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# The Genetic Population Structure of Wild Western Lowland Gorillas (*Gorilla gorilla gorilla*) Living in Continuous Rain Forest

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# Abstract

To understand the evolutionary histories and conservation potential of wild animal species it is useful to assess whether taxa are genetically structured into different populations and identify the underlying factors responsible for any clustering. Landscape features such as rivers may influence genetic population structure, and analysis of structure by sex can further reveal effects of sex-specific dispersal. Using microsatellite genotypes obtained from noninvasively collected fecal samples we investigated the population structure of 261 western lowland gorillas (WLGs) (*Gorilla gorilla gorilla*) from seven locations spanning an approximately 37,000km<sup>2</sup> region of mainly continuous rain forest within Central African Republic (CAR), Republic of Congo and Cameroon. We found our sample to consist of two or three significantly differentiated clusters. The boundaries of the clusters coincided with courses of major rivers. Moreover, geographic distance detoured around rivers better-explained variation in genetic distance than straight line distance. Together these results suggest that major rivers in our study area play an important role in directing WLG gene flow. The number of clusters did not change when males and females were

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analyzed separately, indicating a lack of greater philopatry in WLG females than males at this scale.

#### Keywords

Gorilla gorilla; population structure; sex-specific dispersal; philopatry

## INTRODUCTION

Although many researchers have investigated the pattern of genetic variation in organisms living in multiple disjunct populations or in fragmented habitats [e.g., Haag et al., 2010; Martínez-Cruz et al., 2007; Ruell et al., 2012], relatively few studies have investigated the structure exhibited by animal populations living in mainly continuous, apparently uniform habitat [Neaves et al., 2009; Piertney et al., 1998; reviewed in Storfer et al., 2010]. Even in areas of continuous habitat, rivers may hinder the movement of terrestrial animals [Storfer et al., 2010], thus potentially inhibiting gene flow and affecting patterns of genetic differentiation. However, because water levels vary by season and river courses can change over time, it can be difficult to assess whether a river serves as complete barrier to gene flow or acts as an occasionally permeable hindrance. The effects of rivers on the genetic structure of large mammal populations have accordingly varied. For example, indications that rivers reduce gene flow were found in white-tailed deer (Odocoileus virginianus) [Blanchong et al., 2008] and lowland tapir (Tapirus terrestris) [De Thoisy et al., 2010], whereas in bobcats (Lynx rufus) the large Mississippi River was not significantly associated with genetic differentiation [Reding et al., 2012]. In red deer (Cervus elaphus) from the Scottish Highlands, rivers were even suggested to facilitate gene flow [Pérez-Espona et al., 2008]. Studies of great apes found that in chimpanzees (Pan troglodytes) [Gonder et al., 2006] and Bornean orangutans (Pongo pygmaeus) [Arora et al., 2010; Jalil et al., 2008], rivers apparently act as barriers to individual movement and thereby increase genetic differentiation between subpopulations on either side of the river. No barrier effect of rivers was detected in other smaller-bodied, more arboreal primates like spider monkeys (Ateles) [Collins & Dubach, 2000], long-tailed macaques (Macaca fascicularis) [Ruiter & Geffen, 1998] and a number of lemur species (Lemuriformes) [Craul et al., 2008].

In addition to rivers, postglacial expansions from forest refugia that became isolated during the Pleistocene [Haffer, 1969] may be expected to influence the distribution of present-day genetic variation in geographically continuous populations of forest dwelling species. For example, although forest elephants (*Loxodonta cyclotis*) were found to be only weakly structured [Roca et al., 2001], it was suggested they were restricted to isolated patches during the Pleistocene [Brandt et al., 2012]. While an early study in bonobos (*Pan paniscus*) attributed patterns of genetic variation to the effect of river barriers on migration [Eriksson et al., 2004], a recent study by Kawamoto et al. [2013] suggested that present day rivers have only a weak effect on gene flow among bonobo populations and that their current population genetic structure was shaped by Pleistocene forest refugia [Haffer, 1969; Hewitt, 1996]. For gorillas (*Gorilla*) it has been suggested that both Pleistocene refugia and rivers played a role in structuring genetic diversity [Anthony et al., 2007]. This study found signs

of demographic expansion consistent with postglacial expansion from formerly isolated refugia as well as admixture along the borders of mtDNA haplogroups. Nonetheless, some rivers acted as barriers between different haplogroups of gorillas.

