



Published in final edited form as:

*Semin Immunol.* 2014 October ; 26(5): 380–388. doi:10.1016/j.smim.2014.04.001.

## Stress Regulates Endocannabinoid-CB1 Receptor Signaling

Cecilia J. Hillard

Neuroscience Research Center and Department of Pharmacology and Toxicology, Medical College of Wisconsin

### Abstract

The CB1 cannabinoid receptor is a G protein coupled receptor that is widely expressed throughout the brain. The endogenous ligands for the CB1 receptor (endocannabinoids) are *N*-arachidonylethanolamine and 2-arachidonoylglycerol; together the endocannabinoids and CB1R subserve activity dependent, retrograde inhibition of neurotransmitter release in the brain. Deficiency of CB1 receptor signaling is associated with anhedonia, anxiety, and persistence of negative memories. CB1 receptor-endocannabinoid signaling is activated by stress and functions to buffer or dampen the behavioral and endocrine effects of acute stress. Its role in regulation of neuronal responses is more complex. Chronic variable stress exposure reduces endocannabinoid-CB1 receptor signaling and it is hypothesized that the resultant deficiency in endocannabinoid signaling contributes to the negative consequences of chronic stress. On the other hand, repeated exposure to the same stress can sensitize CB1 receptor signaling, resulting in dampening of the stress response. Data are reviewed that support the hypothesis that CB1 receptor signaling is stress responsive and that maintaining robust endocannabinoid/CB1 receptor signaling provides resilience against the development of stress-related pathologies.

### 1. Introduction: Stress

Physical and psychological threats to the well-being of an individual induce a pattern of physiological responses that are designed to cope with the immediate stress, avoid future threats, and facilitate restoration of homeostasis. Stress exposure results in a broad and significant impact on physiological and psychological function designed to increase chances for escape, and for survival if an injury does occur. As a result of sympathetic nervous system activation, stress exposure increases heart rate, blood pressure and blood flow to muscles. Activation of the sympathetic nervous system (SNS) activation also promotes a pro-inflammatory environment in the periphery and brain [1]. For example, exposure of healthy males to social stress increases circulating T cells and concentrations of pro-inflammatory cytokines, including interleukin-1 $\beta$  [2]. In addition, stress exposure activates the hypothalamic-pituitary-adrenocortical (HPA) axis, and thereby increases circulating

© 2014 Elsevier Ltd. All rights reserved.

Contact information: Cecilia J. Hillard, Ph.D., Neuroscience Research Center, Medical College of Wisconsin, 8701 Watertown Plank Road, Milwaukee, WI 53226, Phone: 414-955-8493, Fax: 414-955-6057, chillard@mcw.edu.

**Publisher's Disclaimer:** This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

glucocorticoid concentrations. The glucocorticoids (cortisol in humans and corticosterone (CORT) in rodents) are steroids released by the adrenal cortex that produce physiological changes across a time range from seconds to hours through non-genomic and genomic mechanisms [3, 4]. Glucocorticoids affect memory and mood and alter the cardiovascular, metabolic and immune systems largely through effects on glucocorticoid receptors (GR).

Although these stress responses can be life-preserving in the face of a threat, they are costly when triggered repeatedly. In fact, chronic exposure to stress is a significant risk factor for psychiatric [5], metabolic [6], and functional pain disorders [7], as well as for neurodegeneration [8]. There is considerable individual vulnerability to the negative consequences of chronic stress. Resilience is associated with coping styles that enable individuals to maintain hedonia and optimism; to continue employing effective behavioral repertoires despite fear; and is associated with positive social interactions [9, 10]. On the other hand, hyperactive HPA responses to stress are associated with increased vulnerability to develop the negative consequences of chronic stress [11]. Importantly, coping styles and HPA axis responsivity are shaped by previous stress exposure and are particularly sensitive to early life stress [12].

## 2. Introduction: CB1 Receptor Mediated Endocannabinoid Signaling

### 2.1 The CB1 Cannabinoid Receptor

<sup>9</sup>-Tetrahydrocannabinol (THC) is the component of *cannabis sativa* that is responsible for the psychoactive effects of the plant [13]. The effects of THC on cognition and mood are mediated by its ability to act as an agonist of a G protein coupled receptor, named the CB1 cannabinoid receptor (CB1R) [14, 15]. The CB1R is present on many neuronal subtypes throughout the brain, and in peripheral nerves as well [16]. Very high density CB1R protein expression occurs in the cingulate gyrus, prefrontal cortex (PFC), hippocampus, cerebellum, basal ganglia and substantia nigra; moderate density is seen in the basal forebrain, amygdala, nucleus accumbens, periaqueductal gray and hypothalamus; and low density occurs in brain stem regions, primary motor cortex and thalamus [17]. The psychoactive effects of *cannabis sativa* in humans are completely antagonized by a selective antagonist of the CB1R [18].

Outside of the brain, CB1R are present in the dorsal horn of the spinal cord; specifically found on interneurons and axon terminals of descending inputs and peripheral afferents [17]. Primary sensory afferents also express the CB1R on terminals in innervated tissues [19]. Functional data indicate that sympathetic nerves express CB1R [20]; and CB1R are distributed throughout the neurons of the enteric nervous system [21].

Non-neuronal cells in the brain also express CB1R, including astrocytes [22], oligodendrocytes [23] and cells of the cerebral vasculature [24, 25]. Outside of the CNS, CB1R are expressed by circulating immune cells [26], adipocytes [27], hepatocytes [28], and adrenal cortex [29]. Thus, CB1R activation by exogenous or endogenous agonists has the potential to modify function of many organs, most particularly the brain.

## 2.2 The Endocannabinoids

In 1992, a low abundance member of the family of *N*-acylethanolamines (NAEs) was shown to function as an endogenous agonist of the CB1R [30]. This lipid, *N*-arachidonylethanolamine (AEA, also called anandamide), functions as a partial agonist of CB1R [31]. A second arachidonate, 2-arachidonoylglycerol (2-AG), was identified a short time later [32, 33]. 2-AG functions as a fully efficacious agonist of the CB1R [34]. Both AEA and 2-AG are considered endocannabinoids (eCBs), although their specific roles as regulators of CB1R signaling are not well understood [35].

AEA is synthesized from a minor phospholipid, *N*-arachidonyl-phosphatidylethanolamine (NAPE) [36], via several possible enzymatic processes [37]. The regulatory mechanisms of these synthetic pathways are not well understood. 2-AG is biosynthesized from diacylglycerol (DAG), through the actions of sn-1 specific DAG lipases (DAGL $\alpha$  and  $\beta$ ) [38]. DAG is produced from phosphatidylinositol-bis-phosphate, which is metabolized by phospholipase C (PLC) to DAG and inositol triphosphate. DAGL $\alpha$ , which is thought to be the rate limiting step in 2-AG synthesis, is present on the postsynaptic membrane of neurons that make synaptic contact with neurons expressing the CB1R [39].

Two amidohydrolases have been identified that hydrolyze and thus, inactivate, AEA. The first, fatty acid amide hydrolase (FAAH), is present in the brain [40] and is an important regulator of brain concentrations of AEA as well as other NAEs [41, 42]. The second amidohydrolase, *N*-acylethanolamine-hydrolyzing acid amidase, hydrolyzes AEA in the periphery [43]. 2-AG is also inactivated by hydrolysis; more than 80% of the serine hydrolase-mediated metabolism of 2-AG in the brain is accomplished by monoacylglycerol lipase (MAGL) [44]. Another important serine hydrolase that metabolizes 2-AG is alpha-beta hydrolase 6 [45]. Both AEA and 2-AG are also substrates for cyclooxygenase 2 (COX2); the oxygenation of the eCBs by COX2 results in inactivation with respect to CB1R signaling [46].

## 2.3 Endocannabinoid Regulation of Synaptic Activity

There are considerable data supporting an important role for eCB/CB1R signaling in the regulation of synaptic plasticity. eCB signaling (ECS) underlies retrograde, activity-dependent, suppression of neurotransmitter release in many regions of the brain [47]. In brief, activation of CB1R on presynaptic terminals produce long- and short-term inhibition of vesicular transmitter release. The short term effects are mediated by CB1R-initiated, G protein-mediated inhibition of the opening of voltage operated calcium channels. The long term effects likely involve changes in the machinery regulating neurotransmitter release. As a result, the activation of CB1R on presynaptic terminals results in inhibition of neurotransmitter release for variable lengths of time.

Activation of presynaptic CB1R signaling occurs as a result of increased eCB mobilization. 2-AG is synthesized by post-synaptic neurons in response to several triggers, including receptors that activate PLC [48]; increased calcium concentrations, secondary to neuronal activity [47] and glucocorticoids (discussed in section 2.4.). The mechanisms and cellular site of AEA synthesis are not well understood. Since both 2-AG and AEA are highly

lipophilic, increased intracellular concentrations drive increased extracellular concentrations; in other words, synthesis and release are coupled. The eCBs can diffuse from the synthetic cells and inhibit neurotransmitter release from nearby neurons that express CB1R. CB1R are present on glutamatergic [49], GABAergic [50], serotonergic [51] and noradrenergic [52] axon terminals in the brain; thus ECS can regulate the release of the primary neurotransmitters involved in processing of stress and fear.

