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CASE CONTROL STUDY

Circulating microRNA, miR-122 and miR-221 signature in Egyptian patients with chronic hepatitis C related hepatocellular carcinoma

Hassan El-Garem, Ayman Ammer, Hany Shehab, Olfat Shaker, Mohammed Anwer, Wafaa El-Akel, Heba Omar

Hassan El-Garem, Ayman Ammer, Hany Shehab, Mohammed Anwar, Wafaa El-Akel, Heba Omar, Endemic Medicine and Hepatology Department, Faculty of Medicine, Cairo University, Cairo 12613, Egypt

Olfat Shaker, Medical Biochemistry and Molecular Biology Department, Faculty of Medicine, Cairo University, Cairo 12613, Egypt

Author contributions: El-Garem H contributed to the study design, revision; Ammer A contributed to the study design; Shehab H contributed to the study design and drafting the manuscript; Shaker O contributed to the study design, biochemical analysis of samples; Anwer M contributed to the drafting manuscript and revision; El-Akel W contributed to the study design and statistical analysis; Omar H contributed to the study design, samples and data collection, manuscript drafting.

Correspondence to: Heba Omar, Msc, PhD, Endemic Medicine and Hepatology Department, Faculty of medicine, 16 Azam Street, Helwan, Cairo 12613,

Egypt. hebaomar1202@hotmail.com

Telephone: +20-2-01007032146

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Abstract

AIM: To explore the potential usefulness of serum miR-122 and miR-221 as non-invasive diagnostic markers of hepatitis C virus (HCV)-related hepatocellular carcinoma (HCC).

METHODS: This prospective study was conducted on 90 adult patients of both sex with HCV-related chronic liver disease and chronic hepatitis C related HCC. In addition to the 10 healthy control individuals, patients were stratified into; interferon-naïve chronic hepatitis C (CH) (n = 30), post-hepatitis C compensated cirrhosis (LC) (n = 30) and treatment-naïve HCC (n = 30). All patients and controls underwent full clinical assessment

and laboratory investigations in addition to the evaluation of the level of serum miRNA expression by RT-PCR.

RESULTS: There was a significant fold change in serum miRNA expression in the different patient groups when compared to normal controls; miR-122 showed significant fold increasing in both CH and HCC and significant fold decrease in LC. On the other hand, miR-221 showed significant fold elevation in both CH and LC groups and significant fold decrease in HCC group (P = 0.01). Comparing fold changes in miRNAs in HCC group *vs* non HCC group (CH and Cirrhosis), there was non-significant fold elevation in miR-122 (P = 0.21) and significant fold decreasing in miR-221 in HCC *vs* non-HCC (P = 0.03). ROC curve analysis for miR-221 yielded 87% sensitivity and 40% specificity for the differentiation of HCC patients from non-HCC at a cutoff 1.82.

CONCLUSION: Serum miR-221 has a strong potential to serve as one of the novel non-invasive biomarkers of HCC.

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Key words: MiRNA; Hepatocellular carcinoma; Serum

Core tip: In the current study a signature of circulating miRNAs (miR-122 and miR-221) was evaluated. miR-221 was differentially expressed between patients with hepatocellular carcinoma and those without (chronic hepatitis and liver cirrhosis) with lower serum level of miR-221 in former group of patients in comparison to later one. miR-221 yielded 87% sensitivity and 40% specificity in differentiating between both groups at a cutoff 1.82 folds. The present study emphasizes that circulating miR-221 deserves further attention as a potential non-invasive biomarkers for hepatocellular



carcinoma.

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INTRODUCTION

Hepatocellular carcinoma (HCC) is the most common primary liver cancer, and the fifth most common cancer worldwide^[1]. The last decade has witnessed a significant rise in the incidence of HCC^[2-4] with a specially high incidence reported in Egypt^[5]. A direct role of hepatitis C virus (HCV) in hepatocarcinogenesis has been suggested^[6]. However, it seems that cirrhosis is the common route through which several risk factors act and induce carcinogenesis^[7].

Currently available blood tumor markers are far from optimal. Alfa-fetoprotein (AFP), Lens culinaris agglutinin-reactive AFP (AFP-L3) and des-carboxyprothrombin (DCP) perform poorly in the surveillance mode and early detection of HCC^[8]. The Practice Guidelines of the American Association for the Study of Liver Diseases (AASLD) (July 2010) rejected AFP whether for the surveillance or the diagnosis of HCC. This highlights the need for other methods that would be minimally-invasive, simple and reliable for the early detection of HCC.