If population genetic structure is found to be mainly caused by rivers or other hindrances to movement then these barriers will have a greater impact on the dispersing sex. Dispersal is male-biased in most mammals [Greenwood, 1980; Lawson Handley & Perrin, 2007]. However, there are also male-philopatric species (e.g., chimpanzee, bonobo) and species in which both sexes disperse (e.g., African wild dog (*Lycaon pictus*), gray wolf (*Canis lupus*)). Differences in dispersal distance are typical for species in which both sexes disperse [reviewed in Mabry et al., 2013]. If the rate of dispersal is similar, then the sex that travels further is less likely to be structured into differentiated clusters.

Gorillas are among the relatively few mammal species for which both sexes usually disperse from their natal groups [Stoinski et al., 2009; Stokes et al., 2003]. The typical reproductive unit in western lowland gorillas (WLGs) (*Gorilla gorilla gorilla*) is composed of a single adult silverback male accompanied by several females [Breuer et al., 2010; Gatti et al., 2004; Parnell, 2002; Robbins et al., 2004]. Adult females transfer directly between breeding groups during inter-group encounters and do not range on their own [Stokes et al., 2003]. Males, in contrast, leave their natal groups after maturation and range several years as solitaries before eventually obtaining a harem [Breuer et al., 2010]. Bradley et al. [2004] found evidence that WLG silverbacks leading social groups were often related to nearby males, suggesting low-distance dispersal of many males. In contrast, results of a recent study by Inoue et al. [2013] suggested that closely related WLG males do not stay close to each other after maturation arguing for relatively long dispersal distances. Thus, dispersal patterns of male WLGs may vary between different sites, as the former study was in the Central African Republic and the latter was in Gabon.

Research on WLGs as well as on mountain gorillas (*Gorilla beringei beringei*) suggested greater population genetic structure in females as compared to males, which was attributed to a tendency of females to disperse shorter distances than males [Douadi et al., 2007; Guschanski et al., 2008]. However, these studies were conducted over limited spatial scales (WLGs: 70km×85 km; mountain gorillas: 28km×18 km). In addition, the study of WLGs relied on sex-specific markers, namely mitochondrial DNA and the Y-chromosome. Although rapidly evolving and hence likely to reveal any underlying genetic structure, it is difficult to directly compare results from these female-and male-transmitted systems. This is because the Y-chromosome is likely to exhibit lower variation than mtDNA [Burrows & Ryder, 1997; Hellborg, 2003] and differences in genetic differentiation estimates between mtDNA and the non-recombining portion of the Y-chromosome cannot be tested for significance. In addition, differences in reproductive skew between the sexes will alter the relative effective population sizes of males and females. Thus it is difficult to make inferences about dispersal rate from the extent of genetic differentiation observed from sex-specific markers [Lawson Handley & Perrin, 2007].

For the present study WLG samples from key locations across a large expanse of their range were collected. Genetic characterization of those samples may effectively address questions

on (i) population structure including the effects of rivers and (ii) the relative roles of male and female dispersal in shaping genetic structure. Samples were collected in a relatively large, roughly 160km×230km region consisting of continuous rainforest with several notable rivers and few roads (Fig. 1).

# **METHODS**

#### **Study Area and Sample Collection**

Fecal samples from wild WLGs were obtained from three areas within Central Africa (Fig. 1): (i) two sampling sites east of Odzala NP, Republic of Congo; (ii) three sampling sites within and west of Lobéké NP, Cameroon; and (iii) various locations (southern Ndoki and Bai Hokou) within and close to Nouabalé-Ndoki NP, Republic of Congo and Dzanga Ndoki NP, Central African Republic. The latter two national parks are adjacent and only separated by the national border with no physical barrier between them. We will summarize that area as "Ndoki forest". The Ngoko River may serve as natural barrier between Odzala and Lobéké, and the Sangha River separates the Ndoki forest from Lobéké and Odzala. The whole region of approximately 37,000km<sup>2</sup> is covered by continuous rainforest without notable elevation changes and has few public roads. One road runs between the two sampling sites east of Odzala NP, another is located between Odzala NP and the two other major areas. Both are regularly crossed by WLGs (F. Maisels, personal communication). Samples were collected noninvasively between 2003 and 2012 using Nsubuga et al.' s [2004] two-step ethanol-silica storage method (samples from Odzala and the Ndoki forest) or were stored in RNAlater (samples from Lobéké).

