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Location, Location, Location: Macrophage positioning within tumors determines pro-or anti-tumor activity

Lee Rivera¹ and Gabriele Bergers^{1,*}

¹Department of Neurological Surgery, Brain Tumor Research Center, Helen Diller Family Comprehensive Cancer Center, University of California, San Francisco, San Francisco, CA 94158, USA

Abstract

Macrophages infiltrate hypoxic tumor regions where they promote angiogenesis and immunosuppression. In this issue of *Cancer Cell*, Mazzone and colleagues report that tumor-associated macrophage (TAM) entry into avascular tumor areas is regulated by Semaphorin 3A/Neuropilin-1 signaling; interference with this pathway entraps TAMs in oxygenated areas preventing their tumorigenic function.

Tumor-associated macrophages (TAMs) are tissue-resident cells that differentiate from circulating monocytes in peripheral blood. They can constitute the major leukocytic infiltrate found within the stroma of many tumor types. Although macrophages in normal tissues are implicated in phagocytosis of microbes and antigen presentation to T cells, TAMs have two opposing phenotypes, they can either endorse proimmune and tumoricidal processes or promote tumor growth and metastasis by suppressing immunity and promoting angiogenesis. The phenotype of TAMs is regulated by specific tumor-derived chemokines and cytokines that polarize macrophages to a proimmune ‘M1’ or immunosuppressive/proangiogenic ‘M2’ phenotype. The dichotomous TAM phenotypes may explain why TAMs can elicit a poor prognosis in some tumors including glioma and breast cancers and a better prognosis in others such as stomach and colon cancers and some prostate and non-small cell lung cancers (Allavena et al., 2008; Bingle et al., 2002). Macrophage polarization is also in part regulated by intratumoral hypoxia, in which infiltrating myeloid cells accumulate and are stimulated to secrete various immune suppressive and proangiogenic factors (De Palma and Lewis, 2013; Qian and Pollard, 2010).

In this issue of *Cancer Cell* Casazza et al. describe a Neuropilin-1 (Nrp1)-dependent guidance mechanism by which macrophages enter hypoxic areas to elicit proangiogenic and immune suppressive functions (Figure 1) (Casazza et al. 2013). Utilizing elegant genetic tools to interfere with Nrp1 function in TAMs in various mouse tumor models they demonstrated

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*Correspondence: gabriele.bergers@ucsf.edu.

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that Semaphorin3A (Sema3A) mediates Nrp-1-dependent signaling of a PlexinA1/PlexinA4/VEGFR1 holoreceptor complex that leads to VEGFR1 activation in TAMs and their subsequent migration into hypoxic regions. Notably, although Sema3A and VEGF levels are both increased under hypoxic conditions, Sema3A, but not VEGF, was sufficient to attract TAMs. They tested this by generating TAMs with a Sema3A-binding mutant of Nrp1 that was still able to bind to VEGF. These macrophages failed to enter hypoxic regions of the tumor similarly to *Nrp1-KO* TAMs. As soon as TAMs were positioned in the hypoxic environment, Nrp1 expression was repressed; this terminated the migratory response of TAMs to Sema3A. Interestingly, hypoxia-dependent Nrp1 repression was facilitated by HIF2 α -mediated activation of the NF- κ B pathway. The loss of Nrp1 switched Sema3A to mediating a PlexinA1/PlexinA4-mediated TAM arrest antagonizing VEGFR1-induced attraction and entrapping TAMs in hypoxic regions (Figure 1). As TAMs shift from an anti- to a pro-tumoral phenotype upon association with hypoxic environments, the authors then asked how loss of Nrp1 in TAMs and their subsequent differing positioning within tumors would affect tumor propagation and progression.

Casazza et al. explored the function of Nrp1 on TAMs by creating conditional TAM-specific *Nrp1*-knockout (*KO*) mice. Orthotopic lung and pancreatic tumors, and tumors from a transgenic breast cancer mouse model, in *Nrp1-KO* mice grew to only a fraction of the size of tumors in wildtype (WT) mice. Nrp-1 deficiency in TAMs yielded tumors with nearly double the number of TAMs, likely due to increased tumor hypoxia. Surprisingly, however, TAMs solely accumulated within normoxic regions. Moreover, despite the increase in TAMs, endstage tumors exhibited reduced vessel density and perfusion, suggesting that *Nrp1-KO* TAMs were impaired in their angiogenic functions compared to their WT counterparts. Indeed, isolated WT TAMs induced more robust endothelial cell migration and capillary formation compared to *Nrp1-KO* TAMs. In addition, *Nrp1-KO* TAMs secreted more nitric oxide, increased T cell proliferation and were more cytotoxic. Interestingly, Casazza et al. found that the acquired “M1” TAM phenotype was not endorsed by the lack of Nrp1 *per se* because WT and *Nrp1-KO* macrophages obtained from bone marrow were equally able to switch between pro-immune and immune suppressive phenotypes upon appropriate stimulation *in vitro*. Further *Nrp1* deficiency in TAMs neither affected the numbers of circulating or resident monocyte numbers nor changed proliferation and apoptosis of macrophages precluding a Nrp1-dependent regulation of monocyte/TAM recruitment or differentiation. Rather, these elegant studies revealed that blocking Nrp1 in TAMs was sufficient to keep the cells in a tumor-suppressive state by solely entrapping the cells in vascularized normoxic tumor areas.

These studies support the concept of macrophage “reprogramming” as a sufficient and feasible approach to abrogate angiogenesis and restore T cell-mediated antitumor immunity (Coussens et al., 2013). Further, Casazza et al. provide a new therapeutic opportunity to turn TAMs against cancer by modulating their intratumoral location via inhibition of Nrp1. Such an approach is advantageous over those that target total TAM infiltration as it harnesses the tumor suppressing capacities of TAMs.

