

SHORT REPORT

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High lipid storage in vacuolar forms of subtype 6 *Blastocystis* sp. in ostrich

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Abstract

Background: *Blastocystis* sp., a widely prevalent intestinal protozoan parasite is found in a wide range of animals, including humans. The possibility of zoonotic transmission to human from birds especially ostriches led us to investigate on the cross infectivity of *Blastocystis* sp. isolated from the ostrich feces as well as the phenotypic and subtype characteristics. There is a need to investigate this especially with the rising number of ostrich farms due to the growing global ostrich industry.

Findings: 100% of the ostriches were found to be positive for *Blastocystis* sp. using the *in-vitro* cultivation method. Transmission electron microscopy revealed high electron dense material in the central body of the vacuolar forms. The membrane layer of the ostrich isolate was significantly ($p = 0.003$) thicker as compared to human isolate. Sudan staining revealed that this was lipid accumulation. We provide evidence for the first time, the existence of subtype 6 which has been previously reported only in pigs and cattle. Cysts, ranging from 3.0 to 7.0 μm in diameter caused experimental infection in Sprague Dawley rats implicating that *Blastocystis* sp. isolated from ostriches exhibits low host specificity.

Conclusion: The study for the first time demonstrates that *Blastocystis* sp. subtype 6 do exist in ostriches and show high lipid storage in the vacuoles of the parasites. The study further provides evidence for potential zoonotic transmission in ostrich farms as *Blastocystis* subtype 6 can infect rats and the same subtype have been previously reported in humans.

Keywords: *Blastocystis* sp, Ultrastructure, Subtype, Host susceptibility

Findings

The increasing number of ostrich farms led us to investigate on the possibility of cross infectivity of *Blastocystis* sp. isolated from the ostrich feces as well elucidate phenotypic and subtype characteristics. 37 fresh fecal samples of ostriches (*Struthio camelus*) and fresh human fecal sample were collected from a local ostrich farm and from an asymptomatic individual infected with *Blastocystis* sp. respectively from one of the states in Malaysia and cultured using the *in-vitro* culture technique using Jones' medium and sequenced-tagged site (STS) primer-polymerase chain reaction using ten sets of primers for subtype analysis. 100% of the ostriches were found to be positive for *Blastocystis* sp.

using the *in-vitro* cultivation method. Transmission electron microscopy revealed high electron dense material in the central body of the vacuolar forms. We provide evidence for the first time, the existence of subtype 6 in ostriches which showed high lipid storage. Cysts, ranging from 3.0 to 7.0 μm in diameter caused experimental infection in Sprague Dawley rats implicating that *Blastocystis* sp. isolated from ostriches exhibits low host specificity. The study further provides evidence for potential zoonotic transmission in ostrich farms as *Blastocystis* subtype 6 can infect rats and the same subtype have been previously reported in humans.

Background

Blastocystis sp. is a widely prevalent intestinal protozoan parasite seen in a wide of a range of animals, including humans. Previous publications have shown *Blastocystis*

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sp. in non-human primates, birds, chickens, ducks, geese, ostriches, amphibians, reptiles, fish, arthropods and annelids [1-6].

There are of 17 distinct subtypes (ST1-ST17) seen in humans, non-human primates, other mammals and birds [7-9]. Although only some subtypes are described to be host specific especially subtype 6 which are commonly found in pigs and cattle, however most of the subtypes are shown to exhibit low host specificity [10] which may play a role in the cross-transmission between animal and human especially humans with histories of close association to pets or farm animals [11]. Transmission is through the fecal-oral route, though water-borne, food borne and sexual transmission have been reported [12-15].

Despite a few studies showing *Blastocystis* in ostriches [16-19], there has been none that has elucidated the ultrastructural details and subtype characterization. As ostrich farming industry is increasing worldwide including Malaysia due to its economic sale of its meat, feathers, oil and leather, it is vital to ascertain of the possible potential zoonotic transmission that can be transmitted to man. It is also possible that rats can acquire the infection by traveling to human dwelling. The study attempts to further elucidate information pertaining to ultrastructural, subtype and host susceptibility of *Blastocystis* sp. isolated from domestic ostriches.

