

Complete Nucleotide Sequence of a *Citrobacter freundii* Plasmid Carrying KPC-2 in a Unique Genetic Environment

Yancheng Yao,^a Can Imirzalioglu,^a Torsten Hain,^a Martin Kaase,^b Soeren Gatermann,^b Martin Exner,^c Martin Mielke,^d Anja Hauri,^e Yolanta Dragneva,^f Rita Bill,^f Constanze Wendt,^g Angela Wirtz,^h Eugen Domann,^a Trinad Chakraborty^a

Institute for Medical Microbiology, German Centre of Infection Research, Justus-Liebig-University Giessen, Giessen, Germany^a; Institute for Hygiene and Medical Microbiology, Ruhr University, Bochum, Germany^b; Institute for Hygiene and Public Health, University of Bonn, Bonn, Germany^c; Robert Koch Institute, Berlin, Germany^d; Hessisches Landesprüfungs- und Untersuchungsamt im Gesundheitswesen, Dillenburg, Germany^e; Gesundheits- und Pflegezentrum Rüsselsheim, Rüsselsheim, Germany^f; Labor Limbach, Heidelberg, Germany^g; Hessisches Ministerium für Soziales und Integration, Wiesbaden, Germany^h

The complete and annotated nucleotide sequence of a 54,036-bp plasmid harboring a *bla*_{KPC-2} gene that is clonally present in *Citrobacter* isolates from different species is presented. The plasmid belongs to incompatibility group N (IncN) and harbors the class A carbapenemase KPC-2 in a unique genetic environment.

Received 26 September 2014 Accepted 6 October 2014 Published 13 November 2014

Citation Yao Y, Imirzalioglu C, Hain T, Kaase M, Gatermann S, Exner M, Mielke M, Hauri A, Dragneva Y, Bill R, Wendt C, Wirtz A, Domann E, Chakraborty T. 2014. Complete nucleotide sequence of a *Citrobacter freundii* plasmid carrying KPC-2 in a unique genetic environment. *Genome Announc.* 2(6):e01157-14. doi:10.1128/genomeA.01157-14.

Copyright © 2014 Yao et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](http://creativecommons.org/licenses/by/3.0/).

Address correspondence to Eugen Domann, eugen.domann@mikrobio.med.uni-giessen.de.

Carbapenems remain the most effective antibiotics for the treatment of serious infections caused by multi-resistant Gram-negative bacteria producing extended-spectrum β -lactamases (ESBL). The rise of carbapenem-resistant Gram-negative bacteria is increasingly being reported and is now a matter of great clinical concern. Carbapenem resistance in *Enterobacteriaceae* is mainly due to the production of carbapenemases, the most common of which is the *K. pneumoniae* carbapenemase (KPC) family of enzymes (1–3).

Recently, isolation of a cluster of carbapenem-resistant *Citrobacter* species was reported from a single hospital environment in southern Hesse, Germany (4). Most isolates were typed as *C. freundii* but several isolates of carbapenem-resistant *C. amalonaticus*, *C. braakii*, and *C. koseri* were also detected. *Citrobacter* species are environmental pathogens that can colonize the intestinal tract of humans and animals. They are generally considered low-grade pathogens that rarely cause infections. However, these bacteria have been associated with a wide spectrum of infections involving the central nervous system and the gastrointestinal, urinary, and respiratory tracts (5).

Preliminary characterization revealed that all isolates harbored a nonconjugable *bla*_{KPC-2} gene. In order to determine the genetic localization of the KPC-2 we determined the genome sequence of 11 representative strains (8 *C. freundii*, 1 *C. amalonaticus*, 1 *C. braakii*, 1 *C. koseri*). DNA sequencing libraries were prepared using the Nextera XT kit (Illumina, San Diego) according to the manufacturer's instructions. Individually tagged libraries were sequenced as a part of a flowcell as 2×300 base paired-end reads using the Illumina MiSeq platform (Illumina, San Diego). A total of 12,447,167,642 sequences were produced and the sequences from each isolate were separately assembled using CLC Genomics Workbench version 7.0.4. We identified contigs harboring *bla*_{KPC-2} by using ResFinder (<http://cge.cbs.dtu.dk/services/ResFinder/>) and assembled the flanking sequences to generate a closed contig comprised of 54,036 bp with 82 coding sequences (CDS) (6). Open reading frame (ORF) finding and gene annota-

tion was done by using RAST (<http://rast.nmpdr.org/>) and a genetic map of the resulting contigs was generated with MAUVE (7, 8) and with the plasmid reference nucleotide sequence of pKPC_FCF/3SP (accession no. CP004367.2). Further analysis revealed that *bla*_{KPC-2} is located on an IncN plasmid and inserted in a region between the *traI* and *traG* genes (9–11). The *bla*_{KPC-2} gene is part of a 9,571-bp insertion with a unique genetic environment comprising at one end of a Tn4401 element with the ISKpn6 and *Kpc-2* genes and an adjacent Tn3-like segment (12–14), harboring a *bla*_{TEM1B}, *ISCfr1*, and *aac3-IId* genes flanked by a 137 bp direct repeat. All sequenced strains harbor genetically identical plasmids, suggesting its horizontal spread among the different *Citrobacter* species. The plasmid, derived from *C. freundii* isolate Cfr08698 encoding *bla*_{KPC-2}, was designated pCfr-08698KPC-2.

Nucleotide sequence accession number. The nucleotide sequence of the *C. freundii* plasmid carrying *bla*_{KPC-2} has been deposited in the EMBL database under accession no. LN610760.

ACKNOWLEDGMENTS

We thank Christina Gerstmann and Alexandra Amend for excellent technical assistance and Katrin Gentil for reading the manuscript.