#### 2.4. Glucocorticoids Alter Synaptic Activity via ECS

Glucocorticoids rapidly modulate glutamatergic [53, 54] and GABAergic [55] transmission in stress-sensitive brain regions through alteration in the release of neurotransmitters. Recent studies support the hypothesis that glucocorticoids modulate the release of glutamate and GABA as a result of enhanced ECS. In the hypothalamus, 2-AG concentrations are significantly elevated by restraint stress [56] and by CORT treatment [57]. Evidence suggests that glucocorticoids act through a membrane receptor to rapidly mobilize eCBs [58]. The consequence of ECS activation is to reduce the release of glutamate and, thus reduce excitatory drive onto corticotropin releasing hormone (CRH) neurons in the paraventricular nucleus (PVN) [58, 59]. This mechanism is hypothesized to contribute to glucocorticoid-mediated feedback inhibition of HPA axis activation at the level of the hypothalamus. Recent data also extend the glucocorticoid-eCB functional pair to regulation of excitatory transmission onto serotonergic neurons of the dorsal raphe [60]. Stress exposure also increases concentrations of 2-AG in the PFC and hippocampus. Glucocorticoid treatment of slices taken from PFC [61] and hippocampus [62] results in CB1R-mediated inhibition of GABA release.

These studies support the hypothesis that eCBs function as a second messenger for glucocorticoids in stress-responsive brain regions; in particular, allowing glucocorticoid presence to produce rapid changes in synaptic activity [63]. Available data indicate that glucocorticoids increase 2-AG concentrations; however, it seems that different glucocorticoid receptor sub-types and signaling cascades are employed, allowing for the specific pattern and time-course of eCB mobilization to be matched to the function of the glucocorticoids at a particular synapse. COX2 can oxidize both AEA and 2-AG [46] and its inhibition has been shown to increase ECS in the hippocampus [64, 65] and dorsal raphe [60]. There is evidence that COX2 is negatively regulated by glucocorticoids [66] supporting the possibility that glucocorticoids can regulate concentrations of the eCBs available to signal via alterations in the activity of COX2. This mechanism could be important for linking changes in inflammatory state with changes in synaptic activity.

### 3. Regulation of Mood, Cognition and Stress by CB1R Signaling

Preclinical and human data demonstrate that eCB/CB1R signaling is required for regulation of stress responses and mood. This conclusion is primarily supported by studies in which eCB/CB1R signaling is reduced with pharmacologic antagonists or genetic deletion. CB1R blockade and deletion results in excessive [67, 68] and prolonged [61] activation of the HPA axis by stress. These data are in accord with the discussion in section 2.4 that ECS in several brain regions subserves glucocorticoid-mediated feedback inhibition in multiple brain regions.

Inhibition of eCB/CB1R signaling can result in increased anxiety. Rodents treated with a CB1R antagonist demonstrate increased anxiety-like behaviors [69]. Importantly, anxiety was a significant adverse effect in humans treated with a CB1R antagonist for metabolic disorder and obesity [70], and treatment of healthy humans prior to an experimental stress paradigm results in increased anxiety [71]. Reduced eCB/CB1R signaling also results in delayed and ineffective extinction of fearful memories in rodents [72]; is associated with reduced behavioral flexibility [73] and with anhedonia [74]. A subset of humans exposed to CB1R antagonists, particularly those with prior depressive symptoms, exhibited increased depressive symptoms, including suicidality [75]. Furthermore, women with depression exhibit significantly reduced 2-AG concentrations in the circulation [76], further evidence that hypoactive ECS is associated with negative affect.

These and other data strongly support the hypothesis that robust eCB/CB1R signaling is vital for appropriate stress responses and for the maintenance of emotional homeostasis, particularly in the face of chronic stress.

#### 4. Regulation of ECS by Stress

The data described below support the role of ECS as a homeostatic mechanism that both inhibits unnecessary HPA axis activation through actions in the amygdala and promotes the recovery of the HPA axis to baseline after the threat has ended. While neither of these properties is absolutely required for the endocrine response to occur, loss of ECS would be expected to increase the "wear and tear" of stress on the brain because the stress response is activated with less provocation and remains in an active state for longer periods of time.

The ECS stress buffer is reduced by chronic stress exposure. Chronic stress is a significant risk factor for the development of psychopathology, and there are considerable data to suggest that loss of ECS results in symptoms of depression and anxiety [77]. Thus, deficiency of ECS can be induced by chronic stress and likely is a contributor to the negative consequences that follow.

##### 4.1. Acute stress and ECS

There is considerable evidence that acute exposure of rodents and humans to stress results in changes in ECS. In rodent models, studies have focused primarily on changes in CNS ECS, while in humans, eCB concentrations in the circulation have been used as an index of ECS [78]. As described in section 2.4, acute stress exposure increases 2-AG and 2-AG/CB1R signaling in the hypothalamus, hippocampus, PFC and raphe.

On the other hand, acute stress exposure in rodents decreases amygdalar and PFC concentrations of AEA [68, 79–81]. The acute effect of stress to reduce AEA concentrations was accompanied by an increase in the activity of FAAH. The effect on AEA is not likely mediated by glucocorticoids, because exogenous glucocorticoids produce a rapid increase in tissue contents of AEA in the amygdala, hippocampus and hypothalamus [57] and in hypothalamic slices [58]. These changes are transient, returning to baseline even when the concentration of circulating CORT is still elevated. Interestingly, the increase in AEA was not accompanied by increases in two other NAEs, suggesting that reduction in activity of

FAAH is not responsible for the change. It is possible that AEA synthesis has increased, or that an AEA-selective catabolic process has been inhibited. Interesting recent data from Hill and colleagues indicate that CRH, through actions at CRH-R1 receptors, increases FAAH activity and decreases AEA in the amygdala [82]. These data, together with the rapid time course of change, indicate that a reduction in AEA is a very early event in the stress response and is not a glucocorticoid-mediated effect, at least in the amygdala.

The glucocorticoids also produce delayed effects that occur 1–5 hours after stress, particularly in the hippocampus, basolateral amygdala, medial PFC (mPFC) and ventral tegmental area of the midbrain [4]. The delayed effects of the glucocorticoids are hormone-like and are mediated by binding to GR and mineralocorticoid (MR) receptors [83]. GRs are ubiquitously distributed in brain, with very high densities in the PVN of the hypothalamus, hippocampal CA1 and dentate gyrus, lateral septum and the nucleus tractus solitarius (NTS) [83]. The brain distribution of MR is more restricted than GR and overlaps GR distribution in the hippocampal regions and the lateral septum [83]. The affinity of CORT for MR is ten times higher than for GR such that MR are occupied at basal concentrations of CORT [4]. An early study demonstrated that adrenalectomy resulted in increased CB1R mRNA expression in the caudate putamen of male rats [84], suggesting that glucocorticoids exert a negative effect on the transcription of CB1R. In support of this notion, prolonged exogenous CORT treatment reduces CB1R binding site density in the hippocampus [85, 86] and amygdala [86]. However, the changes in binding site density were not accompanied by changes in mRNA [86], so it is possible that genomic GR effects were not involved in the effect. Rats injected with CORT daily for 10 days also exhibit decreased CB1R protein expression and function in primary sensory neurons through a GR mechanism [87]. Similarly, intense activation of the HPA axis significantly reduces CB1R mRNA expression in the frontal cortex measured 7 days later [88]. Although the specific role of GR in these effects was not determined, several of these studies are consistent with glucocorticoids reducing CB1R expression through transcriptional regulation.

## 4.2 Chronic Stress Alters CB1R Expression

Chronic unpredictable (also called chronic mild or chronic variable) stress (CUS) paradigms expose rodents to a variety of stressors presented in a random manner [89]. CUS results in increased basal CORT secretion, hyperactive HPA axis responses to stress, anhedonia and anxiety-like behaviors [89]. In both intact and gonadectomized male rats, CUS exposure results in a 50% decrease in CB1R binding site density and protein in the hippocampus [73, 90]. Interestingly, CUS increases CB1R protein expression in both intact and gonadectomized female rats [90].

Chronic exposure to homotypic (i.e. the stressor stays the same) stress also results in increased anxiety-like behaviors, but is less likely to elevate basal CORT concentrations and to induce body weight loss than CUS [91]. The difference is that animals can habituate to some of the effects of repeated homotypic stress exposure while the unpredictable nature of the CUS model prevents this. Repeated exposure to restraint (21 days of 6 hrs restraint per day) reduces CB1R binding site density in the dentate gyrus of the hippocampus, although the changes are less robust than occur following CUS [92]. Chronic restraint stress impairs

eCB-mediated suppression of GABA signaling in the CA1 region of the hippocampus [93], which is in accord with a loss of CB1R binding sites and protein.

Therefore, chronic stress exposure reduces CB1R expression and ECS in the hippocampus. Decreased ECS mediated regulation of GABA release in the hippocampus could have important implications for spatial learning and flexibility, which are altered by chronic stress exposure [5].

In contrast to the effects seen in the hippocampus, CUS increases CB1R binding site density in the PFC of male rats [94]. Similarly, male mice exposed for 4 days to 2 hours per day of immobilization and acoustic stress exhibit a significant increase in CB1R protein expression in PFC [95]. Chronic treatment of rats with the tricyclic antidepressant, imipramine, reverses the up-regulation of CB1R binding in the mPFC produced by CUS [94]. These findings suggest that the increase in CB1R binding site density in mPFC is secondary to an effect of stress to alter monoamine signaling.

CUS has been shown to reduce CB1R binding site density in the ventral striatum in rats [94] and to produce functional down-regulation of eCB signaling in the nucleus accumbens in mice [96]. These changes in ECS likely contribute to the anhedonic effects of CUS [96].

