

Effects of Thyroid Hormone on Sodium Pump Sites, Sodium Content, and Contractile Responses to Cardiac Glycosides in Cultured Chick Ventricular Cells

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Abstract. Sensitivity of cardiac muscle to digitalis glycosides depends on the thyroid state. The mechanism of this interaction was investigated at the cellular level using spontaneously beating monolayers of cultured chick embryo ventricular cells. Cells were grown for 48 h in serum-free medium containing concentrations of triiodothyronine (T_3) from zero to 10^{-7} M, and the total number of sodium pump sites, sodium content, and contractile amplitude in the presence and absence of various concentrations of ouabain were determined. T_3 caused a concentration-dependent increase in the number of specific ouabain binding sites; the maximal increase to 160% of control was observed in response to 10^{-8} M T_3 . T_3 lowered steady-state cellular sodium content in a concentration-dependent manner, also. Ouabain (1 μ M) exposure elevated cellular sodium content in all cells, but the increase was greatest in cells grown in T_3 -free medium and least in cells grown in 10^{-8} M T_3 . The positive inotropic and toxic effects of ouabain in cells grown in 10^{-8} M T_3 were diminished at any given ouabain concentration, and thus, the dose-response curve was shifted to the right. These results indicate that T_3 causes induction of additional sodium pump sites that are functional. The increased tolerance of hyperthyroid cells and reduced tolerance of hypothyroid cells to cardiac glycosides can be explained by these changes in the number of sodium pump sites and cellular sodium

content, and consequently, calcium influx via sodium-calcium exchange.

Introduction

Changes in thyroid state have been well documented to affect the sensitivity of the heart to digitalis glycosides. Thus, hyperthyroidism and hypothyroidism are associated with increased and decreased tolerance, respectively, to the cardiac effects of digitalis (1-3). The mechanism of altered digitalis sensitivity has been attributed in part to changes in pharmacokinetic disposition of glycosides, since significantly higher serum levels of glycosides were found in patients with hypothyroidism than in patients with hyperthyroidism following administration of identical doses (4, 5). These differences in serum levels of cardiac glycosides appear to be accounted for by changes in distribution volume and in renal clearance (5-7). However, Kokenge et al. (8) observed no significant differences in serum digitoxin levels between hypo- and hyperthyroid patients receiving similar doses. Furthermore, it is a common clinical experience to find the ventricular rate of thyrotoxic patients with atrial fibrillation difficult to control, despite relatively large doses and high serum concentrations of digoxin. Thus, the fundamental basis for the influence of thyroid status on cardiac glycoside effects remains incompletely understood.

At the cellular level, thyroid hormone has been shown to increase NaK-ATPase activity (9-11) and the number of NaK-ATPase complexes in several types of tissues (12-15). If other properties of myocardial cells remained constant, we would expect such changes in the number of sodium pump sites or sodium pump activity to alter intracellular sodium content, and hence, sodium-calcium exchange across sarcolemmal membranes (16-19). We have previously shown that cardiac glycosides cause elevation of intracellular sodium content in cultured chick ventricular cells, and that this increase in $[Na^+]_i$ is associated with an increase in rapidly exchangeable cellular Ca^{++} and the increased contractile state associated with enhanced Na-Ca exchange (20, 21). In cardiac tissues exposed to excessive levels of thyroid hormone, we postulate that intra-

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cellular sodium content may not be elevated to the same extent by cardiac glycosides as a result of increased sodium pump activity compared with tissues exposed to normal or low levels of thyroid hormone. If this occurs, contractile responses to positive inotropic agents that act by elevating intracellular sodium content would be expected to be reduced due to decreased calcium influx via sodium-calcium exchange (17, 19, 22). Therefore, we used cultured chick ventricular cell monolayer preparations, for which the growth conditions can be precisely controlled, to explore the effects of graded amounts of T_3 and ouabain on NaK-ATPase, intracellular sodium content, and contractile state.

