

Video Article

Milk Collection Methods for Mice and Reeves' Muntjac Deer

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Abstract

Animal models are commonly used throughout research laboratories to accomplish what would normally be considered impractical in a pathogen's native host. Milk collection from animals allows scientists the opportunity to study many aspects of reproduction including vertical transmission, passive immunity, mammary gland biology, and lactation. Obtaining adequate volumes of milk for these studies is a challenging task, especially from small animal models. Here we illustrate an inexpensive and facile method for milk collection in mice and Reeves' muntjac deer that does not require specialized equipment or extensive training. This particular method requires two researchers: one to express the milk and to stabilize the animal, and one to collect the milk in an appropriate container from either a Muntjac or mouse model. The mouse model also requires the use of a P-200 pipetman and corresponding pipette tips. While this method is low cost and relatively easy to perform, researchers should be advised that anesthetizing the animal is required for optimal milk collection.

Video Link

The video component of this article can be found at <http://www.jove.com/video/51007/>

Introduction

Animal models provide insight into disease pathology that cannot be gained by *in vitro* analysis. To provide the most efficacious results, it is important to use an animal model that is closely related to the disease and species of interest. For example, the Reeves' muntjac (*Muntiacus reevesi*), a small Asian deer^{1,2}, and transgenic mice expressing the cervid prion protein (CerTgPrP)³ are useful animal models for cervid species. Both species are polyestrous, allowing year-round breeding, and therefore a consistent source for pregnancy-related tissues and fluids to study specific mechanisms in cervid biology. Studies of milk have a vast array of applications that are more simply (and inexpensively) accomplished in animal models than in humans. Researchers could investigate milk and colostrum as a potential source of 1) infectious disease transmission, 2) immunoglobulins transferred from mother to offspring in the development of passive immunity⁴ and 3) lactoferrin, a protein found in human breast milk involved in passive immunity that researchers are currently attempting to commercially produce⁵.

Collecting a substantial amount of milk from small animals can prove to be a difficult task. Rogers proposed an approach to collect milk from rats⁶, which was subsequently used in mice. DePeters and Hovey proposed two methods for milk collection, one using a manually-generated vacuum produced by a rubber pipette bulb attached to a Pasteur pipette, and a second requiring the construction of a milking unit, which is then attached to a vacuum source (such as a faucet) to harvest mouse milk⁷. Here we propose a simple, low cost method for collecting milk from both mice and Reeves' muntjac deer, which requires only readily available laboratory equipment and basic technical skills. Our method yields sufficient volumes of milk for various applications.

Protocol

1. Mouse Milking Protocol

The methods described in this protocol were approved by IACUC.

1. Separation of the Dam from Offspring

1. Select the dam to be milked. Choose a dam with a litter of 4 or more pups that is 8-12 days post parturition to provide maximal milk collection—although collection is possible at any time point post parturition up to 21 days.
2. Separate the dam from her litter at least 2 hr before milking.
Note: Should one desire to milk more than one dam in a given time period, it is acceptable to house the separated dams in the same cage with adequate food and water supply.

2. Administration of Oxytocin, Anesthesia and Eye Lubricant

1. Administer 2 IU/kg of oxytocin intraperitoneally (IP). Oxytocin is a hormone that acts on the mammary glands of lactating females to stimulate the release of milk. Oxytocin can be acquired from a veterinarian.
2. Administer an anesthetic mixture containing 80-100 mg/kg ketamine in combination with 5-10 mg/kg xylazine IP. For example, a 10 ml stock anesthetic mixture could be prepared in advance for mice weighing approximately 30 g and would include 0.1 ml xylazine, 8.9 ml H₂O, and 1.0 ml ketamine. 0.25 ml total anesthetic will sedate a mouse of this size for approximately 20-30 min, which should provide adequate time for milk collection.

Notes: To determine that the animal has been successfully anesthetized, gently place her on her stomach on a solid surface; she should not be moving her feet or attempting to rise to her feet if she is fully anesthetized. In the case of a mouse not being fully anesthetized with the original dosage, the researcher may administer another ¼ or ½ of the original dose of the anesthetic mixture depending on the size of the mouse and/or level of sedation that the mouse has already achieved. However, this could increase the chance of an adverse reaction to anesthesia, and also may result in the mouse being more deeply anesthetized and/or anesthetized for a longer period of time.

Note: If milking more than one dam, it is necessary to stagger oxytocin and anesthetic injections due to the short half-life of oxytocin and sedation times for ketamine and xylazine.

