

## Research Highlight

# The Challenges of Studying the Anaerobic Microbial World

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Studies on strictly anaerobic microorganisms represent one of the most challenging areas of research, because anaerobic conditions (oxygen-free) need to be reconstructed to understand microbial activities and to obtain enrichments and pure culture. It is well-known that the micro/macro anaerobic environments are present everywhere on the Earth, and anaerobes comprise complex communities that play an important role in the carbon, nitrogen, and sulfur cycles on earth (49). Microbial community studies using 16S rRNA gene, as well as various functional genes, have offered new insights into anaerobic microbial ecology. Furthermore, numerous new lines of evidence offered by recent omics-driven and high-throughput sequencing studies provide a new vision of the anaerobic microbial world.

In the current issue of *Microbes and Environments*, Cheng *et al.* (4) reported that different types of sulfate-reducing prokaryotes that can be grown under specific temperatures ranges were detected in sulfate-amended enrichment cultures of muddy fluids taken from a Taiwanese terrestrial hydrocarbon seep, and that indigenous microbial communities might change based on the dynamic environmental fluctuations in volcanic mud ecosystems. Sulfate-reducing prokaryotes are frequently found in sulfate-supplied environment and are capable of growing on a variety of electron donors. In gas seeps and oil fields, the presence of various sulfate-reducing prokaryotes (9, 27, 30, 39) is associated with their potential to degrade anaerobic aromatic compounds and hydrocarbons. In fact, some sulfate-reducing bacteria are known to be decomposers of these compounds (*e.g.*, *Desulfobacula toluolica*, *Desulfogloeba alkanexedens*, *Desulfosarcina* sp.) (5, 11, 12, 48). In addition, a recent study reported that the hyperthermophilic sulfate-reducing archaeon *Archaeoglobus fulgidus* oxidizes long-chain *n*-alkanes (24). Together with these findings, sulfate-reducing bacteria are also known to be an important microbial group as syntrophic partners in anaerobic ecosystems. Consortia of anaerobic methanotrophic archaea and sulfate-reducing bacteria contribute to the global methane consumption in methane-seeps (41). In addition, hydrogen and sulfur-compounds are syntrophically utilized by sulfate-reducing bacteria, sulfur-oxidizing bacteria, fermenters and anoxygenic photosynthetic bacteria in hot springs and hydrothermal fields showing the complexity and importance of syntrophic associations between organisms (10, 25, 36, 38).

Methanogens play a key role in anaerobic ecosystems, and represent the most important member for the effective organic degradation and the recovery of methane as energy in anaerobic digesters treating various types of wastewater (3, 13, 34, 50). Due to their growth under very low redox conditions, their cultivation and physiological analyses require special laboratory techniques and apparatus (8, 22). Methanogens are phylogenetically widespread among the phylum *Euryarchaeota*, and the discovery of new lineages is ongoing. *Methanomassiliicoccus luminyensis* was isolated from human feces (7) and is the first methanogenic representative belonging to the class *Thermoplasmata* (14). The class originally consisted of acidophilic and aerobic archaea (42) and of uncultured lineages retrieved from hydrothermal fields (46), rice fields (23), and so on. Isolation of *Thermoplasmata*-related methanogens within the unexpected lineage suggests that methanogens are phylogenetically more diverse than previously thought, and holds the promise of the discovery of as-yet-unrecognized methanogens (6, 35). The genus *Methanocella* also represents a novel lineage of methanogens, formerly called "Rice Cluster I", and the only cultivated representative belonging to the order *Methanocellales* (44). Sakai *et al.* successfully isolated *Methanocella paludicola* using an elaborate enrichment method: low-hydrogen conditions were created by using *Syntrophobacter fumaroxidans* as a hydrogen-producing fermenter (43) so that methanogens that favor low concentrations of hydrogen was selectively enriched and isolated. This example makes it clear that inventive approaches to cultivation provide opportunities for success. On the other hand, it is also important to easily and efficiently create the conditions for cultivation of fastidious anaerobic microorganisms like methanogens. Carbonero *et al.* (2) reported that improving the culture medium made the colony formation of *Methanosaeta* species successful. Nakamura *et al.* (32) developed a simple technique for the cultivation of anaerobic microorganisms, by using a six-well plate and anaerobic gas-pack system. Subsequently, *Methanothermobacter tenebrarum* was successfully isolated using this technique (33). Clearly, increase in colony forming efficiency would facilitate not only isolation of yet-to-be cultured methanogens but further studies using genetic manipulations.