MicroRNA (miRNA) is a non-coding *RNA* gene product that negatively controls gene expression by altering the stability or translational efficiency of its target mRNAs^[2]. MiRNAs regulate several biological processes, such as cell differentiation, apoptosis and proliferation. miRNAs have been reported to be aberrantly present in cancers whether through up- or down-regulation in neoplastic cells compared with their normal counterparts^[3,4]. What makes miRNAs even more interesting is that several recent studies have demonstrated that miRNAs are detectable and stable in plasma and serum^[4-6].

The goal of the present study was to evaluate circulating serum miRNAs (miR-122 and miR-221) expression levels in Egyptian patients with HCC as well as in patients with HCV-related chronic liver disease to explore their potential as novel non-invasive markers for diagnosis of HCV-related HCC.

MATERIALS AND METHODS

During the period between March and June 2012 serum samples were collected from consecutive HCV-infected patients presenting to our outpatient department: 30 with chronic HCV alone (CH), 30 with HCV-related cirrhosis (LC) and 30 with HCV-related HCC. Serum samples were also collected from 10 age and gender-matched healthy volunteers (defined as those with normal transaminases, normal hepatic ultrasound and negative for HBsAg, HBc-Ab and HCV RNA-PCR). All patients were recruited after a written informed consent and the study protocol was approved by the ethics review committee of Cairo University hospital. Exclusion criteria included: patients with chronic HBV infection or any other identifiable cause for chronic hepatitis other than HCV, previous treatment for HCC or antiviral therapy for HCV and any associated malignancies other than HCC.

RNA extraction

For the real-time PCR RNAs were extracted from serum using TRIzol according to the manufacturer's instruction. The RNA purity was assessed by the RNA concentration and quantified by NanoDrop ND-1000 (Nanodrop, United States). Single-stranded cDNAs were generated using the RT kit (Qiagen, Valencia, CA, United States) according to the manufacturer's directions (miSCript miRNA PCR system, miRneasy mini kit for miRna extraction, miScript RT II for miRna reverse transcription, miSCript Primer Assay and miSCript SYBR Green PCR Kit for PCR amplification.

RNA quantification

PCR quantification experiments were performed with PCR (Applied Biosystems; Foster City, CA) using the SYBR Green PCR Master Mix according to the manufacturer's protocol. The primers for microRNA-122, -221 and housekeeping gene were supplied by Qiagene, Germany (catalog numbers 3416, 3857 and 33712). The housekeeping miRNA SNORD68 was used as the endogenous control. Fluorescence measurements were made in every cycle and the cycling conditions used were: 95 °C for 30 s, and 40 cycles of 95 °C for 5 s and 60 °C for 34 s.

Expression of miRNAs was reported as Δ Ct value. The Δ Ct was calculated by subtracting the Ct values of miRNA SNORD68 from the Ct values of the target miRNAs. As there is an inverse correlation between Δ Ct and miRNA expression level, lower Δ Ct values were associated with increased miRNA. The resulting normalized Δ Ct values were used in calculating relative expression values by using 2- Δ (Ct), these values are directly related to the miRNA expression levels. The 2- $\Delta\Delta$ (Ct) method was used to determine relative-quantitative levels of individual miRNAs.

Statistical analysis

Patients were categorized into 4 groups; normal, CH, Cirrhosis and HCC. Further comparisons were performed between HCC group and Non-HCC (CH and cirrhosis). Quantitative variables were expressed by mean \pm SD or expressed by median and inter quartile range (IQR) for non-parametric data. They were compared by *t*-student or ANOVA test when appropriate. Qualitative variables were compared by χ^2 or Fischer's exact test when appropriate. AFP levels were transformed into their log values