This work was supported by grants from the German Centre of Infection Research (DZIF) to C.I. and T.C. (FKZ: 80 00 701-2 HZI).

REFERENCES

- Munoz-Price LS, Poirel L, Bonomo RA, Schwaber MJ, Daikos GL, Cormican M, Cornaglia G, Garau J, Gniadkowski M, Hayden MK, Kumarasamy K, Livermore DM, Maya JJ, Nordmann P, Patel JB, Paterson DL, Pitout J, Villegas MV, Wang H, Woodford N, Quinn JP. 2013. Clinical epidemiology of the global expansion of *Klebsiella pneumoniae* carbapenemases. *Lancet Infect. Dis.* 13:785–796. [http://dx.doi.org/10.1016/S1473-3099\(13\)70190-7](http://dx.doi.org/10.1016/S1473-3099(13)70190-7).
- Nordmann P, Dortet L, Poirel L. 2012. Carbapenem resistance in *Enterobacteriaceae*: here is the storm!. *Trends Mol. Med.* 18:263–272. <http://dx.doi.org/10.1016/j.molmed.2012.03.003>.
- Tzouveleki LS, Markogiannakis A, Psychogiou M, Tassios PT, Daikos GL. 2012. Carbapenemases in *Klebsiella pneumoniae* and other

- Enterobacteriaceae*: an evolving crisis of global dimensions. Clin. Microbiol. Rev. 25:682–707. <http://dx.doi.org/10.1128/CMR.05035-11>.
4. Robert-Koch-Institut. 23 September 2014. Häufung von KPC-2 produzierenden Stämmen verschiedener Enterobacteriaceae-Spezies in Hessen. Epidemiolog. Bull. <http://dx.doi.org/10.1016/j.khinf.2014.08.009>.
 5. Abbott SL. 2007. *Klebsiella*, *Enterobacter*, *Citrobacter*, *Serratia*, *Plesiomonas*, and other *Enterobacteriaceae*, p 698–715. In Murray PR, Baron EJ, Jorgensen JH, Landry ML, Pfaller MA (ed), Manual of clinical microbiology, 9th ed. ASM Press, Washington, D.C.
 6. Zankari E, Hasman H, Cosentino S, Vestergaard M, Rasmussen S, Lund O, Aarestrup FM, Larsen MV. 2012. Identification of acquired antimicrobial resistance genes. J. Antimicrob. Chemother. 67:2640–2644. <http://dx.doi.org/10.1093/jac/dks261>.
 7. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. BMC Genomics 9:75. <http://dx.doi.org/10.1186/1471-2164-9-75>.
 8. Darling AE, Mau B, Perna NT. 2010. progressiveMauve: multiple genome alignment with gene gain, loss, and rearrangement. PLoS One 5:e11147. <http://dx.doi.org/10.1371/journal.pone.0011147>.
 9. Andrade LN, Curiao T, Ferreira JC, Longo JM, Clímaco EC, Martinez R, Bellissimo-Rodriguez F, Basile-Filho A, Evaristo MA, Del Peloso PF, Ribeiro VB, Barth AL, Paula MC, Baquero F, Cantón R, da Costa Darini AL, Coque TM. 2011. Dissemination of *bla*_{KPC-2} by the spread of *Klebsiella pneumoniae* clonal complex 258 clones (ST258, ST11, ST437) and plasmids (IncFII, IncN, IncL/M) among *Enterobacteriaceae* species in Brazil. Antimicrob. Agents Chemother. 55:3579–3583. <http://dx.doi.org/10.1128/AAC.01783-10>.
 10. Chmelnitsky I, Shklyar M, Leavitt A, Sadovsky E, Navon-Venezia S, Ben Dalak M, Edgar R, Carmeli Y. 2014. Mix and match of KPC-2 encoding plasmids in *Enterobacteriaceae*-comparative genomics. Diagn. Microbiol. Infect. Dis. 79:255–260. <http://dx.doi.org/10.1016/j.diagmicrobio.2014.03.008>.
 11. Eikmeyer F, Hadiati A, Szczepanowski R, Wibberg D, Schneiker-Bekel S, Rogers LM, Brown CJ, Top EM, Pühler A, Schlüter A. 2012. The complete genome sequences of four new IncN plasmids from wastewater treatment plant effluent provide new insights into IncN plasmid diversity and evolution. Plasmid 68:13–24. <http://dx.doi.org/10.1016/j.plasmid.2012.01.011>.
 12. Cain AK, Hall RM. 2012. Evolution of IncHI2 plasmids via acquisition of transposons carrying antibiotic resistance determinants. J. Antimicrob. Chemother. 67:1121–1127. <http://dx.doi.org/10.1093/jac/dks004>.
 13. Cuzon G, Naas T, Nordmann P. 2011. Functional characterization of Tn4401, a Tn3-based transposon involved in *bla*_{KPC} gene mobilization. Antimicrob. Agents Chemother. 55:5370–5373. <http://dx.doi.org/10.1128/AAC.05202-11>.
 14. Pérez-Chaparro PJ, Cerdeira LT, Queiroz MG, de Lima CP, Levy CE, Pavez M, Lincopan N, Gonçalves EC, Mamizuka EM, Sampaio JL, Nunes MR, McCulloch JA. 2014. Complete nucleotide sequences of two *bla*_{KPC-2}-bearing IncN Plasmids isolated from sequence type 442 *Klebsiella pneumoniae* clinical strains four years apart. Antimicrob. Agents Chemother. 58:2958–2960. <http://dx.doi.org/10.1128/AAC.02341-13>.