These studies also have important clinical applications. Although historically successful tumor eradication had been linked with tumor necrosis, various studies have demonstrated

that hypoxia-generating drugs cause more aggressive disease in part by accumulating more immune suppressive innate immune cells that facilitate angiogenesis, tumor invasion and metastasis. Emerging data support the notion that normalization of the tumor vasculature provides beneficial effects enabling better drug delivery and enhanced influx of T cells. A recent study by Klug et al. demonstrated that low-dose irradiation and T cell transfer normalized the tumor vasculature and enhanced the recruitment CD8⁺ T cells and TAMs expressing high levels of the M1 marker iNOS (Klug et al., 2013). Similarly, Casazza et al. found that normoxia enhanced secretion of nitric oxide by TAMs and induced CD8⁺ T cell expansion. Thus oxygenation of the tumor should also help to redirect macrophage differentiation to facilitate anti-tumor immunity. In addition, targeting Nrp1 would restrict the TAMs to oxygenated areas even during hypoxia-inducing therapies including standard chemo- and radiation therapy.

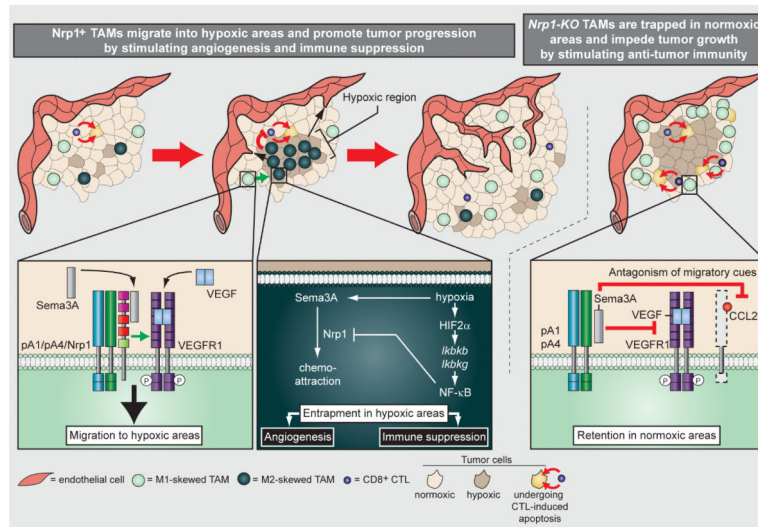
The authors confirm that exposure of TAMs to hypoxia is a requisite for their acquisition of a tumor promoting phenotype; however, whether or not hypoxia directly regulates M2 reprogramming is unclear. A recent study by Laoui et al. suggests hypoxia plays a supportive rather than a direct role in driving M2 functions by TAMs (Laoui et al., 2013). Using prolyl-4 hydroxylase 2-haplodeficient mice, this group found that reduced tumor hypoxia resulted in downregulated TAM expression of genes involved in glycolysis, angiogenesis, and metastasis, and not in typical M2 markers including mannose receptor and arginase. This suggests targeting the Nrp1/Sema3A axis can synergize with reprogramming approaches to provide better TAM-mediated anti-tumor responses.

In support of these hypotheses, blockade of Nrp1 in preclinical tumor models has been encouraging, suppressing both angiogenesis and tumor growth, and clinical trials are currently ongoing (Pan et al., 2007). As Nrp1 is expressed in a variety of cell types besides TAMs, including endothelial cells and tumor cells, it will be pivotal to analyze whether the mechanism proposed by this study is still evident when Nrp1 activity is broadly abrogated in murine tumor models and human tumors. Whether TAM location and activity is similarly regulated in other hypoxia-generating pathologies also warrants further investigation. For example, in a mouse model of cerebral stroke, microglia and macrophages were found to undergo M2 polarization immediately after ischemic insult, but eventually underwent M1 polarization induced by ischemic neurons (Huang and Feng, 2013). The M2 polarized cells were found to have a protective effect on neurons whereas M1 polarized cells promoted neuronal destruction; therefore preventing microglia and macrophages from associating with ischemic areas might maintain their neuronal-protective phenotypes. If validated, manipulation of the Nrp1/Sema3A axis could become a valuable agent for diseases like ischemia and stroke to redirect macrophage function and improve patient outcome.

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Sema3A/Nrp1 signaling regulates TAM entry into hypoxic regions and thereby promotes tumor progression

Intratumoral hypoxia enhances the expression of VEGF and Sema3A. Sema3A binds to the Nrp1/PlexinA1 (pA1)/PlexinA4 (pA4)holoreceptor complex at the TAM surface, resulting in VEGFR1/Nrp1-dependent migration towards the Sema3A-expressing hypoxic area. Hypoxia-associated TAMs experience stabilization of HIF2 α , which induces expression of *Ikbkb* and *Ikbkg*, ultimately leading to phosphorylation of I κ B and nuclear translocation of NF- κ B. NF- κ B then represses expression of *Nrp1*. In the absence of Nrp1, Sema3A antagonizes migration signals through PlexinA1/PlexinA4 signaling, thus retaining and entrapping TAMs within hypoxic areas. Here, TAMs are “educated” to endorse angiogenesis and suppress anti-tumor immunity, thus facilitating tumor progression. Sema3A/PlexinA1/PlexinA4 retention signals entrap *Nrp1-KO* TAMs in normoxic areas by blocking VEGF-mediated migration into hypoxic regions. *Nrp1-KO* TAMs therefore do not attain a tumor promoting phenotype and suppress tumor growth by stimulating anti-tumor immunity.