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Table 1 Demographic parameters of patients							
	HCC $(n = 30)$	Cirrhosis $(n = 30)$	CH (<i>n</i> = 30)	Normal $(n = 10)$	<i>P</i> value		
Age (yr)	$60.27 \pm 8.20^{\circ}$	$55.07 \pm 7.35^{\text{B}}$	$38.20 \pm 8.21^{\text{A}}$	$40.89 \pm 16.85^{\text{A}}$	≤ 0.001		
Mean ± SD							
Gender (male)	25 (83.3)	21 (70)	22 (73.3)	6 (66.7)	0.6		
Hb (g/dL)	11.44 ± 2.85^{B}	10.53 ± 2.00^{B}	$14.23 \pm 1.58^{\text{A}}$	$14.03 \pm 2.48^{\text{A}}$	< 0.001		
WBC $\times 10^3$ /mm ³	$5.73 \pm 2.56^{\text{A}}$	$7.08 \pm 3.77^{\text{A}}$	$6.15 \pm 2.13^{\text{A}}$	$7.38 \pm 2.72^{\text{A}}$	0.217		
Platelets 10/mm ^{3D}	$126.00 \pm 74.05^{\text{B}}$	$116.10 \pm 68.94^{\text{B}}$	$228.80 \pm 59.74^{\text{A}}$	$271.3 \pm 116.7^{\text{A}}$	< 0.001		
Total bilirubin (0.1-1.2 mg/dL)	$1.88 \pm 2.01^{\text{A}}$	$4.29 \pm 6.75^{\text{B}}$	$0.74 \pm 0.26^{\text{A}}$	$0.79 \pm 0.36^{\text{A}}$	< 0.001		
ALT (0-42 IU/L)	66.59 ± 44.59^{B}	$32.15 \pm 23.59^{\text{A}}$	66.78 ± 36.56^{B}	$29.16 \pm 19.96^{\text{A}}$	< 0.001		
AST (0-42 IU/L)	$119.99 \pm 56.12^{\text{B}}$	$63.81 \pm 39.49^{\text{A}}$	$59.67 \pm 45.04^{\text{A}}$	$45.64 \pm 54.32^{\text{A}}$	< 0.001		
ALP (0-290 IU/L) ^D	$394.8 \pm 282.28^{\text{A}}$	$304.89 \pm 191.85^{\text{A}}$	203.58 ± 82.37^{AB}	198.8 ± 139.1^{B}	0.003		
Albumin $(3.5-5.5 \text{ g/dL})^{D}$	$3.16 \pm 0.40^{\circ}$	2.49 ± 0.54^{B}	$4.22 \pm 0.36^{\text{A}}$	$4.09 \pm 0.92^{\text{A}}$	< 0.001		
PC %	$69.63 \pm 16.25^{\circ}$	$51.58 \pm 17.12^{\text{B}}$	$88.24 \pm 10.89^{\text{A}}$	$97.63 \pm 11.77^{\text{A}}$	< 0.001		
AFP log10 ng/dL	$2.50 \pm 1.19^{\text{B}}$	$0.79\pm0.54^{\rm A}$	$0.59 \pm 0.38^{\text{A}}$	NA	< 0.001		

^{AB,CD}Groups with different letters show significant difference, those with similar letters show no significant difference. HCC: Hepatocellular carcinoma; NA: Not applicable. CH: Chronic hepatitis C.

Table 2 Tumor-related characteristics $(n = 30)$				
	Parameter	Number (%)		
AFP level (0-10)	Normal	4 (13.4%)		
	Elevated	26 (86.6%)		
PS	PS 0	24 (80)		
	PS 1-2	4 (13.4)		
	PS > 2	2 (6)		
BCLC	Stage 0	0 (0)		
	Stage A	1 (3.8)		
	Stage B	19 (73.1)		
	Stage C	4 (15.4)		
	Stage D	2 (7.7)		
Number of focal lesions	Single	17 (56.7)		
	Multiple	13 (43.4)		
Site of focal lesions	Right lobe	18 (60)		
	Left lobe	5 (16.7)		
	Both	7 (23.3)		
Tumor size by CT	< 3 cm	1 (3.3)		
	3-5 cm	12 (40)		
	> 5 cm	17 (56.7)		
Portal vein invasion	Yes	7 (23.3)		
	No	23 (76.7)		

PS: Performance status; AFP: Alfa-fetoprotein; CT: Computed tomography.

to undergo parametric statistical tests. Receiver operator characteristic (ROC) curves were constructed to assess the value of miRNA in diagnosing HCC and to assess area under the curve (AUROC). AUROC less than 0.60 with *P* value > 0.05 is considered unreliable for ROC curve. Spearmen and Pearson correlations were done for correlating quantitative variables. In all tests, *P* value was considered significant if less than 0.05.

RESULTS

This study was conducted on 100 participants stratified into: Group1: thirty patients with HCV-related HCC who were diagnosed according to EASL guidelines 2012; Group2: thirty patients with hepatitis C related liver cirrhosis; Group 3: thirty non-cirrhotic patients with chronic hepatitis C viral infection (CH), while Group 4 included ten age and gender-matched healthy volunteers (defined as those with normal hepatic ultrasound and transaminases and negative for hepatitis B and C by PCR) considered as internal reference.