Materials and methods

Ethical approval

All animals used in this study were handled according to Institutional Animal Care and Use Committee (IACUC), University Malaya guidelines with the Reference. No: PAR/29/06/2012/LIL(R) and PAR/23/05/2013/HC(R). Human ethical approval for this study was obtained in accordance with University Malaya Medical Centre research policy with Reference. No: 926.7.

Animals and management

A local ostrich farm from one of the states in Malaysia was selected for the study. The farm practiced intensive type of management where the animals are confined in the same pen. Ostriches were kept in pairs or three's for breeding purpose in well-fenced housing pens. The animals were fed with quail layer mash, napier grass and given ad libitum water.

Source of *Blastocystis* sp. isolates

A total of thirty-seven (n = 37) fresh fecal samples of ostriches (*Struthio camelus*) and fresh human fecal sample were collected from a local ostrich farm and from an asymptomatic individual infected with *Blastocystis* sp. respectively from one of the states in Malaysia. The

samples were collected in stool collection container and were processed as soon after collection.

Laboratory testing

In vitro cultivation of *Blastocystis* sp. isolates

The parasites were isolated from the fecal samples of ostriches and human by *in-vitro* cultivation using Jones' medium supplemented with 10% heat-inactivated horse serum at 37°C. Subsequently after isolation, the parasites were maintained in Jones' medium by consecutive sub-cultures every 3 to 4 days for at least one month prior to phenotypic, subtype and ultrastructural analysis [6,20].

Floatation method

Approximately one gram of fresh feces, emulsified with saturated salt solution was then filtered through gauze into a centrifuge tube. Saturated salt solution was then later added up to the meniscus of test tube before lowering a coverslip onto the top of the tube. Coverslip was then lifted vertically up and placed onto a clean slide. Samples were observed under 10x objective lens of compound microscope for the presence of helminth ova, nematode eggs, coccidia oocysts and other parasites [21].

Transmission electron microscopy

Parasites isolated from ostriches and human were chosen for the ultrastructural studies. The contents of day 3 culture were washed three times using phosphate buffered saline (PBS) pH 7.4. The samples were centrifuged at 3000rpm for 5 minutes. The pelleted cells were re-suspended overnight in 2.5% glutaraldehyde in 0.1 M sodium cacodylate buffer, pH 7.3 at 4°C, washed thoroughly with cacodylate buffer and post fixed for 30 min in 1% osmium tetroxide in cacodylate buffer. The fixed cells were dehydrated for 5 minutes in ascending series of ethanols (30%, 50%, 70%, 80%, 90% and 100%) and embedded in epoxy resin. Semi-thin sections were stained with toluidine blue. Ultrathin sections were cut, contrasted with uranyl acetate and lead citrate and viewed using a transmission electron microscope (LEO Libra120) [22].

Sudan Black B staining

Parasites from day 3 culture of the ostrich and human isolates were smeared on a clean glass slide and immediately dried with a hair dryer at room temperature. Then the cells were fixed with 4% glutaraldehyde fixative solution in borate buffer, pH 7.6 for 1 minute at 2-8°C with gentle agitation followed by thorough rinsing in deionized water. The cells were then stained with Sudan Black B reagent by immersing slides for 5 minutes with intermittent agitation. The cells were then

