

Challenges to targeting epidermal growth factor receptor in glioblastoma: escape mechanisms and combinatorial treatment strategies

Patrick Roth and Michael Weller

Department of Neurology and Brain Tumor Center Zurich, University Hospital Zurich, Zurich, Switzerland (P.R., M.W.)

Corresponding Author: Michael Weller, MD, Department of Neurology, University Hospital Zurich, Frauenklinikstrasse 26, CH-8091 Zurich, Switzerland (michael.weller@usz.ch).

Epidermal growth factor receptor (EGFR) gene amplification and activating mutations are common findings in glioblastomas. EGFR is at the top of a downstream signaling cascade that regulates important characteristics of glioblastoma cells, including cellular proliferation, migration, and survival. Targeting EGFR has therefore been regarded as a promising therapeutic strategy in glioblastoma for decades. However, although various pharmacological inhibitors and anti-EGFR antibodies are available, the antiglioma activity of these agents has been largely limited to preclinical models, whereas their administration to glioblastoma patients was characterized by lack of clinical benefit. Comprehensive efforts have been made within the last years to understand the underlying mechanisms that confer resistance to EGFR inhibition in glioma cells. The absence of well-known mutations that predict response to EGFR tyrosine kinase inhibitors (TKIs) in gliomas as well as the presence of redundant and alternative compensatory pathways are among the most important escape mechanisms that prevent potent antiglioma effects of EGFR-targeting drugs. Accordingly, an increasing number of in vitro and in vivo studies are aimed at overcoming this resistance by combinatorial approaches using anti-EGFR treatment together with one or more additional drugs. Novel insights into the molecular mechanisms mediating resistance to anti-EGFR treatment and promising combinatorial approaches may help to better define a future role for EGFR inhibition in the treatment of glioblastoma.

Keywords: EGFR, EGFRvIII, escape mechanism, therapeutic targeting, therapy resistance.

Background

Gliomas are the most common primary brain tumors in adults. They are classified by the World Health Organization into grades I through IV, with glioblastoma being the most malignant subtype. Despite all efforts, median survival in glioblastoma patients is restricted to approximately 16 months in clinical trial populations.¹ Various therapeutic strategies have been explored within the last years in order to improve the prognosis of glioblastoma patients. Several of these novel strategies aim at targeting specific molecules or signaling pathways that are deregulated in glioma cells. Among the genetic aberrations associated with gliomas, amplification of epidermal growth factor receptor (EGFR, also named HER1 or ERBB1) is a frequent finding, which has been described in approximately 40%–50% of all glioblastomas.² Besides EGFR, the family of HER receptor tyrosine kinases comprises ERBB2 (more frequently known as HER2/neu), ERBB3, and ERBB4. EGFR binds several ligands, including epidermal growth factor (EGF), transforming growth factor- α , heparin-binding EGF-like growth factor, amphiregulin, betacellulin, epigen, and epiregulin.³ Engagement of EGFR results in the activation of a cytoplasmic

tyrosine kinase (TK) domain and subsequent intracellular downstream signaling involving, among others, the mitogen-activated protein kinase and phosphatidylinositol 3-kinase (PI3K) pathways.² Thus, EGFR signaling affects various cellular processes, including proliferation, survival, and metabolism. Amplification of EGFR is frequently associated with the occurrence of a mutant form of EGFR called EGFR variant III (EGFRvIII, also known as Δ EGFR). EGFRvIII is found in approximately 20%–30% of all glioblastomas.^{4,5} However, there are various other mutations in EGFR, some of which predict a response to pharmacological inhibitors (see below). Because of its role as a central regulator of various biological processes in glioma cells as well as its potential contribution to resistance to apoptotic stimuli and alkylating chemotherapy with temozolomide,^{6,7} EGFR has attracted much attention as a therapeutic target.

Resistance to Pharmacological EGFR Inhibitors and Antibodies Targeting EGFR

As outlined above, EGFR has been regarded as a promising point of attack for therapeutic interventions against malignant

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gliomas. However, most approaches used so far have shown disappointing results in the clinic, with virtually no benefit for populations of unselected patients. Thus, a major research focus within the last years has been the deciphering of the molecular mechanisms underlying the resistance of glioma cells to EGFR inhibition. The following section describes EGFR-targeted therapies as well as molecular alterations that may confer resistance to EGFR inhibition.