3. Apply a small amount of eye lubricant to the corner of each eye and spread to prevent the mouse's eyes from drying out while under anesthesia.

3. Milk Collection

The milk collection is most easily performed with two researchers: one researcher to hold the anesthetized mouse while manually expressing the milk (referred to as R1), and one researcher to collect the milk (R2). This method can also be performed with one person, if the mouse is secured on a flat surface to avoid harm.

1. R2 should be equipped with sterile alcohol prep pads, a P-200 pipetman, a clean pipette tip for each mouse, and a container to hold the milk, such as a 1.5 ml Eppendorf tube or a 1.2 ml cryovial (see Table of Materials).
2. As the oxytocin begins to take effect, milk letdown will become visible in the mouse's mammary area (see **Figure 1A**). R1: Unwrap a sterile alcohol prep pad and gently wipe the mammary area of the mouse to clean the area prior to milking. Manually express milk from the teat by using the thumb and forefinger to gently massage and squeeze the mammary tissue in an upward motion until a visible bead of milk begins to form at the base of the teat.
3. Press the P-200 pipetman plunger to its first stop to release air out of the pipette tip and prepare for milk collection.
4. Position the pipette tip at the top of the drop of milk, and gently pull the milk into the pipette tip by slowly releasing pressure on the plunger. Take care not to put the tip too close to the teat or the skin (see **Figure 1B**). It is not necessary to completely release the plunger as this will pull the milk all the way to the top of the pipette tip and will make it difficult to expel the often viscous milk into the holding container.
5. Expel the milk into the container by using the thumb to press the plunger past the first stop (see **Figure 1C**). To facilitate expulsion of small volumes, press the pipette tip against the side of the tube so that no liquid is lost.
6. Continue to manually express milk from each teat, moving in either a clockwise or counterclockwise direction so as not to skip any teats. It is both acceptable and useful to come back to and continually express milk from the same teat should it produce more milk than another.

4. Anesthetic Reversal in the Dam

1. In most cases, it is common for the dam to begin to wake up during the milking process. If this is not the case, place the dam in a cage indirectly underneath a heat source, such as a heated circulating blanket, until she begins to wake up.
Note: As the dam begins to wake up, keep close observation of her activity until she is able to walk about her cage on her own.
2. Once the dam is moving about the cage without struggle, place her back into her original cage with her litter. No ill effects have been noted with respect to the pups' ability to feed after milk collection.

2. Muntjac Milking Protocol

The methods described in this protocol were approved by IACUC.

1. Separation of the Doe from Offspring

1. Select the doe to be milked. Reeves' muntjac deer generally give birth to one fawn². Milk the doe the day after birthing to allow the fawn opportunity to gain access to essential nutrients in colostrum⁸.
Note: The fawn usually hides and does not reappear during the procedure; separation of the mother-offspring pair is not necessary.

2. Administration of Oxytocin, Anesthesia and Eye Lubricant

1. Administer 10 IU/kg of Oxytocin intramuscularly (IM). Oxytocin is a hormone that acts on the mammary glands of lactating females to stimulate the let-down of milk. Oxytocin can be acquired from a veterinarian.
2. Administer the BAM anesthetic IM¹¹ containing butorphanol 0.45 mg/kg, azaperone 0.25 mg/kg, medetomidine 0.07 mg/kg. One dose of the BAM anesthetic mixture will sedate a muntjac for approximately 20-30 min, which provides ample time for complete milk collection. Another option for anesthetic is midazolam at 1-2 mg/kg, which will sedate a muntjac for approximately 10-15 min. While this provides adequate time for milk collection, this anesthetic is metabolized more quickly than BAM and the animal may attempt to escape at the end of the shorter time period.

Note: Minimal response to pain, minor reflexes upon touching the ears, followed by the ability of R1 to hold the MJ without struggle indicates that the animal has been successfully anesthetized.

Note: In the case of a muntjac not being fully anesthetized with the original dosage, the researcher may administer another ¼ or ½ of the original dose of the anesthetic mixture depending on the size of the animal and/or level of sedation that has already been achieved. However,

this could increase the chance of an adverse reaction to anesthesia, and also may result in the muntjac being more deeply anesthetized and/or anesthetized for a longer period of time.

3. Apply a small amount of lubricant to the corner of each eye and spread to prevent the muntjac's eyes from drying out while under anesthesia.