Table 1 List of sequenced-tagged site (STS) primers

Subtype	STS primer	Product size (bp)	Sequence of forward (F) and reverse (R) primer (5' – 3')	Genebank accession no.
1	SB82	462	F-TCTTGCTTCATCGGAGTC R-CCTTCTCGCAGTTCTTTATC	AF166085
1	SB83	351	F-GAAGGACTCTCTGACGATGA R-GTCCAAATGAAAGGCAGC	AF166086
2	SB155	650	F-ATCAGCCTACAATCTCCTC R-ATCGCCACTTCTCCAAT	AF166087
3	SB227	526	F-TAGGATTTGGTGTTTGGAGA R-TTAGAAGTGAAGGAGATGGAAG	AF166088
3	SB228	473	F-GACTCCAGAACTCGCAGAC R-TCTTGTTTCCCCAGTTATCC	AF166089
3	SB229	631	F-CACTGTGTCGTCATTGTTTTG R-AGGGCTGCATAATAGAGTGG	AF166090
4	SB332	338	F-GCATCCAGACTACTATCAACATT R-CCATTTTCAGACAACCACTTA	AF166091
5	SB340	704	F-TGTTCTTGTCCTCTCAGCTC R-TTCTTTCACACTCCCGTCAT	AY048752
6	SB336	317	F-GTGGGTAGAGGAAGGAAAACA R-GAACAAGTCGATGAAGTGAGAT	AY048751
7	SB337	487	F-GTCCTTCCCTGTCTATTCTGCA R-AATTCGGTCTGCTTCTCTG	AY048750

rinsed 3 to 5 times in 70% ethanol followed by a thorough rinsing in distilled water. After rinsing, the cells were counterstained in haematoxylin solution for 5 minutes followed by thorough rinsing in tap water. Slides were then examined under a conventional Olympus microscope equipped with an immersion oil objective lens (100x) for the presence of black droplets in the central vacuole indicating positive reactions. The samples were stained using a commercial solution, Sudan Black B Staining System (Sigma Aldrich, Germany) according to the recommendations of the manufacturer.

Molecular detection

Genomic DNA preparation

DNA was extracted from the culture sample of all 37 ostrich isolates and a human isolate using QIAamp DNA Stool Mini Kit (QIAGEN, Hilden, Germany) according to the manufacturer's protocol [4].

Subtyping of *Blastocystis* sp. isolates

All 37 parasite isolates were subjected to sequenced-tagged site (STS) primer-polymerase chain reaction using the following ten sets of primers [4] (Table 1). Two to five µl of DNA preparations were used to amplify the genomic sequences in a 20 µl reaction containing 0.5 mM of the dNTPs, 0.5 mM of each primer, 1 × PCR buffer (75 mM Tris-HCL, pH 8.8, 20 mM (NH₄)₂SO₄ and 0.01% Tween 20), 2.5 mM MgCl₂ and 1 U Taq

DNA Polymerase (recombinant) (FERMENTAS, USA). PCR conditions consisted of 1 cycle of initial denaturing at 94°C for 3 minutes, followed by 30 cycles including denaturing at 94°C for 30 s, annealing at 57°C for 30 s and extending at 72°C for 1 minute, and an additional cycle with a 10 min chain elongation at 72°C (thermocycler Eppendorf, Germany). The amplification products were electrophoresed in 1.5% agarose gels (PROMEGA USA) and Tris-Borate-EDTA buffer. Gels were stained with ethidium bromide and photographed using ultra-violet gel documentation system (Uvitec, United Kingdom). The PCR amplification for each primer pair was repeated thrice for each isolate [4]. The classification of the subtypes for each *Blastocystis* sp. isolate was based on the standard terminology [8].

In vivo study

The cysts, isolated from fresh fecal material of two respective ostriches, collected in separate tubes by using Ficoll Paque method were made to a concentration of 10⁵ of cysts/ml. This was then inoculated orally, using

Table 2 Prevalence of *Blastocystis* sp. in ostrich isolates

Sex of host	Number of animals	Number of animals detected positive
Male	20	20
Female	17	17