Pharmacological EGFR inhibitors

Pharmacological inhibitors, mostly small molecule TKIs, targeting EGFR have been extensively tested in preclinical glioma models. Similar to their practice with other tumor entities such as lung carcinomas, for which these drugs are well established in clinical practice, most investigators used erlotinib or gefitinib to interfere with EGFR signaling. The EGFR-blocking activity of erlotinib and gefitinib largely depends on the presence of mutations in exons 19 and 21 of the TK domain. These mutations are commonly found in lung cancer and other tumor entities and have led to the approval of several EGFR inhibitors. However, these “sensitizing” mutations are virtually absent in glioblastomas, which may partially explain the lack of activity of standard TKIs in this disease.^{8–11}

Antibodies against EGFR

Antibodies directed against EGFR—with cetuximab, nimotuzumab, and panitumumab as the most prominent candidates—were also investigated for their antiglioma activity in vitro and in vivo. Antibodies may exert their effect by preventing the binding of EGFR ligands to the receptor. Furthermore, antibody binding may result in receptor internalization and degradation.¹² Although antibodies to EGFR have been approved for other cancer types, such as cetuximab for the treatment of Kirsten rat sarcoma viral oncogene homolog wild-type colon cancer, their use against intracranial neoplasms such as glioblastoma represents a challenge due to the presence of the blood–brain barrier, which may preclude the penetration of the antibody to all parts of the tumor. However, small molecule EGFR inhibitors such as erlotinib and gefitinib also did not markedly inhibit EGFR phosphorylation in vivo.¹³ Accordingly, poor tumor perfusion or the blood–brain barrier may represent an important “resistance factor” that limits the activity of EGFR-targeting drugs in the brain.

General mechanisms of resistance to EGFR-targeted therapies

The escape of glioma cells from EGFR-targeted therapy is caused by several characteristics of these cells and particularly by the existence of multiple overlapping and alternative compensatory signaling pathways that allow for a loss of EGFR function without detrimental effects on the cells.^{14,15} One common finding in glioma cells, that is, loss of phosphatase and tensin homolog (PTEN) deleted on chromosome 10, has been identified as a resistance factor to drugs directed against EGFR. PTEN loss promotes resistance to EGFR presumably by dissociating EGFR inhibition from downstream inhibition of the PI3K pathway.^{9,16} Inhibition of mammalian target of rapamycin (mTOR) restores the sensitivity of PTEN-deficient gliomas to EGFR inhibitors.^{9,17,18} Since 40%–50% of glioblastomas lack PTEN expression, these findings were

the rationale for the combined use of EGFR and mTOR inhibitors in human patients (see below). Furthermore, it became obvious that even in glioblastoma specimens with wild-type PTEN expression, resistance to EGFR inhibitors may occur as a result of phosphorylation of PTEN at the conserved tyrosine residue Y240.¹⁹

One of the most comprehensive strategies aiming at identifying molecules and pathways that mediate resistance to EGFR inhibition has been the use of genome-wide small hairpin (sh)RNA screens. The application of such an approach revealed the dopamine receptor D2 signaling pathway as a novel therapeutic target. Combined inhibition of dopamine receptor D2 signaling and EGFR inhibition resulted in synergistic antiglioma activity in in vitro and in vivo models.²⁰

Glioma cells can be characterized by intrinsic resistance to EGFR inhibitors or acquired mechanisms that allow them to escape from EGFR-targeted treatment. Acquired resistance to EGFR inhibition in EGFR-mutant glioma cells is conferred by an induction of platelet-derived growth factor receptor (PDGFR)– β expression. Consequently, the combined targeting of EGFR and PDGFR- β resulted in more potent antitumor activity in preclinical glioma models than either treatment alone.²¹ Furthermore, expression of the promyelocytic leukemia (PML) gene in glioma cells prevents the induction of cell death in response to EGFR inhibition.²² Abrogation of PML expression by siRNA-mediated gene silencing or administration of the PML inhibitor arsenic trioxide restored the susceptibility of experimental gliomas in vivo to EGFR inhibition.

The fact that not all tumor cells share the same molecular makeup may also contribute to resistance to EGFR inhibition. In this regard, a population of cells within gliomas that exhibit stem cell–like properties has been described within the last years. These cells, also known as glioma-initiating cells, have been proposed as a major factor for the ultimately lethal course of the disease due to their contribution to the resistance of gliomas to various treatments. Preclinical data indicate that the resistance of glioma-initiating cells to EGFR inhibition is partially due to focal adhesion kinase–mediated integrin β 1 signaling.²³ A similar study revealed that only cotreatment consisting of erlotinib and the hedgehog pathway inhibitor cyclopamine had an effect on sphere initiation in glioblastoma stem cell cultures.²⁴ These reports demonstrate that anti-EGFR strategies hold promise in targeting the stem cell population within glioblastomas. However, they also suggest that EGFR inhibition alone is insufficient and needs to be combined with the therapeutic targeting of at least one additional pathway. Further studies are required to define which combination may work best and which of the available combinations ultimately succeeds in human patients.