Methods

Tissue Culture. Monolayer cultures of spontaneously contracting chick embryo ventricular cells were prepared as previously described (20). Briefly, hearts of 10-d-old chick embryos were removed and ventricular tissue was cut into small fragments ($<0.5 \text{ mm}^3$); individual cells were isolated by four cycles of trypsinization with 0.025% (wt/vol) trypsin at 37°C . The culture medium contained 6% heat-inactivated fetal calf serum, 40% M199 with Hanks's salts, 0.1% penicillin-streptomycin solution, and 54% balanced salt solution. Balanced salt solution contained 116 mM NaCl, 1.0 mM NaH_2PO_4 , 0.8 mM $\text{MgSO}_4 \cdot \text{H}_2\text{O}$, 1.18 mM KCl, 26.2 mM NaHCO_3 , 0.87 mM CaCl_2 , and 5.5 mM glucose. The final concentrations of K^+ , Na^+ , and Ca^{++} in culture medium were 4.0, 137, and 0.97 mM, respectively. The cell suspension was diluted to 5×10^5 cells/ml and placed in plastic culture dishes (100×20 mm or 60×15 mm, Falcon Labware, Oxnard, CA). The larger culture dishes contained 25-mm circular glass coverslips and were used in ion flux studies whereas the smaller dishes without coverslips were used in [^3H]ouabain-binding studies. Cultures were incubated in a humidified 5% CO_2 :95% air atmosphere at 37°C . Confluent monolayers in which at least 80% of cells exhibited spontaneous synchronous contractions developed by 2–3 d of incubation.

After the cells were allowed to grow in culture medium containing 6% fetal calf serum for 48 h, the medium was replaced with serum-free medium (23) containing 10^{-6} M insulin, 5×10^{-8} M transferrin, 10^{-9} M selenium, and concentrations of triiodothyronine (T_3)¹ varying from zero to 10^{-7} M. The final concentrations of K^+ , Na^+ , and Ca^{++} in serum-free medium were identical to those in serum-containing medium. Cells continued to contract spontaneously and remained viable for at least 2 wk in the serum-free medium.

[^3H]ouabain binding. The number of sodium pump sites in the sarcolemma was estimated from [^3H]ouabain binding to intact, beating cultured ventricular cells. Specific binding of [^3H]ouabain was measured by modification of the methods of McCall (24) and Aiton et al. (25). Briefly, monolayers of ventricular cells attached to culture dishes were incubated in K^+ -free HEPES-buffered solution (pH 7.35) containing 4 mM HEPES, 0.05 mM CaCl_2 , 137 mM NaCl, 0.5 mM MgCl_2 , and various concentrations of [^3H]ouabain (60–1000 nM) for predetermined periods of time (0–20 min) at 37°C . Stock [^3H]ouabain (18 Ci/mmol, New England Nuclear, Boston, MA) was diluted with unlabeled ouabain to give the desired final concentrations of [^3H]ouabain. Following [^3H]ouabain exposure, the cells were quickly scraped from the dish and filtered through a microfiber filter (pore size, 12 μm ;

Whatman Laboratory Products, Inc., Clifton, NJ). Filters were washed three times with ice-cold HEPES-buffered solution (pH 7.35) containing 20 mM KCl and dissolved in Hydrofluor (National Diagnostics, Inc., Somerville, NJ). Radioactivity on the filter was assayed using a liquid scintillation spectrometer. Specific [^3H]ouabain binding was calculated as total binding in the absence of 10^{-3} M nonlabeled ouabain minus the value obtained in its presence, and is expressed as picomoles per milligram protein. To determine protein content on each filter, cells on the plates not exposed to [^3H]ouabain were thoroughly washed to remove all [^{14}C]leucine in the extracellular space by placing the plates in three large volumes (50 ml) of bicarbonate buffer solution for 5 min each at 37°C . The cells were then dissolved, and aliquots of the medium containing dissolved cells were assayed for protein content using the method of Lowry et al. (26).

Contractility measurements. Changes in the contractile state of individual cells in the monolayers were assessed by the use of an optical-video system as previously described (20, 21). A glass coverslip with attached monolayer of cultured cells was continuously superfused with serum-free medium at a rate of 2 ml/min in a chamber with inlet and outlet ports on the stage of an inverted phase-contrast microscope. The pH of the medium was maintained at 7.4 by continuously gassing the chamber with a 95% air:5% CO_2 gas mixture. A constant temperature of 37°C was maintained by enclosing the microscope in a thermostated Lucite box. Following a 15-min equilibration period in control medium, cells were superfused with otherwise identical solution containing ouabain at concentrations varying from zero to 4 μM , and changes in the amplitude of motion of an individual cell were monitored. Only one cell from each coverslip was used and the change in the amplitude of contraction was expressed as a percent of the change observed in response to exposure to 3.6 mM $[\text{Ca}^{++}]_0$, a concentration which produced a maximal contractile response in cells grown under these conditions.