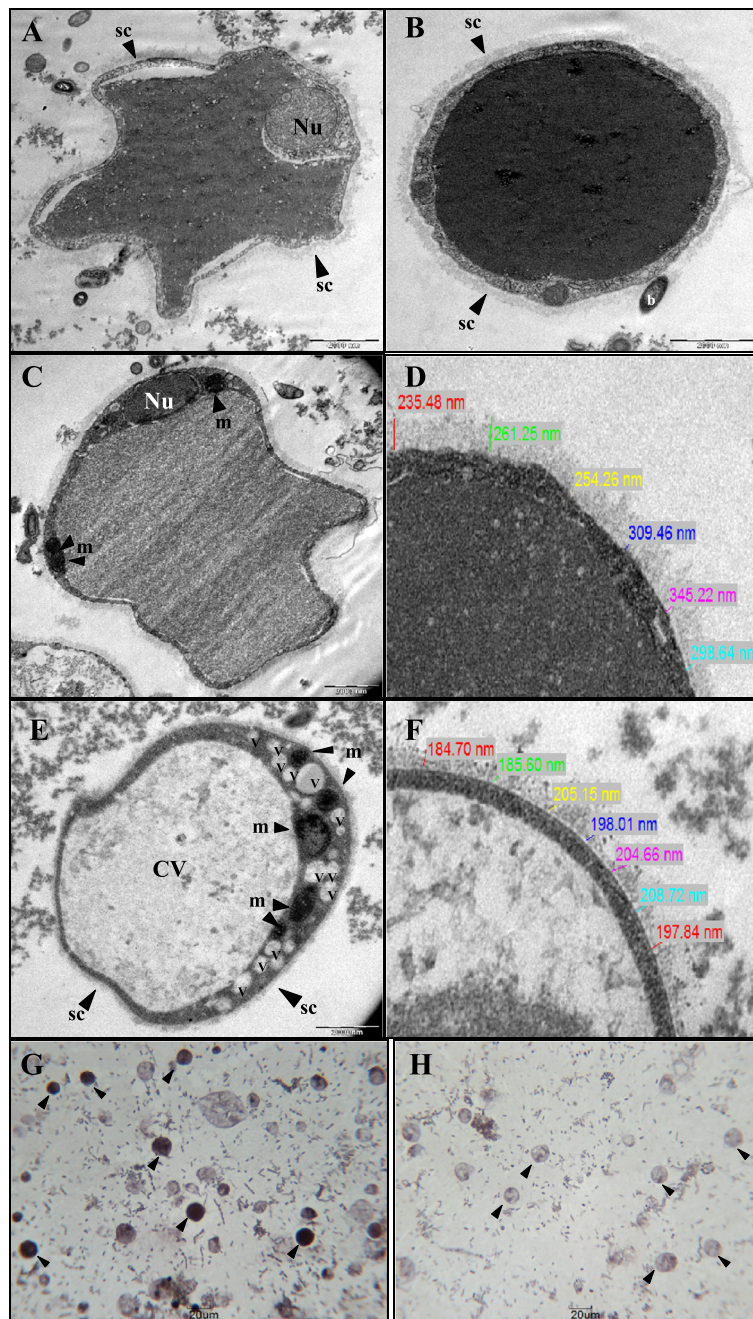


Figure 1 Comparison of Transmission electron microscopy and light microscopy of *Blastocystis* sp. for ostrich isolates and human isolates. **A:** Transmission electron micrograph showing an irregular shape *Blastocystis* sp. with a prominent nucleus (*Nu*) **B:** A thick, compact surface coat (*sc*) is seen to surround the cell when examined by transmission electron microscopy. A high electron dense area was observed in the central vacuole (*CV*). **C:** Numerous mitochondria (*m*) were seen in the *Blastocystis* sp. cells of the ostrich isolates. **D:** Higher magnification of *Blastocystis* sp. membrane in the ostrich faecal culture. **E:** A multi-vacuolated form (*v*) of *Blastocystis* sp. in human faecal culture with multiple mitochondria present in the cytoplasm. **F:** Higher magnification of *Blastocystis* sp. membrane in the human faecal culture. Note: the cell membrane of *Blastocystis* sp. in ostrich and human isolates were 235.48 to 345.22 nm and 184.70 to 208.72 nm, respectively. **G:** Light microscopic images of *Blastocystis* sp. isolated on day 3 of ostrich faecal culture stained with Sudan Black B. Positive reactions are seen as dark droplets in the central vacuole. Note: dark droplets (*arrows*). **H:** Light microscopic images of *Blastocystis* sp. isolated on day 3 of human faecal culture stained with Sudan Black B. No reactions were observed in the central vacuole.