Clinical Trials and Combined Treatment Approaches in Patients With Malignant Gliomas

Treatment of glioma patients with pharmacological EGFR inhibitors or blocking antibodies as single treatment has been largely futile. Several trials using anti-EGFR approaches for gliomas with different World Health Organization grades in the settings of newly diagnosed or recurrent tumors have failed to show signs of activity. However, as outlined above, extensive preclinical work demonstrated that the disruption of converging signaling pathways may help to overcome resistance to EGFR inhibitors. Based on the increasing awareness that combinatorial targeting

of EGFR and one or more additional molecules may exert more robust antitumor activity, various trials were initiated using combinations of EGFR-targeting agents and additional drugs.

Clinical administration of TKIs

Several preclinical reports suggest a sensitizing effect of EGFR inhibition to irradiation.²⁵ Accordingly, clinical trials were designed to explore the combination of radiation therapy with EGFR inhibitors in patients with newly diagnosed glioblastoma. However, such trials failed to show benefit from addition of gefitinib or erlotinib to radiation therapy compared with historical controls. Notably, these trials completed enrollment before the introduction of temozolomide to the standard of care for glioblastoma patients.^{26,27} Addition of erlotinib to temozolomide-based chemoradiation in patients with newly diagnosed glioblastoma resulted in prolonged overall survival compared with historical controls,²⁸ but confirmation of this finding in a randomized trial is lacking and seems not to be further pursued. A phase I/II study explored the combination of the EGFR inhibitor lapatinib and the multikinase inhibitor pazopanib in patients with recurrent malignant glioma. Here, patients were stratified into 2 groups, with either intact PTEN or EGFRvIII expression or without PTEN and EGFRvIII expression. However, the overall limited activity of this regimen did not differ among patients stratified by tumor EGFRvIII or PTEN status. A pharmacokinetic analysis demonstrated that only subtherapeutic levels of lapatinib were reached, which may have precluded sufficient inhibition of EGFR signaling.²⁹ The combined administration of the mTOR inhibitor everolimus and gefitinib did not achieve durable responses in patients with recurrent glioblastoma.³⁰ Similarly, a phase I/II trial exploring the combination of erlotinib with the mTOR inhibitor temsirolimus in patients with recurrent malignant glioma failed to prove relevant antitumor activity. However, dose-limiting toxicity involving rash and mucositis was common.³¹ Low tumor levels of both drugs and the failure to prove target inhibition in the posttreatment tissue of several patients may partially explain the futility of this regimen. Compared with the “first-generation” EGFR inhibitors, second-generation, irreversible TKIs may exert more potent antiglioma activity. However, one of these novel drugs, afatinib, did not show any signs of activity when used as a single agent and did not improve the outcome of patients with recurrent glioblastoma in combination with temozolomide, likely because of negligible blood–brain barrier penetration.³² Other compounds, such as dacomitinib (PF-00299804), are currently being tested in clinical trials enrolling patients with recurrent glioblastoma (NCT01520870 and NCT01112527).

Anti-EGFR antibodies

The anti-EGFR antibody nimotuzumab was assessed in several clinical trials in patients with high-grade gliomas either alone or in combination with other treatment modalities and demonstrated only modest or no signs of activity.^{33,34} The lack of benefit of this regimen may be explained by the recent finding that treatment of glioma cells with an anti-EGFR antibody enhances DNA repair and thereby abrogates the effectiveness of DNA-damaging agents.³⁵ Accordingly, the development of more elaborated strategies that combine irradiation and/or alkylating chemotherapy with anti-EGFR strategies is required. Novel anti-EGFR antibodies such as the monoclonal antibody (mAb) 806 target EGFRvIII

and a subset of the overexpressed wild-type EGFR but do not interact with wild-type EGFR expressed by normal cells. The administration of mAb 806 has shown promising results in preclinical glioma models.^{36,37} However, data on its putative clinical activity are still lacking. Further strategies include the administration of antibody-drug conjugates (ADCs), which consist of an anti-EGFR antibody conjugated to potent cytotoxic drugs. ABT-414 is an ADC that interacts mainly with tumor cells expressing wild-type amplified EGFR or EGFRvIII. The activity of ABT-414 against glioblastoma is currently being tested in clinical trials.³⁸ In a similar approach, administration of a ¹²⁵I-labeled anti-EGFR antibody (¹²⁵I-mAb 425) in combination with radiation therapy did not result in better outcome compared with irradiation alone in patients with anaplastic glioma or glioblastoma.³⁹

Preclinical developments

Novel drug conjugates such as DAB389EGF, a fusion protein composed of diphtheria toxin linked to EGF, have shown activity in experimental glioma models.⁴⁰ Further approaches that have not yet reached the clinic include EGFR gene silencing by RNA interference and ribozyme-mediated cleavage of EGFR mRNA molecules.^{41,42} It needs to be awaited whether these techniques may become available for clinical testing in the future.