Anaerobic ammonium oxidation (anammox) is a microbial process in which ammonium is anaerobically oxidized to nitrogen gas with nitrite as an electron acceptor. Strous *et al.* first reported that this phenomenon occurs with anammox bacteria belonging to the order "*Brocadiales*" in the phylum *Planctomycetes* (45). As this process does not require a supply of oxygen or of organic substrates for stimulation of denitrification, it is expected to serve as an alternative

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to conventional processes used to remove nitrogen from ammonia-rich wastewater. In nature, anammox has been detected in marine and fresh-water sediments, soils, and so on (1, 52), and it is likely that anammox bacteria significantly contribute to the global nitrogen cycle (15, 16, 28). Based on enrichment studies (26, 37, 51), five candidate genera, “*Candidatus Anammoxoglobus*”, “*Candidatus Brocadia*”, “*Candidatus Jettenia*”, “*Candidatus Kuenenia*”, and “*Candidatus Scalindua*”, have been proposed (17), but none of the pure cultures have been so far isolated. Oshiki *et al.* (37) reported that two dominant enrichments, those of “*Candidatus Brocadia sinica*” and of “*Candidatus Scalindua sp.*”, were obtained by using membrane bioreactors. The fluorescence *in situ* hybridization study indicated that anammox bacteria dominated the biomass, as they accounted for more than 90% of its total biomass. Additional ecophysiological and biochemical studies using this dominated and stable enrichment are required to obtain pure anammox bacteria and to fully clarify the anammox process.

As the current issue of *Microbes and Environments* introduces the ecophysiological functions of anaerobes. For example, the intestinal colonization by *Lachnospiraceae* bacterial strain AJ11941 may contribute to the development of metabolic dysfunctions in obese mice (20). The gut environments may represent interesting anaerobic ecosystems to study in association with their hosts (40, 47). Cross-interactions between aerobic and anaerobic microorganisms are important factors with respect to organic matter degradation and material cycles of various types (19, 29, 31). Recent studies reported that anaerobic microorganisms use filaments (flagella and pili) for their communication and respiration, and that they are important functional parts than previously thought (18, 21). Interspecies electron transfer using conductive flagella or inorganic materials will become one of the most intriguing topics in microbial ecology.