Other paradigms that exert a chronic stress have also been shown to alter CB1R protein or mRNA expression. For example, social isolation results in a significant increase in CB1R mRNA throughout the cortical regions of adult male rats compared to those raised in groups [97]. However, autoradiographic analyses did not detect changes in CB1R binding in cortical regions in socially isolated male rats [98]. On the other hand, CB1R binding was decreased in the supraoptic hypothalamic nuclei and ventrolateral thalamus and increased in subregions of the caudate putamen in singly-housed compared to group-housed rats [98]. Repeated episodes of alcohol withdrawal, which produces anxiety in rodents [99], resulted in a significant reduction of CB1R mRNA in the amygdala of male rats [100].

#### 4.3 Chronic Stress Regulates FAAH and MAGL Expression and Activities

FAAH is the primary catabolic enzyme for AEA in the brain and its inhibition increases AEA concentrations throughout the brain [101]. Among other things, increased AEA concentrations are associated with decreased anxiety [69] and dampened HPA axis activation [67, 68] as a result of CB1R activation. Five putative GR binding elements have been identified in the promoter region of the mouse FAAH gene [102], and *in vitro* studies demonstrate that non-liganded, GR represses FAAH expression [102]. Interestingly, the addition of a GR agonist did not increase the repression *in vitro*, suggesting that the GR does not need to be bound by ligand to repress FAAH although ligand binding is necessary for GR translocation to the nucleus *in vivo*. Isolation stress in male rats produces very significant reductions (greater than 50% in most regions) in FAAH mRNA expression in cortical regions and throughout the dorsal and ventral striatum [97]. Interestingly, AEA concentrations measured in adult male rats raised in isolation were significantly elevated in the pyriform cortex compared to group housed rats, but were unchanged in PFC, nucleus accumbens and hippocampus [98]. Thus, the comparison of mRNA and AEA content data indicates that the large reduction in FAAH mRNA does not always result in increased AEA

content, and suggests that isolation stress could reduce AEA synthesis as well as decrease its catabolism in a brain-region specific manner.

FAAH protein is reduced by 40% in dorsal root ganglion (DRG) cells from rats chronically treated with CORT [87], which supports the hypothesis that GR activation decreases FAAH expression.

CORT also appears to have effects on FAAH activity that are independent of changes in FAAH expression. For example, chronic administration of CORT to male rats does not alter mRNA expression but produces an increase in the  $V_{max}$  for FAAH in the amygdala and hippocampus [86]. AEA concentrations in these brain regions are significantly reduced by chronic CORT administration, consistent with enhanced FAAH-mediated hydrolysis. Similarly, repeated restraint in male mice increases the  $V_{max}$  for FAAH and decreases AEA concentrations in amygdala and mPFC [81]. This is a brain region selective effect of stress since repeated restraint decreases FAAH  $V_{max}$  and increases AEA content in ventral striatum [81]. A reciprocal relationship between AEA content and FAAH activity is seen consistently in rodents exposed to chronic stress, suggesting that chronic stress increases catabolism of AEA by FAAH, which could contribute to hypoactive CB1R signaling. The mechanism by which CORT and stress alter the  $V_{max}$  for FAAH is not known, but the lack of clear demonstration that mRNA is reduced suggests post translational modification of the enzyme has occurred.

The predominant effect of social isolation on MAGL mRNA content is to increase expression throughout the cortex and in selected regions of the striatum [97]. In spite of this change, 2-AG contents in the PFC and pyriform cortex of male rats reared in isolated are significantly increased [98]. Among the possible explanations for the discordance of these results is that an increase in MAGL activity is accompanied by an increase in 2-AG synthesis such that no net change occurs. In support of this concept, 10 days of one-hour restraint stress in male mice increases mPFC 2-AG content and increases the  $V_{max}$  for MAGL activity; on the other hand, amygdalar 2-AG is also increased on day 10, but MAGL activity is unchanged from control [81]. These results suggest that repeated restraint exposure could increase the synthesis of 2-AG in many brain regions while MAGL activity is altered in a brain-region dependent manner, perhaps as a compensatory process.

Repeated exposure of male mice to restraint also increases 2-AG content in the basolateral amygdala and enhances 2-AG mediated synaptic plasticity at inhibitory synapses in that region [103]. Repeated restraint produced a decrease in MAGL protein associated with membranes, where it would presumably have more influence on 2-AG concentrations at the CB1R [103]. Furthermore, chronic inhibition of MAGL mimicked and occluded the effects of repeated restraint, suggesting that reduced MAGL activity is a major mechanism for the increase in 2-AG signaling in this model.

## 5. ECS and Habituation to Stress

Stress-induced HPA axis activation and behavioral response repertoires to repeated and predictable exposure to the same stress (homotypic stress) can habituate. Stress habituation is stress-specific, dependent upon the time between stress exposures, and the intensity of the

initial stress [104]. The ability of an individual to habituate to the effects of stress is one of the factors that confers resilience to the negative consequences of stress [9].

Emerging evidence demonstrates that enhanced ECS contributes to the mechanism of habituation to stress. While chronic exposure to unpredictable and variable stressors reduces CB1R-mediated signaling (section 4), repeated exposure to a short period of restraint sensitizes 2-AG mobilization. In particular, a single exposure to restraint does not alter contents of 2-AG in the limbic forebrain, mPFC, amygdala, or cerebellum of male mice [79]. However, when mice are exposed to increasing numbers of restraint episodes, carried out at the same time of day and for the same duration, 2-AG contents are progressively increased in these brain regions immediately after the stress offset [79, 81]. Similar changes are seen in the amygdala of the male rat [105]. Since chronic treatment with CORT also increases 2-AG content in the amygdala [106], it is possible that CORT mediates this effect. Recent data suggest that the mechanism for this effect is inhibition of MAGL function, perhaps as a result of changes in the subcellular distribution of MAGL such that less of the enzyme is present at the plasma membrane [103]. In contrast to the increase in 2-AG seen with repeated stress in these cortical regions, 2-AG contents are reduced in ventral striatum with repeated restraint [81], suggesting a different mechanism and purpose for this change in the reward system.

There is evidence that enhanced ECS contributes to the habituation of behavioral responses and HPA axis activation to the stressor. In male mice, systemic injection of CB1R antagonist prevents habituation to behavioral activation [79] and anhedonia [107] in response to stress exposure. Inhibition of ECS in the amygdala prevents habituation to HPA axis activation by stress in rats [105] while increased 2-AG concentrations in the amygdala prevent behavioral and synaptic adaptations to repeated restraint exposure in mice [103]. CB1R activation is also required for the habituation of innate fear behaviors in mice to repeated homotypic stress [108–110]. Recent data suggest that the effect of chronic stress exposure to enhance ECS content could also contribute to negative consequences of stress habituation. In particular, chronic treatment with CRH produces an increase in anxiety that is antagonized by CB1R antagonist treatment [111].

Taken together, these data suggest that the plasticity that is afforded by the ECS provides an important mechanism by which the brain can habituate to repeated, reliable stress exposures. As the ability to habituate to a non-threatening stimulus will allow an individual to conserve resources and avoid the consequences of chronic stress, this role of ECS could be one of the most important in the context of human psychiatric pathology.

## 6. ECS and Sympathetic Nervous System Responses to Stress

Exposure to an acute stress or anticipation of danger evoke characteristic physiological changes through activation of the neuronal defense pathway. Stress information is provided to the NTS from the amygdala, infralimbic cortex and the PVN [112]. Excitatory projections from the NTS to the locus coeruleus and ventromedial medulla activate preganglionic sympathetic neurons while inhibitory projections to the dorsal motor nucleus of the vagus and nucleus ambiguus inhibit the parasympathetic nervous system. There are several sites

in the CNS where ECS has been found to regulate activation of the SNS; interestingly, enhanced ECS can both increase and decrease SNS responses to stress.

Mobilization of ECS in the rat dorsal periaqueductal gray (PAG) enhances SNS activation [113, 114]. CB1R are found in the PAG [115] and microinjection of AEA into the dorsal PAG increases renal sympathetic nerve activity within 30 sec of injection [113]. These effects are inhibited by CB1R antagonist pretreatment and are consistent with CB1R-mediated inhibition of GABA release [114]. Microinjection of CB1R antagonist into the dorsal PAG also inhibits SNS responses evoked by hypothalamic stimulation [113]. As acute stress results in a rapid increase in the PAG tissue contents of both AEA and 2-AG [116], these results suggest that ECS in the dorsal PAG enhances or even enables stress-induced SNS activation.

On the other hand, the direct effects of CRH to increase SNS activation are inhibited by i.c.v. administration of CB1R agonists and increased by CB1R antagonists [117], suggesting a stress-inhibitory role for ECS. In support of this hypothesis, injection of AEA directly into the NTS of rats prolongs baroreceptor-induced sympathoinhibition [118]. These apparently contradictory effects of CB1R activity on SNS outflow illustrate the point that ECS is a local process, so it is not surprising that it exerts opposite effects on a circuit.