The demographic and pathologic features of the studied participants are shown in Tables 1 and 2. There was a significant difference between the diseased groups regarding age (P < 0.001). Regarding gender difference; males were predominant in HCV related liver disease patients in the three groups and they represented 83.3%, 70%, 73.3% in HCC, cirrhosis and CH groups respectively with no statistically significant difference between the studied groups (P = 0.60).

miR-122 serum levels

Analysis of median fold change in expression level of miR-122 in patients' sera in comparison to the normal control group showed that miR-122 displayed significant fold decrease in expression in cirrhosis group (0.8) and significant fold increasing in expression level in both CH (2.1) and HCC (2.15) groups ($P \leq 0.01$), Table 3. Comparing serum miR-122 expression level between different studied groups displayed an increasing tendency towards statistical significant fold elevation in expression of miR-122 in serum of HCC patients (2.15) in comparison to liver cirrhosis (0.8) with P value 0.083 (AUC = 0.646), Figure 1. No significant fold change in miR-122 expression was found between either (HCC vs CH groups) or (CH vs cirrhosis groups). MiRNA122 showed nonsignificant up-regulation in HCC patients in comparison to non-HCC patients (CH and Cirrhosis); (P = 0.21).

miR-221 serum levels

Analysis of fold change in expression level of miR-221 in patients' sera in comparison to the normal control group showed significant fold decrease in HCC group and significant fold increase in expression level in CH and cirrhosis groups in comparison to normal control group (< 0.01), Table 4. There was a statistically significant fold decreasing in serum miR-221 levels of HCC patients (0.92) in comparison to cirrhosis (3.4) and in comparison to CH (1.7) (P = 0.05 and 0.06 respectively). On the other



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Table 3 MiR-122 serum levels in the different groups						
	Fold of chai	nge comparing	to normal			
Group	HCC <i>n</i> = 30	$\begin{array}{l} \text{Cirrhosis} \\ n = 30 \end{array}$	CH n= 30	<i>P</i> value		
miR-122 (IQR)	2.15 (7.3)	0.8 (3.7)	2.1 (9)	< 0.01 0.083 ¹ 0.572 ² 0.417 ³		
Group	HCC n = 30	Non HCC (CH + cirrhosis) n = 60		<i>P</i> value		
miR-122 (IQR)	2.15 (7.3)	1.75	(6.8)	0.21 (NS)		

¹HCC *vs* cirrhosis; ²HCC *vs* CH; ³Cirrhosis *vs* CH. HCC: Hepatocellular carcinoma; CH: Chronic hepatitis C; IQR: Inter quartile range; NS: Non significant; PVP: Positive predictive value; NVP: Negative predictive value.

Table 4 MiR-221 serum levels							
	Fold of cha	ange in compari	son to normal				
	HCC <i>n</i> = 30	Cirrhosis n = 30	CH n = 30	<i>P</i> value			
miR-221 (IQR)	0.92 (0.88)	3.4 (19.2)	1.7 (2.6)	> 0.01 0.05 ¹ 0.06 ² 0.214 ³			
	HCC 0.92 (0.88)	Non HCC 1.81 (7.75)		0.03 (S)			

¹HCC *vs* LC; ²HCC *vs* CH; ³LC *vs* CH. HCC: Hepatocellular carcinoma; CH: Chronic hepatitis C; IQR: Inter quartile range; LC: Liver cirrhosis.

hand, there was no statistical significant fold change in serum miR-221 expression level between (CH vs cirrhosis groups) (P = 0.214).

miRNA-221 displayed significant fold decrease in HCC group (0.92) compared to non-HCC patients (CH and cirrhosis) (1.81) (*P* value 0.03). At a cut-off of 1.82 folds, miR-221 yielded 87% sensitivity and 40% specificity in differentiating between both groups. Figure 2, Table 5.

Correlation of miRNA's with features of HCC

There was no statistically significant correlation between serum expression level of studied miRNAs and serum AFP level in the different studied groups of patients. No significant correlation was found between the two miR-NAs and tumor size, Child-pugh grade in HCC group of patients.

Correlation of miRNA's with severity of hepatic dysfunction

Serum miRNA-122 expression level showed statistically significant correlation with serum necro-inflammatory markers of the liver [aspartate transaminase (AST) and alanine transaminase (ALT) levels) in CH group (*P* value 0.034 and 0.030 respectively), Table 1.

DISCUSSION

Over the last 2 decades it has become common practice



Figure 1 Receiver operator characteristic curve for miR-122 as a discriminant of hepatocellular carcinoma vs cirrhosis patients.