## References

- Amano, T., I. Yoshinaga, T. Yamagishi, C.V. Thuoc, P.T. Thu, S. Ueda, K. Kato, Y. Sako, and Y. Suwa. 2011. Contribution of anammox bacteria to benthic nitrogen cycling in a mangrove forest and shrimp ponds, Hai Phong, Vietnam. *Microbes Environ.* 26:1–6.
- Carbonero, F., B.B. Oakley, and K.J. Purdy. 2010. Improving the isolation of anaerobes on solid media: the example of the fastidious *Methanosaeta*. *J. Microbiol. Methods* 80:203–205.
- Chen, C.-L., J.-H. Wu, I.C. Tseng, T.-M. Liang, and W.-T. Liu. 2009. Characterization of active microbes in a full-scale anaerobic fluidized bed reactor treating phenolic wastewater. *Microbes Environ.* 24:144–153.
- Cheng, T.-W., L.-H. Lin, Y.-T. Lin, S.-R. Song, and P.-L. Wang. 2014. Temperature-dependent variations in sulfate-reducing communities associated with a terrestrial hydrocarbon seep. *Microbes Environ.* 29:377–387.
- Davidova, I.A., K.E. Duncan, O.K. Choi, and J.M. Suflita. 2006. *Desulfoglaeba alkanexedens* gen. nov., sp. nov., an *n*-alkane-degrading, sulfate-reducing bacterium. *Int. J. Syst. Evol. Microbiol.* 56:2737–2742.
- Dewi Puspita, I., Y. Kamagata, M. Tanaka, K. Asano, and C.H. Nakatsu. 2012. Are uncultivated bacteria really uncultivable? *Microbes Environ.* 27:356–366.
- Dridi, B., M.-L. Fardeau, B. Ollivier, D. Raoult, and M. Drancourt. 2012. *Methanomassiliococcus luminyensis* gen. nov., sp. nov., a methanogenic archaeon isolated from human faeces. *Int. J. Syst. Evol. Microbiol.* 62:1902–1907.
- Enoki, M., N. Shinzato, H. Sato, K. Nakamura, and Y. Kamagata. 2011. Comparative proteomic analysis of *Methanothermobacter therautotrophicus* ΔH in pure culture and in co-culture with a butyrate-oxidizing bacterium. *PLoS ONE* 6:e24309.
- Gieg, L., T. Jack, and J. Foght. 2011. Biological souring and mitigation in oil reservoirs. *Appl. Microbiol. Biotechnol.* 92:263–282.
- Hamamura, N., J. Meneghin, and A.-L. Reysenbach. 2013. Comparative community gene expression analysis of *Aquificales*-dominated geothermal springs. *Environ. Microbiol.* 15:1226–1237.
- Harms, G., K. Zengler, R. Rabus, F. Aeckersberg, D. Minz, R. Rosselló-Mora, and F. Widdel. 1999. Anaerobic oxidation of *o*-xylene, *m*-xylene, and homologous alkylbenzenes by new types of sulfate-reducing bacteria. *Appl. Environ. Microbiol.* 65:999–1004.
- Higashioka, Y., H. Kojima, and M. Fukui. 2012. Isolation and characterization of novel sulfate-reducing bacterium capable of anaerobic degradation of *p*-xylene. *Microbes Environ.* 27:273–277.
- Iguchi, A., T. Terada, T. Narihiro, T. Yamaguchi, Y. Kamagata, and Y. Sekiguchi. 2009. *In situ* detection and quantification of uncultured members of the phylum *Nitrospirae* abundant in methanogenic wastewater treatment systems. *Microbes Environ.* 24:97–104.
- Iino, T., H. Tamaki, S. Tamazawa, Y. Ueno, M. Ohkuma, K.-i. Suzuki, Y. Igarashi, and S. Haruta. 2013. *Candidatus Methanogramma caenicola*: a novel methanogen from the anaerobic digested sludge, and proposal of *Methanomassiliococcaceae* fam. nov. and *Methanomassiliococcales* ord. nov., for a methanogenic lineage of the class *Thermoplasmata*. *Microbes Environ.* 28:244–250.