#### **DNA Extraction, Quantification and Amplification**

The QIAamp Stool kit (QIAGEN) was used to extract DNA from fecal samples. To estimate the amount of amplifiable DNA in the extracts we used a real-time quantitative PCR with Maxima SYBR<sup>®</sup> Green (Thermo Scientific, Waltham, MA, USA) as Master Mix and c-myc\_E3\_F1U1 plus c-myc\_E3\_R1U as primers [Morin et al., 2001] to amplify a portion of the c-myc gene. Standard curves were constructed from serially diluted human genomic DNA (BIO-35025, Bioline, London, UK). Initially, three independent amplifications from each extract were performed at 12 microsatellite loci (D10s1432, D8s1106, D6s474, D1s2130, D5s1470, D4s1627, D7s817, D1s550, D16s2624, D6s1056, D2s1326, D3s2459), using a two-step multiplex PCR [Arandjelovic et al., 2009]. For sex determination we also amplified a segment of the X-Y homologous amelogenin gene in a one-step PCR [Bradley et al., 2001].

The ABI PRISM 3100 Genetic Analyzer was used for electrophoresis of the PCR products. The sizes of the alleles relative to an internal size standard were determined using GeneMapper Software version 3.7 (Applied Biosystems, Foster City, CA, USA). We calculated allelic dropout rates to assess how many observations from independent reactions are necessary to confirm homozygosity [Morin et al., 2001]. We found that three observations from independent reactions are sufficient to confirm homozygous genotypes with 99% certainty even in the extracts with the lowest DNA concentrations. Heterozygote genotypes were confirmed with 99% certainty by observing each allele two or more times

[Taberlet et al., 1996]. For each extract up to six independent reactions were analyzed per locus.

#### Discrimination of Individuals and Distinguishing Gorilla and Chimpanzee Samples

We used CERVUS 3.0 [Kalinowski et al., 2007] to calculate  $P_{\text{IDsib}}$  [Waits et al., 2001], the probability that samples with matching genotypes come from siblings rather than from the same individual. Matching genotypes were combined into a consensus genotype when the  $P_{\text{IDsib}}$  was <0.01. In the rare cases where two genotypes matched but the  $P_{\text{IDsib}}$  was >0.01 the less complete genotype was removed from further analyses. That approach resulted in a dataset in which the least completely genotyped individuals were typed at a minimum of four loci, because genotypes consisting of fewer loci were in no case informative enough to be found unique.

In the field, fecal samples coming from chimpanzees are occasionally mistakenly identified as gorilla feces, and vice versa. We therefore checked for incorrectly identified samples by genotyping 10 samples of known chimpanzee origin at the 12 microsatellite loci typically analyzed in gorillas (see above) and performing STRUCTURE analysis of them in combination with the purported gorilla samples as in Arandjelovic et al. [2010]. After attributing samples to the correct species, we added those samples to our dataset that were genetically identified as WLGs and removed samples that were genetically identified as chimpanzees.

#### Data Analysis

Using SPAGeDi 1.3 [Hardy & Vekemans, 2002] we tested via permutations (equivalent to carrying out a Mantel test) for a significant correlation between genetic distance and two different measures of geographic distance: (i) straight line Euclidean distance between sampling sites (also called isolation by distance, IBD) and (ii) detoured distance from one sampling site to another around the headwaters of the Sangha and the Ngoko rivers. We chose seven geographically distinct sampling sites (Fig. 1): Odzala- South (Libonga), Odzala-North (Liouesso), Lobéké-West, Lobéké-Central, Lobéké-East, Bai Hokou, and southern Ndoki. We plotted for each of the resulting 21 pairs the natural logarithm of the geographic distance in meters against the genetic differentiation measured as  $F_{\rm ST}/(1-F_{\rm ST})$  and plotted a regression line. The slope of this line was considered significantly positive if the slope was >0 and the probability in 20,000 permutations of obtaining by chance a slope larger than the observed one is less than 5%.