CB1R are expressed on terminals of sympathetic axons innervating blood vessels and there is evidence their activation inhibits the release of norepinephrine [20, 119]. CB1R mRNA has been detected in superior cervical ganglion [20], which is consistent with CB1R protein expression by post-ganglionic, sympathetic neurons. Mice lacking CB1R selectively in sympathetic neurons are lean and resistant to diet-induced obesity, which the authors hypothesize is due to the loss of CB1R-mediated suppression of norepinephrine release, which results in increased lipid oxidation and thermogenesis as a result of increased sympathetic tone [120]. CB1R on sympathetic terminals in bone inhibit norepinephrine release and oppose the effects of the SNS to reduce bone formation [121, 122]. CB1R activation suppresses norepinephrine release evoked by perivascular nerve stimulation in the isolated heart [123]. The source of eCB that provides innervation of these receptors is not known, although the retrograde signaling paradigm of ECS in brain suggests that the tissues receiving the neuronal input are a possible source. The inflammatory molecule, lipopolysaccharide (LPS), is able to mobilize functional ECS at sympathetic terminals, suggesting recruitment of the CB1R during inflammation. It is possible that ECS at the sympathetic terminal functions as a local feedback modulator to protect tissues from excessive SNS activation.

An important component of the sympathetic response to stress is a coordinated effect on the cardiovascular system. Activation of  $\beta$ -adrenergic receptors of the heart and blood vessels by norepinephrine released from sympathetic terminals results in alterations in heart rate, contractile force and blood flow to muscles and skin that are necessary to support the fight or flight response. It is well accepted that chronic stress increases the risk of cardiovascular diseases, particularly those of the heart [124]. CB1R are expressed by non-neuronal cells of the cardiovascular system, including cardiomyocytes, vascular smooth muscle cells and endothelium [24, 125, 126]. While the majority of available evidence indicates that

endogenous CB1R signaling does not contribute to the regulation of cardiovascular function under normal conditions, it is likely that CB1R signaling in this system is recruited under various pathophysiological states, including inflammation [127] and profound hypotension [128]. O'Sullivan and colleagues recently reviewed the roles for the broadly considered endocannabinoid system in the effects of stress on the cardiovascular system and concluded that there are multiple possible sites of interaction at all levels, including the brain, sympathetic nerves, HPA axis and end organs [124].

## 7. CB1R and Immune Responses to Stress

Among the many effects of stress is a profound ability to suppress the immune system [129]. This is the result of both arms of the stress response. Glucocorticoids act through GR receptors to alter the expression of a variety of cytokines and inflammatory mediators, particularly TNF $\alpha$  [130]. Postganglionic fibers of the SNS innervate the spleen and release norepinephrine in response to activation, which reduces inflammatory cytokine production through  $\beta$ -adrenoreceptor activation [131]. In light of the data discussed in section 6 that ECS inhibits release of norepinephrine from sympathetic terminals, a logical hypothesis is that ECS, through CB1R activation, promotes stress-induced inflammation and that CB1R antagonists will be anti-inflammatory. There is evidence to support this hypothesis. In a study of the anti-inflammatory mechanism of CB1R antagonists, Mnich and colleagues demonstrated that the site of action of this effect was at the sympathetic terminals in the spleen [132]. Their data strongly suggest that, at least in the inflamed state, eCBs mobilized in the spleen function to inhibit norepinephrine release and thus, decrease the anti-inflammatory influence of SNS activation.

Periodontitis includes inflammation of the gums and other tissues supporting the teeth [133]. Rettori and colleagues reported that gingival injection of AEA in rats with periodontitis also exposed to restraint stress reduced the elevation of circulating CORT, and reduced gingival tissue necrosis factor alpha (TNF $\alpha$ ) and IL-1 $\beta$  immunoreactivities [134]. These effects were reversed by combined CB antagonist treatment [134]. Thus, general activation of ECS in the inflamed area of the tooth exerts an immune-suppressive effect that extends to a reduction in basal HPA axis activity.

## 8. ECS, Stress and the Gastrointestinal Tract

All components of ECS are found in the gastrointestinal (GI) tract. CB1R have been shown to be present in cholinergic neurons in both the myenteric and submucosal plexi of the ENS [135, 136].

Stress affects multiple functions of the GI tract, including gastric secretion, motility, epithelial permeability and barrier function, and mucosal blood flow [137]. Acute and chronic stress are associated with lower pain thresholds and visceral hypersensitivity to painful stimuli [138]. Male rats exposed to water avoidance stress exhibit GR-mediated decreases in CB1R expression in DRG neurons, a change that is hypothesized to contribute to stress-induced hypersensitivity to colorectal distension [87]. On the other hand, four days of exposure of male rats to partial restraint increases CB1R expression in the colon [139].

This stress protocol also produces visceral hypersensitivity, and the authors suggest that increased colonic CB1R expression is an attempt to normalize pain sensitivity [139].

CB1R<sup>-/-</sup> mice exposed to 4 days of 2-hour immobilization and acoustic stress exposure exhibit increased permeability of the colonic barrier; enhanced inflammation; lower IgA secretion and higher bacterial translocation into the mesenteric lymph nodes than wild type mice [140]. IgA secretion by the GI tract is the first line defense against pathogens, through its ability to neutralize viruses, bind toxins and food antigens, and to reduce bacterial binding to epithelial cells [141]. Thus, the loss of IgA secretion, together with loss of the intestinal barrier function and reduced visceral hypersensitivity to pain, suggest an important homeostatic role of CB1R in the intestine.

## 9. Conclusions

Overwhelming data support the hypothesis that the ECS is a critical component of homeostatic regulation of the body. Endocannabinoid/CB1R signaling is primarily stress-inhibitory, reducing both the endocrine and neuronal responses to stress. CB1R signaling participates in habituation to stress exposure, which is a protective mechanism designed to dampen responses to a non-threatening stimulus. On the other hand, chronic stress exposure decreases endocannabinoid/CB1R signaling. Given the vital role of CB1R activation in maintaining hedonia and reducing anxiety, the reduction of CB1R signaling is hypothesized to contribute to the negative consequences of stress.

There is some support for the "Endocannabinoid Deficiency" hypothesis in humans. Recent data that proteins of the ECS exhibit polymorphisms in humans suggests that differences in the tone of ECS could contribute to vulnerability or resilience to psychopathology [142]. A very interesting recent study found that CB1R genotype exerts a significant effect on the likelihood that early childhood neglect will result in anhedonia in adulthood [143] and animal studies strongly suggest that early life stress alters ECS [144]. Thus, alterations of ECS could be an important link between early life stress and psychopathology in later life. Finally, emerging data indicate that significant sex differences in the role of ECS in the regulation of stress responsivity [145]. It is also generally acknowledged that most psychiatric disorders exhibit clear sex differences, with substance abuse disorders, antisocial personality and attention deficit disorder being more common in men; while depression, anxiety and eating disorders are more common in women [146].

## Acknowledgments

The author was supported during the writing of this review by NIH grant DA026996 and by the Research and Education Component of the Advancing a Healthier Wisconsin Endowment of the Medical College of Wisconsin.

## Abbreviations

<b>2-AG</b>	2-arachidonoylglycerol
<b>AEA</b>	<i>N</i> -arachidonylethanolamine
<b>CB1R</b>	type 1 cannabinoid receptor

<b>CORT</b>	corticosterone
<b>COX2</b>	cyclooxygenase type 2
<b>CRH</b>	corticotropin releasing hormone
<b>CUS</b>	chronic, unpredictable stress
<b>DAG</b>	diacylglycerol
<b>DAGL</b>	diacylglycerol lipase
<b>DRG</b>	dorsal root ganglion
<b>eCBs</b>	endocannabinoids
<b>ECS</b>	endocannabinoid signaling
<b>FAAH</b>	fatty acid amide hydrolase
<b>GR</b>	glucocorticoid receptor
<b>HPA</b>	hypothalamic-pituitary-adrenal
<b>MAGL</b>	monoacylglycerol lipase
<b>mPFC</b>	medial prefrontal cortex
<b>MR</b>	mineralocorticoid receptor
<b>NAE</b>	<i>N</i> -acylethanolamine
<b>NAPE</b>	<i>N</i> -acyl-phosphatidylethanolamine
<b>NTS</b>	nucleus tractus solitarius
<b>PAG</b>	periaqueductal gray
<b>PFC</b>	prefrontal cortex
<b>PLC</b>	phospholipase C
<b>PVN</b>	paraventricular nucleus
<b>SNS</b>	sympathetic nervous system
<b>THC</b>	<sup>9</sup> -tetrahydrocannabinol

## References

1. Dantzer R, O'Connor JC, Freund GG, Johnson RW, Kelley KW. From inflammation to sickness and depression: when the immune system subjugates the brain. *Nat Rev Neurosci.* 2008; 9:46–56. [PubMed: 18073775]
2. Yamakawa K, Matsunaga M, Isowa T, Kimura K, Kasugai K, Yoneda M, et al. Transient responses of inflammatory cytokines in acute stress. *Biol Psychol.* 2009; 82:25–32. [PubMed: 19446599]
3. McEwen BS. Steroid hormone actions on the brain: when is the genome involved? *Horm Behav.* 1994; 28:396–405. [PubMed: 7729808]
4. Joels M, Sarabdjitsingh RA, Karst H. Unraveling the time domains of corticosteroid hormone influences on brain activity: rapid, slow, and chronic modes. *Pharmacol Rev.* 2012; 64:901–938. [PubMed: 23023031]