Resistance to Therapeutic Approaches Specifically Targeting EGFRvIII

EGFRvIII has been described as a mediator of glioma cell resistance to chemotherapeutic drugs in vitro through upregulation of the anti-apoptotic protein B-cell lymphoma–extra large.⁴³ However, the clinical impact of EGFRvIII expression on progression-free and overall survival remains controversial. A report assessing tumor samples from 73 patients revealed an association of EGFRvIII expression with prolonged overall survival.⁴⁴ These authors also reported that EGFRvIII-negative neurosphere cells are more resistant to temozolomide than EGFRvIII-positive cells, suggesting that expression of EGFRvIII rather acts as a sensitizer to alkylating drugs. Notably, very high concentrations of temozolomide were used for these in vitro studies, which precludes translation into a clinical setting. Indeed, these findings are at odds with other reports. A study by Shinjima and colleagues⁴⁵ revealed an association between EGFRvIII expression and poor overall survival in glioblastoma patients. Ultimately, a comprehensive analysis of more than 180 glioblastoma patients demonstrated that the clinical course of EGFRvIII-expressing glioblastomas was not significantly different from that of patients harboring EGFRvIII-negative tumors. However, long-term survival, defined by overall survival of more than 3 years, was virtually absent in patients with EGFRvIII-positive glioblastoma.⁴⁶

Expression of EGFRvIII also defines a subgroup of tumor cells within a glioblastoma with stem cell characteristics.⁴⁷ It was reported that EGFRvIII is coexpressed with the putative stem cell marker CD133 and that these cells are characterized by the highest degree of self-renewal as well as a pronounced tumorigenicity in vivo. On the cellular level, there is a close oncogenic signaling relationship between wild-type EGFR and EGFRvIII that drives glioblastoma progression.⁴⁸ Furthermore, a minority of cells within a glioblastoma expressing the EGFRvIII mutant may be sufficient to promote tumor growth by inducing the expression of

several cytokines such as interleukin-6 and leukemia inhibitory factor. Subsequently, these cytokines act in a paracrine manner on EGFRvIII-negative cells in the neighborhood by accelerating their proliferation.⁴⁹ Accordingly, the specific targeting of EGFRvIII may have an impact on tumor growth beyond the population of EGFRvIII-positive cells. Proteomic analyses revealed that the expression of EGFRvIII results in the activation of different downstream pathways compared with gliomas that are EGFRvIII deficient.⁵⁰ In this regard, preclinical data suggest a synergistic activity when EGFRvIII inhibition is combined with targeting of an additional pathway, such as c-MET signaling or the urokinase-type plasminogen activator receptor pathway.^{51,52} It was also reported that resistance to anti-EGFR strategies is associated with increased expression of EGFRvIII and an activation of the PI3K pathway. The latter is accompanied by an induction of the expression of the regulatory 110-kDa delta subunit of PI3K (p110δ). Preclinical findings suggest that insulin-like growth factor receptor-1 signaling via PI3K and the presence of major vault proteins, which stabilize EGFR/PI3K signaling, also contribute to resistance to anti-EGFR therapy in glioma cells.^{53,54} Silencing of EGFRvIII resulted in a sensitization to the EGFR inhibitor erlotinib. Similarly, targeting PI3K or the p110δ subunit also restored erlotinib sensitivity.⁵⁵ Considering these findings, selective interference with EGFRvIII signaling may help to overcome the treatment resistance of glioblastomas.

Immunotherapy: Escape From Vaccination Against EGFRvIII

Since EGFRvIII is exclusively expressed on tumor cells, it represents an appealing target for therapeutic interventions. In contrast to wild-type EGFR, which has been used as a point of attack for pharmacological inhibitors or antibodies, EGFRvIII has