To assess which measure of geographic distance better explains variation in genetic distance we ran general linear models in R (R Development Core Team 2012) for calculations of the Akaike Information Criterion (AIC). The response variable for each model was genetic distance and the predictor variables were straight distance and detoured distance. The intercept was excluded from the models. The predictor variable of the model with the smaller AIC better explains the variation in the response variable. AIC is the difference of the AIC values of the two models and the larger it is the less empirical support the model with the higher AIC receives. Due to the non-independence of the data points we ran 5,040 permutations of the genetic distances (total number of possible permutations) and declared

Most programs that output summary statistics require a priori delineation of populations which is a conflicting issue if the main goal of the study is to find the previously unknown number of clusters within the area of interest. However, software like Geneland 4.0.3 [Guillot, 2005, 2008; Guillot et al., 2005, 2008] and STRUCTURE 2.2 [Falush et al., 2003, 2007; Pritchard et al., 2000] are based on Bayesian clustering algorithms for inferring genetic structure and do not need a priori assignment of populations. STRUCTURE employs a nonspatial cluster algorithm that assumes an independent identically distributed (iid) mixture model. The iid mixture model puts equal prior weights on all partitions, whereas the hidden model of spatial organization through Voronoi tessellation implemented in Geneland favors partitions that are spatially organized. This leads to better classification with limited data due to a more informative prior [Guillot, 2005]. We therefore first investigated our data from the whole study area with Geneland to assess the most likely number of underlying clusters for each of the following different data sets: (i) all individuals; (ii) only females; and (iii) only males. To account for differences in sample size we conducted the male-specific analysis with the same number of samples as the female-specific analysis (N=89). For each dataset we conducted five independent runs using the following parameters: maximum rate of Poisson process: 100; uncertainty on coordinate: 5; minimum and maximum number of populations: 1 and 8, respectively; maximum number of nuclei: 300; number of iterations: 100,000; thinning: 100; allele frequency model: correlated; with spatial model; without null allele model. From the five runs we always present the results of the one with the highest mean posterior density.

For comparison with results generated by Geneland we additionally analyzed our data from the whole study area in STRUCTURE and BAPS 5.3 [Corander et al., 2007; Corander &Marttinen, 2006]. As with Geneland, BAPS also makes use of the geographic origin of the genotypes. However, inferences in BAPS are based on another model and different computational techniques [Corander et al., 2007]. BAPS assumes that all individuals have unique coordinates. Therefore we added random noise of up to 100m to each UTM\_X and each UTM Y coordinate that was not unique. We ran the spatial clustering of individuals with 10 replicates for each number of clusters K (K=3, 5, 10, and 20, with the numbers specifying the upper limit of expected clusters). In STRUCTURE we varied K from 1 to 8 clusters and conducted 10 independent iterations at each K using the admixture model, correlated allele frequencies, a burn-in period of 100,000 steps and then 500,000 steps of data collection. Subsequently we used Harvester Web v0.6.93 [Earl & vonHoldt, 2011] to calculate K [Evanno et al., 2005], a measure of second-order rate of change in the likelihood of K which is widely used to determine the optimal value of K. For that calculation we removed runs with extraordinarily high variance of Ln likelihood compared to other runs within the same K.

It is important to check that the inferred groups are significantly differentiated from each other and in Hardy–Weinberg equilibrium (HWE). Geneland further assumes that marker loci are at linkage equilibrium within populations. Using CERVUS 3.0 [Kalinowski et al., 2007] we tested for the existence of null alleles within each cluster inferred by the Geneland

analysis of all individuals. GENEPOP 4.2 [Raymond & Rousset, 1995] was used to perform the exact tests of Guo and Thompson [1992] for deviations from HWE as well as the test of composite linkage disequilibrium [Weir, 1996] within each cluster. Once clusters were identified, pairwise  $F_{ST}$  estimates of population differentiation with associated *P*-values (using 9999 permutations) were obtained using GenAlEx 6.5 [Peakall & Smouse, 2006, 2012].