5. Lupien SJ, McEwen BS, Gunnar MR, Heim C. Effects of stress throughout the lifespan on the brain, behaviour and cognition. *Nat Rev Neurosci*. 2009; 10:434–445. [PubMed: 19401723]
6. van Dijk G, Buwalda B. Neurobiology of the metabolic syndrome: an allostatic perspective. *European journal of pharmacology*. 2008; 585:137–146. [PubMed: 18395710]
7. Maletic V, Raison CL. Neurobiology of depression, fibromyalgia and neuropathic pain. *Frontiers in bioscience : a journal and virtual library*. 2009; 14:5291–5338.
8. Goldstein DS. Stress, allostatic load, catecholamines, and other neurotransmitters in neurodegenerative diseases. *Endocrine regulations*. 2011; 45:91–98. [PubMed: 21615193]
9. Charney DS. Psychobiological mechanisms of resilience and vulnerability: implications for successful adaptation to extreme stress. *Am J Psychiatry*. 2004; 161:195–216. [PubMed: 14754765]
10. Thoits PA. Stress and health: major findings and policy implications. *J Health Soc Behav*. 2010; 51(Suppl):S41–S53. [PubMed: 20943582]
11. Young E, Korszun A. Sex, trauma, stress hormones and depression. *Molecular psychiatry*. 2010; 15:23–28. [PubMed: 19773810]
12. Lai MC, Huang LT. Effects of early life stress on neuroendocrine and neurobehavior: mechanisms and implications. *Pediatr Neonatol*. 2011; 52:122–129. [PubMed: 21703552]
13. Hollister LE, Moore F, Kanter S, Noble E. 1-tetrahydrocannabinol, synhexyl and marijuana extract administered orally in man: catecholamine excretion, plasma cortisol levels and platelet serotonin content. *Psychopharmacologia*. 1970; 17:354–360. [PubMed: 5523370]
14. Howlett AC. Cannabinoid inhibition of adenylate cyclase. Biochemistry of the response in neuroblastoma cell membranes. *Mol Pharmacol*. 1985; 27:429–436. [PubMed: 2984538]
15. Matsuda LA, Lolait SJ, Brownstein MJ, Young AC, Bonner TI. Structure of a cannabinoid receptor and functional expression of the cloned cDNA. *Nature*. 1990; 346:561–564. [PubMed: 2165569]
16. Herkenham M, Lynn AB, Little MD, Johnson MR, Melvin LS, de Costa BR, et al. Cannabinoid receptor localization in brain. *Proc Natl Acad Sci*. 1990; 87:1932–1936. [PubMed: 2308954]
17. Mackie K. Distribution of cannabinoid receptors in the central and peripheral nervous system. *Handb Exp Pharmacol*. 2005:299–325. [PubMed: 16596779]
18. Huestis MA, Gorelick DA, Heishman SJ, Preston KL, Nelson RA, Moolchan ET, et al. Blockade of effects of smoked marijuana by the CB1-selective cannabinoid receptor antagonist SR141716. *Arch Gen Psychiatry*. 2001; 58:322–328. [PubMed: 11296091]
19. Ahluwalia J, Urban L, Capogna M, Bevan S, Nagy I. Cannabinoid 1 receptors are expressed in nociceptive primary sensory neurons. *Neuroscience*. 2000; 100:685–688. [PubMed: 11036202]
20. Ishac EJ, Jiang L, Lake KD, Varga K, Abood ME, Kunos G. Inhibition of exocytotic noradrenaline release by presynaptic cannabinoid CB1 receptors on peripheral sympathetic nerves. *Br J Pharmacol*. 1996; 118:2023–2028. [PubMed: 8864538]
21. Izzo AA, Sharkey KA. Cannabinoids and the gut: new developments and emerging concepts. *Pharmacol Ther*. 2010; 126:21–38. [PubMed: 20117132]
22. Salio C, Doly S, Fischer J, Franzoni MF, Conrath M. Neuronal and astrocytic localization of the cannabinoid receptor-1 in the dorsal horn of the rat spinal cord. *Neurosci Lett*. 2002; 329:13–16. [PubMed: 12161251]
23. Moldrich G, Wenger T. Localization of the CB(1) cannabinoid receptor in the rat brain. An immunohistochemical study\*. *Peptides*. 2000; 21:1735–1742. [PubMed: 11090929]
24. Gebremedhin D, Lange AR, Campbell WB, Hillard CJ, Harder DR. Cannabinoid CB1 receptor of cat cerebral arterial muscle functions to inhibit L-type Ca<sup>2+</sup> channel current. *Am J Physiol*. 1999; 276:H2085–H2093. [PubMed: 10362691]
25. Golech SA, McCarron RM, Chen Y, Bembry J, Lenz F, Mechoulam R, et al. Human brain endothelium: coexpression and function of vanilloid and endocannabinoid receptors. *Brain Res Mol Brain Res*. 2004; 132:87–92. [PubMed: 15548432]
26. Bouaboula M, Rinaldi M, Carayon P, Carillon C, Delpech B, Shire D, et al. Cannabinoid-receptor expression in human leukocytes. *Eur J Biochem*. 1993; 214:173–180. [PubMed: 8508790]
27. Bensaid M, Gary-Bobo M, Esclangon A, Maffrand JP, Le Fur G, Oury-Donat F, et al. The cannabinoid CB1 receptor antagonist SR141716 increases Acrp30 mRNA expression in adipose

- tissue of obese fa/fa rats and in cultured adipocyte cells. *Mol Pharmacol.* 2003; 63:908–914. [PubMed: 12644592]
28. Jeong WI, Osei-Hyiaman D, Park O, Liu J, Batkai S, Mukhopadhyay P, et al. Paracrine activation of hepatic CB1 receptors by stellate cell-derived endocannabinoids mediates alcoholic fatty liver. *Cell Metab.* 2008; 7:227–235. [PubMed: 18316028]
  29. Ziegler CG, Mohn C, Lamounier-Zepter V, Rettori V, Bornstein SR, Krug AW, et al. Expression and Function of Endocannabinoid Receptors in the Human Adrenal Cortex. *Horm Metab Res.* 2010; 42:88–92. [PubMed: 19862666]
  30. Devane WA, Hanus L, Breuer A, Pertwee RG, Stevenson LA, Griffin G, et al. Isolation and structure of a brain constituent that binds to the cannabinoid receptor. *Science.* 1992; 258:1946–1949. [PubMed: 1470919]
  31. Kearns CS, Greenberg MJ, DiCamelli R, Kurzawa K, Hillard CJ. Relationships between ligand affinities for the cerebellar cannabinoid receptor CB1 and the induction of GDP/GTP exchange. *J Neurochem.* 1999; 72:2379–2387. [PubMed: 10349847]
  32. Mechoulam R, Ben-Shabat S, Hanus L, Ligumsky M, Kaminski NE, Schatz AR, et al. Identification of an endogenous 2-monoglyceride, present in canine gut, that binds to cannabinoid receptors. *Biochem Pharmacol.* 1995; 50:83–90. [PubMed: 7605349]
  33. Sugiura T, Kondo S, Sukagawa A, Nakane S, Shinoda A, Itoh K, et al. 2-Arachidonoylglycerol: a possible endogenous cannabinoid receptor ligand in brain. *Biochem Biophys Res Commun.* 1995; 215:89–97. [PubMed: 7575630]
  34. Sugiura T, Waku K. Cannabinoid receptors and their endogenous ligands. *J Biochem (Tokyo).* 2002; 132:7–12. [PubMed: 12097154]
  35. Vaughn LK, Denning G, Stuhr KL, de Wit H, Hill MN, Hillard CJ. Endocannabinoid signalling: has it got rhythm? *Br J Pharmacol.* 2010; 160:530–543. [PubMed: 20590563]
  36. Schmid HHO, Schmid PC, Natarajan. V. N-Acylated glycerophospholipids and their derivatives. *Prog Lipid Res.* 1990; 29:1–43. [PubMed: 2087478]
  37. Simon GM, Cravatt BF. Characterization of mice lacking candidate N-acyl ethanolamine biosynthetic enzymes provides evidence for multiple pathways that contribute to endocannabinoid production in vivo. *Mol Biosyst.* 2010; 6:1411–1418. [PubMed: 20393650]
  38. Bisogno T, Howell F, Williams G, Minassi A, Cascio MG, Ligresti A, et al. Cloning of the first sn1-DAG lipases points to the spatial and temporal regulation of endocannabinoid signaling in the brain. *J Cell Biol.* 2003; 163:463–468. [PubMed: 14610053]
  39. Lafourcade M, Elezgarai I, Mato S, Bakiri Y, Grandes P, Manzoni OJ. Molecular components and functions of the endocannabinoid system in mouse prefrontal cortex. *PLoS ONE.* 2007; 2:e709. [PubMed: 17684555]
  40. Tsou K, Noguero MI, Muthian S, Sanudo-Pena MC, Hillard CJ, Deutsch DG, et al. Fatty acid amide hydrolase is located preferentially in large neurons in the rat central nervous system as revealed by immunohistochemistry. *Neurosci Lett.* 1998; 254:137–140. [PubMed: 10214976]
  41. Patel S, Carrier EJ, Ho WS, Rademacher DJ, Cunningham S, Reddy DS, et al. The postmortal accumulation of brain N-arachidonylethanolamine (anandamide) is dependent upon fatty acid amide hydrolase activity. *J Lipid Res.* 2005; 46:342–349. [PubMed: 15576840]
  42. Bortolato M, Mangieri RA, Fu J, Kim JH, Arguello O, Duranti A, et al. Antidepressant-like activity of the fatty acid amide hydrolase inhibitor URB597 in a rat model of chronic mild stress. *Biol Psychiatry.* 2007; 62:1103–1110. [PubMed: 17511970]
  43. Ueda N, Tsuboi K, Uyama T. N-acylethanolamine metabolism with special reference to N-acylethanolamine-hydrolyzing acid amidase (NAAA). *Prog Lipid Res.* 2010; 49:299–315. [PubMed: 20152858]
  44. Blankman JL, Simon GM, Cravatt BF. A comprehensive profile of brain enzymes that hydrolyze the endocannabinoid 2-arachidonoylglycerol. *Chem Biol.* 2007; 14:1347–1356. [PubMed: 18096503]
  45. Marris WR, Blankman JL, Horne EA, Thomazeau A, Lin YH, Coy J, et al. The serine hydrolase ABHD6 controls the accumulation and efficacy of 2-AG at cannabinoid receptors. *Nat Neurosci.* 2010; 13:951–957. [PubMed: 20657592]