gained additional interest as a target structure for active immunotherapy, that is, vaccination that aims at overcoming the immune evasion of glioma cells.⁵⁶ Currently, a peptide-based vaccine (CDX-110, also known as rindopepimut) is in late-stage clinical development and is being assessed in combination with standard temozolomide-based chemoradiation in patients with newly diagnosed glioblastoma with proven expression of EGFRvIII (ACT IV, NCT01480479) as well as in a phase II study in patients with relapsed EGFRvIII-positive glioblastoma (ReACT, NCT01498328). This vaccine has already shown promising results in smaller trials and prolonged survival compared with matched historical controls. However, it was also reported that the expression of EGFRvIII is lost upon vaccination with rindopepimut when tissue specimens from recurrent tumors were compared with the tumor tissue at initial diagnosis.⁵⁷ On the one hand, this may indicate the activity of the vaccine and the removal of tumor cells expressing the target antigen by the immune system. On the other hand, it also suggests immune evasion by the tumor due to loss of the target structure. This process, also known as “cancer immunoediting,” precludes durable immune responses against glioma cells unless further tumor antigens are recognized by the effector mechanisms of the immune system.⁵⁸ Currently, various strategies aimed at boosting the immune system against cancer are being investigated in clinical trials. Immune checkpoint inhibitors targeting programmed cell death-1 or cytotoxic T lymphocyte antigen-4, as well as drugs that may help to overcome the immunosuppressive environment surrounding glioma (eg, by suppressing transforming growth factor-β signaling), may allow for more powerful immune responses against gliomas. However, whether the combination of any of these novel approaches with a vaccine against EGFRvIII prevents escape from vaccination alone and results in sustained clinical benefit must be determined with clinical trials.

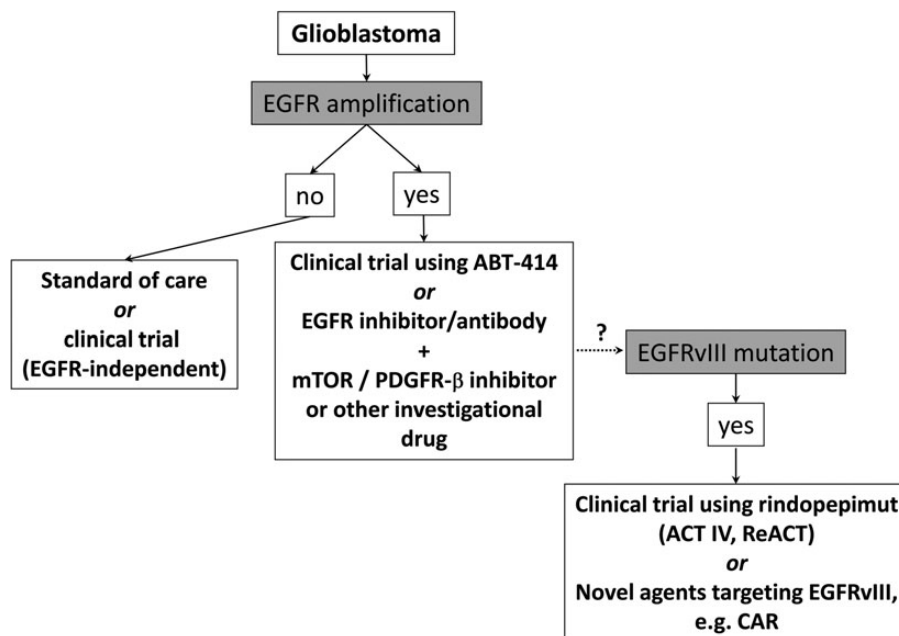


Fig. 1. Treatment approach to glioblastoma based on EGFR stratification. CAR, chimeric antigen receptor. ClinicalTrials.gov identifier: ACT IV (NCT01480479), ReACT (NCT01498328).

Outlook

The Cancer Genome Atlas has reconfirmed the EGFR gene as a principal target of mutation in glioblastoma but also illustrated the variability of alterations. The history of negative trials with EGFR-targeted agents teaches us that any future effort at exploiting this target for therapy must be based on a molecularly defined patient enrichment including at least EGFR status, but potentially also changes in associated pathways (Fig. 1). Advances with the use of anti-EGFR treatments will require stratification based on the presence of EGFR overexpression or amplification as well as the presence of the EGFRvIII mutation. Patients with tumors harboring any of these alterations may be most likely to benefit from EGFR-targeted therapies. While the fate of the immunotherapeutic efforts targeting EGFRvIII will depend on the outcome of the currently ongoing trials, the availability of ADCs, as well as the drug combinations selected upon individual tumor tissue examination, may pave the way for more successful therapies.