Figure 2 Receiver operator characteristic curve for miR-221 as a discriminant between hepatocellular carcinoma vs non-hepatocellular carcinoma.

to use tumour markers, mainly AFP, for the screening of HCC. However, performance of tumor markers has not been optimal with the sensitivity and specificity of AFP and PIVKA-II in the range of 39%-64% and 76%-91%, and 41%-77% and 72%-98%, respectively^[9,10], the quest for an optimal tumor marker hence continues. miRNAs have been implicated in roles affecting cellular proliferation and oncogenesis^[11]. Cellular miRNAs have been linked with HCC^[12]. Their availability in the circulation makes them a tempting target for early tumor detection^[12]. The aim of the present study was to explore the potential usefulness of serum miR-122 and miR-221 as novel noninvasive markers for diagnosis of HCV related hepatocellular carcinoma in Egyptian patients.

Most of our HCC patients were within Child-Pugh A and B classifications (56.7%, 36.7% respectively), 73.1% were stage B on BCLC scoring system^[13]. This could be explained by the fact that most of them were recruited

Table 5 Diagnostic performance of miR-221 for discriminatingpatients with hepatocellular carcinoma from those without							
AUC	P value	Best cutoff	Sensitivity	Specificity	PPV	NPV	
0.655	0.03	< 1.82	0.87	0.40	0.47	0.83	

HCC: Hepatocellular carcinoma; PVP: Positive predictive value; NVP: Negative predictive value.

while being assessed for the possibility of interventional treatment. Another possible explanation for rather good liver condition seen in HCC series could be attributed to that with implementing surveillance programs, allowing detecting tumors at an early stage in well compensated patients. Alfa fetoprotein level was normal (< 10 ng/ dL) in 13.4% of recruited HCC patients. Similar finding was observed by Tateishi *et al*¹⁴ who suggested that not all tumors secrete AFP, and serum levels are normal in up to 40% of small HCCs. It was also showed that α -Fetoprotein alone is not recommended for the diagnosis of HCC and studies showed that its cut off value should be set at 200 ng/mL.

Analysis of fold changes in expression level of miR-122 displayed significant fold increase in expression level in chronic hepatitis C group (2.1) and significant fold decrease in expression in cirrhotics (0.8) in comparison to normal controls. miR-122 is present abundantly in hepatocytes with much lower levels in the circulation in healthy subjects. With hepatocyte injury miR-122 is released in the circulation more readily and serum levels rise. With the eventual loss of hepatocytes and development of fibrosis with proliferation of myelofibroblasts and accumulation of extracellular matrix the circulating miR-122 levels drop again^[15].

In the current study there was significant fold rise in serum expression level of miR-122 in HCC group in comparison to normal control group (*P* value < 0.01). Matching our results, Trebicka *et al*^{15]} who studied hepatic miR-122 expression in 43 HCV related HCC in comparison to 3 healthy liver samples using qRT-PCR; miR-122 was strongly up-regulated in malignant liver nodules in comparison to healthy liver. They suggested that miR-122 might down regulate target mRNA of unknown tumor suppressor genes and thus lead to further tumor growth^[15].

In a study on hepatitis B patients Xu *et al*^{16]} suggested that cancer-induced hepatocyte damage would release the abundant intracellular miR-122 into the circulation, the stability of miRNA would be reflected by easily detectable high blood levels^[17]. In contrast to our results, significant down regulation of miR-122 in HCC compared to normal liver tissue was reported by Meng *et al*^{18]}, Wang *et al*^{19]} and Huang *et al*^{20]} who compared miR-122 expression profile of 3 different pairs of tumor and normal human liver-derived RNA and 20 HCC liver tissues (mixed etiologies) to normal tissues respectively using microarray^[18,19,20]. Similarly a significant down regulation in miR-122 in 19 HBV related HCC liver tissue in comparison to paired healthy liver by next-generation sequencing was reported by Connolly *et al*^[21].</sup>

Ladeiro *et al*^{22]} have established significant down expression of miR-122 in 28 HCC liver tissues (mixed etiologies) in comparison to 4 healthy liver tissues by qRT-PCR.

In our series, no statistically significant correlation could be verified between serum miR122 expression level and patient characters (age), liver synthetic functions tests (Albumin, bilirubin and PC), or serum AFP level in HCC *vs* non HCC group (CH and cirrhosis). However, in the chronic hepatitis groups serum miR-122 was correlated with higher AST and ALT levels, further solidifying the theory regarding the initial rise in miR-122 levels due to hepatocyte inflammation and destruction followed by a drop in the levels with the developing fibrosis. Köberle *et al*^[23] also reported significant correlation between serum miR-122 expression level and necro-inflammatory markers (AST, ALT), and Albumin but no significant correlation was found with bilirubin in HCC patients^[23].