3. Milk Collection

The milk collection is most easily performed with two researchers; one researcher to hold the anesthetized muntjac in the lap while manually expressing the milk (Referred to as R1), and a second researcher to keep the muntjac's head in a comfortable and safe position and to collect the milk (R2).

1. Depending upon the anesthetic used, milking should be performed either 1) in their pens to provide a safe place for them should they wake up during the procedure (midazolam-induced) or 2) in a bedding-free anteroom to maintain cleanliness (BAM-induced).
2. R1 and R2 should be equipped with ethanol-soaked gauze and several 15 ml conical polypropylene tubes to hold the milk.
3. As the oxytocin begins to take effect, milk letdown will become visible in the muntjac's mammary area. R1: Sanitize gloved hands and fingers, as well as the entire udder and each individual teat of the animal with the ethanol-soaked gauze. R2: Hold the muntjac's head in a safe and comfortable position to allow ease of breathing and help prevent rumen regurgitation. Hold the collection tube close to the teat being milked without touching it to prevent potential contamination.
4. Express milk from one teat by using the thumb and forefinger to gently squeeze the mammary tissue in an upward motion until the teat is engorged with milk, and gently roll the milk out of the teat and into the tube that R2 is holding (see **Figure 2**).
5. Note: The milk comes out in a stream, so it is important for R2 to have the collection tube positioned so that little to no milk will be wasted.
6. The muntjac's udder is divided into quadrants. R1: Continue to express milk from each quadrant until the milk no longer flows easily. If milk cannot be expressed from one teat, express from all other teats before returning to the first. It is both acceptable and useful to come back to and continually express milk from the same teat should it produce more milk than another.

4. Anesthetic Reversal in the Doe

It is common for midazolam-induced doe to begin to wake up during the milking process. In most cases, the doe will attempt to escape before all the milk has been expressed. Minor restraint is used in an effort to increase the milk yield; however the safety and comfort of both the researchers and the doe are equally important.

1. Administer atipamezole at 2.5 mg/mg medetomidine subcutaneously for rapid reversal of BAM anesthetized doe.
Note: With both types of anesthetic, it is necessary to maintain close observation of the doe until she is able to walk on her own without stumbling or falling, and is able to eat and drink without fear of aspiration or choking. No ill effects have been noted with respect to the fawn's ability to nurse after milk collection.

Representative Results

Mouse

From our experiments, we have determined that it is possible to collect approximately 100-400 μ l of milk from one laboratory mouse, dependent on several variables. These variables include 1) the amount of time set aside for collection, 2) the dose of oxytocin administered, 3) how many pups the dam is currently nursing and 4) the amount of time post parturition. Our studies have shown that the highest yield is obtained when the dams are separated from their offspring for at least 2 hr, and at least 45 min to an hour is set aside per mouse for milking. Maximum milk volumes were harvested 8-12 days post parturition using 2 IU/kg of oxytocin. Our results are mostly in accordance with other studies where litter size was positively correlated with milk yield⁹ (see **Figure 3**).

Muntjac

Although the amount of milk harvested from the muntjac was variable from day to day, we were able to collect 5 to 30 ml milk/dam/session, depending on the anesthesia used and/or time post parturition. Muntjac deer are polyestrous; milk can be collected at various time points throughout the year² (see **Table 1**). Midazolam-induced doe remained under anesthesia for as little as 10 min, allowing for smaller volume collections (typically 5-30 ml), whereas BAM-induced doe sustained a longer period of anesthetic effect (20-30 min), allowing complete milk expression (15-130 ml). Milk collections were discontinued when milk was no longer being produced in sufficient amounts by the doe, and the corresponding fawn was being nourished on a diet of mostly hay forage.