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References

- Weller M, van den Bent M, Hopkins K, et al. EANO guideline on the diagnosis and treatment of malignant glioma. *Lancet Oncol.* 2014; 15(9):e395–e403.
- Hatanpaa KJ, Burma S, Zhao D, et al. Epidermal growth factor receptor in glioma: signal transduction, neuropathology, imaging, and radioresistance. *Neoplasia.* 2010;12(9):675–684.
- Schneider MR, Wolf E. The epidermal growth factor receptor ligands at a glance. *J Cell Physiol.* 2009;218(3):460–466.
- Weller M, Kaulich K, Hentschel B, et al. Assessment and prognostic significance of the epidermal growth factor receptor vIII mutation in glioblastoma patients treated with concurrent and adjuvant temozolomide radiochemotherapy. *Int J Cancer.* 2014;134(10):2437–2447.
- Heimberger AB, Hlatky R, Suki D, et al. Prognostic effect of epidermal growth factor receptor and EGFRvIII in glioblastoma multiforme patients. *Clin Cancer Res.* 2005;11(4):1462–1466.
- Steinbach JP, Supra P, Huang HJ, et al. CD95-mediated apoptosis of human glioma cells: modulation by epidermal growth factor receptor activity. *Brain Pathol.* 2002;12(1):12–20.
- Steinbach JP, Supra P, Huang HJ, et al. CD95-mediated apoptosis of human glioma cells: modulation by epidermal growth factor receptor activity. *Brain Pathol.* 2002;12(1):12–20.
- Munoz JL, Rodriguez-Cruz V, Greco SJ, et al. Temozolomide resistance in glioblastoma cells occurs partly through epidermal growth factor receptor-mediated induction of connexin 43. *Cell Death Dis.* 2014;5:e1145.
- Marie Y, Carpentier AF, Omuro AM, et al. EGFR tyrosine kinase domain mutations in human gliomas. *Neurology.* 2005;64(8):1444–1445.
- Mellinghoff IK, Wang MY, Vivanco I, et al. Molecular determinants of the response of glioblastomas to EGFR kinase inhibitors. *N Engl J Med.* 2005;353(19):2012–2024.
- Vivanco I, Robins HI, Rohle D, et al. Differential sensitivity of glioma versus lung cancer-specific EGFR mutations to EGFR kinase inhibitors. *Cancer Discov.* 2012;2(5):458–471.
- Brennan CW, Verhaak RG, McKenna A, et al. The somatic genomic landscape of glioblastoma. *Cell.* 2013;155(2):462–477.
- Weiner LM, Surana R, Wang S. Monoclonal antibodies: versatile platforms for cancer immunotherapy. *Nat Rev Immunol.* 2010; 10(5):317–327.
- Lassman AB, Rossi MR, Raizer JJ, et al. Molecular study of malignant gliomas treated with epidermal growth factor receptor inhibitors: tissue analysis from North American Brain Tumor Consortium Trials 01-03 and 00-01. *Clin Cancer Res.* 2005;11(21):7841–7850.
- Stommel JM, Kimmelman AC, Ying H, et al. Coactivation of receptor tyrosine kinases affects the response of tumor cells to targeted therapies. *Science.* 2007;318(5848):287–290.
- Snuderl M, Fazlollahi L, Le LP, et al. Mosaic amplification of multiple receptor tyrosine kinase genes in glioblastoma. *Cancer Cell.* 2011; 20(6):810–817.
- Bianco R, Shin I, Ritter CA, et al. Loss of PTEN/MMAC1/TEP in EGF receptor-expressing tumor cells counteracts the antitumor action of EGFR tyrosine kinase inhibitors. *Oncogene.* 2003;22(18):2812–2822.
- Wang MY, Lu KV, Zhu S, et al. Mammalian target of rapamycin inhibition promotes response to epidermal growth factor receptor kinase inhibitors in PTEN-deficient and PTEN-intact glioblastoma cells. *Cancer Res.* 2006;66(16):7864–7869.
- Fan QW, Cheng CK, Nicolaides TP, et al. A dual phosphoinositide-3-kinase alpha/mTOR inhibitor cooperates with blockade of epidermal growth factor receptor in PTEN-mutant glioma. *Cancer Res.* 2007;67(17):7960–7965.
- Fenton TR, Nathanson D, Ponte de Albuquerque C, et al. Resistance to EGF receptor inhibitors in glioblastoma mediated by phosphorylation of the PTEN tumor suppressor at tyrosine 240. *Proc Natl Acad Sci U S A.* 2012;109(35):14164–14169.
- Li J, Zhu S, Kozono D, et al. Genome-wide shRNA screen revealed integrated mitogenic signaling between dopamine receptor D2 (DRD2) and epidermal growth factor receptor (EGFR) in glioblastoma. *Oncotarget.* 2014;5(4):882–893.
- Akhavan D, Pourzia AL, Nourian AA, et al. De-repression of PDGFR β transcription promotes acquired resistance to EGFR tyrosine kinase inhibitors in glioblastoma patients. *Cancer Discov.* 2013;3(5):534–547.
- Iwanami A, Gini B, Zanca C, et al. PML mediates glioblastoma resistance to mammalian target of rapamycin (mTOR)-targeted therapies. *Proc Natl Acad Sci U S A.* 2013;110(11):4339–4344.
- Srikanth M, Das S, Berns EJ, et al. Nanofiber-mediated inhibition of focal adhesion kinase sensitizes glioma stemlike cells to epidermal growth factor receptor inhibition. *Neuro Oncol.* 2013;15(3):319–329.
- Eimer S, Dugay F, Airiau K, et al. Cyclopamine cooperates with EGFR inhibition to deplete stem-like cancer cells in glioblastoma-derived spheroid cultures. *Neuro Oncol.* 2012;14(12):1441–1451.
- Geoerger B, Gaspar N, Opolon P, et al. EGFR tyrosine kinase inhibition radiosensitizes and induces apoptosis in malignant glioma and childhood ependymoma xenografts. *Int J Cancer.* 2008;123(1):209–216.
- Krishnan S, Brown PD, Ballman KV, et al. Phase I trial of erlotinib with radiation therapy in patients with glioblastoma multiforme: results