- Ishii, S., S. Ikeda, K. Minamisawa, and K. Senoo. 2011. Nitrogen cycling in rice paddy environments: past achievements and future challenges. *Microbes Environ.* 26:282–292.
- Isobe, K., and N. Ohte. 2014. Ecological perspectives on microbes involved in N-cycling. *Microbes Environ.* 29:4–16.
- Jetten, M.S.M., L.v. Niftrik, M. Strous, B. Kartal, J.T. Keltjens, and H.J.M. Op den Camp. 2009. Biochemistry and molecular biology of anammox bacteria. *Crit. Rev. Biochem. Mol. Biol.* 44:65–84.
- Jiang, S., and H.-G. Hur. 2013. Effects of the anaerobic respiration of *Shewanella oneidensis* MR-1 on the stability of extracellular U(VI) nanofibers. *Microbes Environ.* 28:312–315.
- Kaiya, S., S. Utsunomiya, S. Suzuki, N. Yoshida, H. Futamata, T. Yamada, and A. Hiraishi. 2012. Isolation and functional gene analyses of aromatic-hydrocarbon-degrading bacteria from a polychlorinated-dioxin-dechlorinating process. *Microbes Environ.* 27:127–135.
- Kameyama, K., and K. Itoh. 2014. Intestinal colonization by a *Lachnospiraceae* bacterium contributes to the development of diabetes in obese mice. *Microbes Environ.* 29:427–430.
- Kato, S., and K. Watanabe. 2010. Ecological and evolutionary interactions in syntrophic methanogenic consortia. *Microbes Environ.* 25:145–151.
- Kato, S., K. Sasaki, K. Watanabe, I. Yumoto, and Y. Kamagata. 2014. Physiological and transcriptomic analyses of the thermophilic, acetate-oxidizing methanogen *Methanosaeta thermophila* responding to ammonia stress. *Microbes Environ.* 29:162–167.
- Kemnitz, D., S. Kolb, and R. Conrad. 2005. Phenotypic characterization of Rice Cluster III archaea without prior isolation by applying quantitative polymerase chain reaction to an enrichment culture. *Environ. Microbiol.* 7:553–565.
- Khelifi, N., O. Amin Ali, P. Roche, V. Grossi, C. Brochier-Armanet, O. Valette, B. Ollivier, A. Dolla, and A. Hirschler-Rea. 2014. Anaerobic oxidation of long-chain *n*-alkanes by the hyperthermophilic sulfate-reducing archaeon, *Archaeoglobus fulgidus*. *ISME J* 8:2153–2166.
- Kimura, H., K. Mori, H. Nashimoto, S. Hanada, and K. Kato. 2010. *In situ* biomass production of a hot spring sulfur-turf microbial mat. *Microbes Environ.* 25:140–143.
- Kindaichi, T., T. Awata, Y. Suzuki, K. Tanabe, M. Hatamoto, N. Ozaki, and A. Ohashi. 2011. Enrichment using an up-flow column reactor and community structure of marine anammox bacteria from coastal sediment. *Microbes Environ.* 26:67–73.
- Kleindienst, S., F.-A. Herbst, M. Stagars, *et al.* 2014. Diverse sulfate-reducing bacteria of the *Desulfosarcina/Desulfococcus* clade are the key alkane degraders at marine seeps. *ISME J* 8:2029–2044.
- Kubota, K. 2013. CARD-FISH for environmental microorganisms: technical advancement and future applications. *Microbes Environ.* 28:3–12.