Sodium content and ^{42}K -uptake. The procedures for estimating sodium content have been described in detail by Biedert et al. (20) and Barry and Smith (19). For determination of sodium content, glass coverslips with attached monolayers of cells were immersed in bicarbonate-buffered solution (pH 7.35) containing ^{24}Na (5 $\mu\text{Ci/ml}$, $[\text{Na}^+] = 143 \text{ mM}$) and the cells were labeled to asymptote (30 min). After appropriate periods of labeling with ^{24}Na , cells were washed twice for 8 s each by gently agitating the glass coverslips sequentially in two 60 ml volumes of ice-cold balanced salt solution containing no radioisotope. The cells were then dissolved in medium containing 1% sodium dodecyl sulfate and 10 mM sodium borate. Aliquots of the medium containing dissolved cells were assayed for radioactivity in a liquid scintillation spectrometer (model 3330, Packard Instrument Co., Inc., Downers Grove, IL) and for protein content (26).

The rate of monovalent cation transport was estimated by measurement of ^{42}K -uptake. Coverslips with attached monolayers of cells were similarly immersed in bicarbonate buffer solution (pH 7.35) containing 4 mM K^+ and a tracer quantity of ^{42}K (5 $\mu\text{Ci/ml}$) with or without 1 mM ouabain. Specific ^{42}K -uptake was calculated as the difference between values obtained in the presence and absence of 1 mM ouabain. The initial rate of ^{42}K -uptake was determined similarly following a 10-min preexposure of cells to medium containing zero K^+ .

Cell density correction. To normalize for cell density on each culture dish or coverslip, the monolayers were incubated in either L-[^{14}C]leucine (0.05 $\mu\text{Ci/ml}$) or L-[4,5- ^3H (N)]leucine (0.1 $\mu\text{Ci/ml}$) for 24 h before each experiment. [^{14}C]leucine or [^3H]leucine was incorporated into the cell protein and the ^{14}C - or ^3H -counts permitted

1. Abbreviations used in this paper: T_3 , triiodothyronine.

estimation of cell density on each culture dish or glass coverslip, respectively. The relationship between radioactive counts and protein concentration (26) allowed accurate estimation of protein concentration in each culture dish or glass coverslip. Thus, simultaneous counting of ^3H and ^{24}Na , or ^{14}C and ^3H permitted normalization of sodium content or [^3H]ouabain binding per milligram cell protein.

Miscellaneous. Protein concentration was assayed by the method of Lowry et al. (26) using crystalline bovine serum albumin as standard. Hanks' salt solution, M199, and fetal calf serum were purchased from Gibco Laboratories, Grand Island, NY. [^3H]ouabain and $^{24}\text{NaCl}$ were purchased from New England Nuclear, Boston, MA. Insulin, transferrin, and T_3 were purchased from Sigma Chemical Co., St. Louis, MO. Selenium (Spectrapure) was purchased from United Mineral and Chemical Corp., NY. All other chemicals used were of the highest grade commercially available. All solutions were sterilized by filtering through Nalgene sterilization filters (pore size, $0.45\ \mu\text{m}$; Nalge Co., Rochester, NY). Insulin containing solutions were filtered through Millex-GV filter (pore size, $0.22\ \mu\text{m}$; Millipore Corp., Bedford, MA).

Statistical analyses were performed using Student's *t* test and two-way analysis of variance.

Results

[^3H]ouabain binding. Fig. 1 shows the binding of $10^{-7}\ \text{M}$ [^3H]ouabain to intact cells grown in serum-free medium containing either zero or $10^{-8}\ \text{M}$ T_3 . In both types of cells, the specific [^3H]ouabain binding was rapid and reached a steady state within 10 min. Specific [^3H]ouabain binding followed pseudo first order kinetics with an association rate constant of $2.8 \times 10^6\ \text{M}^{-1}\ \text{min}^{-1}$ both in cells grown in zero and in $10^{-8}\ \text{M}$ T_3 , indicating that the kinetic characteristics of [^3H]ouabain binding to intact cultured heart cells are not affected by thyroid hormone in K^+ -free medium. The steady state level of [^3H]ouabain binding was increased, however, by growth in $10^{-8}\ \text{M}$ T_3 from 1.67 ± 0.27 to $2.57 \pm 0.26\ \text{pmol/mg}$ protein ($P < 0.05$).