Perhaps the most significant finding in our study was related to miR-221. Analysis of fold change in expression level of miR-221 in patients' sera of HCV associated liver disease (CH and cirrhosis) in comparison to normal control group showed significant fold increase in expression level in CH and cirrhosis groups in comparison to normal control group (< 0.01). Also a significant fold decrease in serum miR-221 in HCC group (0.92) in comparison to normal control was noticed. We assumed that with the progression of liver disease from chronic hepatitis to cirrhosis the increased activity of hepatic stellate cells was associated with increase miR-221 expression level, such high level stimulated tumorigenesis and increase level of miR-221 in tissue, but as miR-221 is anti apoptotic so serum miR-221 didn't show similar increase. In contrast to our results many studies established up regulation of miR-221 in HCC in relation to normal control, e.g.,^[18,19,20,24]. However, most these studies assessed tissue miR-221 rather than serum levels .The different results could also be explained by technical variations including sampling methods and freezing and RNA isolation procedures. The etiology of liver disease is also variable in different studies including viral and alcoholic. The stage of the disease is also a source of variation especially that it is still not evident how miRNA expression changes with fibrosis progression. Different studies have also used different control samples for normalization, e.g., non-HCC tissue from the same patient, healthy liver tissue from another subject or patients with the same pathology but not HCC, this is especialy relevant to studies assessing tissue miRNA levels^[25]

Similar to what was previously reported by Rong *et* at^{26} , we found no statistically significant correlation could be verified between serum miR-221 expression level and patient characters (age), laboratory values (AST, ALT), liver synthetic functions tests (Albumin, bilirubin and PC), or serum a-fetoprotein level in HCC *vs* non HCC group (CH and cirrhosis) and no statistically significant correlation could be found between the clinic-pathological parameters of hepatic focal lesion, *e.g.*, (number of focal lesions, Child

score, biggest diameter of focal lesion BCLC, and portal vein invasion) and miR-221 expression level $(P \ge 0.05)^{|26|}$.

Circulating miR-221 level is significantly up-regulated in the serum of HCV infected patients. It has some value in the differentiation between HCV patients with hepatocellular carcinoma and those without with 87% sensitivity and 40% specificity. It may be able to serve as a promising non-invasive diagnostic marker for HCC. Better results could be obtained if combined with other markers and testing a panel of miRNA's collectively could ultimately serve as a reliable diagnostic test for HCC.

COMMENTS

Background

Hepatocellular carcinoma (HCC), the most common type of liver cancer, is amongst the top three leading causes of cancer-related deaths worldwide with a median survival of only six to eight months. This poor outcome is related to the late detection, with more than two thirds of patients diagnosed at advanced stages of disease. Thus, surveillance of populations at-risk may detect tumors at an early stage when curative interventions can be implemented. The performance of available circulating biomarkers in the screening and diagnostic settings of HCC is sub-optimal.

Research frontiers

MiRNAs constitute a large class of genes that encode short RNAs (19-24 nucleotides long), which play key roles in development and differentiation, by the post-transcriptional regulation of protein coding genes. At present, miRNAs have a widely recognized role in human carcinogenesis, including hepatocarcinogenesis, and many experimental evidences indicate that they may act as oncogenes or tumor suppressor genes regulating the expression of crucial protein coding genes. MiRNAs have been proposed as possible novel biomarkers for cancer diagnosis.

Innovations and breakthroughs

In the current study a signature of circulating miRNAs (miR-122 and miR-221) was evaluated. MiR-221 was differentially expressed between patients with HCC and those without (chronic hepatitis and cirrhosis) with lower serum level of miR-221 in former group of patients in comparison to later one. MiR-221 yielded 87% sensitivity and 40% specificity in differentiating between both groups at a cutoff 1.82 folds.

Applications

The present study emphasis that circulating miR-221 deserves much attention as potential non invasive biomarkers for HCC in the diagnostic setting.

Terminology

HCC: Hepatocellular carcinoma; Non HCC: Chronic hepatis C group of patients and patients with liver cirrhosis.

Peer review

The manuscript entitled "Circulating microRNA, miR-122 and miR221 Signature in Egyptian Patients with Chronic Hepatitis C Related Hepatocellular Carcinoma". The manuscript is interesting.