- of North Central Cancer Treatment Group protocol N0177. *Int J Radiat Oncol Biol Phys*. 2006;65(4):1192–1199.
28. Chakravarti A, Wang M, Robins HI, et al. RTOG 0211: a phase 1/2 study of radiation therapy with concurrent gefitinib for newly diagnosed glioblastoma patients. *Int J Radiat Oncol Biol Phys*. 2013;85(5):1206–1211.
 29. Prados MD, Chang SM, Butowski N, et al. Phase II study of erlotinib plus temozolomide during and after radiation therapy in patients with newly diagnosed glioblastoma multiforme or gliosarcoma. *J Clin Oncol*. 2009;27(4):579–584.
 30. Reardon DA, Groves MD, Wen PY, et al. A phase I/II trial of pazopanib in combination with lapatinib in adult patients with relapsed malignant glioma. *Clin Cancer Res*. 2013;19(4):900–908.
 31. Kreisl TN, Lassman AB, Mischel PS, et al. A pilot study of everolimus and gefitinib in the treatment of recurrent glioblastoma (GBM). *J Neurooncol*. 2009;92(1):99–105.
 32. Wen PY, Chang SM, Lamborn KR, et al. Phase I/II study of erlotinib and temsirolimus for patients with recurrent malignant gliomas: North American Brain Tumor Consortium trial 04-02. *Neuro Oncol*. 2014;16(4):567–578.
 33. Eisenstat DD, Nabors L, Mason W, et al. A phase II study of daily afatinib (BIBW 2992) with or without temozolomide (21/28 days) in the treatment of patients with recurrent glioblastoma. *J Clin Oncol*. 2011;29(suppl; abstr 2010).
 34. Solomon MT, Selva JC, Figueredo J, et al. Radiotherapy plus nimotuzumab or placebo in the treatment of high grade glioma patients: results from a randomized, double blind trial. *BMC Cancer*. 2013;13:299.
 35. Bartels U, Wolff J, Gore L, et al. Phase 2 study of safety and efficacy of nimotuzumab in pediatric patients with progressive diffuse intrinsic pontine glioma. *Neuro Oncol*. 2014.
 36. Weinandy A, Piroth MD, Goswami A, et al. Cetuximab induces eme1-mediated DNA repair: a novel mechanism for cetuximab resistance. *Neoplasia*. 2014;16(3):207–220.
 37. Johns TG, Perera RM, Vernes SC, et al. The efficacy of epidermal growth factor receptor-specific antibodies against glioma xenografts is influenced by receptor levels, activation status, and heterodimerization. *Clin Cancer Res*. 2007;13(6):1911–1925.
 38. Mishima K, Johns TG, Luwor RB, et al. Growth suppression of intracranial xenografted glioblastomas overexpressing mutant epidermal growth factor receptors by systemic administration of monoclonal antibody (mAb) 806, a novel monoclonal antibody directed to the receptor. *Cancer Res*. 2001;61(14):5349–5354.
 39. Gan HK, Fichtel L, Lassman AB, et al. A phase 1 study evaluating ABT-414 in combination with temozolomide (TMZ) for subjects with recurrent or unresectable glioblastoma (GBM). *J Clin Oncol*. 2014;32:5s(suppl; abstr 2021).
 40. Wygoda Z, Kula D, Bierzynska-Macyszyn G, et al. Use of monoclonal anti-EGFR antibody in the radioimmunotherapy of malignant gliomas in the context of EGFR expression in grade III and IV tumors. *Hybridoma*. 2006;25(3):125–132.
 41. Liu TF, Hall PD, Cohen KA, et al. Interstitial diphtheria toxin-epidermal growth factor fusion protein therapy produces regressions of subcutaneous human glioblastoma multiforme tumors in athymic nude mice. *Clin Cancer Res*. 2005;11(1):329–334.
 42. Yamazaki H, Kijima H, Ohnishi Y, et al. Inhibition of tumor growth by ribozyme-mediated suppression of aberrant epidermal growth factor receptor gene expression. *J Natl Cancer Inst*. 1998;90(8):581–587.