To examine the effect of T_3 concentration in growth media on the total number of sodium pump sites in the sarcolemmal

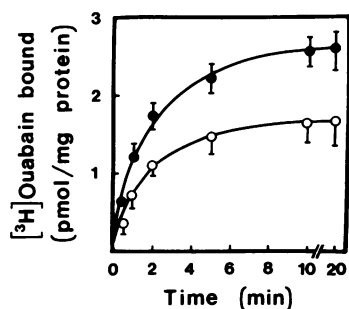


Figure 1. Time-dependency of [^3H]ouabain binding in K^+ -free medium. Cultured heart cells from 10-d-old chick embryos were grown in serum-containing culture medium for 48 h, and subsequently, grown in serum-free medium containing either zero (open circles) or $10^{-8}\ \text{M}$ T_3 (filled circles) for another 48 h. Cells were exposed to

$10^{-7}\ \text{M}$ [^3H]ouabain for 0.5–20 min. Cells were then filtered, washed, and dissolved, and the amount of [^3H]ouabain bound was estimated from the radioactivity present on the filter. Specific binding, the difference in values obtained in the presence and absence of $1\ \text{mM}$ nonlabeled ouabain, is plotted. Each point is the mean of five determinations. Vertical lines are $\pm 1\ \text{SEM}$.

membrane, cultured chick ventricular cells were incubated to equilibrium with concentrations of [^3H]ouabain ranging from 0.06 to $1.0\ \mu\text{M}$, and specifically bound [^3H]ouabain was determined. Fig. 2 shows the concentration dependency of specific [^3H]ouabain binding in K^+ -free medium in cells grown for 48 h in medium containing $10^{-10}\ \text{M}$ T_3 . The binding data indicate that near saturation of ouabain binding sites occurs with $1\ \mu\text{M}$ [^3H]ouabain. Therefore, the total number of sodium pump sites in the cell membrane was determined by assaying the amount of [^3H]ouabain bound following incubation of cells to equilibrium in $1\ \mu\text{M}$ [^3H]ouabain.

To determine the number of sodium pump sites maintained in the absence of T_3 in the growth medium, cells were grown in serum-containing culture medium for 48 h and subsequently grown for several days in serum-free medium containing no added T_3 . The total number of [^3H]ouabain binding sites gradually declined to a new steady state level on the third day of growth in T_3 -free, serum-free medium and remained at this level for another 48 h. As shown in Fig. 3, the total number of sodium pump sites under these conditions was $\sim 80\%$ of the initial value observed just before the replacement of the culture medium by serum-free medium.

As shown in Fig. 1, growth in T_3 elevates the number of sodium pump sites in the sarcolemmal membrane. To determine whether this is a concentration-dependent phenomenon, we studied the effects of graded concentrations of T_3 in the serum-free culture medium on the total number of sodium pump sites. Growth of cells in T_3 concentrations from 10^{-10} to $10^{-7}\ \text{M}$ for 48 h caused induction of additional sodium pump sites in the sarcolemmal membrane (Fig. 4). This effect was concentration-dependent. Increasing concentrations of T_3 to $10^{-8}\ \text{M}$ were associated with increasing numbers of sodium pump sites. The peak level of sodium pump sites was produced by $10^{-8}\ \text{M}$ T_3 and beyond that, the number of sodium pump sites decreased again. This was probably due to toxic effects of $10^{-7}\ \text{M}$ T_3 as indicated by altered cell morphology including the presence of large cytoplasmic vacuoles, unresponsiveness to positive inotropic agents such as ouabain or isoproterenol,

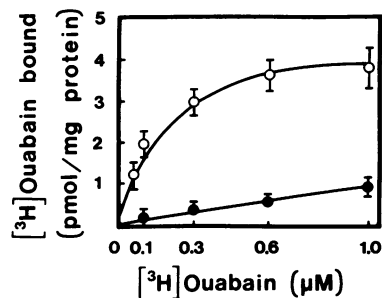


Figure 2. Concentration-dependency of [^3H]ouabain binding in K^+ -free medium. Cells were grown in serum-free medium containing $10^{-10}\ \text{M}$ T_3 for 48 h, then exposed to various concentrations of [^3H]ouabain (0.06 – $1.0\ \mu\text{M}$) for 20 min at 37°C . Cells were then filtered, washed and dissolved, and radioac-

tivity on the filter was assayed to determine bound [^3H]ouabain. The amount of specifically bound [^3H]ouabain (open circles) is the difference between total binding and that observed in the presence of $1\ \text{mM}$ nonlabeled ouabain (filled circles). Each point is the mean of six determinations. Vertical lines indicate $\pm 1\ \text{SEM}$.