REFERENCES

- 1 **Parkin DM**, Bray FI, Devesa SS. Cancer burden in the year 2000. The global picture. *Eur J Cancer* 2001; **37** Suppl 8: S4-S66 [PMID: 11602373 DOI: 10.1016/S0959-8049(01)00267-2]
- 2 El-Serag HB. Hepatocellular carcinoma: an epidemiologic view. J Clin Gastroenterol 2002; 35: S72-S78 [PMID: 12394209 DOI: 10.1097/00004836-200211002-000028]
- 3 Velázquez RF, Rodríguez M, Navascués CA, Linares A, Pérez R, Sotorríos NG, Martínez I, Rodrigo L. Prospective analysis of risk factors for hepatocellular carcinoma in patients with liver cirrhosis. *Hepatology* 2003; 37: 520-527 [PMID: 12601348 DOI: 10.1053/jhep.2003.50093]
- 4 Bruix J, Sherman M. Management of hepatocellular carcinoma. *Hepatology* 2005; **42**: 1208-1236 [PMID: 16250051 DOI: 10.1002/hep.20933]

- 5 El-Attar IA. Cancer databases in the Arab world. *Ethn Dis* 2005; **15**: S1-3-S1-4 [PMID: 15787030]
- 6 El-Nady GM, Ling R, Harrison TJ. Gene expression in HCVassociated hepatocellular carcinoma--upregulation of a gene encoding a protein related to the ubiquitin-conjugating enzyme. *Liver Int* 2003; 23: 329-337 [PMID: 14708893 DOI: 10.1034/j.1478-3231.2003.00862.x]
- 7 Fattovich G, Giustina G, Sanchez-Tapias J, Quero C, Mas A, Olivotto PG, Solinas A, Almasio P, Hadziyannis S, Degos F, de Moura MC, Krogsgaard K, Pantalena M, Realdi G, Corrocher R, Schalm SW. Delayed clearance of serum HBsAg in compensated cirrhosis B: relation to interferon alpha therapy and disease prognosis. European Concerted Action on Viral Hepatitis (EUROHEP) Am J Gastroenterol 1998; 93: 896-900 [PMID: 9647014 DOI: 10.1111/j.1572-0241.1998.00272.x]
- 8 Marrero JA, Feng Z, Wang Y, Nguyen MH, Befeler AS, Roberts LR, Reddy KR, Harnois D, Llovet JM, Normolle D, Dalhgren J, Chia D, Lok AS, Wagner PD, Srivastava S, Schwartz M. Alpha-fetoprotein, des-gamma carboxyprothrombin, and lectin-bound alpha-fetoprotein in early hepatocellular carcinoma. *Gastroenterology* 2009; **137**: 110-118 [PMID: 19362088 DOI: 10.1053/j.gastro.2009.04.005]
- 9 Okuda H, Nakanishi T, Takatsu K, Saito A, Hayashi N, Takasaki K, Takenami K, Yamamoto M, Nakano M. Serum levels of des-gamma-carboxy prothrombin measured using the revised enzyme immunoassay kit with increased sensitivity in relation to clinicopathologic features of solitary hepatocellular carcinoma. *Cancer* 2000; 88: 544-549 [PMID: 10649245 DOI: 10.1002/(SICI)1097-0142(2000201)88]
- 10 Marrero JA, Lok AS. Newer markers for hepatocellular carcinoma. *Gastroenterology* 2004; **127**: S113-S119 [PMID: 15508074 DOI: 10.1053/j.gastro.2004.09.024]
- 11 Bushati N, Cohen SM. microRNA functions. Annu Rev Cell Dev Biol 2007; 23: 175-205 [PMID: 17506695 DOI: 10.1146/annurev.cellbio.23.090506.123406]
- 12 Llovet JM, Burroughs A, Bruix J. Hepatocellular carcinoma. Lancet 2003; 362: 1907-1917 [PMID: 14667750 DOI: 10.1016/ S0140-6736(03)14964-1]
- 13 Chen DS, Sung JL, Sheu JC, Lai MY, How SW, Hsu HC, Lee CS, Wei TC. Serum alpha-fetoprotein in the early stage of human hepatocellular carcinoma. *Gastroenterology* 1984; 86: 1404-1409 [PMID: 6201411]
- 14 Tateishi R, Yoshida H, Matsuyama Y, Mine N, Kondo Y, Omata M. Diagnostic accuracy of tumor markers for hepatocellular carcinoma: a systematic review. *Hepatol Int* 2008; 2: 17-30 [PMID: 19669276 DOI: 10.1007/s12072-007-9038-x]
- 15 Trebicka J, Anadol E, Elfimova N, Strack I, Roggendorf M, Viazov S, Wedemeyer I, Drebber U, Rockstroh J, Sauerbruch T, Dienes HP, Odenthal M. Hepatic and serum levels of miR-122 after chronic HCV-induced fibrosis. *J Hepatol* 2013; 58: 234-239 [PMID: 23085648 DOI: 10.1016/j.jhep.2012.10.015]
- 16 Xu J, Wu C, Che X, Wang L, Yu D, Zhang T, Huang L, Li H, Tan W, Wang C, Lin D. Circulating microRNAs, miR-21, miR-122, and miR-223, in patients with hepatocellular carcinoma or chronic hepatitis. *Mol Carcinog* 2011; 50: 136-142 [PMID: 21229610 DOI: 10.1002/mc.20712]
- 17 Varnholt H, Drebber U, Schulze F, Wedemeyer I, Schirmacher P, Dienes HP, Odenthal M. MicroRNA gene expression profile of hepatitis C virus-associated hepatocellular carcinoma. *Hepatology* 2008; 47: 1223-1232 [PMID: 18307259 DOI: 10.1002/hep.22158]
- 18 Meng F, Henson R, Wehbe-Janek H, Ghoshal K, Jacob ST, Patel T. MicroRNA-21 regulates expression of the PTEN tumor suppressor gene in human hepatocellular cancer. *Gastroenterology* 2007; **133**: 647-658 [PMID: 17681183 DOI: 10.1053/ j.gastro.2007.05.022]
- 19 Wang Y, Lee AT, Ma JZ, Wang J, Ren J, Yang Y, Tantoso E, Li KB, Ooi LL, Tan P, Lee CG. Profiling microRNA expression in hepatocellular carcinoma reveals microRNA-224 up-regulation and apoptosis inhibitor-5 as a microRNA-224-specific