Because our study did not involve animal testing and the samples were collected noninvasively we did not violate any regulations of the Deutsches Tier-schutzgesetz and the US Public Health Service Policy on Humane Care and Use of Laboratory Animals. The study complied with ASP principles for the ethical treatment of non-human primates. All research was conducted in accordance with the laws of Republic of Congo, Cameroon, Central African Republic, and Germany.

# RESULTS

#### Genotype Analysis

We genotyped 411 putative WLG fecal samples at 12 autosomal microsatellite loci. These samples yielded 339 usable genotypes representing 272 unique individuals. An analysis of these genotypes using STRUCTURE in comparison with known chimpanzee DNAs typed at the same loci, showed that 12 of the newly generated genotypes were actually from chimpanzees, leaving a total of 260 different WLGs. Field researchers thus correctly identified WLG dung in 327/339=96% of cases. In a reciprocal analysis of genotypes purportedly from 134 chimpanzees and typed as part of a complementary study, we found three samples that derived from WLGs. These three samples were secondarily genotyped at microsatellite loci used here. One sample failed to produce a usable genotype, one sample was identical to an already existing genotype and one represented an additional individual. The final dataset therefore contained 261 different WLG genotypes, which were on average 84% complete. Two hundred and twenty-six out of these 261 individuals (=86.6%) had complete genotypes at more than eight loci and only 18/261 (=6.9%) had complete genotypes at six or fewer loci. The 261 individuals consisted of 89 females (F) and 168 males (M) while the sex could not be determined for four individuals. Forty-two individuals were from Odzala (20F, 21M, 1 unknown sex), 24 from Lobéké (8F, 16M) and 195 (61F, 131M, 3 unknown sex) from the Ndoki forest.

The average DNA concentration of the 298 quantified extracts was 247.3 pg/µl (SD: 548.8 pg/µl; range: 3.6–5682.5 pg/µl). Considering results from individual PCRs, allelic dropout rate was 0.06 over all loci and all samples and 0.22 over all loci for extracts with the lowest DNA amounts (5 pg/µl). Three independent observations of homozygosity were therefore sufficient to confirm homozygous genotypes with 99% certainty (0.22<sup>3</sup>=0.01) even in samples with very low DNA concentrations.

Using the clusters defined by Geneland (see below), we found null allele frequency estimates >0.05 at one locus within the Ndoki forest cluster, at three loci within the Lobéké cluster and at one locus within the Odzala cluster. However, it is possible that null allele frequency is overestimated when sample size is small, which is the case for the Lobéké and

the Odzala cluster (24 and 42 samples, respectively). We tested for HWE both before and after removal of the above mentioned loci from the respective clusters. In no case did any of the three clusters deviate significantly from HWE (P>0.05). We found no indication for linkage disequilibrium (P>0.05 after Bonferroni correction for multiple tests) between any pair of loci within any of the inferred clusters.

#### **Geographic Distribution of Genetic Variation**

Pairwise comparisons between the seven sampling sites showed that genetic variation increased both with straight line geographic distance and detoured distance (Fig. 2). However, the correlation coefficient of the regression line was smaller in the straight distance (r=0.60) than in the detoured distance plot (r=0.73). Additionally, the GLM with detoured distance as predictor variable was found to better explain variation in genetic distance (AIC=-83.6) than the GLM with straight distance as predictor variable (AIC=-81.7). The difference in AIC was highly significant (P=0.0032).

In an analysis of all genotypes, the Geneland run with the highest mean posterior density (-6399.11) depicted three clusters. All Ndoki forest samples were grouped into the first, all Lobéké samples into the second and all Odzala samples into the third cluster (Fig. 3). The boundaries of the inferred clusters roughly correspond to naturally occurring barriers: the Ngoko River between Odzala and Lobéké and the Sangha River between Ndoki forest and the two other sites. Considering each of the clusters as populations, we found significant pairwise  $F_{\text{ST}}$  estimates between all three localities with the highest differentiation between Odzala and Ndoki forest, an intermediate level between Lobéké and Ndoki forest and the lowest between Odzala and Lobéké (Table I).