Table 3 Statistical comparison of membrane thickness of *Blastocystis* sp. isolated from ostrich and human (p < 0.05)

	Organism	Range	Means ± S.D	p-value
Thickness of membrane (nm)	Ostrich	235.48 - 345.22	284.05 ± 40.91	0.003
	Human	184.70 - 208.72	197.81 ± 9.49	

20G feeding needle of 1.5inch length into nine Sprague Dawley rats. All the 9 rats were divided into two respective groups, Group A of 5 rats and Group B of 4 rats and were anesthetized before inoculation of respective strains. Stools were collected from the rats to examine for the presence of *Blastocystis* sp., two days of post-inoculation [23]. Samples found positive for *Blastocystis* sp. were then subjected to subtyping.

Statistical analysis

Statistical analysis was carried out using IBM® SPSS® Statistics Version 21. Independent Students t-test was used to assess the differences in the membrane thickness of *Blastocystis* sp. isolated from ostrich and human. A value of p < 0.05 is considered statistically significant.

Results

Prevalence

100% of 37 ostriches were found to be positive for *Blastocystis* sp. No other parasites were seen. However all ostriches appeared healthy without any symptoms. Direct microscopy of fecal smears revealed 8-12 *Blastocystis* sp. under 40x magnification (Table 2).

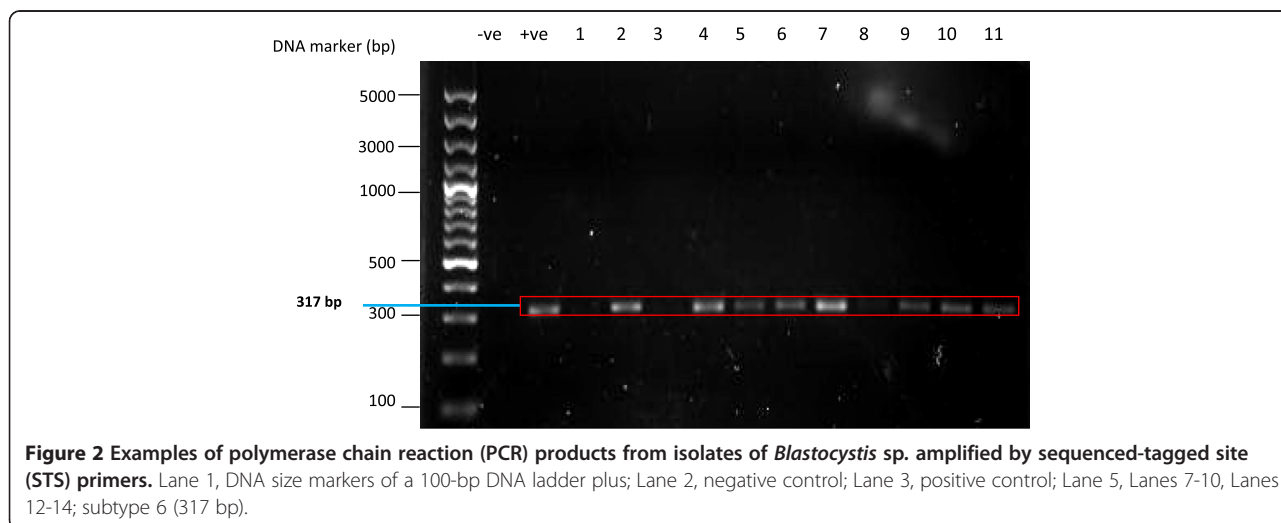
Ultrastructural studies of *Blastocystis* sp.