 43. Kang CS, Pu PY, Li YH, et al. An in vitro study on the suppressive effect of glioma cell growth induced by plasmid-based small interference RNA (siRNA) targeting human epidermal growth factor receptor. *J Neurooncol*. 2005;74(3):267–273.
 44. Nagane M, Levitzki A, Gazit A, et al. Drug resistance of human glioblastoma cells conferred by a tumor-specific mutant epidermal growth factor receptor through modulation of Bcl-XL and caspase-3-like proteases. *Proc Natl Acad Sci U S A*. 1998;95(10):5724–5729.
 45. Montano N, Cenci T, Martini M, et al. Expression of EGFRvIII in glioblastoma: prognostic significance revisited. *Neoplasia*. 2011;13(12):1113–1121.
 46. Shinjima N, Tada K, Shiraishi S, et al. Prognostic value of epidermal growth factor receptor in patients with glioblastoma multiforme. *Cancer Res*. 2003;63(20):6962–6970.
 47. Emllet DR, Gupta P, Holgado-Madruga M, et al. Targeting a glioblastoma cancer stem-cell population defined by EGF receptor variant III. *Cancer Res*. 2014;74(4):1238–1249.
 48. Fan QW, Cheng CK, Gustafson WC, et al. EGFR phosphorylates tumor-derived EGFRvIII driving STAT3/5 and progression in glioblastoma. *Cancer Cell*. 2013;24(4):438–449.
 49. Inda MM, Bonavia R, Mukasa A, et al. Tumor heterogeneity is an active process maintained by a mutant EGFR-induced cytokine circuit in glioblastoma. *Genes Dev*. 2010;24(16):1731–1745.
 50. Johnson H, Del Rosario AM, Bryson BD, et al. Molecular characterization of EGFR and EGFRvIII signaling networks in human glioblastoma tumor xenografts. *Mol Cell Proteomics*. 2012;11(12):1724–1740.
 51. Lal B, Goodwin CR, Sang Y, et al. EGFRvIII and c-Met pathway inhibitors synergize against PTEN-null/EGFRvIII+ glioblastoma xenografts. *Mol Cancer Ther*. 2009;8(7):1751–1760.
 52. Hu J, Jo M, Cavenee WK, et al. Crosstalk between the urokinase-type plasminogen activator receptor and EGF receptor variant III supports survival and growth of glioblastoma cells. *Proc Natl Acad Sci USA*. 2011;108(38):15984–15989.
 53. Chakravarti A, Loeffler JS, Dyson NJ. Insulin-like growth factor receptor I mediates resistance to anti-epidermal growth factor receptor therapy in primary human glioblastoma cells through continued activation of phosphoinositide 3-kinase signaling. *Cancer Res*. 2002;62(1):200–207.
 54. Lotsch D, Steiner E, Holzmann K, et al. Major vault protein supports glioblastoma survival and migration by upregulating the EGFR/PI3K signalling axis. *Oncotarget*. 2013;4(11):1904–1918.
 55. Schulte A, Liffers K, Kathagen A, et al. Erlotinib resistance in EGFR-amplified glioblastoma cells is associated with upregulation of EGFRvIII and PI3Kp110delta. *Neuro Oncol*. 2013;15(10):1289–1301.
 56. Roth P, Eisele G, Weller M. Immunology of brain tumors. *Handb Clin Neurol*. 2012;104:45–51.
 57. Sampson JH, Heimberger AB, Archer GE, et al. Immunologic escape after prolonged progression-free survival with epidermal growth factor receptor variant III peptide vaccination in patients with newly diagnosed glioblastoma. *J Clin Oncol*. 2010;28(31):4722–4729.
 58. Pellegatta S, Cuppini L, Finocchiaro G. Brain cancer immunoediting: novel examples provided by immunotherapy of malignant gliomas. *Expert Rev Anticancer Ther*. 2011;11(11):1759–1774.