In contrast to the results from Geneland, the STRUCTURE analysis inferred two clusters of individuals. Nearly all individuals (244/261) had 75% ancestry from one population. One cluster contained individuals from Odzala and Lobéké and the other contained the individuals from the Ndoki forest. However, there were also three individuals (one female, two males) that were sampled in Lobéké, but assigned to the Ndoki forest cluster and three others (two females, one male) that were sampled in the Ndoki forest but assigned to the Odzala/Lobéké cluster (Fig. 4).

Analysis with BAPS also grouped individuals into two clusters, which can be seen on a map of Voronoi tessellation in which each tile represents one individual (Fig. 5). Of the 244 genotypes that could be allocated to a single cluster by STRUCTURE, BAPS assigned 243 genotypes into the same clusters (99.6% agreement). Of the six individuals assigned by STRUCTURE to a cluster other than the one where they were sampled, five (three females, two males) were also flagged by BAPS.

We also conducted genetic cluster analysis using just male or just female genotypes. Using Geneland, analysis of only females produced three clusters in the run with the highest mean posterior density (-2026.85). However, in that analysis the two females from Lobéké-Central formed one cluster together with the Odzala samples. The second cluster contained all remaining Lobéké females and the third all individuals from the Ndoki forest. A Geneland analysis of only males also showed three clusters in the run with the highest mean

posterior density (-1649.14). The three clusters were identical with the ones that were found by Geneland when all genotypes were included: the Ndoki forest males formed the first, the Lobéké males the second and the Odzala males the third cluster.

## DISCUSSION

#### **Effect of Rivers on Population Structure**

The area under study consists of continuous rain forest and swamps with high uniformity and low fragmentation through human impact. Nevertheless, we found that our WLG sample could be divided into two (STRUCTURE and BAPS) or three (Geneland) different clusters. Apart from very few exceptions, individuals from the Ndoki forest formed one and the remaining individuals the other subpopulation when two clusters were found. That pattern could be explained by the Sangha River which runs between the Ndoki forest and the other areas and might form a substantial although not absolute impediment for migration. The Sangha is around 1,300km long and up to hundreds of meters wide [Stauch, 1963], potentially forming a considerable natural barrier. Although elephants and duikers have been observed crossing the Sangha, WLGs avoid it and do not swim [A. Todd, personal communication]. As all analyses grouped the Ndoki forest into a distinct cluster the Sangha can be considered the greatest obstacle in the studied area. However, using simulations it has been argued that Geneland is more successful in detecting barriers than STRUCTURE and other similar software [Blair et al., 2012]. Analyzing the data with Geneland indeed revealed three instead of two clusters (Odzala, Lobéké and the Ndoki forest). Lobéké and Odzala are separated from each other by the Ngoko River which hence might also function as a barrier to gene flow, albeit probably to a lesser extent than the Sangha. The lesser effect of the Ngoko is also in agreement with its smaller dimensions of approximately 700km length and 120m width [Balarin, 1985]. The varied results obtained using different software highlight the importance of not relying on the output of a single program. Only the comparison of results from Geneland with results from STRUCTURE and BAPS gave us a comprehensive view: If we would have used only Geneland we would not have been able to infer that the Sangha River is a larger barrier to gene flow than the Ngoko. Conversely, using only STRUCTURE or BAPS would have resulted in not recognizing the effect of the Ngoko River at all.

Information about whether a population is continuous or not is also of key importance for conservation. In an action plan for the conservation of chimpanzees and gorillas in western equatorial Africa Tutin et al. [2005] defined seven areas of exceptional priority. One of them was the Sangha Trinational Complex comprising the Lobéké NP and the Ndoki forest and another one a complex including Odzala NP. As we found less evidence for a clustering between Lobéké and Odzala than between Lobéké and the Ndoki forest it might be more reasonable to group Lobéké and Odzala into one area of exceptional priority and the Ndoki forest into another. However, our results indicate that some individuals also migrated across the Sangha. Therefore one should generally be careful in defining distinct conservation units within our study site.