46. Kozak KR, Prusakiewicz JJ, Marnett LJ. Oxidative Metabolism of Endocannabinoids by COX-2. *Curr Pharm Des.* 2004; 10:659–667. [PubMed: 14965328]
47. Freund TF, Katona I, Piomelli D. Role of endogenous cannabinoids in synaptic signaling. *Physiol Rev.* 2003; 83:1017–1066. [PubMed: 12843414]
48. Maejima T, Ohno-Shosaku T, Kano M. Endogenous cannabinoid as a retrograde messenger from depolarized postsynaptic neurons to presynaptic terminals. *Neurosci Res.* 2001; 40:205–210. [PubMed: 11448511]
49. Katona I, Urban GM, Wallace M, Ledent C, Jung KM, Piomelli D, et al. Molecular composition of the endocannabinoid system at glutamatergic synapses. *J Neurosci.* 2006; 26:5628–5637. [PubMed: 16723519]
50. Katona I, Sperlagh B, Sik A, Kafalvi A, Vizi ES, Mackie K, et al. Presynaptically located CB1 cannabinoid receptors regulate GABA release from axon terminals of specific hippocampal interneurons. *J Neurosci.* 1999; 19:4544–4558. [PubMed: 10341254]
51. Hermann H, Marsicano G, Lutz B. Coexpression of the cannabinoid receptor type 1 with dopamine and serotonin receptors in distinct neuronal subpopulations of the adult mouse forebrain. *Neuroscience.* 2002; 109:451–460. [PubMed: 11823058]
52. Oropenza VC, Mackie K, Van Bockstaele EJ. Cannabinoid receptors are localized to noradrenergic axon terminals in the rat frontal cortex. *Brain Res.* 2007; 1127:36–44. [PubMed: 17113043]
53. Karst H, Berger S, Turiault M, Tronche F, Schutz G, Joels M. Mineralocorticoid receptors are indispensable for nongenomic modulation of hippocampal glutamate transmission by corticosterone. *Proceedings of the National Academy of Sciences of the United States of America.* 2005; 102:19204–19207. [PubMed: 16361444]
54. Karst H, Berger S, Erdmann G, Schutz G, Joels M. Metaplasticity of amygdalar responses to the stress hormone corticosterone. *Proceedings of the National Academy of Sciences of the United States of America.* 2010; 107:14449–14454. [PubMed: 20663957]
55. Verkuyl JM, Karst H, Joels M. GABAergic transmission in the rat paraventricular nucleus of the hypothalamus is suppressed by corticosterone and stress. *Eur J Neurosci.* 2005; 21:113–121. [PubMed: 15654848]
56. Evanson NK, Tasker JG, Hill MN, Hillard CJ, Herman JP. Fast feedback inhibition of the HPA axis by glucocorticoids is mediated by endocannabinoid signaling. *Endocrinology.* 2010; 151:4811–4819. [PubMed: 20702575]
57. Hill MN, Karatsoreos IN, Hillard CJ, McEwen BS. Rapid elevations in limbic endocannabinoid content by glucocorticoid hormones in vivo. *Psychoneuroendocrinology.* 2010; 35:1333–1338. [PubMed: 20399021]
58. Di S, Malcher-Lopes R, Marcheselli VL, Bazan NG, Tasker JG. Rapid glucocorticoid-mediated endocannabinoid release and opposing regulation of glutamate and GABA inputs to hypothalamic magnocellular neurons. *Endocrinology.* 2005; 146:4292–4301. [PubMed: 15994343]
59. Di S, Malcher-Lopes R, Halmos KC, Tasker JG. Nongenomic glucocorticoid inhibition via endocannabinoid release in the hypothalamus: a fast feedback mechanism. *J Neurosci.* 2003; 23:4850–4857. [PubMed: 12832507]
60. Wang J, Shen RY, Haj-Dahmane S. Endocannabinoids mediate the glucocorticoid-induced inhibition of excitatory synaptic transmission to dorsal raphe serotonin neurons. *J Physiol.* 2012; 590:5795–5808. [PubMed: 22946098]
61. Hill MN, McLaughlin RJ, Pan B, Fitzgerald ML, Roberts CJ, Lee TT, et al. Recruitment of prefrontal cortical endocannabinoid signaling by glucocorticoids contributes to termination of the stress response. *J Neurosci.* 2011; 31:10506–10515. [PubMed: 21775596]
62. Wang M, Hill MN, Zhang L, Gorzalka BB, Hillard CJ, Alger BE. Acute restraint stress enhances hippocampal endocannabinoid function via glucocorticoid receptor activation. *J Psychopharmacol.* 2012; 26:56–70. [PubMed: 21890595]
63. Hill MN, McEwen BS. Endocannabinoids: The silent partner of glucocorticoids in the synapse. *Proc Natl Acad Sci U S A.* 2009; 106:4579–4580. [PubMed: 19293387]
64. Kim J, Alger BE. Inhibition of cyclooxygenase-2 potentiates retrograde endocannabinoid effects in hippocampus. *Nat Neurosci.* 2004; 7:697–698. [PubMed: 15184902]

65. Straiker A, Wager-Miller J, Hu SS, Blankman JL, Cravatt BF, Mackie K. COX-2 and FAAH can regulate the time course of depolarization induced suppression of excitation. *Br J Pharmacol*. 2011; 164:1672–1683. [PubMed: 21564090]
66. Ma Y, Matsuwaki T, Yamanouchi K, Nishihara M. Cyclooxygenase-2-related signaling in the hypothalamus plays differential roles in response to various acute stresses. *Brain Res*. 2013; 1508:23–33. [PubMed: 23458502]
67. Patel S, Roelke CT, Rademacher DJ, Cullinan WE, Hillard CJ. Endocannabinoid signaling negatively modulates stress-induced activation of the hypothalamic-pituitary-adrenal axis. *Endocrinology*. 2004; 145:5431–5438. [PubMed: 15331569]
68. Hill MN, McLaughlin RJ, Morrish AC, Viau V, Floresco SB, Hillard CJ, et al. Suppression of amygdalar endocannabinoid signaling by stress contributes to activation of the hypothalamic-pituitary-adrenal axis. *Neuropsychopharmacology*. 2009; 34:2733–2745. [PubMed: 19710634]
69. Patel S, Hillard CJ. Pharmacological evaluation of cannabinoid receptor ligands in a mouse model of anxiety: further evidence for an anxiolytic role for endogenous cannabinoid signaling. *J Pharmacol Exp Ther*. 2006; 318:304–311. [PubMed: 16569753]
70. Moreira FA, Grieb M, Lutz B. Central side-effects of therapies based on CB1 cannabinoid receptor agonists and antagonists: focus on anxiety and depression. *Best Pract Res Clin Endocrinol Metab*. 2009; 23:133–144. [PubMed: 19285266]
71. Bergamaschi MM, Queiroz RH, Chagas MH, Linares IM, Arrais KC, de Oliveira DC, et al. Rimonabant effects on anxiety induced by simulated public speaking in healthy humans: a preliminary report. *Hum Psychopharmacol*. 2014; 29:94–99. [PubMed: 24424711]
72. Marsicano G, Wotjak CT, Azad SC, Bisogno T, Rammes G, Cascio MG, et al. The endogenous cannabinoid system controls extinction of aversive memories. *Nature*. 2002; 418:530–534. [PubMed: 12152079]
73. Hill MN, Patel S, Carrier EJ, Rademacher DJ, Ormerod BK, Hillard CJ, et al. Downregulation of endocannabinoid signaling in the hippocampus following chronic unpredictable stress. *Neuropsychopharmacology*. 2005; 30:508–515. [PubMed: 15525997]
74. Rademacher DJ, Meier SE, Shi L, Ho W-SV, Jarrhian A, Hillard CJ. Effects of acute and repeated restraint stress on endocannabinoid content in the amygdala, ventral striatum and medial prefrontal cortex in mice. *Neuropharmacol*. 2008; 54:108–116.
75. Christensen R, Kristensen PK, Bartels EM, Bliddal H, Astrup A. Efficacy and safety of the weight-loss drug rimonabant: a meta-analysis of randomised trials. *Lancet*. 2007; 370:1706–1713. [PubMed: 18022033]
76. Hill MN, Miller GE, Ho WS, Gorzalka BB, Hillard CJ. Serum endocannabinoid content is altered in females with depressive disorders: a preliminary report. *Pharmacopsychiatry*. 2008; 41:48–53. [PubMed: 18311684]
77. Hillard CJ, Liu QS. Endocannabinoid Signaling in the Etiology and Treatment of Major Depressive Illness. *Curr Pharm Des*. 2013
78. Dlugos A, Childs E, Stuhr KL, Hillard CJ, de Wit H. Acute stress increases circulating anandamide and other N-acyl ethanolamines in healthy humans. *Neuropsychopharmacology*. 2012; 37:2416–2427. [PubMed: 22763622]
79. Patel S, Roelke CT, Rademacher DJ, Hillard CJ. Inhibition of restraint stress-induced neural and behavioural activation by endogenous cannabinoid signalling. *Eur J Neurosci*. 2005; 21:1057–1069. [PubMed: 15787710]
80. McLaughlin RJ, Hill MN, Bambico FR, Stuhr KL, Gobbi G, Hillard CJ, et al. Prefrontal cortical anandamide signaling coordinates coping responses to stress through a serotonergic pathway. *Eur Neuropsychopharmacol*. 2012; 22:664–671. [PubMed: 22325231]
81. Rademacher DJ, Meier SE, Shi L, Ho WS, Jarrhian A, Hillard CJ. Effects of acute and repeated restraint stress on endocannabinoid content in the amygdala, ventral striatum, and medial prefrontal cortex in mice. *Neuropharmacol*. 2008; 54:108–116.
82. Gray, M.; Veccharelli, H.; Kim, A.; Hassan, K.; Hermanson, D.; McLaughlin, R.J., et al. Corticotropin-releasing hormone signaling drives anandamide hydrolysis to promote anxiety. International Cannabinoid Research Society Annual Meeting; Vancouver, BC. 2013. p. 40

