A: mice inoculated with parental cells; B: mice inoculated with vector control cells; C: mice inoculated with TGIF expressing cells.

after inoculation of cells in female athymic mice. As shown in Figure 6, TGIF transfectants revealed no difference in tumor growth as compared with vector control and parental cells throughout the observation period. The mean tumor weights in mice transfected parental, vector control cells and TGIF were 0.85 ± 0.09 , 0.87 ± 0.13 and 0.87 ± 0.27 g, respectively. In addition, the mean tumor volumes were 0.99 ± 0.08 , 1.01 ± 0.11 and 1.10 ± 0.12 cm³, respectively. However, the differences had no statistical significance ($P > 0.05$). The animals were killed in accordance with the institutional tumor burden guidelines. After tumors were excised, there were necrotic tissues effused from tumors in parental and vector control groups but not in TGIF transfectants.

DISCUSSION

TGIF is a transcription factor that has been implicated in a number of distinct pathways. TGIF was first identified as a competitor of retinoic acid receptor to bind to retinoic acid response elements^[2]. Subsequently TGIF interacts with Smads and is an inhibitor of TGF- β signaling pathway^[7-13]. Recently Lo *et al.*^[11] revealed that MAPK signaling pathway had a cross-talk with TGF- β signaling pathway. MAPK transducing pathway can phosphorylate TGIF, prolong its half-life and raise its protein level. The enhancement of TGIF function might inhibit negative regulation of cell cycle by TGF- β . Its role in tumorigenicity is worthy of attention. Several reports have indicated that TGIF probably implicates in carcinogenesis. Nakakuki *et al.*^[14] discovered that TGIF gene was overexpressed in esophageal carcinoma. Voorter *et al.*^[15] revealed that there was gene amplification at 18p11 where TGIF locates in bladder transitional cell carcinoma using comparative genome hybridization. Luo *et al.*^[16] found that autologous antibody against TGIF existed in serum of patients with ovarian carcinoma.

Our experiment revealed that the growth rate of SGC-7901

cells had no distinct difference after transfection with TGIF (Figure 3), and the distribution of cell cycle had no obvious change either (Table 1). This result is not consistent with Edwards' report showing that overexpression of TGIF overcame the checkpoint of yeast G1 phase^[17]. The distinction may attribute to the difference of cell types. There were no differences in plating efficiency and nude mice tumorigenicity among parental, vector control and TGIF transfected cells (Figures 4 and 6). All these indicate that TGIF cannot worsen the biological behavior of SGC-7901 cells. Conversely, the number of cell organelles in TGIF transfected cells increased compared to blank and negative control cells (Figure 5), and the tumor tissues of TGIF transfectant group exhibited no distinct necrosis compared to control groups. This data indicates that TGIF may induce differentiation of SGC-7901 cells, at least in part.

After the treatment with TGF- β 1, parental and vector control cells showed distinct reduction in cell growth, whereas TGIF transfectants revealed no obvious difference (Figure 3). Flow cytometry also showed similar results (Table 1). Our finding is coincident with Lo's report that HaCaT cell line could resist the growth inhibition mediated by TGF- β after stable transfection of TGIF^[11]. Thus overexpression of TGIF protein can inhibit the negative regulation of TGF- β in cell cycle. It also implies that tumor cells may escape the growth inhibition by TGF- β via this mechanism.

Cells transfected with TGIF showed no distinct difference in apoptosis compared to the controls (Table 1). After incubation with TGF- β 1 for 72 h, there was no distinct difference among the three groups, indicating that TGIF may not interfere with TGF- β -mediated cell apoptosis. However, we cannot rule out the possibility that the insensitivity to TGF- β -mediated apoptosis resulting from the disturbance of TGF- β signaling pathway in SGC-7901 cells, contributes to cells transfected with TGIF resisting TGF- β mediated apoptosis.

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Edited by Ma JY and Wang XL Proofread by Zhu LH