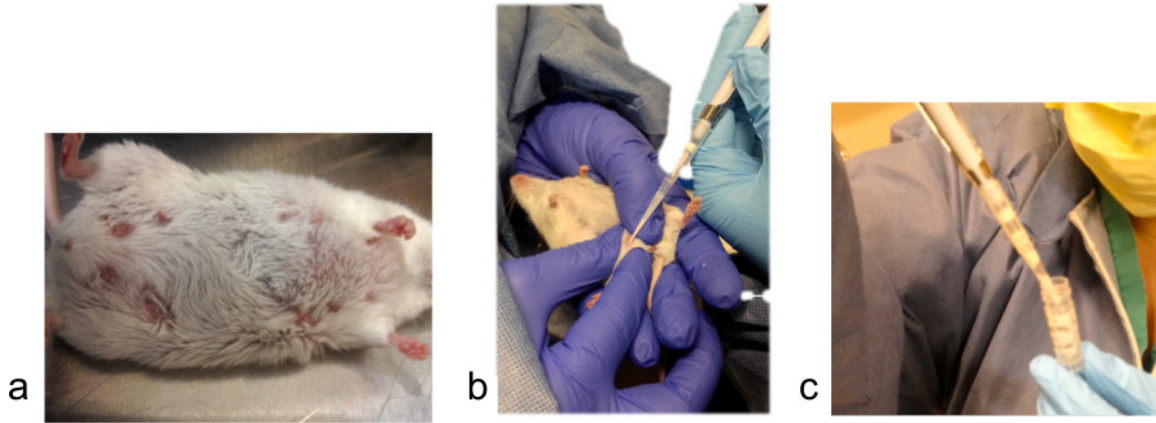


Figure 1. (A) The dam's teats become engorged with milk upon oxytocin injection. (B) Collection of milk from an anesthetized dam using a P-200 pipetman. (C) Ejection of milk into a 1.2 ml cryovial.



Figure 2. Muntjac milk is collected into a 15 ml conical tube.

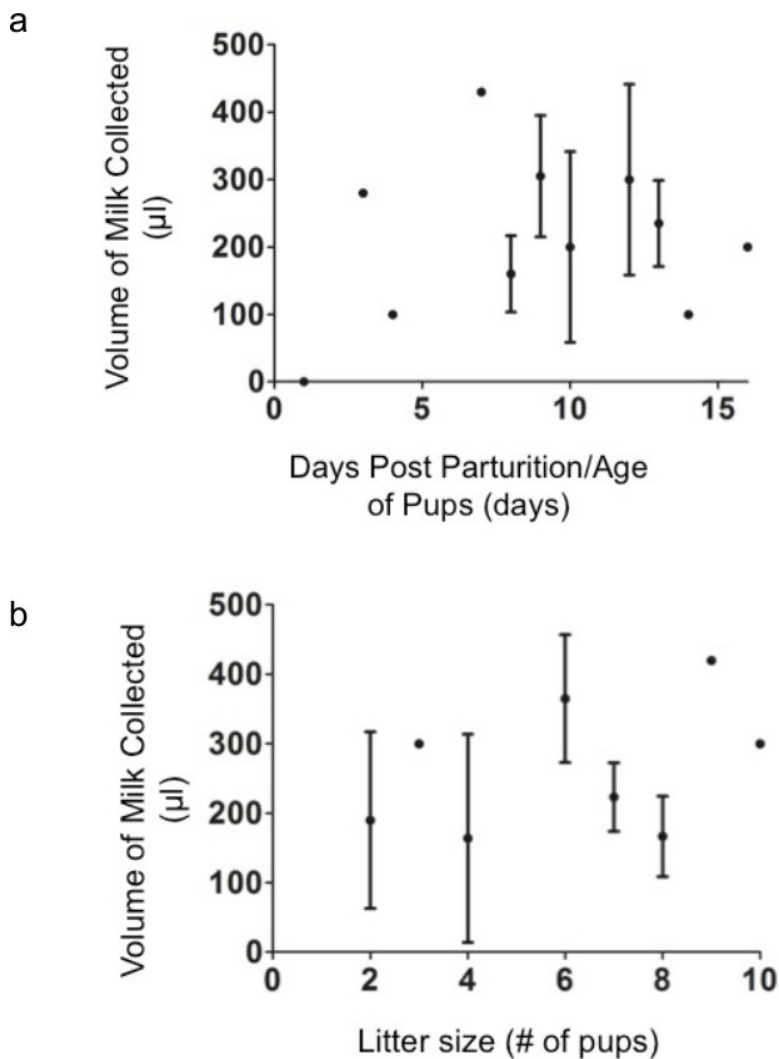


Figure 3. Approximate milk yield collected from mice based on litter size, and age of pups.

Muntjac #	Date started	Date Finished	Total days
19	06.10.08	11.06.08	150
16	07.02.08	11.06.08	128
11	05.05.09	06.25.09	52
19	04.01.10	05.20.10	50
44	04.06.10	04.23.10	18
45	04.06.10	05.20.10	44
77	01.05.12	04.19.12	106
75	01.03.12	04.19.12	108
73	01.03.12	04.19.12	108
76	02.08.12	04.19.12	71
111	09.13.12	10.30.12	48
Average			80.27

Table 1. Milk collection start and finish dates for lactating Reeves' muntjac dams.