83. Reul JM, de Kloet ER. Two receptor systems for corticosterone in rat brain: microdistribution and differential occupation. *Endocrinology*. 1985; 117:2505–2511. [PubMed: 2998738]
84. Mailloux P, Vanderhaeghen JJ. Glucocorticoid regulation of cannabinoid receptor messenger RNA levels in the rat caudate-putamen. An in situ hybridization study. *Neurosci Lett*. 1993; 156:51–53. [PubMed: 8414189]
85. Hill MN, Carrier EJ, Ho WS, Shi L, Patel S, Gorzalka BB, et al. Prolonged glucocorticoid treatment decreases cannabinoid CB1 receptor density in the hippocampus. *Hippocampus*. 2008; 18:221–226. [PubMed: 18058925]
86. Bowles NP, Hill MN, Bhagat SM, Karatsoreos IN, Hillard CJ, McEwen BS. Chronic, noninvasive glucocorticoid administration suppresses limbic endocannabinoid signaling in mice. *Neuroscience*. 2012; 204:83–89. [PubMed: 21939741]
87. Hong S, Zheng G, Wu X, Snider NT, Owyang C, Wiley JW. Corticosterone mediates reciprocal changes in CB 1 and TRPV1 receptors in primary sensory neurons in the chronically stressed rat. *Gastroenterology*. 2011; 140:627–637. [PubMed: 21070780]
88. Campos AC, Ferreira FR, da Silva WA Jr, Guimaraes FS. Predator threat stress promotes long lasting anxiety-like behaviors and modulates synaptophysin and CB1 receptors expression in brain areas associated with PTSD symptoms. *Neurosci Lett*. 2013; 533:34–38. [PubMed: 23178193]
89. Willner P. Chronic mild stress (CMS) revisited: consistency and behavioural-neurobiological concordance in the effects of CMS. *Neuropsychobiology*. 2005; 52:90–110. [PubMed: 16037678]
90. Reich CG, Taylor ME, McCarthy MM. Differential effects of chronic unpredictable stress on hippocampal CB1 receptors in male and female rats. *Behav Brain Res*. 2009; 203:264–269. [PubMed: 19460405]
91. Marin MT, Cruz FC, Planeta CS. Chronic restraint or variable stresses differently affect the behavior, corticosterone secretion and body weight in rats. *Physiology & behavior*. 2007; 90:29–35. [PubMed: 17023009]
92. Hill MN, Hunter RG, McEwen BS. Chronic stress differentially regulates cannabinoid CB(1) receptor binding in distinct hippocampal subfields. *Eur J Pharmacol*. 2009; 614:66–69. [PubMed: 19426726]
93. Hu W, Zhang M, Czeh B, Zhang W, Flugge G. Chronic restraint stress impairs endocannabinoid mediated suppression of GABAergic signaling in the hippocampus of adult male rats. *Brain Res Bull*. 2011; 85:374–379. [PubMed: 21527320]
94. Hill MN, Carrier EJ, McLaughlin RJ, Morrish AC, Meier SE, Hillard CJ, et al. Regional alterations in the endocannabinoid system in an animal model of depression: effects of concurrent antidepressant treatment. *J Neurochem*. 2008; 106:2322–2336. [PubMed: 18643796]
95. Zoppi S, Perez Nieves BG, Madrigal JL, Manzanares J, Leza JC, Garcia-Bueno B. Regulatory role of cannabinoid receptor 1 in stress-induced excitotoxicity and neuroinflammation. *Neuropsychopharmacology*. 2011; 36:805–818. [PubMed: 21150911]
96. Wang W, Sun D, Pan B, Roberts CJ, Sun X, Hillard CJ, et al. Deficiency in endocannabinoid signaling in the nucleus accumbens induced by chronic unpredictable stress. *Neuropsychopharmacology*. 2010; 35:2249–2261. [PubMed: 20664582]
97. Robinson SA, Loiacono RE, Christopoulos A, Sexton PM, Malone DT. The effect of social isolation on rat brain expression of genes associated with endocannabinoid signalling. *Brain Res*. 2010; 1343:153–167. [PubMed: 20430015]
98. Sciolino NR, Bortolato M, Eisenstein SA, Fu J, Oveisi F, Hohmann AG, et al. Social isolation and chronic handling alter endocannabinoid signaling and behavioral reactivity to context in adult rats. *Neuroscience*. 2010; 168:371–386. [PubMed: 20394803]
99. Koob GF. Brain stress systems in the amygdala and addiction. *Brain research*. 2009; 1293:61–75. [PubMed: 19332030]
100. Serrano A, Rivera P, Pavon FJ, Decara J, Suarez J, Rodriguez de Fonseca F, et al. Differential effects of single versus repeated alcohol withdrawal on the expression of endocannabinoid system-related genes in the rat amygdala. *Alcohol Clin Exp Res*. 2012; 36:984–994. [PubMed: 22141465]
101. Fegley D, Gaetani S, Duranti A, Tontini A, Mor M, Tarzia G, et al. Characterization of the fatty acid amide hydrolase inhibitor cyclohexyl carbamic acid 3'-carbamoyl-biphenyl-3-yl ester

- (URB597): effects on anandamide and oleylethanolamide deactivation. *J Pharmacol Exp Ther.* 2005; 313:352–358. [PubMed: 15579492]
102. Waleh NS, Cravatt BF, Apte-Deshpande A, Terao A, Kilduff TS. Transcriptional regulation of the mouse fatty acid amide hydrolase gene. *Gene.* 2002; 291:203–210. [PubMed: 12095693]
103. Sumislawski JJ, Ramikie TS, Patel S. Reversible gating of endocannabinoid plasticity in the amygdala by chronic stress: a potential role for monoacylglycerol lipase inhibition in the prevention of stress-induced behavioral adaptation. *Neuropsychopharmacology.* 2011; 36:2750–2761. [PubMed: 21849983]
104. Patel S, Hillard CJ. Adaptations in endocannabinoid signaling in response to repeated homotypic stress: A novel mechanism for stress habituation. *Eur J Neurosci.* 2008; 27:2821–2829. [PubMed: 18588527]
105. Hill MN, McLaughlin RJ, Bingham B, Shrestha L, Lee TT, Gray JM, et al. Endogenous cannabinoid signaling is essential for stress adaptation. *Proc Natl Acad Sci U S A.* 2010; 107:9406–9411. [PubMed: 20439721]
106. Hill MN, Ho WS, Meier SE, Gorzalka BB, Hillard CJ. Chronic corticosterone treatment increases the endocannabinoid 2-arachidonylethanolamide in the rat amygdala. *Eur J Pharmacol.* 2005; 528:99–102. [PubMed: 16324692]
107. Rademacher DJ, Hillard CJ. Interactions between endocannabinoids and stress-induced decreased sensitivity to natural reward. *Prog Neuropsychopharmacol Biol Psychiatry.* 2007; 31:633–641. [PubMed: 17258369]
108. Kamprath K, Marsicano G, Tang J, Monory K, Bisogno T, Di Marzo V, et al. Cannabinoid CB1 receptor mediates fear extinction via habituation-like processes. *J Neurosci.* 2006; 26:6677–6686. [PubMed: 16793875]
109. Kamprath K, Plendl W, Marsicano G, Deussing JM, Wurst W, Lutz B, et al. Endocannabinoids mediate acute fear adaptation via glutamatergic neurons independently of corticotropin-releasing hormone signaling. *Genes Brain Behav.* 2009; 8:203–211. [PubMed: 19077175]
110. Kamprath K, Romo-Parra H, Haring M, Gaburro S, Doengi M, Lutz B, et al. Short-term adaptation of conditioned fear responses through endocannabinoid signaling in the central amygdala. *Neuropsychopharmacology.* 2011; 36:652–663. [PubMed: 20980994]
111. Kupferschmidt DA, Newman AE, Boonstra R, Erb S. Antagonism of cannabinoid 1 receptors reverses the anxiety-like behavior induced by central injections of corticotropin-releasing factor and cocaine withdrawal. *Neuroscience.* 2012; 204:125–133. [PubMed: 21784132]
112. Ulrich-Lai YM, Herman JP. Neural regulation of endocrine and autonomic stress responses. *Nature reviews Neuroscience.* 2009; 10:397–409.
113. Dean C. Endocannabinoid modulation of sympathetic and cardiovascular responses to acute stress in the periaqueductal gray of the rat. *Am J Physiol Regul Integr Comp Physiol.* 2011; 300:R771–R779. [PubMed: 21228344]
114. Dean C. Cannabinoid and GABA modulation of sympathetic nerve activity and blood pressure in the dorsal periaqueductal gray of the rat. *Am J Physiol Regul Integr Comp Physiol.* 2011; 301:R1765–R1772. [PubMed: 21940402]
115. Tsou K, Brown S, Sanudo-Pena MC, Mackie K, Walker JM. Immunohistochemical distribution of cannabinoid CB1 receptors in the rat central nervous system. *Neurosci.* 1998; 83:393–411.
116. Hohmann AG, Suplita RL, Bolton NM, Neely MH, Fegley D, Mangieri R, et al. An endocannabinoid mechanism for stress-induced analgesia. *Nature.* 2005; 435:1108–1112. [PubMed: 15973410]
117. Shimizu T, Lu L, Yokotani K. Possible inhibitory roles of endogenous 2-arachidonylethanolamide during corticotropin-releasing factor-induced activation of central sympatho-adrenomedullary outflow in anesthetized rats. *Eur J Pharmacol.* 2010; 641:54–60. [PubMed: 20519139]
118. Seagard JL, Dean C, Patel S, Rademacher DJ, Hopp FA, Schmeling WT, et al. Anandamide content and interaction of endocannabinoid/GABA modulatory effects in the NTS on baroreflex-evoked sympathoinhibition. *Am J Physiol Heart Circ Physiol.* 2004; 286:H992–H1000. [PubMed: 14615281]