target. J Biol Chem 2008; **283**: 13205-13215 [PMID: 18319255 DOI: 10.1074/jbc.M707629200]

- 20 Huang XH, Wang Q, Chen JS, Fu XH, Chen XL, Chen LZ, Li W, Bi J, Zhang LJ, Fu Q, Zeng WT, Cao LQ, Tan HX, Su Q. Bead-based microarray analysis of microRNA expression in hepatocellular carcinoma: miR-338 is downregulated. *Hepatol Res* 2009; **39**: 786-794 [PMID: 19473441 DOI: 10.1111/j.1872-034X.2009.00502.x]
- 21 Connolly E, Melegari M, Landgraf P, Tchaikovskaya T, Tennant BC, Slagle BL, Rogler LE, Zavolan M, Tuschl T, Rogler CE. Elevated expression of the miR-17-92 polycistron and miR-21 in hepadnavirus-associated hepatocellular carcinoma contributes to the malignant phenotype. *Am J Pathol* 2008; **173**: 856-864 [PMID: 18688024 DOI: 10.2353/ajpath.2008.080096]
- 22 Ladeiro Y, Couchy G, Balabaud C, Bioulac-Sage P, Pelletier L, Rebouissou S, Zucman-Rossi J. MicroRNA profiling in hepatocellular tumors is associated with clinical features and oncogene/tumor suppressor gene mutations. *Hepatology* 2008;

47: 1955-1963 [PMID: 18433021 DOI: 10.1002/hep.22256]

- 23 Köberle V, Kronenberger B, Pleli T, Trojan J, Imelmann E, Peveling-Oberhag J, Welker MW, Elhendawy M, Zeuzem S, Piiper A, Waidmann O. Serum microRNA-1 and microRNA-122 are prognostic markers in patients with hepatocellular carcinoma. *Eur J Cancer* 2013; 49: 3442-3449 [PMID: 23810247 DOI: 10.1016/j.ejca.2013.06.002]
- 24 **Zekri A**, Youssef AS, Ahmed O. Serum MicroRNAs as differential markers and a molecular therapeutic target for HCV associated liver disease. *AASLD* 2013; poster
- 25 **Borel F**, Konstantinova P, Jansen PL. Diagnostic and therapeutic potential of miRNA signatures in patients with hepatocellular carcinoma. *J Hepatol* 2012; **56**: 1371-1383 [PMID: 22314424 DOI: 10.1016/j.jhep.2011.11.026]
- 26 Rong M, Chen G, Dang Y. Increased miR-221 expression in hepatocellular carcinoma tissues and its role in enhancing cell growth and inhibiting apoptosis in vitro. *BMC Cancer* 2013; **13**: 21 [PMID: 23320393 DOI: 10.1186/1471-2407-13-21]

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