Transmission electron micrographs of *Blastocystis* sp. showed slight irregular in shape with a thick and compact surface coat seen to be surrounding the cell (Figure 1A). Bacteria were occasionally seen to be

adhering to the surface coat (Figure 1B). High electron dense material was observed in the central vacuole with the presence of two prominent nuclei in most of the parasites (Figure 1B). Numerous mitochondria were seen in the *Blastocystis* sp. cells of the ostrich isolates (Figure 1C). Meanwhile, *Blastocystis* sp. in human fecal culture (Figure 1E) illustrates a multi-vacuolated form of this organism with multiple mitochondria present in the cytoplasm and a clear, large central vacuole (CV). The membrane layer of the ostrich isolate (Figure 1D) was significantly (p = 0.003) thicker, 284.05 ± 40.91 nm (range, 235.48 to 345.22 nm) as compared to human isolate (Figure 1F), 197.81 ± 9.49 nm (range, 184.70 to 208.72 nm) (Table 3). Parasites from day 3 cultures showed ostrich isolate stained with Sudan Black B, positive reactions were observed in the central vacuole of *Blastocystis* sp. (Figure 1G) meanwhile no reactions were observed in the central vacuole of the *Blastocystis* sp. from the day 3 human isolate (Figure 1H). Positive reactions are seen as dark droplets in the central vacuole (CV).

Subtyping of *Blastocystis* sp.

Using sequenced-tagged site primer-PCR, 14 of the ostrich isolates were confirmed to be subtype 6 (Figure 2). Meanwhile, the subtype of remaining 23 isolates when amplified with PCR using sequence-tagged site (STS) revealed no bands when assessed using primers for subtypes 1 to 7.



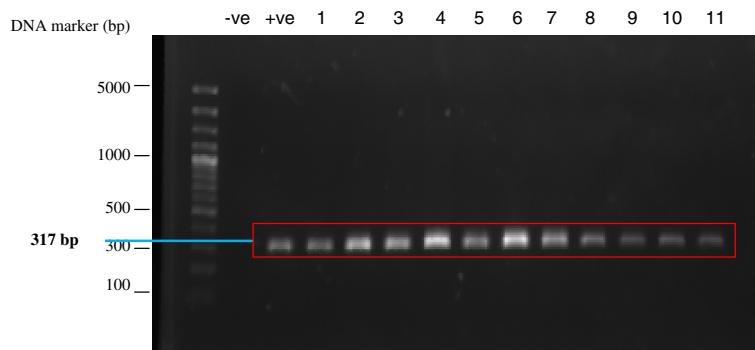


Figure 3 Examples of polymerase chain reaction (PCR) products from isolates of *Blastocystis* sp. from the cross-infection *in vivo* study amplified by sequenced-tagged site (STS) primers. Lane 1, DNA size markers of a 100-bp DNA ladder plus; Lane 2, negative control; Lane 3, positive control; Lanes 4-5, *Blastocystis* sp. cyst of ostrich isolate (inoculum); Lane 6-14, *Blastocystis* sp. isolates of infected rats; subtype 6 (317 bp).

In vivo study

The *Blastocystis* cysts isolated from fresh fecal samples of ostriches ranged from 3.0 to 7.0 μm in diameter. These cysts caused experimental infection in Sprague Dawley rats. *Blastocystis* sp. was detected in feces of experimentally infected Sprague Dawley rats, two days of post-inoculation. The rats infected with ostrich isolate were identified to be subtype 6 which was similar to the strain of the inoculum when amplified with PCR using sequence-tagged site (STS) primers (Figure 3).

Discussion

A high prevalence of *Blastocystis* infection with 100% positive in the ostrich population indicates that this organism is commonly found in this avian. Ultrastructural studies concurred with the previous studies [24,25]. Most of the cells appeared to be rounded or slightly irregular in shape with a thick and compact surface coat seen to be surrounding the cell. One or more nuclei with numerous mitochondria were commonly observed in the cytoplasm of the organism [5]. The outer membrane of ostrich isolate was observed to be significantly thicker when compared to human isolate possibly conferring greater resistance in non-conductive environments. The most distinguishing characteristic was the presence of high electron dense material within the vacuolar forms of parasites on day 3 cultures. Sudan Black B revealed in more than 50% of the parasites dark stains in portions of the central vacuole indicating the presence of neutral lipid [26]. The study confirms that *Blastocystis* sp. from ostrich isolates uses the vacuolar forms to store lipids.