Might there be additional causes for the population structure we observe besides the occurrence of riverine barriers? The Pleistocene refuge theory [Haffer, 1969] posits that

present-day continuous rain forests have repeatedly been fragmented due to the cold and arid climate occurring during glacial maxima. Fragmentation might have led to isolation of populations of taxa that are restricted to forests. As western gorillas (*Gorilla gorilla*) are one of such taxon and do not occur in savannah habitats [Tutin et al., 1997], their widespread present day occurrence could be the consequence of recolonization during periods of warmer climate. Examining western gorilla populations across their entire range, Anthony et al. [2007] found highly different mtDNA haplogroups with the boundaries of these haplogroups only rarely occurring along the courses of major rivers. However, both the Sangha and the Ngoko Rivers demarcated different haplogroups, consistent with these rivers acting as barriers to gene flow. They concluded that both rivers and the occurrence of Pleistocene refugia shaped present-day genetic variation in WLGs.

It is difficult to know the exact locations of putative Pleistocene refugia in west central Africa. Although we cannot rule out that the patterns we observe are partially due to sampling of populations corresponding to different Pleistocene refugia we suggest that our results strongly support the occurrence of a barrier effect on gene flow by major rivers in our study area. This is consistent not only with the results from the clustering analyses, but also with the correlations of genetic distance with geographic distances. We found significant increase of genetic distance both when correlated with straight line and detoured distance. However, the significantly lower AIC of the GLM that used detoured distance as predictor variable suggests that detoured distance better explains the variation in genetic distance than straight line distance. Considering our results in combination with those from Anthony et al. [2007], suggests that gorillas temporarily restricted to certain refugia began to admix when their formerly fragmented patches were reconnected. The admixture, however, seems to have been impeded by certain riverine barriers. The Sangha and to a lesser extent the Ngoko apparently constituted such barriers.

While the rivers within our study site have only a relatively subtle barrier effect, other watercourses have been found to act as borders between subspecies or even species of great apes. The most striking example comes from the genus *Pan*, consisting of two species, the common chimpanzee (*P. troglodytes*) and the bonobo (*P. paniscus*) which range on either side of the Congo River [Osman-Hill, 1969]. That river probably existed since well before the Pleistocene [Beadle, 1981] and possibly constituted an impermeable barrier to gene flow thereby facilitating allopatric speciation in the genus *Pan*. Rivers as barriers between subspecies can be found within the common chimpanzees. They consist of the four subspecies *P.t.verus*, *P.t.ellioti*, *P.t.troglodytes* and *P.t.schweinfurthii* [Schwarz, 1934; Gonder et al., 1997] which are separated from each other through the Dahomey Gap, the Sanaga River and the Ubangi River, respectively. These examples illustrate that gene flow of great ape populations is sometimes profoundly affected through the existence of rivers acting as obstacles to individual movement.

#### Patterns of Female and Male Population Structuring

Clustering analyses in Geneland revealed three clusters of WLGs, irrespective if all individuals, only females or only males were incorporated. Therefore our results do not support a quantitative sex-bias in migration. Observational studies report that both sexes