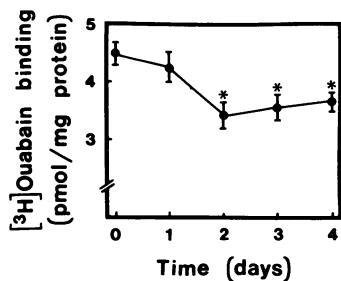


Figure 3. Effects of growth of cultured chick ventricular cells in T_3 -free, serum-free medium on the number of ouabain binding sites. Cells were first grown in standard medium containing 6% fetal calf serum for 48 h. The medium was then replaced with serum-free, T_3 -free medium (day 0) and cells were allowed to continue to grow for four more

days. At each day, the total number of ouabain binding sites was determined by incubating the cells with 10^{-6} M $[^3H]$ ouabain and assaying for specific $[^3H]$ ouabain binding. Each point and vertical line represent mean \pm SE of six determinations. *Significantly different from the value observed at day 0 ($P < 0.05$).

and a marked reduction in the number of spontaneously contracting cells. Cells grown in medium containing 10^{-8} M or lower concentrations of T_3 did not show any such signs of toxicity. Synchronous contraction of normal frequency (100–140 beats/min) were present in at least 80% of cells on the plate.

Since T_3 might in principle modify the growth rate, cell volume, and protein content of cultured chick ventricular cells, the number of sodium pump sites expressed per milligram protein might not reflect the change in sites per cell. To clarify this issue, identical numbers of cells were plated as previously described. After 48 h of growth in serum-containing medium, incubation was continued in serum-free media containing either 10^{-8} M or zero T_3 . After 48 h, cells were suspended by gentle trypsinization and numbers of cells and protein content per plate were determined. T_3 did not significantly alter either the number of cells or protein content per plate. The mean cell numbers per plate ($n = 20$) were $557,000 \pm 11,000$ and $591,000 \pm 15,000$, and mean protein contents per plate were 1.05 ± 0.04 and 1.06 ± 0.03 mg in cells exposed to 10^{-8} M and zero T_3 , respectively. In cells grown in 10^{-10} M T_3 , the density of sodium pump sites was $\sim 3.7 \times 10^6$ sites per cell. Others have reported values of 8×10^5 sites per cell in HeLa cells

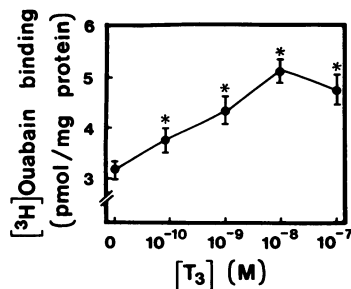


Figure 4. Effects of growth of cells in graded concentrations of T_3 on the number of ouabain binding sites. Following growth of cells in serum-containing medium for 48 h, medium was replaced with a serum-free medium containing 0, 10^{-10} , 10^{-9} , 10^{-8} , or 10^{-7} M T_3 and cells were allowed to grow for another

48 h. The total number of ouabain binding sites in the cell membrane was then determined as above. Each point and vertical line represent mean \pm SE of six determinations. *Significantly different from the value observed with zero T_3 ($P < 0.05$).

(25) and 1.6×10^6 sites per cell in cultured rat heart cells (24). These results indicate that the number of ouabain binding sites expressed per milligram protein permits a valid comparison under our experimental conditions.

The possibility that growth in T_3 altered the affinity of NaK-ATPase for ouabain was next examined. As indicated by the parallel slopes in the Scatchard plots of $[^3H]$ ouabain binding data obtained with cells grown in medium containing 10^{-10} or 10^{-8} M T_3 (Fig. 5), the change in the number of $[^3H]$ ouabain binding sites produced by T_3 exposure was not due to altered affinity of these sites to ouabain. These results indicate that T_3 concentrations spanning the physiologic range modulate the number of sodium pump sites in the sarcolemmal membrane of cultured chick ventricular cells.

Intracellular sodium content. The effect of T_3 concentration in growth media on intracellular sodium content was examined to determine the functional role of the additional sodium pump sites in the sarcolemmal membrane. Cells grown for 48 h in medium containing various concentrations of T_3 were incubated to asymptote in medium containing $^{24}Na^+$ for 30 min and the cellular sodium content was determined. As shown in Fig. 6, increasing concentrations of T_3 in the culture medium caused graded reductions in intracellular sodium content. Mean cellular sodium contents in cells grown in T_3 -free medium and in cells grown in medium containing 10^{-8} M T_3 were 82.0 ± 7.5 and 68.5 ± 4.6 nmol/mg protein, respectively, a 20% difference in sodium content. Cellular Na contents in cells grown in serum-free, T_3 -free medium for 48 and 72 h were 82.0 ± 7.5 and 80.8 ± 8.6 nmol/mg protein, respectively, indicating that growth in serum-free medium alone had no significant effect on cellular Na content over this time period. For comparison, cells grown in the presence of conventional serum-containing media had a mean Na content of 80.6 ± 5.2 nmol/mg protein. When cells were exposed to 1 μ M ouabain for 30 min at the end of 48 h growth in media containing various T_3 concentrations and cellular sodium content was similarly determined, $[Na^+]_i$ increased by $\sim 40\%$ in groups of cells grown in each concentration of T_3 . Thus, growth in T_3 also produced concentration-dependent decreases in $[Na^+]_i$ in