Discussion

Mouse

There are several factors to take into consideration when collecting milk from a mouse, including 1) the amount of time set aside for collection, 2) the dose of oxytocin administered, 3) how many pups the dam is currently nursing and 4) the amount of time that has passed since parturition at the time of collection. Using previous studies as guidelines, we set out to optimize conditions for milk yield.

Previous research has directly correlated milk yield with litter size⁸. Unfortunately, litter size is a variable that cannot be controlled. However, the time of milking post parturition can be controlled. According to *The Laboratory Mouse* maximum milk secretion is at 8-10 days post parturition⁹. Our experience is in congruence with this statement.

It has been shown that oxytocin plays an irreplaceable role in milk production post parturition¹⁰. Oxytocin doses ranging from 0.1 IU/kg⁹ to 4 IU/kg⁶ have been reported for collection of milk from rodents^{6,7,9}. We began our study by administering the lowest dose suggested, 0.1 IU/kg⁹. Using this dosage, the amount of milk produced by the mouse was too small to measure or collect. Upon increasing the oxytocin dose to 2 IU/kg^{6,7}, milk yield was considerably higher at approximately 120 μ l. Because we wanted to use the lowest dosage of oxytocin to achieve optimal milk collection, we did not administer doses higher than 2 IU/kg.

Milk yield is also dependent upon the amount of time set aside for collection. We have determined that a minimum of 3 hr is necessary for optimal collection—2 hr for the pups to be separated from the dam and 1 hr for material preparation and collection. Collecting milk from several dams in one session will decrease the overall time necessary to harvest a specific volume of milk, *i.e.*, 2 hr to separate pups, 30 min to setup materials and 20-30 min to collect milk from each dam. Each dam must be monitored until they are awake before being placed back with their pups. We have noted that the more time set aside for milking leads to higher milk yields, because the mouse can be milked until she regains consciousness.

Instances do arise with animal models that will affect milk yield. One common issue is the presence of blood or exudate in the milk of older dams that have been bred repeatedly and have large litters, or in dams that are nursing their first litter. Should this occur, we suggest discontinuing milk collection from that particular teat. It is advisable to switch pipet tips at this time, and discard any exudate or blood.

Researchers carrying out this method should also be aware that, as is often the case when using animal models, results vary (**Figure 3**). If a less than optimal amount of milk is collected in one session, it is preferable to repeat the procedure at a later time (*e.g.* 24-48 hr) rather than alter the experimental conditions to force milk release.

Muntjac

The volume of milk collected from Reeves' muntjac dams was dependent upon the anesthetic used. Our initial anesthetic of choice was midazolam. Midazolam provided sufficient sedation to allow restraint for short periods of time—*i.e.*, if R1 had a calm disposition and could keep the muntjac calm, R2 could take over the milking to increase the yield before the muntjac would awaken—the safety and comfort of both animal and handler were of utmost importance and we abandoned the milking effort if we felt either was compromised. We discovered that muntjac doe quickly become tolerant to midazolam and that higher doses would be necessary after repeated exposure, making milk collection twice weekly increasingly difficult.

The shortage of midazolam, coupled with a building tolerance, led us to research an alternative anesthetic. A mixture of Butorphanol, Azaperone and Medetomidine (BAM) is effectively used for the immobilization of white tail deer¹¹. In our hands, the BAM cocktail provided sufficient anesthesia to allow collection of acceptable volumes of milk in a safe manner, with less drug tolerance after repeated use, and with better control in reversing the effects of the anesthetic.

Some muntjac dams suffered from sensitive udders and/or were first-time mothers. These muntjac developed bruising on the teats during the milking process, and the milk flow slowed or stopped completely, prompting us to discontinue the ongoing session. Should this occur, one should use a more gentle milking technique in an attempt to prevent discomfort or bruising in the teats.

Milk production appeared to wax and wane on several occasions. Low milk harvests may be caused by 1) the fawns having nursed just prior to collections or 2) a decline in milk production due to the natural process of fawn weaning.

While it can be a trying process to obtain a sizeable volume of milk from small animal models, our work illustrates that it is indeed a possibility for both mouse and muntjac models; neither requiring extensive skill nor deep pockets, simply perseverance.

Disclosures

The authors have nothing to disclose.

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