Although subtype 6 was seen in livestock animals [9] especially in pigs and cattle [27] this is the first study to demonstrate subtype 6 seen in ostrich isolates when amplified with polymerase chain reaction using sequence-tagged site (STS) primers (Figure 2). Roberts *et al.* [19] showed that 6 out of 10 ostriches

examined for *Blastocystis* sp. were subtype 7. In the present study, the *Blastocystis* cysts of ostrich isolate were able to cause experimental infection in Sprague Dawley rats (Figure 3) as evidenced by the subtype 6 seen in the stools of infected rats which were prior negative for *Blastocystis* sp. Subtype 6 have been shown to be seen in humans [28] and therefore can be postulated that the ostrich farms with wild rats can be a reservoir for human infections since *Blastocystis* sp. exhibits low host specificity.

Conclusion

The present study is the first to elucidate the ultra-structural, subtype, and host susceptibility of *Blastocystis* sp. isolated from ostriches in order to determine the true pathogenicity of this zoonotic parasite which is known to be a potential source for cross-transmission between animal and humans especially when in close association.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

HC, SKG, CP, PB, KR, and GT was involved in all phases of the study, including the conceptual framework and the design of the experiment, data collection and analysis, interpretation of results. HC wrote up the manuscript and all authors read and approved the finalized manuscript.