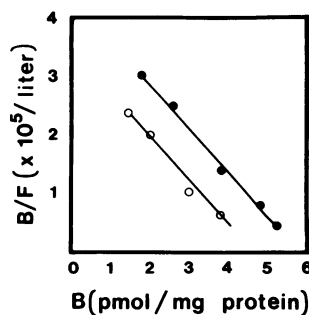


Figure 5. Scatchard plots of $[^3H]$ ouabain binding data. Cells were incubated with graded concentrations of $[^3H]$ ouabain in zero $[K^+]_o$ medium for 20 min and $[^3H]$ ouabain bound to the cells was assayed. The specific $[^3H]$ ouabain binding, defined as the difference in binding observed in the presence and absence of 1 mM nonlabeled ouabain, was used to calculate

bound (B) and free (F) ouabain. Open circles and filled circles are plots of ouabain binding data observed in cells grown in medium containing 10^{-10} and 10^{-8} M T_3 , respectively.

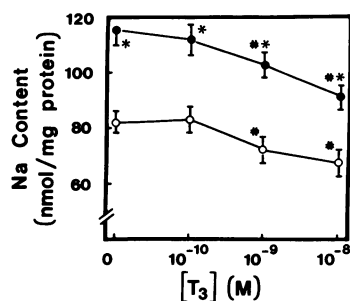


Figure 6. Effect of growth of cells in graded concentrations of T_3 on cellular sodium content. See legend to Fig. 2 for growth conditions. Coverslips with attached monolayer of cells were immersed in medium containing $^{24}\text{Na}^+$ for 30 min (open circles) or in medium containing $1 \mu\text{M}$ ouabain for 10 min, and then, transferred to medium con-

taining $1 \mu\text{M}$ ouabain and $^{24}\text{Na}^+$ for 30 min (filled circles). After washing briefly to remove extracellular $^{24}\text{Na}^+$, cells were dissolved and radioactivity in the cells was assayed. The sodium content was calculated for each coverslip and expressed as nanomoles per milligram protein. Each point and vertical line represent mean \pm SE of eight observations. *, Significantly different from the corresponding control values ($P < 0.05$). #, Significantly different from the value observed in cells grown in T_3 -free medium within the same experimental group ($P < 0.05$).

the presence of $1 \mu\text{M}$ ouabain. These observations indicate that the T_3 content of growth media controls intracellular sodium content by modulating the number of active sodium pump sites in the sarcolemmal membrane.

Contractility measurements. Given the known relationship between sodium pump inhibition and inotropic and toxic effects of cardiac glycosides (17, 22, 27), one would predict that cells with higher numbers of active sodium pump sites would be less responsive and more tolerant to the effects of ouabain compared with cells with lower numbers of sodium pumps, assuming that the affinity of pump sites for ouabain did not change. To examine the effects of altered numbers of sodium pumps produced by growth of cells in varying concentrations of T_3 on the inotropic and toxic effects of cardiac glycosides, dose-response curves for ouabain were determined by measuring percent change in amplitude of cell motion using an optical-video system (19). For each cell, the maximal changes produced by a given concentration of ouabain were determined as the percent of change induced in the same cell by $3.6 \text{ mM } [\text{Ca}^{++}]_0$, a concentration of Ca^{++} that produced maximal increases in amplitude of cell motion in both T_3 -deficient and T_3 -supplemented cells.