29. Lv, X.-M., M.-F. Shao, C.-L. Li, J. Li, X.-L. Gao, and F.-Y. Sun. 2014. A comparative study of the bacterial community in denitrifying and traditional enhanced biological phosphorus removal processes. *Microbes Environ.* 29:261–268.
30. Magot, M., B. Ollivier, and B.C. Patel. 2000. Microbiology of petroleum reservoirs. *Antonie Van Leeuwenhoek* 77:103–116.
31. Makhdoumi-Kakhki, A., M.A. Amoozegar, B. Kazemi, L. Pašić, and A. Ventosa. 2012. Prokaryotic diversity in Aran-Bidgol salt lake, the largest hypersaline playa in Iran. *Microbes Environ.* 27:87–93.
32. Nakamura, K., H. Tamaki, M.S. Kang, H. Mochimaru, S.-T. Lee, K. Nakamura, and Y. Kamagata. 2011. A six-well plate method: less laborious and effective method for cultivation of obligate anaerobic microorganisms. *Microbes Environ.* 26:301–306.
33. Nakamura, K., A. Takahashi, C. Mori, H. Tamaki, H. Mochimaru, K. Nakamura, K. Takamizawa, and Y. Kamagata. 2013. *Methanothermobacter tenebrarum* sp. nov., a hydrogenotrophic, thermophilic methanogen isolated from gas-associated formation water of a natural gas field. *Int. J. Syst. Evol. Microbiol.* 63:715–722.
34. Narihiro, T., T. Terada, K. Kikuchi, *et al.* 2009. Comparative analysis of bacterial and archaeal communities in methanogenic sludge granules from upflow anaerobic sludge blanket reactors treating various food-processing, high-strength organic wastewaters. *Microbes Environ.* 24:88–96.
35. Narihiro, T., and Y. Kamagata. 2013. Cultivating yet-to-be cultivated microbes: the challenge continues. *Microbes Environ.* 28:163–165.
36. Nunoura, T., M. Hirai, M. Miyazaki, *et al.* 2013. Isolation and characterization of a thermophilic, obligately anaerobic and heterotrophic marine *Chloroflexi* bacterium from a *Chloroflexi*-dominated microbial community associated with a Japanese shallow hydrothermal system, and proposal for *Thermomarinilinea lacunofontalis* gen. nov., sp. nov. *Microbes Environ.* 28:228–235.
37. Oshiki, M., T. Awata, T. Kindaichi, H. Satoh, and S. Okabe. 2013. Cultivation of planktonic anaerobic ammonium oxidation (anammox) bacteria using membrane bioreactor. *Microbes Environ.* 28:436–443.
38. Otaki, H., R.C. Everroad, K. Matsuura, and S. Haruta. 2012. Production and consumption of hydrogen in hot spring microbial mats dominated by a filamentous anoxygenic photosynthetic bacterium. *Microbes Environ.* 27:293–299.
39. Priha, O., M. Nyssönen, M. Bomberg, A. Laitila, J. Simell, A. Kapanen, and R. Juvonen. 2013. Application of denaturing high-performance liquid chromatography for monitoring sulfate-reducing bacteria in oil fields. *Appl. Environ. Microbiol.* 79:5186–5196.
40. Qi, W., C.-L. Chen, and J.-Y. Wang. 2011. Reducing sugar-producing bacteria from guts of *Tenebrio Molitor Linnaeus* (yellow mealworm) for lignocellulosic waste minimization. *Microbes Environ.* 26:354–359.
41. Raghoebarsing, A.A., A. Pol, K.T. Van de Pas-Schoonen, *et al.* 2006. A microbial consortium couples anaerobic methane oxidation to denitrification. *Nature* 440:918–921.
42. Rekysenbach, A.-L. 2001. Class IV. *Thermoplasmata* class nov., p. 335–340. In D.R. Boone, R.W. Castenholtz, and G.M. Garrity (ed.), *Bergey's Manual of Systematic Bacteriology*, 2nd edn, vol. 1. Springer, New York.
43. Sakai, S., H. Imachi, Y. Sekiguchi, A. Ohashi, H. Harada, and Y. Kamagata. 2007. Isolation of key methanogens for global methane emission from rice paddy fields: a novel isolate affiliated with the clone cluster rice cluster I. *Appl. Environ. Microbiol.* 73:4326–4331.
44. Sakai, S., H. Imachi, S. Hanada, A. Ohashi, H. Harada, and Y. Kamagata. 2008. *Methanocella paludicola* gen. nov., sp. nov., a methane-producing archaeon, the first isolate of the lineage 'Rice Cluster I', and proposal of the new archaeal order *Methanocellales* ord. nov. *Int. J. Syst. Evol. Microbiol.* 58:929–936.
45. Strous, M., J.A. Fuerst, E.H. Kramer, S. Logemann, G. Muyzer, K.T. van de Pas-Schoonen, R. Webb, J.G. Kuenen, and M.S. Jetten. 1999. Missing lithotroph identified as new planctomycete. *Nature* 400:446–449.
46. Takai, K., and K. Horikoshi. 1999. Genetic diversity of archaea in deep-sea hydrothermal vent environments. *Genetics* 152:1285–1297.
47. Thong-On, A., K. Suzuki, S. Noda, J.-i. Inoue, S. Kajiwara, and M. Ohkuma. 2012. Isolation and characterization of anaerobic bacteria for symbiotic recycling of uric acid nitrogen in the gut of various termites. *Microbes Environ.* 27:186–192.
48. Wöhlbrand, L., J.H. Jacob, M. Kube, *et al.* 2013. Complete genome, catabolic sub-proteomes and key-metabolites of *Desulfobacula toluolica* Tol2, a marine, aromatic compound-degrading, sulfate-reducing bacterium. *Environ. Microbiol.* 15:1334–1355.
49. Whitman, W.B., D.C. Coleman, and W.J. Wiebe. 1998. Prokaryotes: the unseen majority. *Proc. Natl. Acad. Sci. U.S.A.* 95:6578–6583.
50. Yamada, C., S. Kato, Y. Ueno, M. Ishii, and Y. Igarashi. 2014. Inhibitory effects of ferrihydrite on a thermophilic methanogenic community. *Microbes Environ.* 29:227–230.
51. Yasuda, T., M. Waki, I. Yoshinaga, T. Amano, K. Suzuki, Y. Tanaka, T. Yamagishi, and Y. Suwa. 2011. Evidence of exponential growth of an anammox population in an anaerobic batch culture. *Microbes Environ.* 26:266–269.
52. Yoshinaga, I., T. Amano, T. Yamagishi, K. Okada, S. Ueda, Y. Sako, and Y. Suwa. 2011. Distribution and diversity of anaerobic ammonium oxidation (anammox) bacteria in the sediment of a eutrophic freshwater lake, Lake Kitaura, Japan. *Microbes Environ.* 26:189–197.