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References

1. Stenzel DJ, Boreham PF: *Blastocystis hominis* revisited. *Clin Microbiol Rev* 1996, **9**:563–584.
2. Konig G, Muller HE: *Blastocystis hominis* in animals: incidence of four serogroups. *Zentralbl Bakteriol* 1997, **286**:435–440.
3. Belova LM, Krylov MV: The distribution of *Blastocystis* according to different systematic groups of hosts. *Parazitologija* 1998, **32**:268–276.
4. Yoshikawa H, Wu Z, Kimata I, Iseki M, Ali IK, Hossain MB, Zaman V, Haque R, Takahashi Y: Polymerase chain reaction-based genotype classification among human *Blastocystis hominis* populations isolated from different countries. *Parasitol Res* 2004, **92**:22–29.
5. Stenzel DJ, Cassidy MF, Boreham PFL: Morphology of *Blastocystis* sp. from domestic birds. *Parasitol Res* 1994, **80**:131–137.
6. Farah HMT, Chandrawathani P, Mohd ZSN, Suresh KG, Hemalatha C, Premaalatha B: A preliminary study of *Blastocystis* sp. isolated from chicken in Perak and Selangor, Malaysia. *MJVR* 2014, **5**:21–25.
7. Noel C, Dufernez F, Gerbod D, Edgcomb VP, Delgado VP, Ho LC, Viscogliosi E: Molecular phylogenies of *Blastocystis* isolates from different hosts-implications for genetic diversity, identification of species, and zoonosis. *J Clin Microbiol* 2005, **43**:348–355.
8. Stensvold CR, Suresh KG, Tan KS, Thompson RC, Traub RJ, Viscogliosi E, Clark CG: Terminology for *Blastocystis* subtypes—a consensus. *Trends Parasitol* 2007, **23**:93–96.
9. Alfellani MA, Taner MD, Jacob AS, Imeede CA, Yoshikawa H, Stensvold CR, Clark CG: Genetic diversity of *Blastocystis* in livestock and zoo animals. *Protist* 2013, **164**:497–509.
10. Yoshikawa H, Abe N, Iwasawa M, Kitano S, Nagano I, Wu Z, Takahashi Y: Genomic analysis of *Blastocystis hominis* strains isolated from two long-term health care facilities. *J Clin Microbiol* 2000, **38**:1324–1330.
11. Doyle PW, Helgason MM, Mathias RG, Proctor EM: Epidemiology and pathogenicity of *Blastocystis hominis*. *J Clin Microbiol* 1990, **28**:116–121.
12. Nimri L, Raymond B: Intestinal colonization of symptomatic and asymptomatic schoolchildren with *Blastocystis hominis*. *J Clin Microbiol* 1994, **32**:2865–2866.
13. Casemore DP: Foodborne protozoal infection. *Lancet* 1990, **336**:1427–1432.
14. Garavelli P, Orsi P, Scaglione L: *Blastocystis hominis* infection during AIDS. *Lancet* 1988, **332**:1364.
15. Llibre JM, Tor J, Manterola JM, Carbonell C, Foz M: *Blastocystis hominis* chronic diarrhoea in AIDS patients. *Lancet* 1989, **333**:221.
16. Yamada M, Yoshikawa H, Tegoshi T, Matsumoto Y, Yoshikawa T, Shiota T, Yoshida Y: Light microscopical study of *Blastocystis* spp. in monkeys and fowls. *Parasitol Res* 1987, **73**:527–531.
17. Pakandl M: *Blastocystis* sp. from pigs-ultrastructural changes occurring during polyxenic cultivation in Iscove's modified Dulbecco's medium. *Parasitol Res* 1999, **85**:743–748.
18. Ponce GF, Herrera S, Castro AT, Garcia DB, Martinez DRA: Parasites from farmed ostriches (*Struthio camelus*) and rheas (*Rhea Americana*) in Europe. *Vet Parasitol* 2002, **107**:137–160.
19. Roberts T, Stark D, Harkness J, Ellis J: Subtype distribution of *Blastocystis* isolates from a variety of animals from New South Wales, Australia. *Vet Parasitol* 2013, **196**:85–89.
20. Suresh K, Ng GC, Ho LC, Yap EH, Singh M: Differentiation of the various stages of *Blastocystis hominis* by acridine orange staining. *Int J Parasitol* 1994, **24**:605–606.
21. Norakmar I, Chandrawathani P, Nurulaini R, Zawida Z, Premaalatha B, Imelda Lynn V, Adnan M, Jamnah O, Zaini CM: Prevalence of helminthiasis in relation to climate in small ruminants in Perak in 1998 and 2008. *MJVR* 2010, **1**:37–44.
22. Tan TC: Predominance of amoeboid forms of *Blastocystis hominis* in isolates from symptomatic patients. *Parasitol Res* 2006, **98**:189–193.
23. Suresh K, Ng GC, Ramachandran NP, Ho LC, Yap EH, Singh M: In vitro encystment and experimental infections of *Blastocystis hominis*. *Parasitol Res* 1993, **79**:456–460.
24. Tan KS: *Blastocystis* in humans and animals: new insights using modern methodologies. *Vet Parasitol* 2004, **126**:121–144.
25. Yoshikawa H, Wu Z, Howe J, Hashimoto T, Geok-Choo NG, Tan KS: Ultrastructural and phylogenetic studies on *Blastocystis* isolates from cockroaches. *J Eukaryot Microbiol* 2007, **54**:33–37.
26. Yoshikawa H, Satoh J, Enose Y: Light and electron microscopic localization of lipids in *Blastocystis hominis*. *J Electron Microsc* 1995, **44**:100–103.
27. Stensvold CR, Alfellani MA, Norskov LS, Prip K, Victory EL, Maddox C, Clark CG: Subtype distribution of *Blastocystis* isolates from synanthropic and zoo animals and identification of a new subtype. *Int J Parasitol* 2009, **39**:473–479.
28. Stensvold CR, Lewis HC, Hammerum AM, Porsbo LJ, Nielsen SS, Olsen KEP, Molbak K: *Blastocystis*: unravelling potential risk factors and clinical significance of a common but neglected parasite. *Epidemiol Infect* 2009, **137**:1655–1663.

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