Over the range from 0.1 to $4 \mu\text{M}$, increasing concentrations of ouabain produced graded positive inotropic effects in cultured heart cells. As shown in Fig. 7, for each concentration of ouabain, cells grown in T_3 -free medium showed a significantly greater positive inotropic effect than cells grown in medium containing $10^{-8} \text{ M } T_3$. Thus, the concentration-effect curve for ouabain was shifted to the right by growth of cells in high levels of T_3 . The maximal positive inotropic effect in cells grown in T_3 -free medium was produced by $2 \mu\text{M}$ ouabain, whereas that in cells grown in medium containing $10^{-8} \text{ M } T_3$ occurred at $4 \mu\text{M}$ ouabain. The maximal augmentation of inotropic state achieved in both types of cells was not signifi-

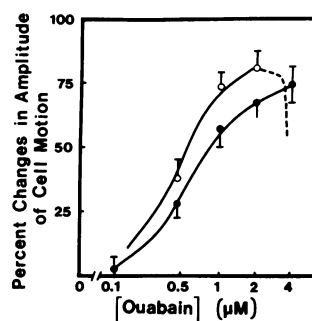


Figure 7. Effects of growth of cells in medium containing zero or $10^{-8} \text{ M } T_3$ on the concentration-effect relation for ouabain. See legend to Fig. 2 for growth conditions. Contractile responses of cells to several concentrations of ouabain (0.1 – $4 \mu\text{M}$) were determined using the optical-video system. The responses of a single cell to ouabain are expressed as percentages of the response of that same cell to $3.6 \text{ mM } \text{Ca}^{++}$.

Open circles and filled circles represent responses of cells grown in medium containing 0 and $10^{-8} \text{ M } T_3$, respectively. Dotted line represents the development of toxicity.

cantly different. For each type of cell, higher concentrations of ouabain than those which produced maximal positive inotropic effects were toxic as manifested by a marked negative inotropic effect, spontaneous arrhythmic contractions, and/or a concomitant shift in resting base line (contracture). Thus, these results show that sensitivity of cardiac myocytes to positive inotropic and toxic effects of ouabain is a function of the concentration of thyroid hormone in which the cells are grown.

Discussion

Sensitivity of the heart to digitalis glycosides *in vivo* depends in part on the thyroid state, since hyperthyroidism is associated with increased tolerance to cardiac glycosides and hypothyroidism with decreased tolerance (1–3). The results of the present study show that a direct effect of thyroid hormone on cultured cardiac myocytes significantly alters responsiveness to cardiac glycosides. Specifically, the effects of thyroid hormone on the sarcolemmal NaK-ATPase and the resulting changes in ion transport across the cell membrane appear to be important mechanisms responsible for the dependence of digitalis' action on thyroid status.

The effects of thyroid hormone in increasing membrane NaK-ATPase have been well documented in a variety of species and tissues with respect to both enzyme activity (9, 11, 28, 29) and the number of sodium pump sites as judged by ouabain binding (12, 13, 15, 28). In cultured chick ventricular cells, T_3 also caused an elevation of sodium pump sites in the sarcolemmal membrane in a concentration-dependent manner. A maximal increase to 160% of the control number of sodium pump sites was produced by growing the cells in medium containing $10^{-8} \text{ M } T_3$ for a period of 48 h. It has been reported that the half-life of T_3 in hepatocyte cultures containing serum is 3.5 h (29). Therefore, it is possible that this concentration of T_3 (10^{-8} M) did not maximally stimulate NaK-ATPase induction due to metabolism of T_3 . However, our finding that a higher T_3 concentration (10^{-7} M) was toxic to the cells, and the fact that the culture medium was serum free, which may

prolong the half-life of T_3 , suggest that the 1.6-fold increase in the number of sodium pump sites is near maximal under the present experimental conditions. The exact mechanism of this induction process is not known, although the studies of Lo and Edelman (30) and Lo and Lo (31) indicate that the increase in sodium pump site density in renal cortex is due to an increase in synthesis rate rather than a change in degradation rate of pump sites.

The initial rates of specific (i.e., ouabain-sensitive) ^{42}K -uptake into cells loaded with Na by 15-min preincubation in zero K^+ medium were 102 ± 5 and 71 ± 4 nmol/mg protein per min in T_3 -treated (10^{-8} M) and untreated cells, respectively, indicating a 44% increase in maximal sodium pump activity in cells grown in 10^{-8} M T_3 . This suggests that the increase in the number of ouabain binding sites is roughly proportional to the increase in maximal monovalent cation active transport activity. ^{42}K -uptake studies with cultured heart cells not loaded with Na^+ prior to ^{42}K -uptake measurements showed that cells grown in 10^{-8} M T_3 as compared with zero T_3 had a 26% greater monovalent cation active transport activity (26.7 ± 1.1 vs. 21.3 ± 0.9 nmol/mg protein per min). This is likely related, at least in part, to the 20% increase in beating rate of cells grown in the presence of 10^{-8} M T_3 (see below). Since the number of ouabain binding sites was increased 1.6-fold by growth in 10^{-8} M T_3 under identical conditions, the turnover rate of individual sodium pumps in T_3 -treated cells was $\sim 80\%$ of that in cells grown in the absence of T_3 . However, the total monovalent cation transport per cell was clearly elevated and our finding that growth of cells in medium containing high concentrations of T_3 was associated with lower intracellular sodium content indicates that the additional pump sites induced by T_3 are indeed functional and contribute to the lowering of intracellular sodium content. Similar results have been observed in rat diaphragm and rat heart (32), although contrasting results have also been reported for mouse diaphragm muscle in which T_3 was found to have no effect on intracellular Na^+ - or K^+ -content (33) and for rat erythrocytes in which T_3 produced inconsistent changes in intracellular Na^+ concentration (32). The reasons for these differing results are not clear, but may be due to the species difference or to differing experimental techniques.