119. Pfitzer T, Niederhoffer N, Szabo B. Search for an endogenous cannabinoid-mediated effect in the sympathetic nervous system. *Naunyn Schmiedebergs Arch Pharmacol.* 2005; 371:9–17. [PubMed: 15660243]
120. Quarta C, Bellocchio L, Mancini G, Mazza R, Cervino C, Brulke LJ, et al. CB(1) signaling in forebrain and sympathetic neurons is a key determinant of endocannabinoid actions on energy balance. *Cell Metab.* 2010; 11:273–285. [PubMed: 20374960]
121. Bab I, Zimmer A. Cannabinoid receptors and the regulation of bone mass. *Br J Pharmacol.* 2008; 153:182–188. [PubMed: 18071301]
122. Tam J, Trembovler V, Di Marzo V, Petrosino S, Leo G, Alexandrovich A, et al. The cannabinoid CB1 receptor regulates bone formation by modulating adrenergic signaling. *FASEB J.* 2008; 22:285–294. [PubMed: 17704191]
123. Kurihara J, Nishigaki M, Suzuki S, Okubo Y, Takata Y, Nakane S, et al. 2-Arachidonoylglycerol and anandamide oppositely modulate norepinephrine release from the rat heart sympathetic nerves. *Jpn J Pharmacol.* 2001; 87:93–96. [PubMed: 11676206]
124. O'Sullivan SE, Kendall PJ, Kendall DA. Endocannabinoids and the cardiovascular response to stress. *J Psychopharmacol.* 2012; 26:71–82. [PubMed: 21708837]
125. Kunos G, Batkai S, Offertaler L, Mo F, Liu J, Karcher J, et al. The quest for a vascular endothelial cannabinoid receptor. *Chem Phys Lipids.* 2002; 121:45–56. [PubMed: 12505689]
126. Mukhopadhyay P, Batkai S, Rajesh M, Czifra N, Harvey-White J, Hasko G, et al. Pharmacological inhibition of CB1 cannabinoid receptor protects against doxorubicin-induced cardiotoxicity. *J Am Coll Cardiol.* 2007; 50:528–536. [PubMed: 17678736]
127. Pacher P, Gao B. Endocannabinoids and Liver Disease. III. Endocannabinoid effects on immune cells: implications for inflammatory liver diseases. *Am J Physiol Gastrointest Liver Physiol.* 2008; 294:G850–G854. [PubMed: 18239059]
128. Wagner JA, Hu K, Bauersachs J, Karcher J, Wiesler M, Goparaju SK, et al. Endogenous cannabinoids mediate hypotension after experimental myocardial infarction. *J Am Coll Cardiol.* 2001; 38:2048–2054. [PubMed: 11738314]
129. Khansari DN, Murgu AJ, Faith RE. Effects of stress on the immune system. *Immunology Today.* 1990; 11:170–175. [PubMed: 2186751]
130. Baschant U, Lane NE, Tuckermann J. The multiple facets of glucocorticoid action in rheumatoid arthritis. *Nat Rev Rheumatol.* 2012; 8:645–655. [PubMed: 23045254]
131. Severn A, Rapson NT, Hunter CA, Liew FY. Regulation of tumor necrosis factor production by adrenaline and beta-adrenergic agonists. *Journal of immunology.* 1992; 148:3441–3445.
132. Mnich SJ, Hiebsch RR, Huff RM, Muthian S. Anti-inflammatory properties of CB1-receptor antagonist involves beta2 adrenoceptors. *J Pharmacol Exp Ther.* 2010; 333:445–453. [PubMed: 20164299]
133. Semenoff-Segundo A, Porto AN, Semenoff TA, Cortelli JR, Costa FO, Cortelli SC, et al. Effects of two chronic stress models on ligature-induced periodontitis in Wistar rats. *Arch Oral Biol.* 2012; 57:66–72. [PubMed: 22119224]
134. Rettori E, De Laurentiis A, Zorrilla Zubilete M, Rettori V, Elverdin JC. Anti-inflammatory effect of the endocannabinoid anandamide in experimental periodontitis and stress in the rat. *Neuroimmunomodulation.* 2012; 19:293–303. [PubMed: 22777139]
135. Kulkarni-Narla A, Brown DR. Localization of CB1-cannabinoid receptor immunoreactivity in the porcine enteric nervous system. *Cell Tissue Res.* 2000; 302:73–80. [PubMed: 11079717]
136. Coutts AA, Irving AJ, Mackie K, Pertwee RG, Anavi-Goffer S. Localisation of cannabinoid CB(1) receptor immunoreactivity in the guinea pig and rat myenteric plexus. *J Comp Neurol.* 2002; 448:410–422. [PubMed: 12115703]
137. Konturek PC, Brzozowski T, Konturek SJ. Stress and the gut: pathophysiology, clinical consequences, diagnostic approach and treatment options. *J Physiol Pharmacol.* 2011; 62:591–599. [PubMed: 22314561]
138. Lightman SL. The neuroendocrinology of stress: a never ending story. *J Neuroendocrinol.* 2008; 20:880–884. [PubMed: 18601712]

139. Shen L, Yang XJ, Qian W, Hou XH. The role of peripheral cannabinoid receptors type 1 in rats with visceral hypersensitivity induced by chronic restraint stress. *J Neurogastroenterol Motil.* 2010; 16:281–290. [PubMed: 20680167]
140. Zoppi S, Madrigal JL, Perez-Nievas BG, Marin-Jimenez I, Caso JR, Alou L, et al. Endogenous cannabinoid system regulates intestinal barrier function in vivo through cannabinoid type 1 receptor activation. *Am J Physiol Gastrointest Liver Physiol.* 2012; 302:G565–G571. [PubMed: 22135307]
141. Woof JM, Mestecky J. Mucosal immunoglobulins. *Immunol Rev.* 2005; 206:64–82. [PubMed: 16048542]
142. Hillard CJ, Weinlander KM, Stuhr KL. Contributions of endocannabinoid signaling to psychiatric disorders in humans: genetic and biochemical evidence. *Neuroscience.* 2012; 204:207–229. [PubMed: 22123166]
143. Agrawal A, Nelson EC, Littlefield AK, Bucholz KK, Degenhardt L, Henders AK, et al. Cannabinoid receptor genotype moderation of the effects of childhood physical abuse on anhedonia and depression. *Arch Gen Psychiatry.* 2012; 69:732–740. [PubMed: 22393204]
144. Llorente-Berzal A, Fuentes S, Gagliano H, Lopez-Gallardo M, Armario A, Viveros MP, et al. Sex-dependent effects of maternal deprivation and adolescent cannabinoid treatment on adult rat behaviour. *Addict Biol.* 2011; 16:624–637. [PubMed: 21521421]
145. Roberts CJ, Stuhr KL, Hutz MJ, Raff H, Hillard CJ. Endocannabinoid signaling in hypothalamic-pituitary-adrenocortical axis recovery following stress: effects of indirect agonists and comparison of male and female mice. *Pharmacol Biochem Behav.* 2014; 117:17–24. [PubMed: 24316201]
146. Palanza P. Animal models of anxiety and depression: how are females different? *Neurosci Biobehav Rev.* 2001; 25:219–233. [PubMed: 11378178]

### Highlights

CB1 receptor signaling is mobilized by acute stress and serves to dampen the endocrine responses

Chronic variable stress down-regulates CB1 receptor signaling in the hippocampus and ventral striatum

Increased CB1 receptor signaling contributes to habituation of endocrine and some behavioral responses to repeated stress

Reduced CB1 receptor signaling exacerbates the effects of stress and may contribute to its negative consequences