disperse when reaching sexual maturity [Breuer et al., 2009; Robbins et al., 2004; Stokes et al., 2003]. However, it is much more difficult to use observational data for drawing conclusions about the distance that each sex travels at dispersal. Adult females do not range on their own and mostly appear to transfer directly between breeding groups, whereas most males live solitarily after leaving their natal group and before creating their own group [Breuer et al., 2010; Robbins et al., 2004]. It has been suggested that at least some males travel very long distances at this phase [Levréro, 2005; Douadi et al., 2007]. Indirect evidence for long-distance dispersal in WLG males also comes from a recent study [Inoue et al., 2013] which did not find kin relationships between neighboring silverbacks. Given these points one would expect to find females to be more philopatric and therefore also genetically more structured than males, as has been previously reported for WLGs and mountain gorillas [Douadi et al., 2007; Guschanski et al., 2008]. However, the study by Douadi et al. [2007] utilized sex-specific markers which are difficult to compare, represent single loci and might be biased by sex-specific differences in effective population size. Therefore, corroboration of their results using the same markers for both sexes is necessary for confirmation of their findings. Our study fills that gap and is the first that utilizes autosomal markers to look at sex-specific differences in WLG population clustering over a large scale. BAPS and STRUCTURE both detected possible migrants that originated from another cluster than the one where they were sampled. Three of these migrants were females. Also the individual that was sampled most far away from the cluster to which it was assigned was a female. Thus our results do not support previous findings of females dispersing less far than males. Instead, it seems that both sexes migrate over similar distances. Although females may only transfer to neighboring groups, multiple dispersal events can occur through adulthood [Stokes et al., 2003], which increases the total potential distance that a female may migrate. Such a pattern of dispersal resembles a stepping-stone model and can produce a pattern of isolation by distance (IBD). Our test of IBD by sex was significant for both females and males. This supports the previously proposed pattern of migration for females, but raises questions about the impact of any long-distance dispersal by males, because it means that males are more related to other males born nearby than to males born further away. This is in line with Bradley et al. [2004] who described dispersed male networks in WLGs in which silverbacks were usually related to nearby males. As that study, however, was conducted on a very small scale (9km×6 km) it could not assess whether some males might have migrated large distances. In sum, it seems that the pattern of genetic variation is quite similar in male and female WLGs, pointing to similar frequencies and extent of dispersal by males and females.

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#### Fig. 1.

Sampled region with red dots depicting sampling locations. Rivers mentioned in the text are in blue, national parks in yellow and roads in white. The three major sampling areas are: (i) Odzala: two sites east of Odzala NP (Libonga and Liouesso); (ii) Lobéké: three sites west of and within LobékéNP (Lobéké-West, Lobéké-Central, and Lobéké-East); and (iii) Ndoki forest: two sites within or close by Dzanga Ndoki and Nouabalé-Ndoki NP (southern Ndoki and Bai Hokou).



### Fig. 2.

Genetic distances measured as Fst/(1–Fst) plotted against natural logarithm of straight distance and detoured distance in meters, respectively. Each point represents one pairwise comparison between the seven sampling sites. The linear equations for the regression lines were y=0.0324x-0.3341 with P=0.0343 for straight distance (solid line) and y=0.0199 x -0.2142 with P=0.0096 for detoured distance (dashed line).



Estimated cluster membership

#### Fig. 3.

A Voroni tesssellation map depicting the sorting of all genotypes into three clusters. The clusters are represented by the colors white (Odzala), green (Lobéké) and yellow (Ndoki forest). The map was produced by the Geneland run with the highest mean posterior density. GPS points at which samples were collected are depicted as black dots.



# Fig. 4.

STRUCTURE plot for all genotypes at K=2. Analysis included WLGs sampled in Odzala (bars 1–42), Lobéké (bars 43–66) and the Ndoki forest (bars 67–261). The 17 individuals marked by a yellow asterisk have less than 75% ancestry from a single population. Individuals indicated by numbers from 1 to 6 were assigned to another cluster than the one in which they were sampled.



# Fig. 5.

The spatial genetic structure of all generated genotypes of WLGs from Odzala, Lobéké, and the Ndoki forest depicted as a Voronoi tessellation by BAPS. Each tile refers to one genotype. The tiles indicated by numbers belong to individuals that were sampled in another cluster than the one to which they were allocated. Note that equal numbers in Figure 4 and Figure 5 refer to the same individuals.

# TABLE I

 $F_{ST}$  Estimates (Below the Diagonal) and Associated *P*-values After 9999 Permutations (Above the Diagonal) for Pairwise Comparisons Between Odzala, Lobéké, and Ndoki Forest Including All Genotypes (Number of Included Samples From Each Locality in Brackets)

	Odzala (42)	Lobéké (24)	Ndoki forest (195)
Odzala (42)	_	< 0.0001	< 0.0001
Lobéké (24)	0.027	_	< 0.0001
Ndoki forest (195)	0.042	0.031	—