A concentration of ouabain ($1 \mu\text{M}$) that causes a positive inotropic effect without evidence of toxicity (20, 21) elevated $[\text{Na}^+]_i$ by $\sim 40\%$ in all cells included in the present study, irrespective of the T_3 level in growth media. Hence, as shown in Fig. 6, the response to thyroid hormone was a concentration-dependent decrease in steady-state $[\text{Na}^+]_i$ in the presence of $1 \mu\text{M}$ ouabain. These ouabain-induced changes in intracellular sodium were associated with altered responses of the cell to ouabain. Cells grown in medium containing 10^{-8} M T_3 were inotropically less responsive to ouabain than cells grown in T_3 -free medium (Fig. 7). In cells grown in medium containing an intermediate range of concentrations of T_3 , the inotropic responsiveness to ouabain was also intermediate. Furthermore,

the cells exhibited a greater tolerance to toxic effects of ouabain when the concentration of T_3 was increased in the culture medium. Thus, the most straightforward interpretation of our findings is that cells with increased numbers of sodium pump sites in response to growth in 10^{-8} M T_3 have altered sensitivity to the inotropic and toxic effects of cardiac glycosides as a result of reduced intracellular sodium levels at any given cardiac glycoside concentration.

In addition to causing changes in the number of sodium pump sites and $[\text{Na}^+]_i$, growth in T_3 augmented the spontaneous contraction rate of the cultured heart cells. In cells grown in medium containing no T_3 or 10^{-8} M T_3 , the frequency of spontaneous contractions was 108 ± 2.4 or 124 ± 4.0 /min ($n = 10$), respectively, indicating significant T_3 -induced changes in the electrophysiology of the cells. In addition to common clinical experience with thyrotoxic patients, similar findings have been reported in animals in which thyroid hormone treatment was associated with a faster rate of beating in isolated rabbit atria and intact dog heart (34). These alterations in beating rate, however, are unlikely to invalidate our interpretation of the mechanism of T_3 -induced change in digitalis tolerance in myocardial cells because the peak positive inotropic effect of glycosides has been shown to be either independent of (35–37) or enhanced by increases in frequency of stimulation (38–39) over this range.

The results reported here support the proposed mechanism of action of cardiac glycosides (17, 20, 22, 40) in which inhibition of sodium pump activity is thought to be followed by elevation of intracellular sodium and consequent augmentation of cellular calcium content via sodium-calcium exchange. Our results appear to be inconsistent with the alternative proposal that binding of digitalis to NaK-ATPase causes the release of calcium from enzyme-associated sarcolemmal sites (41–44). A lesser rather than greater inotropic response to a given ouabain concentration was observed in cells with increased numbers of sodium pump sites, despite the fact that a greater absolute number of NaK-ATPase sites would be occupied because of the unchanged affinity of sites for ouabain (Fig. 5).

In an in vivo situation, changes in sensitivity to digitalis of cardiac tissue may be further modified by T_3 -induced alterations in the tissue responsiveness to catecholamines or insulin levels in the circulation, since both of the latter hormones are known to affect sodium pump activity. Therefore, additional studies are clearly required to elucidate the effects of interactions of hormones on digitalis sensitivity in vivo.

In conclusion, growth in the presence of T_3 caused a concentration-dependent elevation of the total number of functional sodium pump sites in the sarcolemmal membrane of cultured chick ventricular cells. The increase in sodium pump sites was associated with a decrease in cellular sodium content, and with reduced positive inotropic and toxic responses to ouabain. These changes appear to occur as a result of active extrusion of intracellular sodium by the additional sodium

pumps, and consequently, reduced calcium uptake via sodium-calcium exchange (16, 17, 27, 34). Altered tolerance to cardiac glycosides in clinical states of hypothyroidism or hyperthyroidism may be explained, at least in part, by this mechanism.

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